



Global patterns, biases, and advances in phylogeographic and genetic structure studies in Drosophilidae (Insecta: Diptera)

Henrique R. M. Antonioli¹ , Maríndia Deprá¹ and Lizandra Jaqueline Robe²

¹Universidade Federal do Rio Grande do Sul (UFRGS), Instituto de Biociências, Programa de Pós-Graduação em Genética e Biologia Molecular, Porto Alegre, RS, Brazil.

²Universidade Federal de Santa Maria (UFSM), Programa de Pós-Graduação em Biodiversidade Animal, Santa Maria, RS, Brazil.

Abstract

Phylogeographic and genetic structure studies involving Drosophilids have clarified numerous processes that have shaped the evolution of biodiversity over time. In this review, we aim to (i) assess the main biases, gaps, and advances in the scientific literature on this topic; (ii) synthesize the major findings emerging from these studies; and (iii) identify congruencies and discrepancies in the phylogeographical histories of different species and regions. To achieve these goals, we conducted a comprehensive review of peer-reviewed literature on phylogeographic and genetic structure studies of Drosophilidae published between 1987 and 2024. After identifying and filtering relevant studies, we extracted and analyzed key information related to each topic. Overall, we have detected a straightforward predominance of studies involving species of the *Drosophila* genus, especially within the *melanogaster*, *obscura*, and *repleta* groups. Interestingly, most studies employed nuclear DNA markers, either alone or in combination with mitochondrial markers, and were conducted across more than one biogeographical region, primarily in the Palearctic and Nearctic. Thus, our synthesis underscores the importance of broader taxonomic sampling and increased attention to understudied regions to enhance our understanding of biodiversity dynamics in response to environmental changes at both local and global scales.

Keywords: *Drosophila*, model species, molecular clock, molecular markers, phylogeography.

Received: November 13, 2025; Accepted: December 12, 2025.

Introduction

Phylogeography emerged as a new discipline nearly 40 years ago, when Avise *et al.* (1987) highlighted the lack of a field of study that connects population genetics with phylogenetic systematics. The term “phylogeography” was later formalized to describe research that integrates genealogies and biogeography across temporal and spatial dimensions, emphasizing historical factors that explain present-day distributions of lineages (Avise, 2009). In this way, it aims to evaluate patterns of distribution of genetic diversity across the species’ full geographic range, in contrast to studies that assess population structure among individual populations representing only part of the species’ overall range. Nevertheless, both approaches can substantially contribute to understanding the processes that have shaped the evolutionary history of the focal species, at global or more local scales. Moreover, when applied in a comparative framework, they may help address questions about the origin and evolution of biodiversity in specific biogeographical contexts.

Notably, these approaches have been applied to issues in speciation and conservation biology across various taxa,

including animals (Garcez *et al.*, 2018; Kjartanson *et al.*, 2023) and plants (Zhao *et al.*, 2019). They also contribute to current assessments of diversity and conservation by enabling the discovery of new species or lineages (Alfaro *et al.*, 2018) and the accurate assignment of species distributions (Carvalho *et al.*, 2023). Additionally, they may contribute to understanding ecological (Bonebrake *et al.*, 2010) and evolutionary relationships (Rosauer *et al.*, 2016) among populations or species, insights that are critical for explaining speciation, diversification, and the factors influencing survival and extinction. Finally, they may shed light on the general patterns of genetic diversity [such as intra- and interpopulation structure (Eizirik *et al.*, 2001) and underlying demographic history (Zhang *et al.*, 2017)], and help uncover historical processes that have shaped biodiversity [such as vicariance or fragmentation into refugia (Song *et al.*, 2018), range expansion and secondary contact (Gum *et al.*, 2005), introgression (Jeratthitikul *et al.*, 2013) and coevolution (Cuthill and Charleston, 2015)].

To accomplish all these tasks, suitable molecular markers should be carefully selected. Ideally, they should exhibit several properties that maximize their informative power, including sufficient variability without saturation, reliable orthology assignment, and clocklike evolution, combined with technical practicality (i.e. their characterization should be scalable, cost-effective, and reproducible). They may also vary in their inheritance patterns, which contrasts maternally

Send correspondence to Lizandra Jaqueline Robe, Universidade Federal de Santa Maria (UFSM), Programa de Pós-Graduação em Biodiversidade Animal, Av. Roraima, 1000, Prédio 17 / Sala 1140A – Camobi, 97105-900, Santa Maria, RS, Brazil. E-mail: e-mail.lizandra.robe@ufsm.br.

inherited mitochondrial DNA (mtDNA) with biparental nuclear DNA (nDNA) markers, as well as dominant and co-dominant markers [such as microsatellites (satDNA) and Single Nucleotide Polymorphisms (SNPs)]. Mitochondrial genes and satDNA have the advantage of faster mutation rates than nuclear genes, allowing us to infer relatively recent evolutionary events (Avise, 2004). On the other hand, nDNA exhibit biparental inheritance, providing a more comprehensive view of population genetic patterns and offering insights into long-term evolutionary processes. Historically, mtDNA has been widely used as the preferred marker for phylogeographic and population structure analyses (Beheregaray, 2008; Turchetto-Zolet *et al.*, 2013), although marker choice may differ among taxa.

Drosophilidae is a diverse and broadly distributed family of Diptera, comprising more than 4,000 species that inhabit environments ranging from tundra to tropical regions (Bächli, 2025; Throckmorton, 1975) and exploit a wide range of substrates (Carson, 1971; Powell, 1997). The family is subdivided into 77 extant genera distributed across distinct lineages within the Drosophilidae phylogeny (Kim *et al.*, 2024). In several cases, this arrangement results in the paraphyly of *Drosophila*, the “most famous” genus of the family. In fact, different *Drosophila* species, particularly *D. melanogaster*, have been widely used as model organisms across biomedical, ecological, and evolutionary research (Kohler, 1993; Markow and O’Grady, 2006). Notably, eight Nobel Prizes have been awarded to scientists for discoveries using *Drosophila* (Clancy *et al.*, 2023). Their short generation time, ease of laboratory culture, and well-characterized genetics make them ideal candidates for such investigations. In the field of phylogeography, Drosophilidae species have contributed to understanding both shared and idiosyncratic evolutionary histories (Hurtado *et al.*, 2004; Franco and Manfrin, 2013; De Ré *et al.*, 2014; Yukilevich *et al.*, 2018) and to assessing general diversification patterns (Markow, 2019).

In this paper, we reviewed the scientific literature on studies of phylogeography and genetic structure in Drosophilidae species, aiming to identify biases, gaps, and recent advances, and to synthesize the main findings and highlight congruences and discrepancies in phylogeographic patterns across biogeographic regions. For this task, we first identified and filtered our dataset. We then summarized each study according to its main characteristics, including the identity of the focal taxa, sampling design, molecular markers employed, main findings, and general implications. This allowed us to provide a comprehensive overview of the phylogeographic landscape of Drosophilidae for each biogeographic region.

Material and Methods

We conducted the literature surveys in March 2025, in the Web of Science (Clarivate Analytics) database, searching for papers published from 1987 to 2024 using the string TS=(drosophil*) AND TS=(“phylogeograph*” OR “population structure” OR “genetic structure” OR “genetic diversity” OR “demographic history” OR biogeograph* OR diversification OR dispersal OR colonization), that were searched in the

titles, abstracts and keywords. We then filtered the initial set of results to include only original papers.

We employed the ASReview tool (van de Schoot *et al.*, 2021), a machine learning-aided pipeline that applies active learning, for an initial automated screening. To train the model, we first reviewed the titles and abstracts of a random set of 200 articles, labeling each paper as either “relevant” or “non-relevant” to our search focus. The ASReview pipeline then prioritized the remaining studies based on their predicted likelihood of relevance. Screening was stopped after 300 consecutive non-relevant records, when we had already manually inspected 2,050 of the 3,602 papers. ASReview input and output files are presented in the Supplementary Material (Tables S1 and S2, respectively).

After recording the papers selected by ASReview in a Microsoft Excel spreadsheet, we extracted basic information from each study, as follows: (i) year and journal of publication; (ii) number and identity of species and species group(s) evaluated in the study, following the taxonomic data available on the TaxoDros database (Bächli, 2025); (iii) number of individuals and populations assessed, as well as their respective biogeographic regions; (iv) number, identity and inheritance patterns of the employed genetic markers; (v) whether the study included phenotype data; (vi) whether the study employed molecular clock to estimate divergence/diversification times; and (vii) whether the study employed Ecological Niche Modeling (ENM) as a complementary approach. After this, we performed a final filter step to exclude studies that either did not use molecular markers or included fewer than two populations. Biogeographical regions of the sampled species followed the regionalization proposed by Morrone (2015), and geological periods followed the scale proposed by Walker and Geissman (2022). Graphs were plotted in R 4.4.2 (R Core Team 2025) using the *ggplot2* package (Wickham, 2016).

Results and Discussion

General overview

In the initial Web of Science survey, we retrieved 4,169 articles, of which 3,602 were original papers (Table S1). When this dataset was submitted to ASReview screening, labeling of relevant papers clearly asymptoted after approximately 800 rounds (Figure S1a). However, we continued our manual inspection until round 2,050, in which the threshold of 300 consecutive non-relevant records was reached (Figure S1b). This process allowed us to reduce the number of papers to 312 (Table S2), of which 167 employed at least two populations and one molecular marker (Table S3). This dataset was then examined to evaluate temporal and journal-based patterns in the distribution of papers.

The number of publications per year ranged from 0 (in 1988) to 10 (in 2004 and 2007) and showed a steady increase over the first 30 years (Figure 1), following the trend reported by Beheregaray (2008). After that, the number of studies per year stabilized at approximately five. The 167 papers evaluated were published across 48 journals, most of which have high impact factors, with a weighted Journal Citation Reports average score of 3.65 (Clarivate Analytics, 2024) (Table 1).

