CALLUS INDUCTION IN LEAF SEGMENTS OF Croton urucurana BAILL.

Indução de calos em segmentos foliares de sangra d'água (Croton urucurana Baill.)

Ednabel Caracas Lima¹, Renato Paiva², Raírys Cravo Nogueira³, Fernanda Pereira Soares⁴, Eduardo Bucsam Emrich⁵, Álvaro Augusto Naves Silva⁶

RESUMO

A sangra d'água, espécie pertencente à família *Euphorbiaceae*, apresenta potencial na recuperação de matas ciliares degradadas e é extensamente utilizada na medicina tradicional, como cicatrizante e no tratamento de reumatismos. Entretanto, suas sementes apresentam dormência e baixa viabilidade, dificultando a propagação desta espécie. Avaliou-se o efeito de diferentes concentrações de 2,4-D e BAP ou TDZ e de ANA e BAP, com suas possíveis combinações, na indução de calos em segmentos foliares de sangra d'água. Quarenta e cinco dias após a inoculação, foi avaliado o peso fresco dos calos. O uso de BAP isoladamente e a combinação entre ANA e BAP não promoveram calogênese em segmentos foliares de sangra d'água. Embora a combinação de 2,4-D e BAP ou TDZ tenha induzido a formação de calos, o uso isolado de 2,4-D proporcionou o maior peso fresco destes.

Termos para indexação: Micropropagação, calogênese, cultura de tecidos.

ABSTRACT

Croton urucurana Baill., a species belonging to the family Euphorbiaceae, can be useful in the recovery of degraded riparian areas. In the traditional medicine, it is widely used as cicatrizant and in the treatment of rheumatism. However, its seeds present dormancy and low viability, making the propagation of this species a challenge. With the objective of establishing an alternative route for the propagation, the effect of different concentrations of 2,4-D (2,4- dichlorophenoxyacetic acid) with BAP (6-benzylaminopurine) or TDZ (thidiazuron) and of NAA (1-naphthaleneacetic acid) with BAP and their possible combinations were evaluated for callus induction in leaf segments. Callus fresh mass was evaluated forty-five days after inoculation. The isolated use of BAP and the combination of NAA with BAP did not promote calogenesis in leaf segments. Even though the combination of 2,4-D with BAP or TDZ had induced the formation of callus, it was the isolated use of 2,4-D that provided the highest callus fresh mass.

Index terms: Micropropagation, calogenesis, tissue culture.

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INTRODUCTION

Croton urucurana Baill., a tree from Brazilian savanna, holds a great medicinal potential. It can be used in the treatment of rheumatisms and cancer as well as cicatrizant. From the restoration ecology point of view, it plays a very important role as it can be used to recover degraded areas.

This species is endemic to tropical and subtropical regions of South America. In Brazil, it occurs in the States of Bahia, Rio de Janeiro, Mato Grosso do Sul and Rio Grande do Sul, surpassing this country border and occurring also in Uruguay and Argentina (CORDEIRO, 1985).

Its fruits produce seeds in abundance, but shortly after the fruit dehiscence, coleopteras from the genus *Apion*

(*Curculionidae*; *Apioninae*) can be seen in the fruits, causing damage to the embryo and, therefore, drastically affecting the germination. Moreover, *C. urucurana* seeds present dormancy and low viability, being viable for no longer than four months (LORENZI, 2000).

The aspects above mentioned act as obstacles to the production of seedlings via sexual reproduction, which makes the tissue culture technique a viable alternative to overcome the problems that are inherent to the natural propagation of this species. Multiplication of *C. urucurana* can be achieved by micropropagation, either by organogenesis or somatic embryogenesis. In each case, by previously forming calli (GRATTAPAGLIA & MACHADO, 1998).

¹Engenheira Agrônoma, Mestre em Fisiologia Vegetal – Universidade Federal de Lavras/UFLA – Cx. P. 3037 – 37200-000 – Lavras, MG – belcaracas@yahoo.com.br.

