

Use of *sodC* versus *ctrA* for real-time polymerase chain reaction-based detection of *Neisseria meningitidis* in sterile body fluids

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We evaluated the use of a newly described *sodC*-based real-time-polymerase chain reaction (RT-PCR) assay for detecting *Neisseria meningitidis* in normally sterile sites, such as cerebrospinal fluid and serum. The *sodC*-based RT-PCR assay has an advantage over *ctrA* for detecting nongroupable *N. meningitidis* isolates, which are commonly present in asymptomatic pharyngeal carriage. However, in our study, *sodC*-based RT-PCR was 7.5% less sensitive than *ctrA*. Given the public health impact of possible false-negative results due to the use of the *sodC* target gene alone, *sodC*-based RT-PCR for the diagnosis of meningococcal meningitis should be used with caution.

Key words: *sodC*-based RT-PCR - *N. meningitidis* diagnostics - meningitis

Meningococcal disease (MD) is a serious and often fatal infection. In Brazil, approximately 3,200 new cases of sporadic and outbreak-associated MD are reported each year, making this disease an important public health concern. Our laboratory at Adolfo Lutz Institute (IAL) uses a multiplex real-time-polymerase chain reaction (RT-PCR) assay developed and performed at IAL (Sacchi et al. 2011). The assay is composed of three sets of primers and three probes targeting the *ctrA* gene of *Neisseria meningitidis* (Mothershed et al. 2004), the *lytA* gene of *Streptococcus pneumoniae* (Carvalho et al. 2007) and the *bexA* gene of *Haemophilus influenzae* (Corless et al. 2001). *ctrA* is a frequently targeted gene used to detect *N. meningitidis* using RT-PCR (Corless et al. 2001, Mothershed et al. 2004). However, the capsule locus, including *ctrA*, is subject to rearrangement (Swartley et al. 1997, Clauds et al. 2002, Dolan-Livengood et al. 2003, Sadler et al. 2003) and 16% or more of carried meningococci have been shown to lack *ctrA* altogether (Clauds et al. 2002, Dolan-Livengood et al. 2003).

It has been suggested that invasive meningococci can undergo similar rearrangements of the capsule region. For these reasons, a new target based on the *sodC* gene in meningitidis RT-PCR has been proposed as an alternative for clinical specimens, such as cerebrospinal fluid (CSF) and serum (Thomas et al. 2011, WHO 2011).

The possibility of false-negative results due to the use of the *ctrA* target gene alone led us to investigate the sensitivity of *sodC*-based RT-PCR by including it in our routine diagnosis of bacterial meningitis for eight months. We analysed 1,538 clinical samples [957 (62%) CSF and 581 (38%) sera] that were submitted for diag-

nostic testing for bacterial meningitis between January-August 2011. The research protocol was approved by the IAL Institutional Ethical Committee.

CSF and blood culture is the “gold standard method” for diagnosing bacterial meningitis. In Brazil, culture is performed at each hospital and the isolates are sent to IAL for confirmation. For several reasons, including previous antibiotic treatment, most of the cases are not culture confirmed and, for many samples, culture is not performed. Because it is difficult to evaluate a diagnostic test without comparing it to a “gold standard”, we tested all *ctrA*-positive samples for the presence of a second meningococcal gene target (genogrouping).

We used a *ctrA*-based multiplex RT-PCR (Sacchi et al. 2011) and a *sodC*-based monoplex RT-PCR (Thomas et al. 2011, WHO 2011) to detect *N. meningitidis* and a capsule gene-based RT-PCR for meningococcal genogrouping (Mothershed et al. 2004). DNA was extracted using a MagNA Pure LC and the DNA Isolation Kit III according to the manufacturer’s instructions (Roche Diagnostic GmbH, Mannheim, Germany). All extracted DNA was stored at -20°C. The assays were performed in a 25 µL reaction volume using TaqMan Universal Master Mix (Applied Biosystems, Foster City, CA) with 5 µL of extracted DNA. Forward primer, reverse primer and the probe for each gene target were used at previously described concentrations.

