Allelic Diversity at the Merozoite Surface Protein-1 (MSP-1) Locus in Natural Plasmodium falciparum Populations: a Brief Overview

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The merozoite surface protein-1 (MSP-1) locus of Plasmodium falciparum codes for a major asexual blood-stage antigen currently proposed as a major malaria vaccine candidate. The protein, however, shows extensive polymorphism, which may compromise its use in sub-unit vaccines. Here we compare the patterns of allelic diversity at the MSP-1 locus in wild isolates from three epidemiologically distinct malaria-endemic areas: the hypoendemic southwestern Brazilian Amazon (n = 54), the mesoendemic southern Vietnam (n = 238) and the holoendemic northern Tanzania (n = 79). Fragments of the variable blocks 2, 4a, 4b and 6 or 10 of this single-copy gene were amplified by the polymerase chain reaction, and 24 MSP-1 gene types were defined as unique combinations of allelic types in each variable block. Ten different MSP-1 types were identified in Brazil, 23 in Vietnam and 13 in Tanzania. The proportion of genetically mixed infections (isolates with parasites carrying more than one MSP-1 version) ranged from 39% in Brazil to 44% in Vietnam and 60% in Tanzania. The vast majority (90%) of the typed parasite populations from Brazil and Tanzania belonged to the same seven most frequent MSP-1 gene types. In contrast, these seven gene types corresponded to only 61% of the typed parasite populations from Vietnam. Non-random associations were found between allelic types in blocks 4a and 6 among Vietnamese isolates, the same pattern being observed in independent studies performed in 1994, 1995 and 1996. These results suggest that MSP-1 is under selective pressure in the local parasite population. Nevertheless, the finding that similar MSP-1 type frequencies were found in 1994 and 1996 argues against the prominence of short-term frequency-dependent immune selection of MSP-1 polymorphisms. Non-random associations between MSP-1 allelic types, however, were not detected among isolates from Brazil and Tanzania. A preliminary analysis of the distribution of MSP-1 gene types per host among isolates from Tanzania, but not among those from Brazil and Vietnam, shows significant deviation from that expected under the null hypothesis of independent distribution of parasites carrying different gene types in the human hosts. Some epidemiological consequences of these findings are discussed.

Key words: Plasmodium falciparum - malaria - allelic diversity - merozoite surface protein-1 - population genetics - vaccine candidate

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The polymorphic *merozoite surface protein-1* (MSP-1) of Plasmodium falciparum is a major asexual blood-stage malaria vaccine candidate (Holder 1996). Comparisons of nucleotide sequences led to the identification of seven variable blocks in the gene, which are interspersed with five conserved and five semi-conserved blocks (Fig. 1). There are essentially two versions of each block, named after the representative isolates MAD20 and K1 (Tanabe et al. 1987). A major exception to this dimorphic rule is the variable block 2, that has a third version originally described in the isolate RO33 (Certa et al. 1987). Most allelic diversity is generated by intragenic recombination between these representative sequences at the 5' end of the gene, within blocks 3, 4 and 5. Minor differences also exist between homologous versions of the same variable block, and nucleotide substitutions (most of which are dimorphic) occur in semi-conserved and conserved blocks (Tanabe et al. 1987).

Major MSP-1 gene types may be defined as unique combinations of: (a) one of three versions of block 2 (MAD20, K1 or RO33), (b) one of four possible versions of block 4, because recombination within this region generates MAD20/K1 and K1/MAD20 hybrids in addition to the 'pure' allelic types MAD20 and K1 (Conway et al. 1991b, Kaneko et al. 1996), and (c) one of two versions (MAD20 or K1) of the segment between the variable blocks 6 and 16, that comprises about 60% of the gene. Recombination events have not been described in this portion of the gene (Tanabe et al. 1987, 1989, Peterson et al. 1988, Conway et al. 1991b, Jongwutiwes et al. 1991, Kaneko et al. 1996, 1997). Therefore, the 24 MSP-1 gene types shown in Table I may theoretically be observed in natural parasite populations (Kaneko et al. 1997).