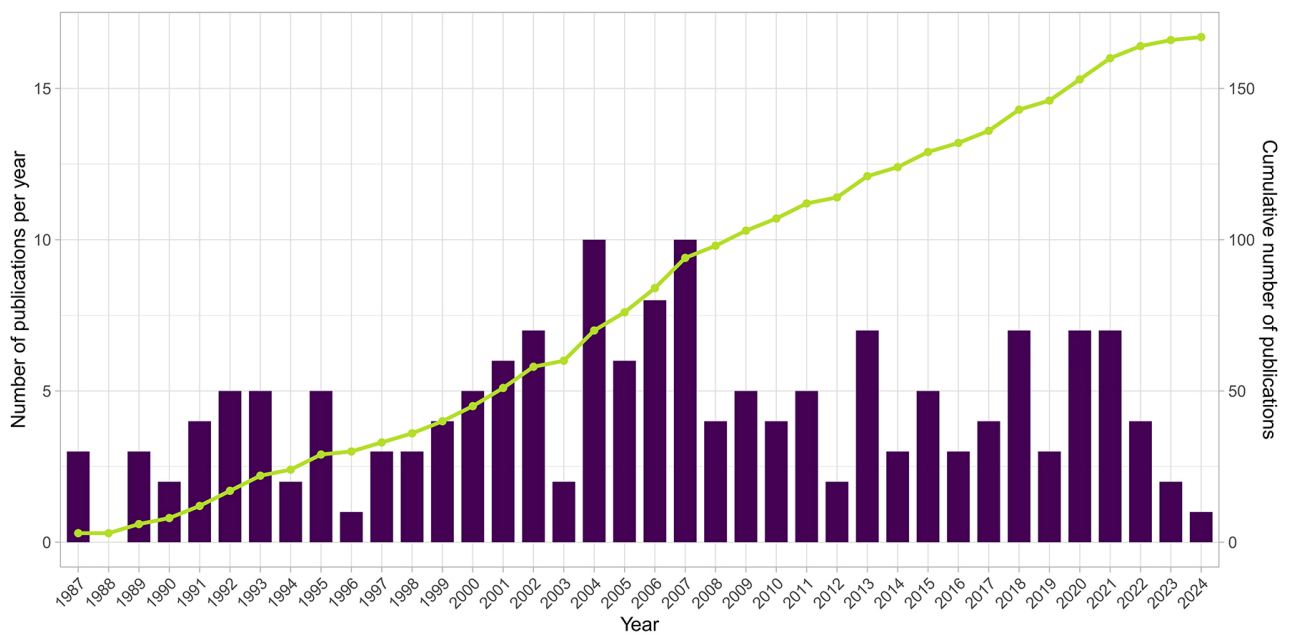


Figure 1 – Number (left Y axis, purple bar plot) and cumulative number (right Y axis, green line plot) of phylogeographic and genetic structure articles employing *Drosophilidae* species per year, from 1987 to 2024.

Table 1 – List of journals that published the retrieved articles, with the corresponding number of recovered publications and Journal Citation Reports (Clarivate Analytics, 2024) scores.

Journal	No of articles	JCR (2024)
Genetics	25	5.1
Molecular Ecology	23	3.9
Molecular Biology and Evolution	13	5.3
Journal of Evolutionary Biology	8	2.3
Evolution	7	2.6
Genetica	7	1.3
Journal of Heredity	7	2.5
Biological Journal of the Linnean Society	6	1.5
Heredity	6	3.9
Molecular Phylogenetics and Evolution	5	3.6
Journal of Pest Science	4	4.1
PloS One	4	2.6
Biological Invasions	3	2.6
Ecology and Evolution	3	2.3
Mitochondrial DNA Part A	3	0.6
Biochemical Genetics	2	1.6
Genetics and Molecular Biology	2	1.3
Genome Biology and Evolution	2	2.8
Hereditas	2	2.5
Journal of Biogeography	2	3.6
Journal of Molecular Evolution	2	1.8
Journal of Zoological Systematics and Evolutionary Research	2	2.6
Proceedings of the National Academy of Sciences of the United States of America	2	9.1
Proceedings of the Royal Society B – Biological Sciences	2	3.5
Zoological Studies	2	1.4
Anais da Academia Brasileira de Ciências	1	1.1

Table 1 – Cont.

Journal	No of articles	JCR (2024)
Annales de la Societe Entomologique de France	1	0.7
Annals of the Entomological Society of America	1	1.8
Biotropica	1	1.7
BMC Ecology and Evolution	1	2.6
BMC Genetics	1	2.9
BMC Genomics	1	3.7
European Journal of Entomology	1	1.2
G3-Genes Genomes Genetics	1	2.2
Genetics Research	1	2.1
Genetics Selection Evolution	1	3.1
Genome	1	1.8
Genome Research	1	5.5
Genomics	1	3.0
Journal of Economic Entomology	1	2.4
Journal of Genetics	1	1.2
Journal of Insect Science	1	2.0
Nature	1	48.5
Nucleic Acids Research	1	13.1
PloS Genetics	1	3.7
Russian Journal of Genetics	1	0.5
Canadian Entomologist	1	1.1
Zoological Science	1	1.0
TOTAL	167	3.65*

*Weighted average of impact factor.

The most frequent publication venues – “Genetics” and “Molecular Ecology” – accounted for more than one-quarter of all papers. Nevertheless, most journals have published only a single paper during the period analyzed, accounting for 13.77 % of the total retrieved publications.

Sampling bias at several taxonomic levels

Of the 77 *Drosophilidae* genera (Bächli, 2025), only four have been studied in a phylogeographic or genetic structure context: *Drosophila* (158 studies), *Mycodrosophila* (1 study), *Scaptodrosophila* (1 study), and *Zaprionus* (7 studies) (Table S3). There is also a strong bias toward certain *Drosophila* species groups, with only 16 of the 59 recognized groups (Bächli, 2025) represented in these studies (Figure 2A; Table S4). The *melanogaster* group stands out as the most extensively studied (80 articles), followed by the *repleta* (29 articles) and *obscura* (27 articles) groups of *Drosophila* (Figure 2A). The three groups also include the highest numbers of evaluated species – 26, 10, and 8, respectively (Table S4). Therefore, at least 43 *Drosophila* species groups remain to be studied in a phylogeographical context, in addition to the thousands of species belonging to the different *Drosophilidae* genera. This pronounced bias is likely explained by a disproportionate research interest in *D. melanogaster* and related species, as well as by difficulties in sampling and identifying non-*Drosophila* species. Many of these species are, in fact, typically collected in small numbers or through occasional sampling and can be

reliably identified only from male specimens by taxonomic specialists (Machado *et al.*, 2017).

A marked bias was also observed in the identity of the investigated species, with only 28 being represented in more than one study. In this context, more than half (~57 %) of the recorded studies focused on just four species: 49 on *D. melanogaster*, 18 on *D. subobscura*, 15 on *D. simulans*, and 12 on *D. suzukii* (Figure 2B; Table S4). Among these, the strong interest in *D. melanogaster* clearly stems from its fundamental role in advancing our understanding of genetics and evolution (Markow and O’Grady, 2006). *Drosophila simulans*, in turn, is not only closely related to *D. melanogaster* (Finet *et al.*, 2021) but has also achieved a worldwide distribution through a similar evolutionary history. Consequently, it is often used to compare genetic patterns with those observed for *D. melanogaster* (Singh *et al.*, 1987; Begun and Aquadro, 1993; Verspoor and Haddrill, 2011; Machado *et al.*, 2016). *Drosophila subobscura* is among the most extensively studied *Drosophila* species in Europe. Over the last 50 years, it has rapidly spread from the Palearctic into Nearctic and Neotropical regions, where it has developed clinal patterns for some chromosomal arrangements that parallel those observed in Old World populations (Prevosti *et al.*, 1988). Finally, *D. suzukii* (also known as the spotted-wing *Drosophila*) is native to East and Southeast Asia but has become a major agricultural pest in the Americas and Europe since the late 2000s (Attallah *et al.*, 2014), causing significant economic losses to fruit crops.

