Diversity of *Campylobacter* Isolates from Three Activated Sludge Systems

AL Lauria-Filgueiras⁺, E Hofer

Laboratório de Zoonoses Bacterianas, Instituto Oswaldo Cruz, Av. Brasil 4365, 21045-900 Rio de Janeiro, RJ, Brasil

Thermophilic campylobacters were isolated from three sewage plants in Rio de Janeiro, RJ, Brazil and identified. Laboratory analysis of 390 sewage samples showed the presence of 169 thermophilic strains. The results demonstrated that human and animal pathogenic biotypes could be isolated from activated sludge during the initial processing steps. The aeration tank could be considered a barrier to Campylobacter survival. C. jejuni was the prevalent species isolated (40.8%). The most common biotypes were C. jejuni biotype I (21.3%), C. coli biotype I (16%) and C. jejuni biotype II (14.8%).

Key words: thermophilic campylobacters - sewage plants - incidence

During the past decade thermophilic campylobacters have been shown to be responsible for enteritis in human and animals (Smibert 1974. 1984, Ketley 1997). Campylobacter is a microaerophilic microorganism which can not proliferate in the environment but can survive fairly well in aqueous environments at low temperature (Stelzer et al. 1991). Studies in the United Kingdom, Germany, Italy, the Netherlands and Libya have shown that sewage is usually heavily contaminated with this pathogen; these papers have also detailed the importance of efflux from sewage plants as a major vehicle for the spread of pathogenic biotypes of C. jejuni and C. coli in the environment (Höller 1988, Betaieb & Jones 1990, Jones et al. 1990a, Stelzer et al. 1991, Stampi et al. 1992, Höller & Schomakers-Revaka 1994, Koenraad et al. 1994).

Based on the above facts and because there is no study in our country on this subject, we investigated the incidence and distribution of these bacteria during biological sewage treatment by the activated sludge process at three sewage plants in Rio de Janeiro, RJ, Brazil.

MATERIALS AND METHODS

Sample collection - Samples were collected in Rio de Janeiro, RJ, Brazil (Table I).

The Penha sewage treatment plant serves a permanent local population of approximately 700,000 inhabitants. In addition to domestic sewage, the plant receives considerable amounts of industrial

Samples were collected at four different stages of biological treatment: influent, entry to the aeration tank, exit from the aeration tank, and effluent. An amount of 120 ml of sewage was collected from each stage into sterile bottles, transferred to the laboratory within 1 hr and immediately centrifuged at 3000 rpm for 30 min.

TABLE I
Distribution of sewage collection

Sewage plant	Period	No. of samples
Penha	15 May 1990 to 21 May 1991	120
Governador Island	12 September 1990 to 21 May 1991	140
International Airport of Rio de Janeiro	12 February 1990 to 26 November 1991	
Total		390

^{*}Corresponding author. Fax: + 55-21-280.0754/ 270. 6565. E-mail: analu@gene.dbbm.fiocruz.br Received 10 November 1997 Accepted 18 March 1998

and hospital wastes, with an estimated discharge volume of 1,400 l/sec. Two treatments are applied at the plant, i.e., activated sludge and biological filters. The Governador Island plant serves about 90,000 persons and mainly treats domestic sewage using the activated sludge process. The Airport plant receives sewage from administration buildings, from the passenger terminal, from the support area and also from all aircrafts. The last type of sewage contains disinfectants at the levels required by law for the removal of polluting loads. The plant also uses the activated sludge process which has a mean efficiency of about 90%. This influent receives material from airplanes and therefore from a fluctuating population consisting of passengers from all over the world.

The Airport samples were collected at only two stages, i.e., influent and effluent. Moore's swab adaptations Hofer (1974) were kept submerged for 72 hr in these residual waters. After this time, the swabs were placed in plastic bags, taken to the laboratory within less than 2 hr of collection and analyzed immediately.

Isolation - Aliquots of 0.1 ml of 0.5 ml PBS resuspension of the centrifuged material as well as 0.1 ml of material squeezed from the swabs were inoculated onto selective plate medium (blood agar base, Columbia, Difco) containing 4 g% of activated charcoal (Group Quimica), 5% reducing solution FBP (George et al. 1978) and 0.5% of a mixture of the following antibiotics: trimethoprim lactate (Roche), cephalothin (Libra), vancomycin (Lilly), cyclohexamide (Upjohn), and colistin (Lafi) according to Filgueiras and Hofer (1989).

Plates were incubated at 43°C for 48 hr in a candle jar prepared by a modification of the copper passivation technique using steel wool, antiacid tablets and an acidulated (pH 2) copper solution (Filgueiras & Hofer 1989).

Identification - Campylobacter growth was confirmed on the basis of colony morphology (gray, small, mucous or flat) and microscopic appearance. If the Gram-stained smears from suspected colonies showed Gram-negative bacteria morphologically similar to the genus Campylobacter, the material was submitted to the oxidase and catalase reactions. The colonies were then isolated in duplicate, with one isolate being maintained at low temperature (-70°C) in Brucella broth with 0.16 g% agar, and the other being used for biochemical tests (glucose fermentation, growth in 3.5% sodium chloride, and in 1% glycine, reduction of nitrate to nitrite and H₂S production) and to demonstrate sensitivity to nalidixic acid (30 mg- Cecon) and resistance to cephalothin (30 mg- Cecon).

