

Metabolic and ultrastructural renal changes in adult Wistar rats fed by a cafeteria diet

Priscila Fernandes dos Santos¹ , Diogo Benchimol de Souza¹ , Eduardo José Lopes Torres² , Waldemar Silva Costa¹ , Francisco José Barcellos Sampaio¹ , Bianca Martins Gregorio^{1*} 

1. Universidade do Estado do Rio de Janeiro  – Biomedical Center – Department of Anatomy – Rio de Janeiro (RJ) – Brazil.
2. Universidade do Estado do Rio de Janeiro  – Biomedical Center – Department of Microbiology, Immunology and Parasitology – Rio de Janeiro (RJ) – Brazil.

ABSTRACT

Purpose: To evaluate, by quantitative and qualitative methods, the glomerular ultrastructure in Wistar rats fed a cafeteria diet. **Methods:** Male Wistar rats were divided into two groups at 21 days of age: control (C, n = 10) and cafeteria diet (CAF, n = 8). The animals were followed up until 5 months of age, followed by euthanasia. The blood, kidneys, and fat deposits—epididymal, retroperitoneal, and subcutaneous—were extracted and analyzed. Data were analyzed by Student's t test, and $p < 0.05$ was considered statistically significant. **Results:** The cafeteria diet promoted glucose intolerance, hyperglycemia ($p < 0.0001$), and deposition of retroperitoneal fat ($p < 0.005$). Scanning electron microscopy revealed that the length of the foot process was similar in both groups. The quantitative analyses by transmission electron microscopy revealed that the cafeteria diet reduced the thickness of the glomerular basement membrane ($p < 0.05$). **Conclusion:** The intake of lipids and simple carbohydrates were found to be associated with alteration in the glomerular ultrastructure. However, more studies are needed to evaluate not only the effects of high-protein and high-fat diets on components of the glomerular filtration barrier, but also renal physiology.

Key words: Diet. Rats. Kidney. Microscopy, Electron, Scanning. Microscopy, Electron, Transmission.

Introduction

Dietary patterns can be analyzed to elucidate the relationship between diet and health problems. Scientific studies show that eating patterns are quite variable among the people. The consumption of vegetables, fruits, fiber, and fish is beneficial for human health, while the intake of red meat and processed foods is harmful¹. In addition to the body modifications promoted by this nutritional imbalance, an increase in the development of chronic diseases, such as overweight / obesity, diabetes mellitus, hypertension, and chronic kidney disease has been observed².

Research involving experimental models and nutrition has been widely carried out in order to elucidate the pathophysiology of several non-transmittable chronic degenerative diseases. Among these diets is cafeteria diet, which is a high source of energy and palatable³. Its lipid and simple carbohydrate content favors the development of glucose intolerance, besides causing morphological changes in the reproductive system, increase of body mass and abdominal fat, hyperinsulinemia, and hyperglycemia^{3,4}. In animals, the intake of cafeteria diet can trigger morphological changes in kidneys, such as interstitial fibrosis and tubular atrophy⁵. However, the influence of this diet on the renal ultrastructure has not yet been elucidated.

*Corresponding author: biancamgregorio.uerj@gmail.com

Received: Nov 1, 2023 | Accepted: Feb 14, 2024

Section editor: Norbert Nemeth 

Research performed at Postgraduate Program in Pathophysiology and Surgical Sciences, Universidade do Estado do Rio de Janeiro, Rio de Janeiro (RJ), Brazil.



It is already known that high-protein diets can dilate the afferent glomerular arteriole, resulting in hyperfiltration and subsequent glomerular damage (inflammation and fibrosis)⁶. Similarly, high-fat diets alter renal physiology, generating glomerular hypertrophy, proteinuria, increased desmin expression, and reduced nephrine expression in the glomerulus⁷. The glomerular filtration barrier is extremely important in the filtration process of the plasma. It consists of a fenestrated glomerular endothelium⁸, supported by a basement membrane⁹.

In this study, we examined specialized epithelial cells of the glomerular filtration barrier (podocytes) with their long cellular processes involving the blood capillaries (foot processes). Therefore, the detailed study of this component, in view of the various macronutrients present in our diet, is necessary, considering the increase in the consumption of such foods of low nutritional value (rich in lipids and simple carbohydrates) in society.

The aim of this study was to evaluate the glomerular ultrastructure, especially the podocytes and the glomerular basement membrane, of rats fed with cafeteria diet.