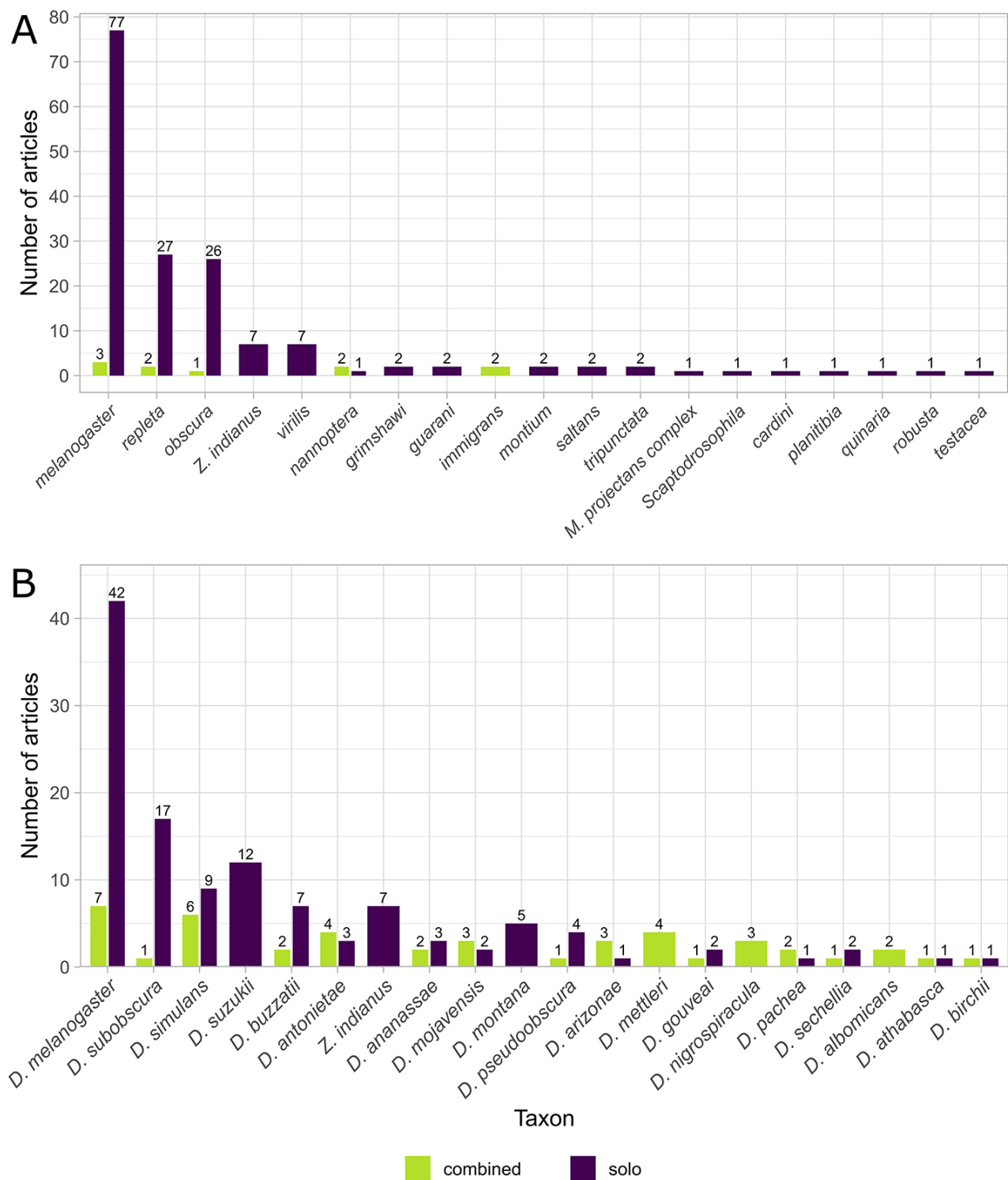


Figure 2 – Total number of articles studying different *Drosophilidae* species groups (A) and the top 20 most studied species in the family (B). Green bars represent studies that focused on two or more taxa; purple bars represent studies that focused on a single species.

Molecular markers

Considering inheritance patterns, most studies addressing phylogeography and genetic structure in *Drosophilidae* relied exclusively on nuclear markers (~64.1 %, 107 out of 167) (Figure 3; Table S5). Otherwise, only ~21 % (35 out of 167) of the studies focused solely on mitochondrial markers, and ~14.9 % (25 out of 167) combined both nuclear and

mitochondrial markers. This pattern differs from that reported by Beheregaray (2008) and Turchetto-Zolet *et al.* (2013), whose literature surveys found that ~72 % and ~55 % of the studies, respectively, employed only mitochondrial markers. Although this incongruence may partially reflect the historical context of each survey, showing a gradual shift in marker preference, it probably also reflects taxon-specific patterns.

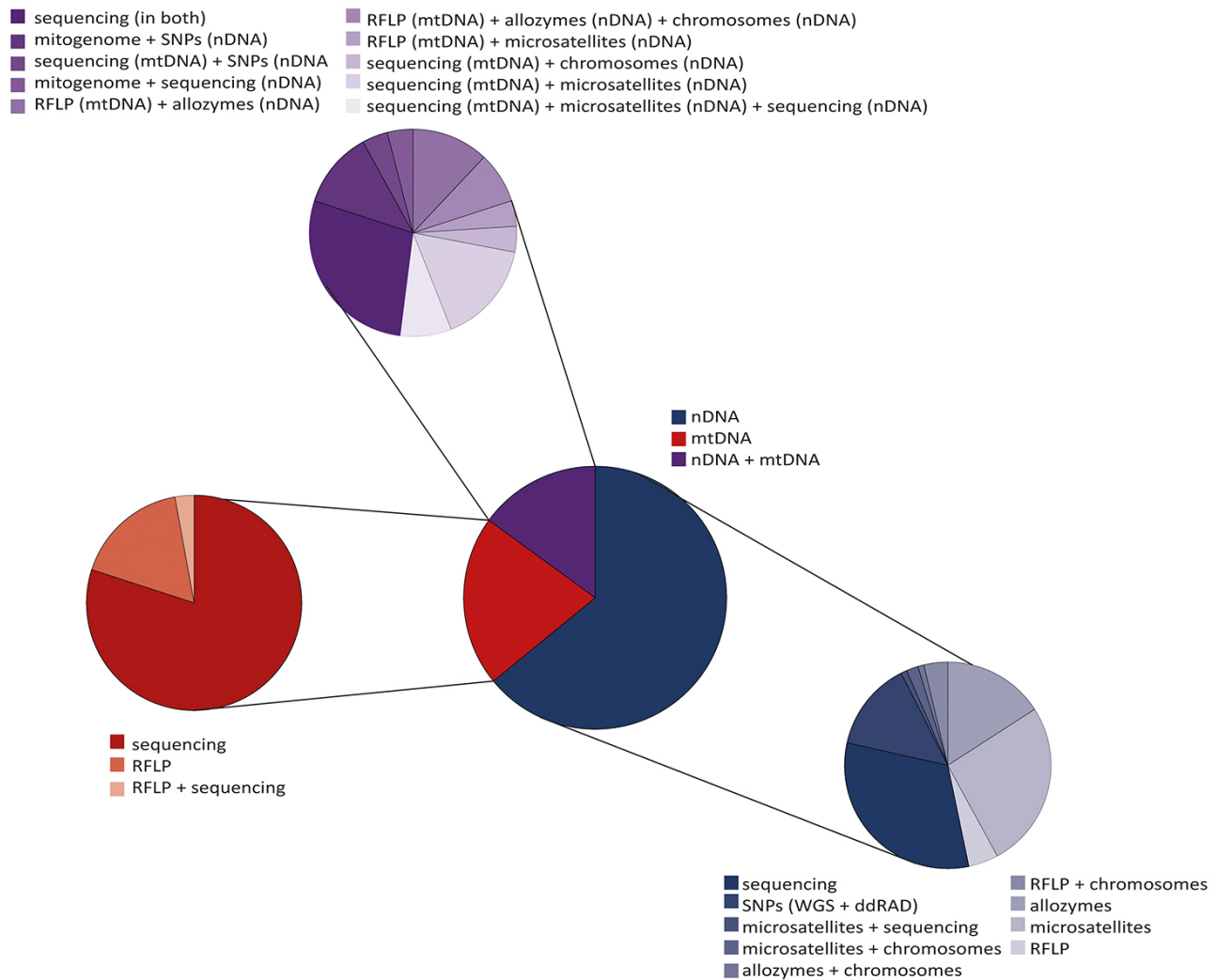


Figure 3 – Proportion of articles employing different genetic markers, according to their corresponding genetic compartment. Blue, red, and purple pie charts represent studies using nuclear, mitochondrial, and combined genetic markers, respectively.

Indeed, evolutionary research in *Drosophilidae* has a long tradition of using allozyme and RFLP markers, which together account for ~24.2 % of the studies using nuclear markers (32 out of 132). There is also a differential preference for microsatellite markers, employed in ~28.8 % of these studies (38 out of 132), likely reflecting the wide availability of microsatellite primers for some of the best-studied species in the *melanogaster*, *obscura*, and *repleta* groups, as well as their heterologous applications.

Concerning nuclear markers, phylogeography and genetic structure studies have, over the past decade, transitioned from using a few nuclear loci to exploring entire genomes – a shift that has given rise to the field of genomic phylogeography (reviewed in Brito and Edwards, 2009). This pattern can also be seen in *Drosophilidae* studies, where single-nucleotide polymorphisms (SNPs) characterized using Whole Genome Shotgun (WGS) strategies or reduced-representation approaches such as RAD-seq already represent the fourth most widely used class of nuclear markers. In fact, although Schlötterer and Harr (2002) were the first to employ SNPs to infer phylogeographic or population structure patterns in *Drosophilidae*, the first genome-wide SNP mapping conducted

with this purpose was performed by Pool *et al.* (2012), followed by 17 additional studies over the last 12 years (Table S5). Only two of these cases used reduced-representation approaches based on RAD-seq (Koch *et al.*, 2020; Nelson *et al.*, 2023).

As for nuclear markers, both historical traditions and recent advances likely contribute to explaining the observed patterns for mitochondrial markers. Among the 60 studies employing mitochondrial markers, 13 used RFLP to characterize restriction sites or fragment size from either the whole mitogenome or from specific mitochondrial genes (Table S5). Of the 43 articles analyzing mitochondrial gene sequences, cytochrome oxidase subunit I (COI) and cytochrome oxidase subunit II (COII) were the most commonly used genes, appearing in 26 and 13 studies, respectively. Only four studies employing mitochondrial markers used whole mitogenome sequences.

Frequently, the patterns obtained for molecular markers were complemented or compared with those observed at the chromosomal and phenotypic levels. Specifically, 10 of the recovered articles examined chromosome rearrangements, whereas 16 analyzed phenotypic traits (Table S3). In the last case, evaluations involved external and internal morphology

patterns, as well as behavioral and physiological data, ranging from geometric morphometry of wing landmarks (Moraes and Sene, 2007; Koch *et al.*, 2020; Barrios-Leal *et al.*, 2021; Machado *et al.*, 2022) to mate choice tests (Schug *et al.*, 2008; Yukilevich *et al.*, 2018). Nevertheless, although strategies associated with Environmental Niche Modelling (ENM) have become common as a complementary approach to addressing biogeographic patterns and demographic history across time in phylogeographic studies using information independent of genetic data (Knowles, 2009), they were employed in only six of the evaluated studies.