²Engenheiro Agrônomo, PhD, Professor Adjunto do Setor de Fisiologia Vegetal – Departamento de Biologia/DBI – Universidade Federal de Lavras/UFLA – Cx. P. 3037 – 37200-000 – Lavras, MG – renpaiva@ufla.br

³Bióloga, Doutora em Fisiologia Vegetal – Cidade Nova 8, 431, WE: 42 – Coqueiro – Ananindua, PA – 67133-250 – rairys@yahoo.com.br

[&]quot;Engenheira Agrônoma, Doutoranda em Fisiologia Vegetal – Departamento de Biologia/DBI – Universidade Federal de Lavras/UFLA – Cx. P. 3037 – 37200-000 – Lavras, MG – Bolsista CAPES

⁵Engenheiro Agrônomo – Departamento de Biologia/DBI – Universidade Federal de Lavras/UFLA – Cx. P. 3037 – 37200-000 – Lavras, MG – bucsan_emrich@yahoo.com.br – Bolsista CNPq

Biólogo – Departamento de Biologia/DBI – Universidade Federal de Lavras/UFLA – Cx. P. 3037 – 37200-000 – Lavras, MG – silvaan@hotmail.com

Calli are defined as tissues constituted by differentiated cells, which develop in response to a chemical or physical lesion, under determinate hormonal conditions (MANTELL et al., 1994). They can be obtained from a tissue fragment and have the ability to differentiate into tissues, organs and even embryos, being able to regenerate whole plants (PAIVA & PAIVA, 2001; PIERIK, 1990; TORRES & CALDAS, 1990).

The exogenous supply of growth regulators is frequently necessary in calogenesis (VIETEZ & SAN-JOSÉ, 1996). This necessity refers to the type, concentration, relation auxin/cytokinin, genotype of the donor plant and the endogenous content of hormones.

According to George & Sherrington (1984), the combination of auxins and cytokinins promote cellular differentiation and also organogenesis. Among the growth regulators used in callus induction, 2,4-D (2,4-dichlorophenoxyacetic acid), NAA (1-naphthaleneacetic acid), BAP (6-benzylaminopurine) and TDZ (thidiazuron) are the most important.

The objective of this work was to assess the interaction of different concentrations of 2,4-D with TDZ or BAP and of NAA with BAP in the callus induction in leaf segments of *C. urucurana*.

MATERIAL AND METHODS

Origin and disinfestation of plant material

Source plants of *C. urucurana* about six months old were obtained from Centrais Elétricas de Minas Gerais (CEMIG) plant nursery, located at Represa de Camargos, in the county of Itutinga and were maintained in growth room at the Plant Physiology Sector of the Biology Department/Federal University of Lavras.

Young leaves collected from source plants were washed in running water and neutral detergent for 12 hours and then disinfested with 70% alcohol (v/v) for one minute in a laminar flux chamber followed by immersion in a 50% sodium hypochlorite solution (1% active chlorine) for 15 minutes and rinsed five times in distilled autoclaved water. Disks of approximately 1 cm² were excised from the leaves.

Effect of the combinations of 2,4-D with TDZ or BAP

The leaf explants were inoculated in WPM culture medium (LLOYD & MCCOWN, 1980), supplemented with different concentrations of 2,4-D (0.0, 0.5, 1.0, 2.0, 3.0, 4.0 and 5.0 mg $\rm L^{-1}$) as well as TDZ (0.0, 0.5, 1.0 and 2.0 mg $\rm L^{-1}$) or BAP (0.0, 0.5, 1.0 and 2.0 mg $\rm L^{-1}$), with their possible combinations and enriched with 30.0 g $\rm L^{-1}$ sucrose and 7.0

g L^{-1} agar. The pH was adjusted to 5.8 and then autoclaved for 15 minutes at 121°C.

After inoculation, the explants were kept in a growth room under irradiance of 36 μ mol m⁻² s⁻¹, photoperiod of 16 hours and temperature of 27 \pm 2°C.

The explants were evaluated at 45 days after inoculation, using the callus fresh mass as the parameter. Treatments were arranged in a completely randomized block design with each treatment replicated 20 times. Each parcel was constituted by one tube, each one containing one explant per tube. The data were submitted to variance analysis and Tukey test was used at a 5% significance level to evaluate the results.

Effect of the combinations of NAA with BAP

The leaf explants were inoculated onto WPM culture medium supplemented with different concentrations of NAA (0.0, 1.0, 2.0, 3.0, 4.0 and 5.0 mg L^{-1}) as well as BAP (0.0, 1.0, 2.0, 3.0, 4.0 and 5.0 mg L^{-1}), with their possible combinations and enriched with 30.0 g L^{-1} sucrose and 7.0 g L^{-1} agar. The pH was adjusted to 5.8 and then autoclaved for 15 minutes at 121°C.