■ Methods

Experimental animal procedures

All procedures were performed with the approval of the Universidade do Estado do Rio de Janeiro Animal Care and Use Committee. Male Wistar rats (n = 18) were obtained from Urogenital Research Unit and housed in cages at 21 ± 2°C with a 12-h light/dark cycle. At weaning, the animals were randomly selected to receive either control diet (C, n = 10) or a cafeteria diet (CAF, n = 8) up to 5 months of age, followed by euthanasia. Both groups were fed *ad libitum* with fresh food daily. C diet consisted of commercial food (Nuvilab)–430 kcal/100 g–, and CAF diet was manipulated in the laboratory, having following constituents: commercial food 60 g/100 g, condensed milk (Nestlé) 25 g/100 g, and hydrogenated vegetable fat (Primor) 15 g/100 g, totaling 550 kcal/100 g. Food intake and body mass were assessed daily and weekly, respectively.

After being fed for 20 weeks with the assigned diet, rats were fasted for 12 h, anaesthetized with 100 mg/kg of sodium pentobarbital and killed, with blood taken directly from the right atrium. After collection, the blood was centrifuged, and plasma was obtained. The kidney and the different white adipose tissue depots (epididymal, retroperitoneal, and subcutaneous) were removed, weighed, and fixed until further analysis. Fat deposits were fixed in buffered formalin (4%) for 48 h. After this period, these tissues were processed, and 5-µm thick sections were cut.

Carbohydrate metabolism and biochemical assays

The oral glucose tolerance test was performed at 5 months of age. Following an overnight fast (12 h), a baseline blood glucose level was measured, and then all animals were administered a dose of hypertonic glucose serum 50% (2 g/kg body weight) by orogastric gavage. Blood glucose concentrations were measured 15, 30, 60, and 120 min after the glucose administration by glucometer (Accu-Chek, Roche, São Paulo, SP, Brazil).

Moreover, plasma glucose, triacylglycerol (TAG), high-density lipoprotein (HDL-C), and total cholesterol (TC) levels were measured using enzymatic colorimetric kits (BioSystems–Cat. 11506–Barcelona, Spain). Insulin levels were analyzed with a rat insulin enzyme-linked immunosorbent assay (ELISA) kit (Millipore–Cat. EZRMI-13K–St. Charles, Missouri, United States of America), according to the manufacturer's protocol.

Kidney collection and morphometry

For scanning electron microscopy (SEM), the kidney was removed, and fragments of approximately 1 mm³ were collected from the cortical region. These fragments were fixed by immersion in glutaraldehyde (2.5% in phosphate buffer, pH = 7.3) during 24 h and post fixed with 1% osmium tetroxide and 0.8% potassium ferrocyanide for 40 min

at room temperature. Then, they were dehydrated in ethanol, critical point-dried with CO₂, sputter-coated with gold-palladium, and observed using a Auriga Compact SEM at a magnification of approximately 27,000x and with a 2 kV beam acceleration voltage.

For transmission electron microscopy (TEM), after post fixation, the fragments were dehydrated in acetone and embedded in Epon. Ultra-thin sections (50 nm) were obtained and contrasted with 1% lead citrate and 5% uranyl acetate. The sections were observed on a JEOL-JEM-1011 TEM of the Rudolf Barth Electronic Microscopy Platform (Fundação Oswaldo Cruz), at a magnification of 50,000x and beam acceleration voltage of 80 kV.

SEM images were used to measure the length of the foot process, while in TEM images the thickness of the glomerular basement membrane (GBM) and the linear length of the foot process in contact with the GBM were measured. For both microscopy techniques, five animals/group and an ImageJ Software (Image Processing and Analysis in Java) were used.

Statistical analysis

Values were expressed as mean \pm standard deviation. Student's t test was used to compare the two groups using GraphPad Prism 5. In all cases, $p < 0.05$ was considered as statistically significant.

Results

Food intake was similar between the groups. Corroborating with this result, the groups showed no difference in weight gain throughout the experiment. However, the CAF diet increased the deposits of retroperitoneal fat when compared to the control (Table 1).