The extent of allelic diversity in different malaria-endemic areas should be evaluated if the variable domains of MSP-1, that are highly immunogenic (Holder & Riley 1996), are to be included in subunit malaria vaccines. A novel polymerase chain reaction (PCR)-based strategy was recently developed to group clinical isolates of P. falciparum into the 24 MSP-1 gene types defined in Table I (Kaneko et al. 1997). This strategy has been successfully applied to type wild isolates from the mesoendemic southern Vietnam (Kaneko et al. 1997, Ferreira et al. 1998b), the hypoendemic Brazilian Amazon (Ferreira et al. 1998a), and the holoendemic Tanzania (Ferreira et al. 1998c). In this communication we analyze available data regarding complete MSP-1 typing of isolates from these three malaria-endemic areas.

MATERIALS AND METHODS

Table II summarizes basic information regarding typed *P. falciparum* isolates in each malaria-endemic area. Genomic DNA was extracted directly from the blood of *P. falciparum*-infected patients, without previous *in vitro* cultivation of parasites. Locations of the oligonucleotide primers are shown in Fig. 1. Primer sequences and PCR protocols are given elsewhere (Kaneko et al. 1997). The basic PCR-based typing procedure developed by Kaneko et al. (1997) may be described as it follows:

First step - Block 2 was typed in three separate reactions with the allelic specific forward primers M2F, K2F and R2F and the common reverse primer C3R.

TABLE I

Merozoite surface protein-1 (MSP-1) gene types defined as unique combinations of allelic types in each variable block

	Variable block			
Gene type	2	4a	4b	10
1	K1	K1	K1	K1
2	MAD20	K1	K1	K1
3	R033	K1	K1	K1
4	K1	MAD20	K1	K1
5	MAD20	MAD20	K1	K1
6	RO33	MAD20	K1	K1
7	K1	K1	MAD20	K1
8	MAD20	K1	MAD20	K1
9	RO33	K1	MAD20	K1
10	K1	MAD20	MAD20	K1
11	MAD20	MAD20	MAD20	K1
12	RO33	MAD20	MAD20	K1
13	K1	K1	K1	MAD20
14	MAD20	K1	K1	MAD20
15	RO33	K1	K1	MAD20
16	K1	MAD20	K1	MAD20
17	MAD20	MAD20	K1	MAD20
18	RO33	MAD20	K1	MAD20
19	K1	K1	MAD20	MAD20
20	MAD20	K1	MAD20	MAD20
21	RO33	K1	MAD20	MAD20
22	K1	MAD20	MAD20	MAD20
23	MAD20	MAD20	MAD20	MAD20
24	RO33	MAD20	MAD20	MAD20

a: each gene type is defined as a unique combination of allelic types detected in the variable blocks 2, 4a (5' segment of block 4), 4b (3' segment of block 4) and 6-16 of the MSP-1 gene. Since there is no recombination at the central and C-terminal portions of this gene, the allelic type detected in block 10 is considered to be the same for the variable blocks 6, 8, 14 and 16. Allelic types are named after the reference isolates MAD20, K1 and RO33.

Second step - The gene fragments between the conserved block 5 and the variable block 6 were amplified in two separate reactions with the common forward primer C5F and the type-specific reverse primers M6R or K6R. Alternatively, block 10 was typed with the semi-conserved forward primer C9F and the type-specific reverse primers M10R and K10R. Since there is no recombination between blocks 6 and 16, the allelic type found in blocks 6 or 10 is the same for variable blocks 8, 14 and 16.