Biogeographic regions

The Palearctic region was represented in 80 of the 167 studies, making it the most frequently investigated biogeographical region in the dataset (Figure 4). The Nearctic and Neotropics follow as the second and third-most-studied regions, with 72 and 62 papers, respectively. In contrast, the Ethiopian, Oriental, and Australasian regions appear in 47, 25, and 22 studies, respectively. Interestingly, in most studies (87 out of 167), species were sampled across more than one biogeographic region, reflecting a prevalence of studies involving broadly distributed species rather than endemic ones.

Palearctic region

According to Morrone (2015), the Palearctic region comprises Europe, North Africa, and most of temperate Asia, extending from the Atlantic coasts to eastern Siberia and the Japanese archipelago. Of the 80 studies involving the Palearctic region, 38 were dedicated to solving different aspects of the evolutionary history of *D. melanogaster*, especially those related to its origin and demographic history (see, for example, Baudry *et al.*, 2004; Arguello *et al.*, 2019; Sprengelmeyer *et al.*, 2020), or population structure and admixture levels (see, for example, Pool *et al.*, 2012; Kao *et al.*, 2015). Demographic analysis performed by Sprenhelmeyer *et al.* (2020) suggests that populations of this species expanded from south-central Africa ~13 kya, but reached Europe much later, at 1.8 kya.

In addition, 14 papers have focused on Palearctic populations of *D. subobscura*, most of which investigated the species' introduction history into other biogeographic regions (Rozas *et al.*, 1995; Pinto *et al.*, 1997; Pascual *et al.*, 2007; Araújo *et al.*, 2011). According to Araújo *et al.* (2011), the colonization of the American continent by *D. subobscura* was most probably a single event involving colonizers from the western Mediterranean region. Otherwise, Brehm *et al.* (2004) evaluated population structure across Moroccan and Iberian populations of the species, rejecting the hypothesis that the Strait of Gibraltar has acted as a barrier to gene flow, and dating radiation to the Canary Islands to approximately 1.1 Mya.

Drosophila suzukii is another well-studied species in the Palearctic, where considerable effort has been devoted to investigating its colonization history (Lavrinienko *et al.*, 2017). In this context, Adrion *et al.* (2014) supported that invasions of Europe and the continental United States are independent demographic events associated with strong bottleneck signatures. Otherwise, Lewald *et al.* (2021) showed that European and Nearctic populations of *D. suzukii* encompass

two distinct clades, suggesting independent migrations from Asia. They also detected signals of population admixture from Western United States populations back to Asia.

In addition to the aforementioned *Drosophila* species, considerable attention (seven recorded studies) has also been given to Palearctic populations of *D. montana* and *D. virilis*, both belonging to the *virilis* group. According to Mirol *et al.* (2007), North American and European populations are clearly differentiated, with divergence dating back to the Pleistocene. Otherwise, Mirol *et al.* (2008) suggest a recent worldwide exponential expansion of the species associated either with post-glacial colonization or with domestication. Another interesting study involved a comparative phylogeographical study between three species of *Drosophila* [*D. albomicans* (*immigrans* group), *D. bipectinata* and *D. takahashi* (*melanogaster* group)], and one species of a *Drosophila*-parasitoid wasp (*Leptopilina ryukyuensis*). Specific phylogeographical patterns were found for each species, thus suggesting no correlation between host and parasite evolutionary histories. In this case, highly differentiated lineages were detected for *D. formosana* and *D. immigrans*, whose divergence was associated with Pliocene and Pleistocene interglacial events.

Nearctic region

The Nearctic region comprises most of North America, including Canada, the United States, and northern Mexico (Morrone, 2015). In total, phylogeographical studies in this region included 36 species belonging to the *melanogaster*, *nannoptera*, *obscura*, *quinaria*, *repleta*, *testacea*, *tripunctata*, and *virilis* groups of *Drosophila*, as well as *Zaprionus indianus* (Table S3). Nevertheless, once again, cosmopolitan, exotic, and invasive species such as *D. melanogaster*, *D. simulans*, *D. subobscura*, *D. suzukii*, and *Z. indianus* accounted for the majority of studies (45 out of 72).

Regarding *Z. indianus*, Comeault *et al.* (2021) suggested a single expansion from Africa to the western hemisphere, since Nearctic and Neotropical populations are genetically more similar to each other than to African lineages. Nonetheless, they also showed that populations from ancestral and introduced ranges exhibit shared patterns of genetic diversity that are consistent with parallel selection. Interestingly, signals of parallel differentiation have also been reported between northern and southern populations of Australian and North American populations of *Drosophila simulans* (Sedghifar *et al.*, 2016), underscoring the significant influence of regional environmental heterogeneity on the evolutionary dynamics of Drosophilidae species. In this sense, clines of latitudinal variation have already been reported for this region in different species, such as *D. melanogaster* and *D. simulans* (Costa *et al.*, 1992; Machado *et al.*, 2016) and *D. montana* (Wiberg *et al.*, 2021).

Studies on native cactophilic species of the *repleta* group – including *D. mojavensis*, *D. arizonae*, *D. mettleri*, *D. nigrospiracula*, *D. navojoa*, *D. mainlandi*, *D. hamatofila*, *D. aldrichi*, and the *D. anceps* species complex – have also revealed shared patterns of population differentiation and demographic history (Markow *et al.*, 2002; Hurtado *et al.*, 2004; Machado *et al.*, 2007). These studies consistently identified common phylogeographical patterns across the

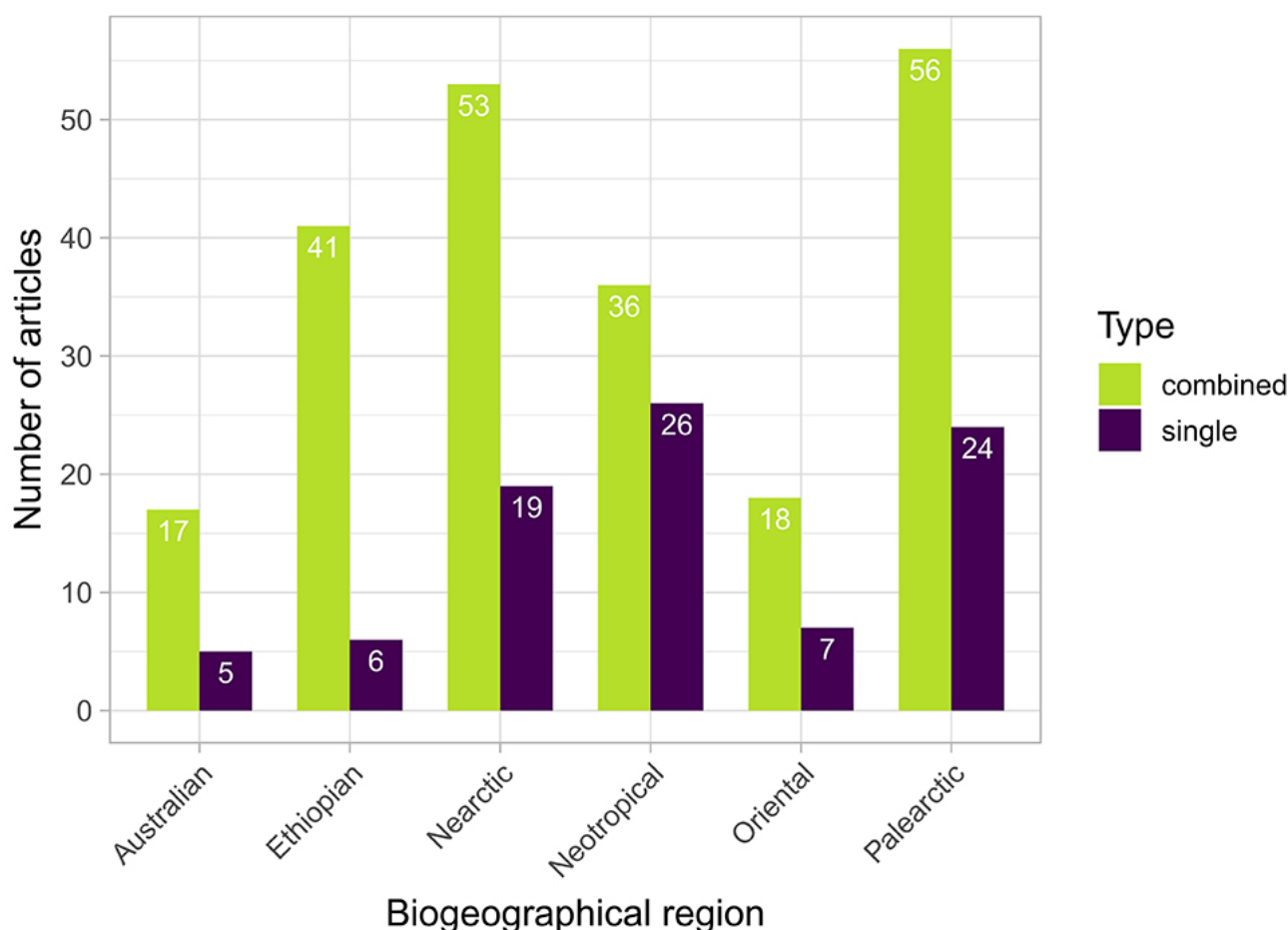


Figure 4 – Distribution of retrieved articles according to the biogeographical region of sampled populations, following Morrone (2015). Green bars represent studies including specimens sampled in two or more regions; purple bars represent studies that sampled specimens in a single region.