All reactions were run in duplicate using an Applied Biosystems 7500 Real-Time PCR System (Applied Biosystems Foster City, CA, USA) with the following cycling parameters: 50°C for 2 min, 95°C for 10 min and 45 cycles of 95°C for 15 s and 60°C for 1 min. Extension at 55°C for 1 min was used for RT-PCR genogrouping for serogroups B, W135 and Y. Positive and negative controls were included in each run.

A review of the literature showed that no standards exist for determining what criteria indicate a positive result. Some researchers established a “cut-off” cycle threshold (Ct) value for a particular assay, while others considered any amplification signal, regardless of the Ct

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value, indicative of a positive result. However, the rationale for determining such parameters was not always clear. In this study, a positive result was defined as a Ct ≤ 38 and a negative result as a Ct value of zero or ≥ 39. All inconclusive results and inconsistent replicates were repeated. The lower limit of detection (LLD) for *sodC* and *ctrA* RT-PCR assays at a Ct of 38 was 200 fg. The efficiency of both reactions was similar: 92.15% for *ctrA* and 90.42% for *sodC*.

The results were classified into four groups: Group A, positive for both *ctrA* and *sodC*, Group B, positive for *ctrA* and negative for *sodC*, Group C, negative for *ctrA* and positive for *sodC*, and Group D, negative for both *ctrA* and *sodC* (Table). All samples that were positive for *sodC* were also positive for *ctrA* (Group A), but 24 samples that were positive for *ctrA* were negative for *sodC* (Group B). The Ct values of *ctrA* and *sodC* for each sample in Group A were mostly similar, varying from 23-38. To compare our results with the original paper that described the use of *sodC* for detecting *N. meningitidis* (Thomas et al. 2011), we also used a second Ct value of ≤ 35 to define a positive result. We were only able to obtain culture results for 75 samples (51 from Group A and 24 from Group B).

Group A - Among the 295 Group A samples, 87.1% (257/295) had a Ct ≤ 35 for both *sodC* and *ctrA*, 7.8% (23/295) had a *ctrA* Ct ≤ 35 and a *sodC* Ct ≥ 36 (10 that were culture-confirmed) and 5.1% (15/295) had a Ct ≥ 36 (8 that were culture-confirmed) for both *sodC* and *ctrA*. Therefore, 38 Group A samples (18 culture-confirmed) were considered negative using a *sodC* Ct ≤ 35 to define a positive result.

Group B - Among the 24 Group B samples, seven had *ctrA* Ct values ≤ 35: Ct = 29, 31, 32 and 33 (1 of each) and Ct = 35 (n = 3). Additionally, 42% (10/24) were isolated from the CSF and 58% (14/24) were isolated from the serum. No samples had less than the ideal volume. Three of 24 samples that were culture confirmed had *ctrA* Ct values of 35, 36 and 37 and the whole-cell suspensions made from the three isolates were positive for *ctrA* and *sodC*.

We were able to determine the genogroup of all 319 *ctrA*-positive samples (Groups A and B): 17.2% were genogroup B (n = 55), 76.5% were genogroup C (n = 244), 3.8% were genogroup W (n = 12) and 2.5% were genogroup Y (n = 8).

A *sodC*-based RT-PCR assay has an advantage over *ctrA* in detecting nongroupable *N. meningitidis* isolates,

which are common in asymptomatic pharyngeal carriage. However, in our study, *sodC*-based RT-PCR was 7.5% less sensitive than *ctrA* in normally sterile body fluids. Our data are discordant from those previously reported by Thomas et al. in 2011. According to our data, there was no suggestion that these differences may be due to sample type or volume or differences in the LLD. Unfortunately, we could not determine the reasons for these differences. However, we analysed a much larger and more diverse collection representing 220 health care units distributed throughout the state of São Paulo.

Given the public health impact of possible false-negative results due to the use of the *sodC* target gene alone, using *sodC*-based RT-PCR to detect *N. meningitidis* should be performed with caution.

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TABLE

Real-time-polymerase chain reaction results for *ctrA* and *sodC*

Group	<i>ctrA</i>	<i>sodC</i>	n (%)
A	Positive	Positive	295 (19.2)
B	Positive	Negative	24 (1.6)
C	Negative	Positive	0 (0)
D	Negative	Negative	1,219 (79.2)
Total			1,538 (100)