Third step - Segments between blocks 2 and 6 were amplified in three separate reactions with the type-specific forward primers M2F, K2F or R2F, and the type-specific reverse primers M6R or K6R. The PCR fragments amplified in this step were used as template in the next step. As an alternative, this

TABLE II
Recent polymerase chain reaction-based studies involving complete typing of the merozoite surface protein-1
gene in natural <i>Plasmodium falciparum</i> populations

Area	Malaria endemicity	No. of typed isolates	Reference
Brazilian Amazon	Low	54	Ferreira et al. 1998a
Southern Vietnam	Intermediate	136	Kaneko et al. 1997
Southern Vietnam	Intermediate	102	Ferreira et al. 1998b
Northern Tanzania	High	79	Ferreira et al. 1998c

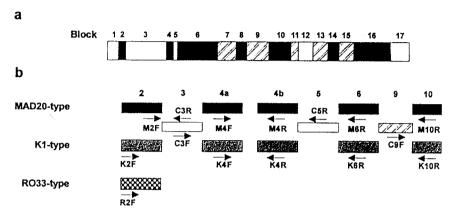


Fig. 1-a: structure of the *merozoite surface protein-1* gene of *Plasmodium falciparum*. Conserved, semi-conserved and variable blocks of the gene are shown as open, hatched and closed boxes, respectively. Block numbers are after Tanabe et al. (1987); b: locations and directions of the oligonucleotide primers used to type blocks 2, 4a, 4b, 6, and 10 are also indicated. Redrawn from Kaneko et al. 1997.

template may be prepared with the conserved forward primer C3F and the conserved reverse primer C5R.

Forth step - Block 4 was typed by nested PCR in four separate reactions with the type-specific forward primers M4F or K4F and type-specific reverse primers M4R or K4R. As an alternative, the first step may be eliminated, and block 2 may be typed by detecting allelic-specific fragments in the second step (Ferreira et al. 1998b).

The detection of PCR products in the expected size ranges after 1.5-2% agarose gel electrophoresis defined the presence of each allelic type in blocks 2, 6 or 10, 4a and 4b. As *MSP-1* is a single-copy gene in the haploid genome of blood-stage parasites, we consider that isolates harboring more than one gene type have mixed infections with genetically distinct *P. falciparum* subpopulations. Each subpopulation may be separately typed by this approach (Kaneko et al. 1997).

RESULTS AND DISCUSSION

Are all theoretically possible MSP-1 gene types found in natural Plasmodium falciparum populations? - As shown in Fig. 2, all but one of the 24 possible MSP-1 gene types were detected in

mesoendemic Vietnam. Only gene type 11 was absent in that area at both occasions (Kaneko et al. 1997, Ferreira et al. 1998b). This suggests that almost all possible combinations of MSP-1 allelic types may be found in parasites that are able to infect human hosts. In contrast, only 10 and 13 MSP-1 types were found in hypoendemic Brazil and holoendemic Tanzania, respectively. Moreover, essentially the same MSP-1 gene types were found to predominate in both countries, and about 90% of the typed parasite populations belonged to the seven most common gene types, namely the types 13, 16, 17, 18, 22, 23 and 24 as defined in Table I (Ferreira et al. 1998c). Nevertheless, these seven gene types were found in only 61% of the typed parasite populations in Vietnam.

Is there any association between the extent of MSP-1 diversity and the intensity of malaria transmission? - If we compare the proportions of genetically mixed infections (that is, patients harboring more than one MSP-1 gene type) and the average number of MSP-1 gene types found per patient, an apparent positive association is found between malaria endemicity and MSP-1 diversity (Table III). However, if we compare the number of different MSP-1 gene types found in each en-

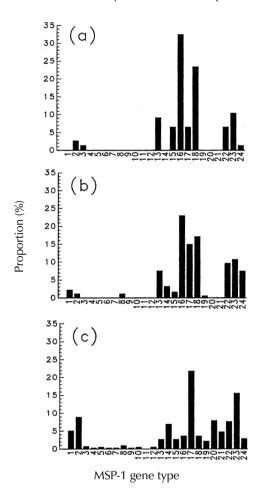


Fig. 2-a: frequency distribution of the *merozoite surface protein-1* (MSP-1) gene types in 54 Plasmodium falciparum isolates collected in July 1995 in the city of Porto Velho, State of Rondônia, southwestern Brazilian Amazon (Ferreira et al. 1998a); b: frequency distribution of the MSP-1 gene types in 79 P. falciparum isolates collected between July and September 1996 in the city of Tanga and the nearby village of Pangani, in northern Tanzania (Ferreira et al. 1998c); c: frequency distribution of the MSP-1 gene types in 238 P. falciparum isolates collected between July 1994 and July 1996 from malaria patients belonging to the ethnic majority Kinh and the minority K'ho living in the towns of Bao Loc and Phu Rieng and nearby areas in southern Vietnam (Kaneko et al. 1997, Ferreira et al. 1998b). The 24 MSP-1 gene types are numbered as in Table I.