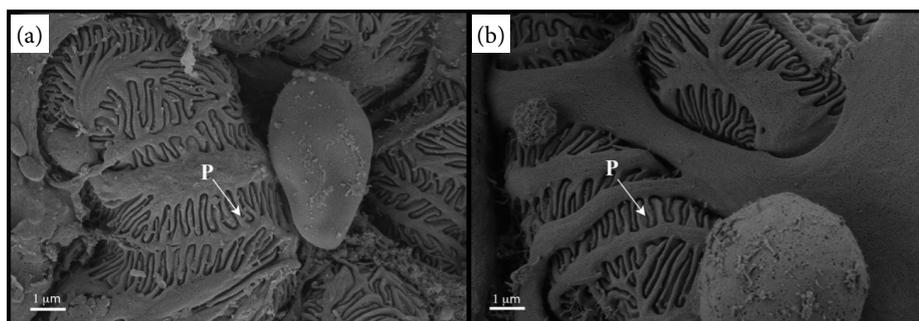
Table 1 – Biometric and kidney data.

| Parameters | C | CAF | p-value |
|--|--------------------|---------------------|----------|
| Food intake (g) | 8.24 \pm 0.46 | 6.98 \pm 0.54 | 0.0943 |
| Weight gain (g) | 350.20 \pm 25.72 | 350.40 \pm 39.84 | 0.9867 |
| Retroperitoneal fat (g) | 5.02 \pm 1.26 | 8.91 \pm 3.40 a | 0.0036 |
| Subcutaneous fat (g) | 3.14 \pm 0.71 | 4.37 \pm 1.38 | 0.0626 |
| Epididymal fat (g) | 5.43 \pm 1.15 | 7.41 \pm 1.77 | 0.0750 |
| OGTT After-resveratrol (u.a.) | 745.00 \pm 42.84 | 869.50 \pm 67.21a | < 0.0001 |
| Serum glucose (mmol/L) | 10.47 \pm 4.32 | 16.22 \pm 5.62a | 0.0361 |
| Cholesterol total (mg/dL) | 59.89 \pm 5.64 | 60.75 \pm 8.01 | 0.7995 |
| Triacylglycerol (mg/dL) | 85.60 \pm 29.51 | 103.80 \pm 38.69 | 0.2625 |
| High-density lipoprotein (mg/dL) | 38.90 \pm 7.01 | 40.44 \pm 7.00 | 0.6376 |
| Insulin (ng/mL) | 3.92 \pm 1.58 | 6.37 \pm 0.75 | > 0.05 |
| Foot process length (μ m) | 0.19 \pm 0.09 | 0.14 \pm 0.06 | 0.2200 |
| Glomerular basement membrane thickness (nm) | 147.40 \pm 10.41 | 126.50 \pm 6.76a | < 0.0001 |
| Length of foot process/glomerular basement membrane segment (nm) | 328.30 \pm 30.50 | 273.60 \pm 12.69 | 0.1359 |

Data were presented as mean \pm standard deviation. Differences were tested by Student's t test; $p < 0.05$; *statistical difference for the C group. Source: Elaborated by the authors.

The cafeteria diet promoted glucose intolerance and hyperglycemia at 5 months of age. The CAF group showed an elevation in the area under the glucose curve ($\uparrow 17\%$) when compared to C group ($p < 0.0001$). Moreover, serum glucose levels remained significantly higher in the CAF group compared to the control. However, cafeteria diet was not able to significantly modify the serum levels of TC, HDL-C, TAG, and insulin of the CAF group compared to the control group (Table 1).

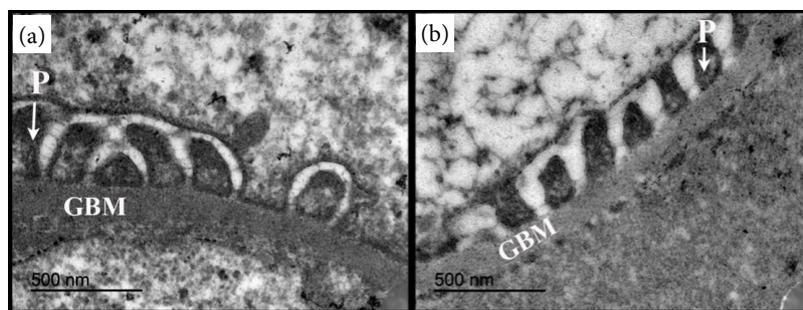
The qualitative analysis of the length of the foot process by SEM revealed it to be comparable between the groups studied (Fig. 1), which was corroborated by the statistical analysis (Table 1).



Source: Elaborated by the authors.

Figure 1 – Scanning electron Microscopy images of glomerulus. We observed that the lengths of foot processes (**P**) were similar between animals from (a) control and (b) cafeteria diet groups. Both images were obtained at the magnification of 27,000 (approximately) and a beam acceleration voltage of 2 kV.