Sonoran Desert (southwestern USA and northwestern Mexico), shaped by host specialization, recent population expansions, and geographic differentiation. Geographic barriers across the Nearctic appear to play a key role in structuring genetic diversity in different species. Notable examples include the separation of Baja California from the mainland, which occurred 3-6 Mya (Lonsdale, 1989) and constrained the dispersal of *D. pachea* (Markow *et al.*, 2002; Hurtado *et al.*, 2004); and the Sierra Madre Occidental mountain range and the Trans-Mexican volcanic belt, which divide western and eastern, as well as northern and southern populations of *D. arizonae* (Machado *et al.*, 2007; Rampasso *et al.*, 2017). In some species, high levels of population structure have suggested the presence of geographic host races and incipient speciation (Rampasso *et al.*, 2017). Evidence for recent population expansions (Hurtado *et al.*, 2004; Pfeiler *et al.*, 2007; Pfeiler, 2019) has been linked to climatic oscillations that affected the Sonoran Desert during the Pleistocene (Van Devender, 2002).

Neotropical region

The Neotropics — encompassing southern Mexico, Central America (with the Caribbean islands), and South America (Morrone, 2015) — are widely recognized as the most biodiverse region on the planet (Raven *et al.*, 2020). This

diversity is reflected in the distribution of phylogeographic studies across various *Drosophila* groups, with representatives of the *cardini*, *guarani*, *melanogaster*, *obscura*, *repleta*, *saltans*, *tripunctata*, and *virilis* groups, as well as the genera *Mycodrosophila* and *Zaprionus*, being the focus of at least one recorded study. Notably, native species of the cactophilic *repleta* group account for ~37 % of the studies (23 out of 62), whereas those from the *melanogaster* group account for ~30 % (19 out of 62). Consequently, only 20 studies are distributed among the other lineages.

Several studies have shown that the current distribution of Drosophilidae populations in this region has been severely influenced by the climatic oscillations of the Pleistocene and Holocene, as well as by complex geographical and ecological barriers that fostered isolation, diversification, and the formation of regional refugia. Within this historical and environmental context, several interesting outcomes have emerged from studies on cactophilic species, such as the *D. buzzatii* cluster (Barrios-Leal *et al.*, 2019; Santos *et al.*, 2023) and the *D. serido* and *D. antonietae* haplogroups (Franco and Manfrin, 2013). In both cases, vicariant events, demographic fluctuations, and migratory routes were widely affected by Quaternary paleoclimatic changes on the spatial dynamics of the Seasonally Dry Tropical Forests (SDTFs) (Santos *et al.*, 2023). This has led to shared patterns of

distribution and divergence, as detected, for example, in the *D. antonietae* and *D. serido* clades, whose diversification was associated with the fragmentation of the SDTF during interglacial periods (Franco and Manfrin, 2013). Otherwise, whereas Pleistocene paleoclimatic oscillations appear to have led to a range expansion from Chaco to Cerrado and Catinga in *D. buzzatii* (Barrios-Leal *et al.*, 2019), they were associated with a deep population structure among mainland and coastal populations of *D. meridionalis* (Barrios-Leal *et al.*, 2018). In fact, responses of the *repleta* group species to climatic oscillations varied among species, possibly due to differences in ecology, distribution, and evolutionary potential imposed by genetic constraints.

Variations in ecological requirements also shaped distinct responses to Pleistocene climatic oscillations: whereas dry environment species expanded their populations during glacial periods, those of humid habitats exhibited the opposite pattern. A complementary perspective on this pattern comes from studies involving *D. maculifrons* and *D. ornatifrons*, two humid forest species that exhibit signs of population expansion during Pleistocene interglacial periods (De Ré *et al.*, 2014; Gustani *et al.*, 2015). These expansions coincide with increases in temperature and humidity in southern and southeastern Brazil, which promoted the expansion of forested areas (Carnaval *et al.*, 2009). Interestingly, several of these conclusions were reached through the combined use of molecular markers and Environmental Niche Models (ENM), whose concordant results certainly provided greater confidence to the inferred patterns (De Ré *et al.*, 2014; Barrios-Leal *et al.*, 2019).

Ethiopian region

The Ethiopian region comprises sub-Saharan Africa, the island of Madagascar, the northern Arabian Peninsula, and the West Indian Ocean islands (Morrone, 2015). Studies in this region included species belonging to the *melanogaster* and *obscura* groups of *Drosophila*, and *Zaprionus indianus* (Table S4). Nevertheless, the bias toward studies involving the *melanogaster* group is even more pronounced in this region, where these species were the focus of approximately 94 % of the recorded papers (44 out of 47 studies). In fact, the African continent has been consistently identified as the ancestral region of the cosmopolitan *D. melanogaster* (Baudry *et al.*, 2004; Arguello *et al.*, 2019) and the invasive *Z. indianus* (Markow *et al.*, 2014; Comeault *et al.*, 2021), which has motivated most phylogeographic and genetic structure studies in this region.

Despite the agreement involving the origin of *D. melanogaster* in the Ethiopian region, the precise location on the continent remains a matter of debate. Some authors have suggested that this occurred either in the Eastern (Benassi and Veuille, 1995; Baudry *et al.*, 2004) or the Western Africa (Dieringer *et al.*, 2005). Nevertheless, recent studies using genomic data have provided strong support for the hypothesis that *D. melanogaster* originated in the region of Austral Africa, in the territories currently comprising Zambia and Zimbabwe (Verspoor and Haddrill, 2011; Pool *et al.*, 2012; Sprengelmeyer *et al.*, 2020). The range expansion within Africa is estimated to have begun at least 72 kya, as dated for the divergence between Austral and Western populations (Kapopoulou *et al.*, 2018). In

contrast, the spread of *D. melanogaster* beyond the Ethiopian region — often referred to as the single “out of Africa” event (Baudry *et al.*, 2004) — appears to have occurred between 12 and 19 kya (Li and Stephan, 2006; Arguello *et al.*, 2019), and was followed by severe bottlenecks that substantially reduced the genetic diversity of non-African populations (Begun and Aquadro, 1993; Haddrill *et al.*, 2005). Finally, there is also compelling evidence suggesting that some populations subsequently migrated back to Africa (Benassi and Veuille, 1995; Pool *et al.*, 2012; Arguello *et al.*, 2019; Sprengelmeyer *et al.*, 2020; Coughlan *et al.*, 2022).

Concerning *Z. indianus*, Comeault *et al.* (2021) showed that introduced populations in South America and North America carry a genomic signature of a common range expansion, and were colonized by the western African lineage following a major admixture event between western and eastern African lineages. According to the best-fit demographic scenario, this split occurred ~600 generations (~35–60 years ago), which agrees with the first records of the species in the American continent around 1990 (van der Linde *et al.*, 2006). Accordingly, Mattos Machado *et al.* (2005) have evidenced the absence of genetic structure among Brazilian populations, supporting the hypothesis of a single colonization event. Otherwise, these authors also suggested a moderately sized propagule was introduced to Brazil, since invading populations were only slightly less diverse than ancestral ones. In America, northward migration of individuals from Brazil would have led to the spread of the species into Central America and Mexico (Markow *et al.*, 2014) or even into Argentina (Fernandez Goya *et al.*, 2020).

Oriental region

The Oriental region encompasses the tropical areas of Eurasia and Southeast Asia, including India, the Himalayas, Myanmar, Malaysia, Indonesia, the Philippines, and the islands of Micronesia, Polynesia, and Hawaii (Morrone, 2015). Phylogeographical studies of Drosophilidae in this region have also been strongly focused on the *melanogaster* group (18 of 25 studies), although some studies also evaluated endemic species (e.g., the *grimshawi* and *planitibia* groups).

Several studies have revealed patterns of population expansion and genetic structure in this biogeographic region (Das *et al.* 2004; Schug *et al.* 2007, 2008). Das *et al.* (2004), for example, traced the migration route of *D. ananassae* to a landmass called “Sundaland” in Southeast Asia during the Pleistocene (about 18 kya), when sea levels were lower. This region harbors five central populations of the species that exhibit high levels of nucleotide diversity and low linkage disequilibrium (Das *et al.* 2004), which are typical of ancestral populations. Migration from Sundaland appears to have occurred in a radial pattern, while subsequent patterns of dispersion in peripheral populations (in Asia and the South Pacific) possibly reflect historical human migratory routes (Das *et al.* 2004; Schug *et al.* 2007). In some cases, this history may have led to the evolution of distinct lineages or (incipient) cryptic species (Schug *et al.* 2007, 2008), some of which even exhibit some level of mating discrimination (Schug *et al.* 2008).

Interesting studies have also been conducted with Hawaiian species of the *grimshawi* group. For instance, Muir and Price (2008) compared the patterns of genetic diversity between populations of *D. engyochracea*, known from only two locations, and *D. hawaiiensis*, which is widespread across the archipelago. Both species were sampled from two isolated forest patches (*kipuka*). As expected, the more widely distributed *D. hawaiiensis* exhibited higher genetic diversity than the narrowly distributed *D. engyochracea*, although higher levels of gene flow were suggested for the latter. Moreover, there were some signals of mitochondrial introgression, evidenced by individuals morphologically identified as one species that carried haplotypes diagnostic of the other.

Australian region

According to Morrone (2015), the Australian region comprises Australia, New Caledonia, New Guinea, New Zealand, and other Oceanian islands. Studies in this region included species belonging to the *immigrans*, *melanogaster*, *montium*, *obscura*, and *repleta* groups of *Drosophila*, as well as the *Scaptodrosophila* genus (Table S3). Nevertheless, ~64 % of these studies focused on species of the *melanogaster* group.

Among cosmopolitan species, the peripheral populations of *D. ananassae* present in Australia and other South Pacific locations exhibit evidence of migration from Southeast Asia during the last glacial maximum (Das *et al.*, 2004; Schug *et al.*, 2008). Strong support comes from microsatellite data, which revealed a complex pattern involving isolation-by-distance, pronounced population structure, range expansion, and multiple colonization events in some receptor populations (Schug *et al.*, 2007). On the other hand, neither population structure nor isolation-by-distance was observed in Australian populations of *D. melanogaster*, likely reflecting their shared history of recent colonization (Agis and Schlötterer, 2001).