demic area, no such association can be detected. Therefore, despite the fact that most infected hosts in Tanzania carry two or more parasite clones which may be ingested by the vector and recombine during meiosis, the resulting repertoire of MSP-1 variants seems to be relatively restricted in human hosts, if compared with the situation found in Vietnam. Strong selective pressure related to the sequential use of several different antimalarials in a few years, in the context of multi-drug resistance, may have resulted in increased genetic diversity of *P. falciparum* populations in Vietnam.

Are the patterns of MSP-1 *diversity temporally* stable in a given malaria-endemic area? - Fig. 3 compares the distribution of MSP-1 gene types in parasite populations sampled in the same communities in southern Vietnam at intervals of 12 months (Fig. 3a) and 18-24 months (Fig. 3b). There is no significant difference when both pairs of frequency distributions are compared. The stability in the frequencies of MSP-1 gene types over periods of 12-24 months does not imply that long-term changes can be ruled out. Under the present conditions of malaria transmission in southern Vietnam, just a few infections with parasites carrying distinct versions of the MSP-1 antigen are expected per host at a one-year or two-year interval. As a consequence, natural acquisition of effective anti-MSP-1 immunity may occur at a rather slow rate. We have now examined this issue in relation to the Brazilian Amazon by typing MSP-1 variable blocks in *P. falciparum* isolates collected over a period of 12 years (LA Silveira & MU Ferreira, unpublished data).

Are there non-random associations between MSP-1 variable blocks in natural parasite populations? - If intragenic recombination occurs frequently at the MSP-1 locus in the absence of major selective constraints, the distribution of MSP-1 gene types would be described by a simple probability model analogous to those used in population genetics to estimate expected frequencies of multiple-locus genotypes (Tibayrenc 1995). For instance, the expected frequency of gene type 1 (as defined in Table I) is given by multiplying the

TABLE III

The extent of allelic diversity at the *merozoite surface protein-1 (MSP-1)* locus in natural *Plasmodium falciparum* populations from areas with different levels of malaria endemicity

Area	Malaria endemicity	No. of <i>MSP-1</i> gene types detected by PCR	Proportion (%) of isolates with > 1 <i>MSP-1</i> type	Average no. of <i>MSP-1</i> types per patient
Brazil	Low	10	39	1.42
Vietnam	Intermediate	23	44	1.76
Tanzania	High	13	60	2.37

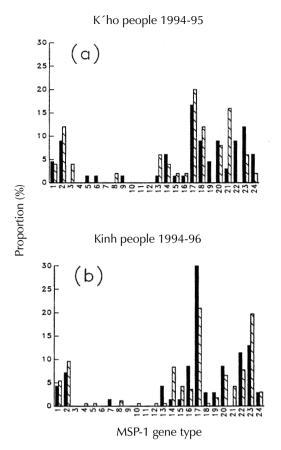


Fig. 3-a: frequency distribution of *merozoite surface protein-I(MSP-I)* gene types in isolates from K'ho people living in hill areas surrounding Bao Loc, southern Vietnam, collected between July-August 1994 (n=34) (closed bars) and in August 1995 (n=28) (striped bars) (redrawn from Kaneko et al. 1997); b: frequency distribution of *MSP-I* gene types in isolates from Kinh people living in the town of Bao Loc, southern Vietnam, collected between July-August 1994 (n=44) (closed bars) and between January-July 1996 (n=95) (striped bars) (redrawn from Ferreira et al. 1998b).