TEM measurements showed a marked reduction of GBM thickness in the CAF group (Fig. 2) and tendency to reduce the length of foot process by GBM segment when compared to control (Table 1).



Source: Elaborated by the authors.

Figure 2 – Transmission electron microscopy images of glomerular filtration barrier components: glomerular basement membrane (GBM) and foot processes (**P**). (a) Control group: the GBM thickness is higher when compared to the animals fed with (b) cafeteria diet. Cafeteria diet also led to the reduction in the foot process length (**P**) compared to the control group. Both images were obtained at the magnification of 50,000 and a beam acceleration voltage of 80 kV.

Discussion

The intake of cafeteria diets has previously been correlated with obesity^{8,9} and important morphological changes in the rodent kidneys, such as glomerulosclerosis⁵, glomerular hypertrophy followed by excess mesangial matrix^{10,11}, and glomerular hyperfiltration¹². In our study, similar to the findings of Gomez-Smith et al.¹³ and Higa et al.¹⁴, cafeteria diet did not promote an increase in the body mass of the animals, although it caused glucose intolerance and an elevation of the retroperitoneal fat deposits.

The exacerbated consumption of foods rich in simple carbohydrates and lipids, together with sedentary lifestyle, promotes hypertrophy of adipose tissue, mainly the visceral fat deposit^{15,16}. The visceral adiposity accentuates lipolysis and leads to increased release of free fatty acids from adipose tissue. Thus, there are reduction in glucose mobilization and a greater risk of the development of glucose intolerance and insulin resistance, which justify our findings^{17,18}. The limitations of this study were the use of rats and a diet with low lipid content, which was not indicative of variation in body mass.

In relation to the kidney, excessive visceral adiposity in humans promotes renal compression, with increased intrarenal pressure¹⁹⁻²¹, and is associated with coronary artery calcification in patients with chronic kidney disease²². However, we did not find the presence of hypertension in the CAF group. It is possible that the physical compression of the kidneys in humans, rabbits, and dogs is different from that in rodents. Anatomically, the kidneys of these mammals appear to float more than adipose tissue and, therefore, are not compressed, preventing the reduction of renal blood inflow and maintaining the blood pressure at normal levels^{21,23}.

The redistribution of body fat caused by intake of the cafeteria diet may also affect the renal ultrastructure²⁴. Previous studies indicated that lipotoxicity leads to changes in the cytoskeleton of podocytes and in the proteins of the slit diaphragm²⁵. Similarly, the toxicity mediated by saturated fatty acids, such as palmitic acid, induces endoplasmic reticulum stress and podocyte death²⁶. In our study, the increase in retroperitoneal fat in the CAF group did not promote significant alterations in this structure. Our qualitative and quantitative analyses by SEM and TEM, respectively, showed that the cafeteria diet caused a slight reduction in podocyte length (not significant). It has been assumed that, at this stage, this diet promoted a remodeling of the podocyte cytoskeleton, neutralizing renal injury²⁷.

The podocytes are differentiated cells that cover the outer surface of the GBM, and contain three main segments: the cell body, the primary processes, and the foot process, and they are indispensable for the normal maintenance of glomerular filtration. Previous literature reports that high-protein and high-fat diets (without the association of sucrose) promote podocyte effacement, with apparent impairment of renal function^{7,28}. However, in the present study, reduction of GBM thickness was verified in the CAF group, regardless of the reduction of foot process length. It is believed that sucrose, the main macronutrient present in the diet used in this study, is not involved neither in the initiation and progression of glomerulosclerosis nor, consequently, in the manifestation of renal injury.

■ Conclusion

It was noticed that macronutrients, except sucrose, are directly associated to the loss of renal function. Even though we did not work with diets rich in proteins and lipids, we saw that simple carbohydrates modified only the GBM thickness, without compromising the length of the foot process. However, more studies are needed to evaluate not only GBM components, but also renal physiology.

■ Conflict of interest

Nothing to declare.

■ Author's contributions

Substantive scientific and intellectual contributions to the study: Gregorio BM e Souza DB; **Conception and design:** Gregorio BM, Souza DB and Sampaio FJB; **Acquisition of data:** Santos PF and Torres EJJ; **Analysis and interpretation of data:** Costa WS and Souza DB; **Technical procedures:** Santos PF and Torres EJJ.

■ Data availability statement

All data sets were generated or analyzed in the current study.

■ Funding

Fundação Carlos Chagas Filho de Amparo à Pesquisa do Estado do Rio de Janeiro 

Grant No: E-26/010.002569/2014

■ About the authors

Santos PF is biologist.