Some interesting comparative phylogeography studies were also performed for species occurring in this biogeographic region. For example, in the tropical forests of eastern Australia, contrasting phylogeographic patterns were evidenced for the pair of sister species *D. serrata* and *D. birchii* (*melanogaster* group). The first — a generalist, widely distributed species — harbors two highly divergent and geographically structured mtDNA lineages, whereas the second — a specialist species, restricted to rainforests — shows no significant population structure (Kelemen and Moritz, 1999). Two main hypotheses, not mutually exclusive, were proposed to explain the phylogeographical break in *D. serrata*: the splitting of populations into two distinct refugia during glacial periods, followed by secondary contact; or the formation of a pair of cryptic species. As for *D. birchii*, the low diversity and the lack of a geographical structure allowed hypothesizing a population expansion, which was later confirmed by Schiffer *et al.* (2007). Another interesting comparative study examined the endemics *Scaptodrosophila hibisci* and *S. acinata*, which reproduce in flowers of *Hibiscus* (McEvey and Barker, 2001) and appear to have diverged following a strong bottleneck in eastern Australia about 40 kya (Barker, 2005). In this case, *S. acinata* exhibited lower genetic variability, whereas

S. hibisci exhibited signals of population expansion (Kelemen and Moritz, 1999; Barker, 2005).

Another interesting set of studies involved the neotropical cactophilic *Drosophila buzzatii*, a notorious invasive Drosophilidae species in the Australian region. This invasion occurred in the decade of 1930, when the host plant *Opuntia* was introduced in eastern Australia, leading *D. buzzatii* populations into a strong bottleneck event, with an estimation of 30–40 founders (Piccinali *et al.*, 2007; Barker *et al.*, 2009; Barker, 2013). Subsequent reductions in the distribution of the invasive plant resulted in new bottlenecks and population differentiation (Barker *et al.*, 2009; Barker, 2013). Microsatellites and the α -esterase5 gene, in particular, revealed notable changes in haplotype frequencies between colonizing and Argentine populations, confirming the strong founder effect (Frydenberg *et al.*, 2002; Piccinali *et al.*, 2007).

Conclusions and perspectives

Our comprehensive review of phylogeographic and genetic structure studies in Drosophilidae identified substantial research gaps, particularly regarding understudied taxa and biogeographical regions. The straightforward focus on certain species or species groups underscores the potential of phylogeographic approaches to shed light on overlooked dimensions of biodiversity. Moreover, the uneven distribution of research efforts across biogeographic regions highlights the opportunity to uncover novel phylogeographic patterns and evolutionary processes. After all, these regions may hold critical insights not only into the evolutionary histories of focal species but also into broader patterns shaping regional biodiversity. Finally, our findings reinforce the importance of Drosophilidae as model organisms in evolutionary and biogeographic research. Continued use of drosophilids in such efforts offers a powerful framework for advancing our understanding of biodiversity dynamics at both local and global scales.

Acknowledgements

This study was funded by Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) (Finance Code 001) and Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) (L.J.R. has a research fellow – process 310308/2025-9).

Conflict of Interest

The authors declare there are no competing interests.

Author Contributions

HRMA and LJR designed and performed the study and wrote the first draft; MD and LJR supervised, reviewed and edited the final draft; all authors read and approved the final version.

Data Availability

The datasets analyzed and/or generated during the current study are published in the article and available in the Supplementary Material section.

References

- Adrion JR, Kousathanas A, Pascual M, Burrack HJ, Haddad NM, Bergland AO, Machado H, Sackton TB, Schlenke TA, Watada M *et al.* (2014) *Drosophila suzukii*: The genetic footprint of a recent, worldwide invasion. *Mol Biol Evol* 31:3148-3163.
- Agis M and Schlötterer C (2001) Microsatellite variation in natural *Drosophila melanogaster* populations from New South Wales (Australia) and Tasmania. *Mol Ecol* 10:1197-1205.
- Alfaro FM, Muñoz-Ramírez CP, Zúñiga-Reinoso Á, Trewick SA and Méndez MA (2018) Phylogeography of the Chilean red cricket *Cratomelus armatus* (Orthoptera: Anostostomatidae) reveals high cryptic diversity in central Chile. *Biol J Linn Soc* 123:712-727.
- Araúz PA, Peris-Bondia F, Latorre A, Serra L and Mestres F (2011) Molecular evidence to suggest the origin of a colonization: *Drosophila subobscura* in America. *Genetica* 139:1477-1486.
- Arguello JR, Laurent S and Clark AG (2019) Demographic history of the human commensal *Drosophila melanogaster*. *Genome Biol Evol* 11:844-854.
- Atallah J, Teixeira L, Salazar R, Zaragoza G and Kopp A (2014) The making of a pest: The evolution of a fruit-penetrating ovipositor in *Drosophila suzukii* and related species. *Proc Biol Sci* 281:20132840.
- Avise JC (2004) *Molecular Markers, Natural History, and Evolution*. 2nd edition. Sinauer Associates, Massachusetts.
- Avise JC (2009) Phylogeography: Retrospect and prospect. *J Biogeogr* 36:3-15.
- Avise JC, Arnold J, Ball RM, Bermingham E, Lamb T, Neigel JE, Reeb CA and Saunders NC (1987) Intraspecific phylogeography: The mitochondrial DNA bridge between population genetics and systematics. *Annu Rev Ecol Syst* 18:489-522.
- Barrios-Leal DY, Franco FF, Silva ECC, Santos CKB, Sene FM and Manfrin MH (2018) Deep intraspecific divergence in *Drosophila meridionalis*, a cactophilic member of the New World *Drosophila repleta* group. *Biol J Linn Soc* 123:163-178.
- Barrios-Leal DY, Neves-da-Rocha J and Manfrin MH (2019) Genetics and Distribution Modeling: The Demographic History of the Cactophilic *Drosophila buzzatii* Species Cluster in Open Areas of South America. *J Hered* 110:22-33.
- Barrios-Leal DY, Menezes RST, Ribeiro JV, Bizzo L, Sene FM, Rocha JN and Manfrin MH (2021) A holocenic and dynamic hybrid zone between two cactophilic *Drosophila* species in a coastal lowland plain of the Brazilian Atlantic Forest. *J Evol Biol* 34:1737-1751.
- Barker JSF (2005) Population structure and host-plant specialization in two *Scaptodrosophila* flower-breeding species. *Heredity* 94:129-138.
- Barker JSF (2013) Genetic history of a colonizing population: *Drosophila buzzatii* (Diptera: Drosophilidae) in Australia. *Biol J Linn Soc* 109:682-698.
- Barker JS, Frydenberg J, González J, Davies HI, Ruiz A, Sørensen JG and Loeschcke V (2009) Bottlenecks, population differentiation and apparent selection at microsatellite loci in Australian *Drosophila buzzatii*. *Heredity* 102:389-401.
- Baudry E, Viginier B and Veuille M (2004) Non-African populations of *Drosophila melanogaster* have a unique origin. *Mol Biol Evol* 21:1482-1491.
- Begun DJ and Aquadro CF (1993) African and North American populations of *Drosophila melanogaster* are very different at the DNA level. *Nature* 365:548-550.
- Beheregaray LB (2008) Twenty years of phylogeography: The state of the field and the challenges for the Southern Hemisphere. *Mol Ecol* 17:3754-3774.
- Benassi V and Veuille M (1995) Comparative population structuring of molecular and allozyme variation of *Drosophila melanogaster* *Adh* between Europe, West Africa and East Africa. *Genet Res* 65:95-103.
- Bonebrake TC, Ponisio LC, Boggs CL and Ehrlich PR (2010) More than just indicators: A review of tropical butterfly ecology and conservation. *Biol Conserv* 143:1831-1841.
- Brehm A, Harris DJ, Hernández M, Perez JA, Larruga JM, Pinto FM and González AM (2004) Phylogeography of *Drosophila subobscura* from North Atlantic islands inferred from mtDNA A+T rich region sequences. *Mol Phylogenet Evol* 30:829-834.
- Brito PH and Edwards SV (2009) Multilocus phylogeography and phylogenetics using sequence-based markers. *Genetica* 135:439-455.
- Carson HL (1971) *The ecology of Drosophila breeding sites*. University of Hawaii Foundation Lyon Arboretum Fund, Honolulu.
- Carnaval AC, Hickerson MJ, Haddad CF, Rodrigues MT and Moritz C (2009) Stability predicts genetic diversity in the Brazilian Atlantic forest hotspot. *Science* 323:785-789.
- Carvalho TL, Cordeiro J, Nornberg B, Schmitz HJ and Robe LJ (2023) Integrative taxonomy and evolutionary ecology of the anthophilous *Drosophila lutzii* species complex (Diptera, Drosophilidae) provide evidence for range expansion of *Drosophila alei*. *Insect Syst Evol* 54:378-401.
- Clancy D, Chtarbanova S and Broughton S (2023) Editorial: Model organisms in aging research: *Drosophila melanogaster*. *Front Aging* 3:1118299.
- Comeault AA, Kautt AF and Matute DR (2021) Genomic signatures of admixture and selection are shared among populations of *Zaprionus indianus* across the western hemisphere. *Mol Ecol* 30:6193-6210.
- Costa R, Peixoto AA, Barbuji G and Kyriacou CP (1992) A latitudinal cline in a *Drosophila* clock gene. *Proc Biol Sci* 250:43-49.
- Coughlan JM, Dagilis AJ, Serrato-Capuchina A, Elias H, Peede D, Isbell K, Castillo DM, Cooper BS and Matute DR (2022) Patterns of population structure and introgression among recently differentiated *Drosophila melanogaster* populations. *Mol Biol Evol* 39:msac223.
- Cuthill JFH and Charleston M (2015) Wing patterning genes and coevolution of Müllerian mimicry in *Heliconius* butterflies: Support from phylogeography, cophylogeny, and divergence times. *Evolution* 69:3082-3096.
- Das A, Mohanty S and Stephan W (2004) Inferring the population structure and demography of *Drosophila ananassae* from multilocus data. *Genetics* 168:1975-1985.
- De Ré FC, Gustani EC, Oliveira APF, Machado LPB, Mateus RP, Loreto ELS and Robe LJ (2014) Brazilian populations of *Drosophila maculifrons* (Diptera: Drosophilidae): Low diversity levels and signals of a population expansion after the Last Glacial Maximum. *Biol J Linn Soc* 112:55-66.
- Dieringer D, Nolte V and Schlötterer C (2005) Population structure in African *Drosophila melanogaster* revealed by microsatellite analysis. *Mol Ecol* 14:563-573.
- Eizirik E, Kim J-H, Menotti-Raymond M, Crawshaw Jr PG, O'Brien SJ and Johnson WE (2001) Phylogeography, population history and conservation genetics of jaguars (*Panthera onca*, Mammalia, Felidae). *Mol Ecol* 10:65-79.
- Fernandez Goya L, Imberti M, Rodriguero MS, Fanara JJ, Risso G and Lavagnino NJ (2020) Mitochondrial genetic diversity of the invasive drosophilid *Zaprionus indianus* (Diptera: Drosophilidae) in South America. *Biol Invasions* 22:3481-3486.
- Finet C, Kassner VA, Carvalho AB, Chung H, Day JP, Day S, Delaney EK, De Ré FC, Dufour HD, Dupim E *et al.* (2021) DrosoPhyla: Resources for drosophilid phylogeny and systematics. *Genome Biol Evol* 13:evab179.