observed frequencies of the allelic type K1 in blocks 2, 4a, 4b and 6-16 in a given population. Fig. 4 shows expected frequencies of *MSP-1* gene types under the null hypothesis of random association of allelic types (MAD20, K1 or RO33) in each variable block in Brazil and Tanzania. No significant difference between expected and observed frequencies was detected by the c² test for goodness of fit in both cases (Ferreira et al. 1998a, c). In contrast, significant differences between expected and observed frequencies of *MSP-1* gene types were found in two surveys in southern Vietnam (Fig. 5). Non-random associations were found to occur, in both cases, between blocks 4a and 6-16: *MSP-1* gene types with concordant allelic fami-

lies (either MAD20 or K1) in blocks 4a and 6 or 10 were found more frequently than expected (Kaneko et al. 1997, Ferreira et al. 1998b). The reasons why similar results were not found in holoendemic Tanzania and hypoendemic Brazil remain to be elucidated.

Non-random associations between allelic types may result from: (a) geographic isolation leading to random genetic drift, (b) limited chances for intragenic recombination during meiosis in the mosquito vector due to the presence of few different *MSP-1* versions and the low prevalence of mixed infections in human hosts, and (c) biological constrains which bias for particular associations. As

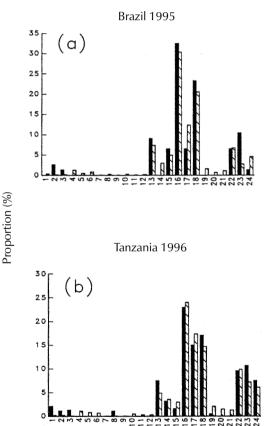
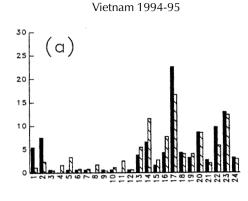


Fig. 4-a: expected (closed bars) and observed (striped bars) frequencies of *merozoite surface protein-1 (MSP-1)* gene types in Porto Velho, southwestern Brazilian Amazon (n=54) (data from Ferreira et al. 1998a); b: expected (closed bars) and observed (striped bars) frequencies of MSP-1 gene types in Tanga and Pangani, northern Tanzania (n=79) (data from Ferreira et al. 1998c). Expected frequencies were computed under the null hypothesis of random associations of allelic types in variable blocks of the gene (see the text for details). There is no significant difference between expected and observed frequencies by c^2 tests of goodness of fit.

MSP-1 gene type

Proportion (%)



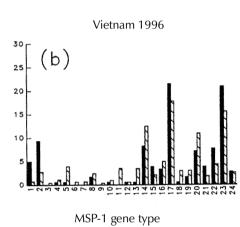


Fig. 5-a: expected (closed bars) and observed (striped bars) frequencies of *merozoite surface protein-1 (MSP-1)* gene types in isolates from Bao Loc, Vietnam, collected between July-August 1994 from both Kinh and K'ho people (n=108) (data from Kaneko et al. 1997); b: expected (closed bars) and observed (striped bars) frequencies of *MSP-1* gene types in isolates from Bao Loc, Vietnam, collected between January-July 1996 from both K'ho and Kinh people (n=102) (data from Ferreira et al. 1998b). Expected frequencies were computed under the null hypothesis of random associations of allelic types in variable blocks of the gene (see the text for details). In both cases significant differences between expected and observed frequencies were detected by c^2 tests of goodness of fit.

discussed elsewhere, the first two possible explanations do not match available data from Vietnam, and speculations regarding the third hypothesis are limited by the fact that the function of *MSP-1* remains unknown (Ferreira et al. 1998b).