Sousa DB, Torres E JL, Costa WS, Sampaio FJB and Gregorio BM are professors.

■ Acknowledgements

We would like to thank Makoto Enoki Caracciolo (PhD) and Ludmila Rocha Lima (PhD student) for their assistance during the processing of samples for SEM and TEM.

■ References

1. Suliga E, Głuszek S. The relationship between diet, energy balance and fertility in men. *Int J Vitam Nutr Res.* 2020;90(5-6):514–26. <https://doi.org/10.1024/0300-9831/a000577>
2. Bettiga A, Fiorio F, Di Marco F, Trevisani F, Romani A, Porrini E, Salonia A, Montorsi F, Vago R. The Modern Western Diet Rich in Advanced Glycation End-Products (AGEs): An Overview of Its Impact on Obesity and Early Progression of Renal Pathology. *Nutrients.* 2019;11(8):1748. <https://doi.org/10.3390/nu11081748>
3. De Oliveira F, Costa WS, Sampaio FJ, Gregorio BM. Resveratrol attenuates metabolic, sperm and testicular changes in adult Wistar rats fed a cafeteria dietary. *Asian J Androl.* 2019;21(2):201–7. https://doi.org/10.4103/aja.aja_67_18
4. Pons Z, Margalef M, Bravo FI, Arola-Arnal A, Muguera B. Chronic administration of grape-seed polyphenols attenuates the development of hypertension and improves other cardiometabolic risk factors associated with the metabolic syndrome in cafeteria diet-fed rats. *Br J Nutr.* 2017;117(2):200–8. <https://doi.org/10.1017/S0007114516004426>
5. Zeeni N, Dagher-Hamalian C, Dimassi H, Faour WH. Cafeteria diet-fed mice is a pertinent model of obesity-induced organ damage: a potential role of inflammation. *Inflamm Res.* 2015;64(7):501–12. <https://doi.org/10.1007/s00011-015-0831-z>
6. Hostetter TH, Olson JL, Rennke HG, Venkatachalam MA, Brenner BM. Hyperfiltration in remnant nephrons: a potentially adverse response to renal ablation. *Am J Physiol.* 1981;241(1):F85–93. <https://doi.org/10.1152/ajprenal.1981.241.1.F85>
7. Chen JY, Jian DY, Lien CC, Lin YT, Ting CH, Chen LK, Hsu TC, Huang HM, Wu YT, Kuan TT, Chao YW, Wu LY, Huang SW, Juan CC. Adipocytes play an etiological role in the podocytopathy of high-fat diet-fed rats. *J Endocrinol.* 2016;231(2):109–20. <https://doi.org/10.1530/JOE-16-0064>
8. Jarad G, Miner JH. Update on the glomerular filtration barrier. *Curr Opin Nephrol Hypertens.* 2009;18(3):226–32. <https://doi.org/10.1097/mnh.0b013e3283296044>
9. Alkhedaide A, Soliman MM, Salah-Eldin AE, Ismail TA, Alshehri ZS, Attia HF. Chronic effects of soft drink consumption on the health state of Wistar rats: a biochemical, genetic and histopathological study. *Mol Med Rep.* 2016;13(6):5109–17. <https://doi.org/10.3892/mmr.2016.5199>
10. Dai HY, Zheng M, Tang RN, Ma KL, Ni J, Liu BC. Inhibition of integrin-linked kinase by angiotensin II receptor