- Franco FF and Manfrin MH (2013) Recent demographic history of cactophilic *Drosophila* species can be related to Quaternary palaeoclimatic changes in South America. *J Biogeogr* 40:142-154.
- Frydenberg J, Pertoldi C, Dahlgaard J and Loeschcke V (2002) Genetic variation in original and colonizing *Drosophila buzzatii* populations analysed by microsatellite loci isolated with a new PCR screening method. *Mol Ecol* 11:181-190.
- Garcez DK, Barbosa C, Loureiro M, Volcan MV, Loebmann D, Quintela FM and Robe LJ (2018) Phylogeography of the critically endangered neotropical annual fish, *Austrolebias wolterstorffi* (Cyprinodontiformes: Aplocheilidae): Genetic and morphometric evidence of a new species complex. *Environ Biol Fishes* 101:1503-1515.
- Gum B, Gross R and Kuehn R (2005) Mitochondrial and nuclear DNA phylogeography of European grayling (*Thymallus thymallus*): Evidence for secondary contact zones in central Europe. *Mol Ecol* 14:1707-1725.
- Gustani EC, Oliveira APF, Santos MH, Machado LPB and Mateus RP (2015) Demographic Structure and Evolutionary History of *Drosophila ornatifrons* (Diptera, Drosophilidae) from Atlantic Forest of Southern Brazil. *Zool Sci* 32:141-150.
- Hadrill PR, Thornton KR, Charlesworth B and Andolfatto P (2005) Multilocus patterns of nucleotide variability and the demographic and selection history of *Drosophila melanogaster* populations. *Genome Res* 15:790-799.
- Hurtado LA, Erez T, Castrezana S and Markow TA (2004) Contrasting population genetic patterns and evolutionary histories among sympatric Sonoran Desert cactophilic *Drosophila*. *Mol Ecol* 13:1365-1375.
- Jeratthitikul E, Hara T, Yago M, Itoh T, Wang M, Usami S and Hikida T (2013) Phylogeography of Fischer's blue, *Tongia fischeri*, in Japan: Evidence for introgressive hybridization. *Mol Phyl Evol* 66:316-326.
- Kao JY, Zubair A, Salomon MP, Nuzhdin SV and Campo D (2015) Population genomic analysis uncovers African and European admixture in *Drosophila melanogaster* populations from the south-eastern United States and Caribbean Islands. *Mol Ecol* 24:1499-1509.
- Kapopoulou A, Pfeifer SP, Jensen JD and Laurent S (2018) The demographic history of African *Drosophila melanogaster*. *Genome Biol Evol* 10:2338-2342.
- Kelemen L and Moritz C (1999) Comparative phylogeography of a sibling pair of rainforest *Drosophila* species (*Drosophila serrata* and *D. birchii*). *Evolution* 53:1306-1311.
- Kim BY, Gellert HR, Church SH, Suvorov A, Anderson SS, Barmina O, Beskid SG, Comeault AA, Crown KN, Diamond SE *et al.* (2024) Single-fly genome assemblies fill major phylogenomic gaps across the Drosophilidae Tree of Life. *PLoS Biol* 22:e3002697.
- Kjartanson SL, Haxton T, Wozney K, Lovejoy NR and Wilson CC (2023) Conservation genetics of lake sturgeon (*Acipenser fulvescens*): Nuclear phylogeography drives contemporary patterns of genetic structure and diversity. *Diversity* 15:385.
- Knowles LL (2009) Statistical Phylogeography. *Annu Rev Ecol Syst* 40:593-612.
- Koch JB, Dupuis JR, Jardeleza MK, Ouedraogo N, Geib SM, Follett PA and Price DK (2020) Population genomic and phenotype diversity of invasive *Drosophila suzukii* in Hawai'i. *Biol Invasions* 22:1753-1770.
- Kohler RE (1993) *Drosophila*: A life in the laboratory. *J Hist Biol* 26:281-310.
- Lavrinenko A, Kesäniemi J, Watts PC, Serga S, Pascual M, Mestres F and Kozeretska I (2017) First record of the invasive pest *Drosophila suzukii* in Ukraine indicates multiple sources of invasion. *J Pest Sci* 90:421-429.
- Lewald KM, Abrieux A, Wilson DA, Lee Y, Conner WR, Andreatza F, Beers EH, Burrack HJ, Daane KM, Diepenbrock L *et al.* (2021) Population genomics of *Drosophila suzukii* reveal longitudinal population structure and signals of migrations in and out of the continental United States. *G3: Genes Genomes Genet* 11:jkab343.
- Li H and Stephan W (2006) Inferring the demographic history and rate of adaptive substitution in *Drosophila*. *PLoS Genet* 2:e166.
- Lonsdale P (1989) Geology and tectonic history of the Gulf of California. In: Winterer EL, Hussong DM and Decker RW (eds) *The Geology of North America*. The Geological Society of America, Boulder, vol. N, pp 499-521.
- Machado CA, Matzkin LM, Reed LK and Markow TA (2007) Multilocus nuclear sequences reveal intra- and interspecific relationships among chromosomally polymorphic species of cactophilic *Drosophila*. *Mol Ecol* 16:3009-3024.
- Machado HE, Bergland AO, O'Brien KR, Behrman EL, Schmidt PS and Petrov DA (2016) Comparative population genomics of latitudinal variation in *Drosophila simulans* and *Drosophila melanogaster*. *Mol Ecol* 25:723-740.
- Machado S, Junges J, Fonseca P, Bolzan A, David JR, Loreto ELS, Gottschalk MS and Robe LJ (2017) Neotropical mycophilic drosophilids (Diptera: Drosophilidae): DNA barcoding as a way of overcoming the taxonomic impediment. *Insect Conserv Divers* 10:271-281.
- Machado S, Bessa MH, Nornberg B, Gottschalk MS and Robe LJ (2022) Unveiling the *Mycodrosophila projectans* (Diptera, Drosophilidae) species complex: Insights into the evolution of three Neotropical cryptic and syntopic species. *PLoS One* 17(5): e0268657.
- Markow TA (2019) Ecological and Evolutionary Genomics: The Cactophilic *Drosophila* Model System. *J Hered* 110:1-3.
- Markow TA, Castrezana S and Pfeiler E (2002) Flies across the water: Genetic differentiation and reproductive isolation in allopatric desert *Drosophila*. *Evolution* 56:546-552.
- Markow TA and O'Grady PM (2006) *Drosophila*: A Guide to Species Identification and Use. Academic Press, Cambridge.
- Markow TA, Hanna G, Riesgo-Escovar JR, Tellez-Garcia AA, Richmond MP, Nazario-Yepiz NO, Laclette MRL, Carpinteyro-Ponce J and Pfeiler E (2014) Population genetics and recent colonization history of the invasive drosophilid *Zaprionus indianus* in Mexico and Central America. *Biol Invasions* 16:2427-2434.
- Mattos Machado T, Solé-Cava AM, David JR and Bitner-Mathé BC (2005) Allozyme variability in an invasive drosophilid, *Zaprionus indianus* (Diptera: Drosophilidae): Comparison of a recently introduced Brazilian population with Old World populations. *Ann Soc Entomol Fr* 41:7-13.
- McEvey SF and Barker JSF (2001) *Scaptodrosophila acclinata*: A new *Hibiscus* flower-breeding species related to *S. hibisci* (Diptera: Drosophilidae). *Rec Aust Mus* 53:255-262.
- Mirol PM, Schafer MA, Orsini L, Routtu J, Schlötterer C, Hoikkala A and Butlin RK (2007) Phylogeographic patterns in *Drosophila montana*. *Mol Ecol* 16:1085-1097.
- Mirol PM, Routtu J, Hoikkala A and Butlin RK (2008) Signals of demographic expansion in *Drosophila virilis*. *BMC Evol Biol* 8:59.
- Moraes EM and Sene FM (2007) Microsatellite and morphometric variation in *Drosophila gouveai*: The relative importance of historical and current factors in shaping the genetic population structure. *J Zool Syst Evol Res* 45:336-344.
- Morrone JJ (2015) Biogeographical regionalisation of the world: A reappraisal. *Aust Syst Bot* 28:81-90.
- Muir C and Price DK (2008) Population structure and genetic diversity in two species of Hawaiian picture-winged *Drosophila*. *Mol Phyl Evol* 47:1173-1180.