Are parasite populations carrying different MSP-1 gene types independently distributed in the host population? - A basic assumption of recent mathematical models of malaria transmission is that infections by different 'strains' are independent. This means that, in genetically mixed infections, a patient infected by a parasite carrying a given MSP-1 gene type (for instance type 1) is as likely to be co-infected with a given second type (for instance type 2) as someone infected with any other MSP-1 type. This does not take into account the possibilities of: (a) frequent multiple-clone infections by vectors carrying two or more gene types including recombinant gene types resulting from the union of two different clones from one previous host (Hill & Babiker 1995), and (b) either facilitation or competition between parasites carrying different versions of a polymorphic antigen which co-infect the same host (Gilbert et al. 1998). We applied here a simple statistical analysis to test this assumption.

The expected distribution of MSP-1 gene types per host under the hypothesis of independent distribution of MSP-1 gene types may be described as the sum of N independent binomial distributions, where N is the number of different MSP-1 gene types observed in host population. The variance of this summed binomial distribution, or expected variance s², was calculated and compared to the observed variance s² using a c² test as described (Lotz & Font 1991). The difference between the expected and observed variances was statistically significant in Tanzania (Table IV), suggesting that the MSP-1 gene types are not independently distributed in the host population. Therefore, we tested 21 possible pairwise associations between gene types in 2 ^ 2 contingency tables using either standard c² or Fisher's exact tests when appropriate, with the significance level adjusted for multiple

TABLE IV
Statistical comparison of expected (s^2) and observed (s^2) variances of the distribution of *merozoite surface* protein-1 (MSP-1) gene types among human hosts living in areas with different levels of malaria endemicity

Area	Malaria endemicity	Expected variance (s ²)	Observed variance (s^2)	c^2 (d. f.) ^a	P
Brazil	Low	1.04	0.32	16.45 (53)	> 0.05
Vietnam	Intermediate	1.35	1.42	105.65 (101)	> 0.05
Tanzania	High	1.59	2.10	103.08 (78)	< 0.05

a: degrees of freedom.

comparisons with the Bonferroni's correction (Lord et al. 1997). At least 1 pair of MSP-1 gene types (18 and 24) was found to be positively associated (P=0.0008 by c^2 test). With the Bonferroni's correction applied to these data, an association is statistically significant at the 5% level if P<0.0024. This means that the genetically similar types 18 and 24, that differ only in the block 4b allelic type (Table I), tend to co-occur more frequently than expected under the null hypotheses that they are independently transmitted. Nevertheless, departures from the null hypothesis of independent transmission were not detected in areas of lower endemicity such as Brazil and Vietnam (Table IV).

Mathematical models have recently regarded malaria as a heterogeneous disease caused by several independently transmitted and antigenically distinct parasite subpopulations or 'strains' that do not interact within the human hosts and are able to elicit 'strain'-specific protective immunity. These models estimate the basic reproduction number R_0 of malaria, defined as the average number of secondary infections generated by one primary infection in a fully susceptible population, as a weighted average of R_0 values for each 'strain'. This estimate is substantially lower than R_0 values obtained by conventional methods, suggesting that malaria eradication in Africa may be quite feasible (Gupta et al. 1994). Nevertheless, the finding that genetically and antigenically distinct parasite populations are not independently distributed in the human hosts in areas of high endemicity, such as northern Tanzania (Ferreira et al. 1998c) and the Gambia (Conway et al. 1991a), implies R_0 values considerably higher than those provided by the weighted average approach (Lord et al. 1997).

In conclusion, this study provides examples of the use of simple molecular and statistical approaches to investigate the extent of antigenic diversity in malaria parasites and to test hypotheses regarding the patterns of transmission and interaction of genetically distinct parasite subpopulations in endemic areas.

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REFERENCES

Certa U, Rotmann D, Matile H, Reber-Liske RA 1987. A naturally occuring gene encoding the major surface antigen precursor p190 of *Plasmodium falciparum* lacks tripeptide repeats. *EMBO J* 6: 4137-4142.