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RESULTS AND DISCUSSION

The interaction of the auxin NAA with the cytokinin BAP in the concentrations tested was not effective to obtain calli in the leaf explants of *C. urucurana*. However, in the presence of 1.0, 2.0, 3.0, 4.0 and 5.0 mg L⁻¹ NAA and in the absence of BAP, root emission was observed in the explants. This also happened when they were inoculated in the presence of 4.0 and 5.0 mg L⁻¹ NAA with 1.0 mg L⁻¹ BAP.

Regarding the different combinations of 2,4-D with TDZ, the highest callus formation in the leaf explants of *C. urucurana* was observed with the isolated addition of 2,4-D to the culture medium. There was no callus formation in the absence of this growth regulator.

TDZ was effective only in the calogenesis, when used in the concentration of 1.0 mg $L^{\text{-1}}$, associated with 2,4-D, at the concentration of 0.5 mg $L^{\text{-1}}$ (Figure 1).

Table 1 shows that 2,4-D at the concentrations of 3.0 and 5.0 mg L^{-1} promoted the highest callus fresh matter, followed by the concentrations of 4.0 and 1.0 mg L^{-1} . The lowest callus fresh matter was observed with the use of 2.0 mg L^{-1} 2,4-D and with the combination of 2.0 mg L^{-1} 2,4-D + 0.5 mg L^{-1} TDZ.

Based on the fact that the concentrations of 2,4-D at 3.0 and 5.0 mg L $^{-1}$ produced results that did not differ statistically from each other, we suggest the use of this growth regulator at 3.0 mg L $^{-1}$ for callus induction in leaf segments of *C. urucurana*.

Concerning the effect of combination of 2,4-D with BAP, it was observed that the concentrations of 2,4-D at 1.0, 2.0, 3.0, 4.0 and 5.0 mg L⁻¹ and the combination of 4.0 mg L⁻¹ 2,4-D with 2.0 mg L⁻¹ BAP promoted high weight variability, whereas the results from the other treatments remained at a similar level. Some combinations of

2,4-D with BAP did not present callus formation. Calogenesis was also absent in the control treatment as well as in the treatments containing only BAP (Figure 2).

Table 2 shows the average of the treatments where callus formation was present.

The auxin 2,4-D at the concentration of 4.0 mg L⁻¹ and without the presence of BAP was the most effective in promoting callus induction, leading to the highest formation of fresh mass (0.374 g).

Similar results were achieved by Santos et al. (2005), who recommends 2,4-D at a concentration of 4.0 mg L^{-1} for the highest production of callus in *Salix humboldtiana* Willd.

On the other hand, Sahoo et al. (1997) observed that the concentrations of 2,4-D between 0.5 and 4.0 mg L⁻¹ were not effective on inducing calogenesis in leaf explants of mulberry (*Morus indica* L.).

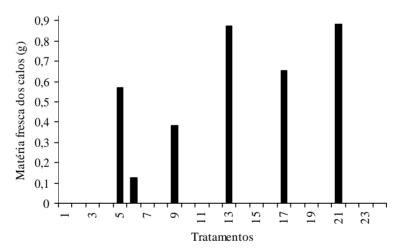


Figure 1 – *Box-plot* of callus fresh matter obtained from leaf segments of *C. urucurana* inoculated in WPM medium, supplemented with combinations of 2,4-D and TDZ.

Table 1 – Callus fresh matter obtained from leaf segments of *C. urucurana* inoculated in WPM medium, supplemented with 2,4-D and TDZ.

Treatments	Average of callus fresh matter (g)*
$(T21) 5.0 \text{ mg L}^{-1} 2,4-D$	0.8815 a
$(T13) 3.0 \text{ mg L}^{-1} 2,4-D$	0.8756 a
$(T17) 4.0 \text{ mg L}^{-1} 2,4-D$	0.6537 b
$(T5) 1.0 \text{ mg L}^{-1} 2,4-D$	0.5706 b
$(T9) 2.0 \text{ mg L}^{-1} 2,4-D$	0.3833 c
(T6) $1.0 \text{ mg L}^{-1} 2,4-D + 0.5 \text{ mg L TDZ}$	0.1274 d

^{*} Same letters in the column do not significantly differ at the 5% level of probability using Tukey test.