- antagonist, irbesartan attenuates podocyte injury in diabetic rats. *Chin Med J (Engl)*. 2012;125(5):888–93.
11. Zhang GY, Wang DD, Cao Z, Wei T, Liu CX, Wei QL. Sitagliptin ameliorates high glucose-induced cell proliferation and expression of the extracellular matrix in glomerular mesangial cells. *Exp Ther Med*. 2017;14(4):3862–7. <https://doi.org/10.3892/etm.2017.5002>
 12. Tobar A, Ori Y, Benchetrit S, Milo G, Herman-Edelstein M, Zingerman B, Lev N, Gafter U, Chagnac A. Proximal tubular hypertrophy and enlarged glomerular and proximal tubular urinary space in obese subjects with proteinuria. *PLoS One*. 2013;8(9):e75547. <https://doi.org/10.1371/journal.pone.0075547>
 13. Gomez-Smith M, Karthikeyan S, Jeffers MS, Janik R, Thomason LA, Stefanovic B, Corbett D. A physiological characterization of the Cafeteria diet model of metabolic syndrome in the rat. *Physiol Behav*. 2016;167:382–91. <https://doi.org/10.1016/j.physbeh.2016.09.029>
 14. Higa TS, Spinola AV, Fonseca-Alaniz MH, Evangelista FS. Comparison between cafeteria and high-fat diets in the induction of metabolic dysfunction in mice. *Int J Physiol Pathophysiol Pharmacol*. 2014;6(1):47–54.
 15. Francisqueti FV, do Nascimento AF, Corrêa CR. Obesity, inflammation and metabolic complications. *Nutrire*. 2015;40(1):81–9.
 16. Francisqueti FV, Chiaverini LC, Santos KC, Minatel IO, Ronchi CB, Ferron AJ, Ferreira AL, Corrêa CR. The role of oxidative stress on the pathophysiology of metabolic syndrome. *Rev Assoc Med Bras*. 2017;63(1):85–91. <https://doi.org/10.1590/1806-9282.63.01.85>
 17. Pereira LO, De Francischi RP, Lancha Jr AH. Obesity: dietary intake, sedentarism and insulin resistance *Arq Bras Endocrinol Metab*. 2003;47(2):111–27. <https://doi.org/10.1590/S0004-27302003000200003>
 18. Ortega-Loubon C, Fernández-Molina M, Singh G, Correa R. Obesity and its cardiovascular effects. *Diabetes Metab Res Rev*. 2019;35(4):e3135. <https://doi.org/10.1002/dmrr.3135>
 19. Maric-Bilkan C. Obesity and diabetic kidney disease. *Med Clin North Am*. 2013;97(1):59–74. <https://doi.org/10.1016/j.mcna.2012.10.010>
 20. Chade AR, Hall JE. Role of the renal microcirculation in progression of chronic kidney injury in obesity. *Am J Nephrol*. 2016;44(5):354–67. <https://doi.org/10.1159/000452365>
 21. Luo K, Bian J, Wang Q, Wang J, Chen F, Li H, Jin D. Association of obesity with chronic kidney disease in elderly patients with nonalcoholic fatty liver disease. *Turk J Gastroenterol*. 2019;30(7):611–5. <https://doi.org/10.5152/tjg.2019.18343>
 22. Cordeiro AC, Qureshi AR, Lindholm B, Amparo FC, Tito-Paladino-Filho A, Perini M, Lourenço FS, Pinto IM, Amodeo C, Carrero JJ. Visceral fat and coronary artery calcification in patients with chronic kidney disease. *Nephrol Dial Transplant*. 2013;28(Suppl.4):iv152–9. <https://doi.org/10.1093/ndt/gft250>
 23. Hall JE, do Carmo JM, da Silva AA, Wang Z, Hall ME. Obesity-induced hypertension: interaction of neurohumoral and renal mechanisms. *Circ Res*. 2015;116(6):991–1006. <https://doi.org/10.1161/CIRCRESAHA.116.305697>
 24. Izquierdo-Lahuerta A, Martínez-García C, Medina-Gómez G. Lipotoxicity as a trigger factor of renal disease. *J Nephrol*. 2016;29(5):603–10. <https://doi.org/10.1007/s40620-016-0278-5>
 25. Martínez-García C, Izquierdo-Lahuerta A, Vivas Y, Velasco I, Yeo TK, Chen S, Medina-Gomez G. Renal lipotoxicity-associated inflammation and insulin resistance affects actin cytoskeleton organization in podocytes. *PLoS One*. 2015;10(11):e0142291. <https://doi.org/10.1371/journal.pone.0142291>
 26. Kampe K, Sieber J, Orellana JM, Mundel P, Jehle AW. Susceptibility of podocytes to palmitic acid is regulated by fatty acid oxidation and inversely depends on acetyl-coa carboxylases 1 and 2. *Am J Physiol Renal Physiol*. 2014;306(4):F401–9. <https://doi.org/10.1152/ajprenal.00454.2013>
 27. Barisoni L. Podocyte biology in segmental sclerosis and progressive glomerular injury. *Adv Chronic Kidney Dis*. 2012;19(2):76–83. <https://doi.org/10.1053/j.ackd.2012.02.018>
 28. Jesudason DR, Pedersen E, Clifton PM. Weight-loss diets in people with type 2 diabetes and renal disease: a randomized controlled trial of the effect of different dietary protein amounts. *Am J Clin Nutr*. 2013;98(2):494–501. <https://doi.org/10.3945/ajcn.113.060889>