- Nelson TD, Uriel Y, Abram PK, Moffat CE, Sherwood J, Rankema JM, Moreau D and Franklin MT (2023) Spotted-wing drosophila, *Drosophila suzukii* (Diptera: Drosophilidae), exhibits large-scale spatial genetic structure across Canada but not fruit host-associated genetic structure. *Can Entomol* 155:e37.
- Pascual M, Chapuis MP, Mestres F, Balanyà J, Huey RB, Gilchrist GW, Serra L and Estoup A (2007) Introduction history of *Drosophila subobscura* in the New World: A microsatellite-based survey using ABC methods. *Mol Ecol* 16:3069-3083.
- Pfeiler E (2019) Genetic diversity and demographic history in the cactophilic *Drosophila repleta* species group (Diptera: Drosophilidae) in North America inferred from mitochondrial DNA barcodes. *J Hered* 110:34-45.
- Pfeiler E, Erez T, Hurtado LA and Markow TA (2007) Genetic differentiation and demographic history in *Drosophila packae* from the Sonoran Desert. *Hereditas* 144:63-74.
- Piccinali RV, Mascord LJ, Barker JS, Oakeshott JG and Hasson E (2007) Molecular population genetics of the alpha-esterase5 gene locus in original and colonized populations of *Drosophila buzzatii* and its sibling *Drosophila koepferae*. *J Mol Evol* 64:158-170.
- Pinto FM, Brehm A, Hernandez M, Larruga JM, Gonzalez AM and Cabrera VM (1997) Population genetic structure and colonization sequence of *Drosophila subobscura* in the canaries and madeira Atlantic islands as inferred by autosomal, sex-linked and mtDNA traits. *J Hered* 88:108-114.
- Pool JE, Corbett-Detig RB, Sugino RP, Stevens KA, Cardeno CM, Crepeau MW, Duchen P, Emerson JJ, Saelao P, Begun DJ *et al.* (2012) Population genomics of sub-Saharan *Drosophila melanogaster*: African diversity and non-African admixture. *PLoS Genet* 8:e1003080.
- Powell JR (1997) Progress and prospects in evolutionary biology: The *Drosophila* model. Oxford University Press, New York.
- Prevosti A, Ribo G, Serra L, Aguade M, Balaña J, Monclus M and Mestres F (1988) Colonization of America by *Drosophila subobscura*: Experiment in natural populations that supports the adaptive role of chromosomal-inversion polymorphism. *Proc Natl Acad Sci U S A* 85:5597-5600.
- Rampasso AS, Markow TA and Richmond MP (2017). Genetic and phenotypic differentiation suggests incipient speciation within *Drosophila arizonae* (Diptera: Drosophilidae). *Biol J Linn Soc* 122:444-454.
- Raven PH, Gereau RE, Phillipson PB, Chatelain C, Jenkins CN and Ulloa Ulloa C (2020) The distribution of biodiversity richness in the tropics. *Sci Adv* 6:eabc6228.
- Rosauer DF, Blom MPK, Bourke G, Catalano S, Donnellan S, Gillespie G, Mulder E, Oliver PM, Potter S, Pratt RC *et al.* (2016) Phylogeography, hotspots and conservation priorities: An example from the Top End of Australia. *Biol Conserv* 204:83-93.
- Rozas J, Segarra C, Zapata C, Alvarez G and Aguade M (1995) Nucleotide polymorphism at the *rp49* region of *Drosophila subobscura*: Lack of geographic subdivision within chromosomal arrangements in Europe. *J Evol Biol* 8:355-367.
- Santos MM, Barrios-Leal DY and Manfrin MH (2023) Phylogeography of *Drosophila buzzatii* (Diptera, Drosophilidae): responses of the species to Quaternary climates in tropical and subtropical South America. *An Acad Bras Cienc* 95:e20220846.
- Sedghifar A, Saelao P, Begun DJ (2016) Genomic Patterns of geographic differentiation in *Drosophila simulans*. *Genetics* 202:1229-1240.
- Schiffer M, Kennington WJ, Hoffmann AA and Blacket MJ (2007) Lack of genetic structure among ecologically adapted populations of an Australian rainforest *Drosophila* species as indicated by microsatellite markers and mitochondrial DNA sequences. *Mol Ecol* 16:1687-1700.
- Schug MD, Smith SG, Tozier-Pearce A and McEvey SF (2007) The genetic structure of *Drosophila ananassae* populations from Asia, Australia and Samoa. *Genetics* 175:1429-1440.
- Schug MD, Baines JF, Killon-Atwood A, Mohanty S, Das A, Grath S, Smith SG, Zargham S, McEvey SF and Stephan W (2008) Evolution of mating isolation between populations of *Drosophila ananassae*. *Mol Ecol* 17:2706-2721.
- Schlötterer C and Harr B (2002) Single nucleotide polymorphisms derived from ancestral populations show no evidence for biased diversity estimates in *Drosophila melanogaster*. *Mol Ecol* 11:947-950.
- Singh RS, Choudhary M and David JR (1987) Contrasting patterns of geographic variation in the cosmopolitan sibling species *Drosophila melanogaster* and *Drosophila simulans*. *Biochem Genet* 25(1/2):27-40.
- Song W, Cao L-J, Li B-Y, Gong Y-J, Hoffmann AA and Wei S-J (2018) Multiple refugia from penultimate glaciations in East Asia demonstrated by phylogeography and ecological modelling of an insect pest. *BMC Evolution Biol* 18:152.
- Sprengelmeyer QD, Mansourian S, Lange JD, Matute DR, Cooper BS, Jirle EV, Stensmyr MC and Pool JE (2020) Recurrent collection of *Drosophila melanogaster* from wild African environments and genomic insights into species history. *Mol Biol Evol* 37:627-638.
- Throckmorton LH (1975) The Phylogeny, Ecology, and Geography of *Drosophila*. In: King RC (ed) Handbook of Genetics. Plenum Publishing, New York, pp 421-469.
- Turchetto-Zolet AC, Pinheiro F, Salgueiro F and Palma-Silva C (2013) Phylogeographical patterns shed light on evolutionary process in South America. *Mol Ecol* 22:1193-1213.
- Van de Schoot R, Bruin J, Schram R, Zahedi P, Boer J, Weijdemans F, Kramer B, Huijts M, Hoogerwerf M, Ferdinands G *et al.* (2021) An open source machine learning framework for efficient and transparent systematic reviews. *Nat Mach Intell* 3:125-133.
- Van der Linde K, Steck GJ, Hibbard K, Birdsley JS, Alonso L M and Houle D (2006) First records of *Zaprionus indianus* (Diptera: Drosophilidae), a pest species on commercial fruits from Panama and the United States of America. *Fla Entomol* 89:402-404.
- Van Devender TR (2002) Environmental history of the Sonoran desert. In: Fleming TH and Valiente-Banuet A (eds) Columnar Cacti and Their Mutualists. University of Arizona Press, Tucson, pp. 3-24.
- Verspoor RL and Haddrill PR (2011) Genetic diversity, population structure and *Wolbachia* infection status in a worldwide sample of *Drosophila melanogaster* and *D. simulans* populations. *PLoS One* 6:e26318.
- Wickham H (2016) ggplot2: Elegant Graphics for Data Analysis. 2nd edition. Springer Nature, Switzerland.
- Wiberg RAW, Tyukmaeva V, Hoikkala A, Ritchie MG and Kankare M (2021) Cold adaptation drives population genomic divergence in the ecological specialist, *Drosophila montana*. *Mol Ecol* 30:3783-3796.
- Yukilevich R, Maroja LS, Nguyen K, Hussain S and Kumaran P (2018) Rapid sexual and genomic isolation in sympatric *Drosophila* without reproductive character displacement. *Ecol Evol* 8:2852-2867.
- Zhang L-J, Cai W-Z, Luo J-Y, Zhang S, Wang C-Y, Lv L-M, Zhu X-Z, Wang L and Cui J-J (2017) Phylogeographic patterns of *Lygus pratensis* (Hemiptera: Miridae): Evidence for weak genetic structure and recent expansion in northwest China. *PLoS One* 12:e0174712.
- Zhao Y, Pan B and Zhang M (2019) Phylogeography and conservation genetics of the endangered *Tugarinovia mongolica* (Asteraceae) from Inner Mongolia, Northwest China. *PLoS One* 14:e0211696.

Internet Resources

- Bächli G (2025) TaxoDros: The database on Taxonomy of Drosophilidae, <https://www.taxodros.uzh.ch/> (accessed 3 September 2025).
- Clarivate Analytics (2024) Journal Citation Reports, <https://jcr.clarivate.com> (accessed 3 September 2025).
- R Core Team (2025) R: A language and environment for statistical computing, <https://www.R-project.org> (accessed 3 September 2025).
- Walker JD and Geissman JW (2022) GSA Geologic Time Scale Version 6.0, <https://doi.org/10.1130/2022.CTS006C> (accessed 3 September 2025).

Supplementary material

The following online material is available for this article:

Figure S1 – Performance of model training and automatic screenings on ASReview.

Table S1 – Complete set of original articles retrieved by the search string in Web of Science.

Table S2 – Set of articles that passed the automatic screening of ASReview under a machine learning framework.

Table S3 – Final set of articles that passed the last manual screening step, along with their corresponding attributes regarding the studied species and species groups, sampling design (number of populations and evaluated specimens), biogeographical regions, and genetic markers.

Table S4 – Summary of the number of articles retrieved for each species, along with their corresponding taxonomic positioning.

Table S5 – Summary of the number of articles employing different genetic markers, according to their corresponding genetic compartment.

Associate Editor: Klaus Hartfelder

License information: This is an open-access article distributed under the terms of the Creative Commons Attribution License (type CC-BY), which permits unrestricted use, distribution and reproduction in any medium, provided the original article is properly cited.