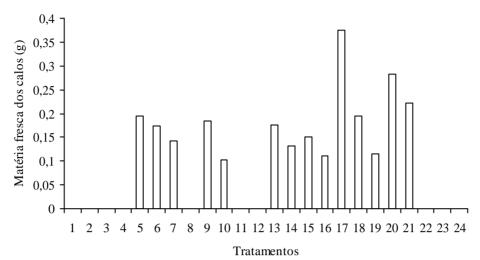


Figure 2 – *Box-plot* of callus fresh matter obtained from leaf segments of *C. urucurana* inoculated in WPM medium, supplemented with combinations of 2,4-D and BAP.

Table 2 – Callus fresh matter obtained from leaf segments of *C. urucurana* inoculated in WPM medium, supplemented with different combinations of 2,4-D with BAP.

Treatments	Callus fresh matter (g)*	
(T17) 4.0 mg L ⁻¹ 2,4-D	0.3740 a	
$(T20) 4.0 \text{ mg L}^{-1} 2,4-D+2 \text{ mg L BAP}$	0.2825 b	
(T21) 5.0 mg L ⁻¹ 2,4-D	0.2229 c	
(T5) 1.0 mg L ⁻¹ 2,4-D	0.1976 c	
$(T18) 4.0 \text{ mg L}^{-1} 2,4-D + 0.5 \text{ mg L BAP}$	0.1955 c	
(T9) 2.0 mg L ⁻¹ 2,4-D	0.1834 c	
(T13) 3.0 mg L ⁻¹ 2,4-D	0.1768 c	
(T6) $1.0 \text{ mg L}^{-1} 2,4-D + 0.5 \text{ mg L BAP}$	0.1728 c	
(T5) $3.0 \text{ mg L}^{-1} 2,4-D+1 \text{ mg L BAP}$	0.1498 d	
(T7) $1.0 \text{ mg L}^{-1} 2,4-D+1 \text{ mg L BAP}$	0.1416 d	
$(T14) 3.0 \text{ mg L}^{-1} 2,4-D + 0.5 \text{ mg L BAP}$	0.1311 d	
$(T16) 3.0 \text{ mg L}^{-1} 2,4-D+2 \text{ mg L BAP}$	0.1123 d	
$(T19) 4.0 \text{ mg L}^{-1} 2,4-D+1 \text{ mg L BAP}$	0.1116 d	
$(T10) 2.0 \text{ mg L}^{-1} 2,4-D + 0.5 \text{ mg L BAP}$	0.1017 e	

^{*} Same letters in the column do not significantly differ at the 5% level of probability using Tukey test.

According to Grattapaglia & Machado (1998), 2,4-D tends to stimulate callus formation, even at low concentrations. This growth regulator shows effect on the RNA metabolism by inducing the transcription

of messenger RNA capable of coding proteins required for the growth and hence, promoting a chaotic cell proliferation, i.e., callus formation (GEORGE, 1996).

Similar results to the ones found in this work with *C. urucurana* were also obtained in *Inga vera* Willd. The optimum concentration of 2,4-D was at 3.0 mg L⁻¹, which induced the highest callus formation (80%) (SOARES, 2003).

Regarding the combination of 2,4-D with TDZ, the results obtained here differ from those found by Deccetti (2000) in *Annona glabra* L. For this species an interaction between the growth regulators TDZ and 2,4-D was necessary for the maximum callus production.

In general, the results found in this work indicate that the combination of 2,4-D with BAP produces the lowest callus fresh mass in leaf explants of *C. urucurana*.

The results of this work support those found by other authors. For instance, Landa et al. (2000) did not observe callus formation in leaf explants of *Caryocar brasiliense* Camb. in the absence of 2,4-D while Vietez & San-José (1996) did not obtain callus in leaf explants of *Fagus silvatica* L. in the presence of BAP.

Conversely, Santiago (2003) concluded that in leaf explants of *Piper hispidinervum* C. DC., the maximum callus production was achieved with the combination of 2,4 D and BAP.

The results here indicate that exogenous supply of the auxin 2,4-D by itself was enough to promote a hormonal balance capable of inducing the highest callus formation in leaf tissues of *C. urucurana*.

CONCLUSIONS

The interaction of the auxins NAA and 2,4-D with the cytokinin BAP, in the tested concentrations, is not enough for callus production in leaf explants of *C. urucurana*.

The presence of 2,4-D is essential for callus induction in leaf explants of this species.

When used by itself in the culture medium, TDZ is not effective in inducing calogenesis in leaf explants of *C. urucurana*.

Maximum production of callus in leaf explants of *C. urucurana* is obtained when they are cultured on WPM medium, supplemented with 3.0, 4.0 or 5.0 mg L⁻¹ 2,4-D.

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