- Conway DJ, Greenwood BM, McBride JS 1991a. The epidemiology of multiple-clone *Plasmodium falciparum* infections in Gambian patients. *Parasitology* 103: 1-6.
- Conway DJ, Rosário V, Oduola AMJ, Salako AL, Greenwood BM, McBride JS 1991b. *Plasmodium falciparum*: intragenic recombination and nonrandom associations between polymorphic domains of the precursor to the major surface antigens. *Exp Parasitol* 73: 469-480.
- Ferreira MU, Liu Q, Kaneko O, Kimura M, Tanabe K, Kimura EAS, Katzin AM, Isomura S, Kawamoto F 1998a. Allelic diversity at the *merozoite surface protein-1* locus of *Plasmodium falciparum* in clinical isolates from the southwestern Brazilian Amazon. *Am J Trop Med Hyg*, in press.
- Ferreira MU, Liu Q, Zhou M, Kaneko O, Kimura M, Thien HV, Isomura S, Tanabe K, Kawamoto F 1998b. Stable patterns of allelic diversity at the merozoite surface protein-1 locus of *Plasmodium falciparum* in clinical isolates from southern Vietnam. *J Euk Microbiol* 45: 131-136.
- Ferreira MU, Liu Q, Kimura M, Ndawi BT, Tanabe K, Kawamoto F 1998c. Allelic diversity in the merozoite surface protein-1 and epidemiology of multiple-clone *Plasmodium falciparum* infections in northern Tanzania. *J Parasitol*, in press.
- Gilbert SC, Plebanski M, Gupta S, Morris J, Cox M, Aidoo M, Kwiatkowski D, Greenwood BM, Whittle HC, Hill ASV 1998. Association of malaria parasite population structure, HLA, and immunological antagonism. Science 279: 1173-1177.
- Gupta S, Trenholme K, Anderson RM, Day KP 1994. Antigenic diversity and transmission dynamics of Plasmodium falciparum. Science 263: 961-963.
- Hill WG, Babiker HA 1995. Estimation of numbers of malaria clones in blood samples. Proc R Soc London B 262: 249-257.
- Holder AA 1996. Preventing merozoite invasion of erythrocytes, p. 77-104. In SL Hoffman, Malaria Vaccine Development. A Multi-immune Response Approach, ASM Press, Washington D.C.
- Holder AA, Riley EM 1996. Human immune response to MSP-1. Parasitol Today 12: 173-174.
- Jongwutiwes S, Tanabe K, Nakazawa S, Uemura H, Kanbara H 1991. Coexistence of gp195 alleles of *Plasmodium falciparum* in a small endemic area. *Am J Trop Med Hyg 44*: 299-305.
- Kaneko O, Jongwutiwes S, Kimura M, Kanbara H, Ishii A, Tanabe K 1996. *Plasmodium falciparum*: variation in block 4 of the precursor to the major surface proteins (MSP1) in natural populations. *Exp Parasitol* 84: 92-95.
- Kaneko O, Kimura M, Kawamoto F, Ferreira MU, Tanabe K 1997. *Plasmodium falciparum*: allelic variation in the merozoite surface protein 1 in wild isolates from southern Vietnam. *Exp Parasitol* 86: 45-57.
- Lord CC, Woolhouse MEJ, Barnard BJH 1997. Transmission and distribution of virus serotypes: African horse sickness in zebra. *Epidemiol Infect 118*: 43-50.

- Lotz JM, Font WF 1991. The role of positive and negative interspecific associations in the organization of communities of intestinal helminths of bats. *Parasitology* 103:127-138.
- Peterson MG, Coppel RL, Moloney MB, Kemp DJ 1988. Third form of the precursor to the major surface antigens of *Plasmodium falciparum*. *Mol Cell Biol 8:* 2664-2667.
- Tanabe K, Murakami K, Doi S 1989. Plasmodium
- falciparum: dimorphism of the p190 alleles. Exp Parasitol 68: 470-403.
- Tanabe K, Mackay M, Goman M, Scaiffe JG 1987. Allelic dimorphism in a surface antigen gene of the malaria parasite *Plasmodium falciparum*. J Mol Biolol 195: 273-287.
- Tibayrenc M 1995. Population genetics of parasitic protozoa and other microorganisms. *Adv Parasitol 36:* 47-115.