To complete the biotyping, according to scheme of Lior (1984), the material was submitted to the hippurate hydrolysis test and to deoxyribonuclease enzyme (DNase) detection.

RESULTS AND DISCUSSION

Sample analysis led to the isolation and identification of 169 thermophilic campylobacters at the three plants studied. Among them, 50% were *Campylobacter* obtained at the Penha and Governador Island plants, probably due to human *Campylobacter* infection in the community and to birds, in particular at the Governador Island plant (Table II). The less frequent isolation obtained at the Airport sewage plant was possibly due to the fact that the sewage from aircraft was mixed with disinfectants. It should also be remembered that this sewage plant serves a fluctuating population whereas the other two receive influent from the permanent local population.

The samples obtained at different stages during biological sewage treatment (Tables III, IV, V) yielded a larger number of isolates during the first steps (influent and entry to the aeration tank), with the detection of the species and biotypes more frequently related to the human and animal enteric process (*C. jejuni* biotypes I and II and *C. coli* biotypes I and II), as reported in the literature (Arimi et al.1988, Filgueiras & Hofer 1989, Stelzer et al. 1991, Stampi et al. 1992). A large amount of untyped strains was obtained after the aeration treatment, perhaps resulting from *Campylobacter* sensitivity to aeration, with a consequent modification in biochemical behavior.

There are no reports in the literature of the detection of *C. jejuni* biotype IV in residual water (Tables IV, V), nor about *C. lari*, a thermophilic species resistant to nalidixic acid. This detection in our study (Table IV) was possibly due to the presence of seagulls at the plant sampled from Governador Island.

The present results agree with data reported by Koenraad et al. (1994), who suggest the disinfection of sewage plant effluxes in order to obstruct the environmental cycles of *Campylobacter*, since the elimination is far from complete. Furthermore, Arimi et al. (1988) and Höller (1988) have reported that the reduction of campylobacters during the

TABLE II
Frequency of thermophilic campylobacters isolated from three sewage plants in Rio de Janeiro, RJ, Brazil

Plants	Campylobacter						
	Positive		Negative		Total		
	No.	%	No.	%	No.	%	
Penha	59	15.2	61	15.6	120	30.8	
Governador Island	72	18.5	68	17.4	140	35.9	
International Airport of Rio de Janeiro	38	9.7	92	23.6	130	33.3	
Total	169	43.4	221	56.6	390	100.0	

TABLE III

Frequency of *Campylobacter* thermophilic species and biotypes according to the classification of Lior (1984), at different stages of biological treatment at the Penha sewage plant Rio de Janeiro, RJ, Brazil

Stages	C. je	juni	C. cc	oli	Campylobacter sp. (untypable)	Total
	I	II	I	II		
Influent	4	4	5	3	6	22
n=30	(6.8)	(6.8)	(8.5)	(5.1)	(10.0)	(37.2)
Entry to the aeration tank	10	3	2	0	3	18
n=30	(16.9)	(5.1)	(3.4)		(5.1)	(30.5)
Exit from the aeration tank	2	3	3	0	4	12
n=30	(3.4)	(5.1)	(5.1)		(6.8)	(20.4)
Effluent	0	0	1	1	5	7
n= 30			(1.7)	(1.7)	(8.5)	(11.9)
Total	16	10	11	4	18	59
n= 120	(27.1)	(17.0)	(18.7)	(6.8)	(30.4)	(100.0)

n: number of samples analized.

TABLE IV
Frequency of *Campylobacter* thermophilic species and biotypes. according to the classification of Lior (1984), at different stages of biological treatment at the Governador Island sewage plant, Rio de Janeiro, RJ, Brazil

Stages	C. jejuni			C. coli		C. lari	Campylobac	ter Total
-	I	II	IV	I	II	I	sp. (untypab	le)
Influent	5	5	1	2	4	1	5	23
n=35	(6.9)	(6.9)	(1.4)	(2.8)	(5.6)	(1.4)	(6.9)	(31.9)
Entry to the aeration tank	2	4	4	7	2	0	4	23
n=35	(2.8)	(5.5)	(5.5)	(9.7)	(2.8)		(5.6)	(31.9)
Exit from the aeration tank	2	0	0	2	2	0	9	15
n=35	(2.8)			(2.8)	(2.8)		(12.5)	(20.9)
Effluent	1	1	1	1	1	0	6	11
n=35	(1.4)	(1.4)	(1.4)	(1.4)	(1.4)		(8.3)	(15.3)
Total	10	10	6	12	9	1	24	72
n=130	(13.9)	(13.9)	(8.3)	(16.7)	(12.5)	(1.4)	(33.3)	(100.0)

n: number of samples analized.

TABLE V

Frequency of *Campylobacter* thermophilic species and biotypes according to the classification of Lior (1984), at different stages of biological treatment at the International Airport of Rio de Janeiro sewage plant, Rio de Janeiro, RJ, Brazil

Stages		C. jejuni			oli	Campylobacter	Total
	I	II	IV	I	II	sp. (untypable)	
Influent	7	3	2	2	1	8	23
n=65	(18.4)	(7.9)	(5.3)	(5.3)	(2.6)	(21.0)	(60.5)
Effluent	3	2	0	2	0	8	15
n=65	(7.9)	(5. 3)		(5.3)		(21.0)	(39.5)
Total	10	5	2	4	1	16	38
n=130	(26.3)	(13.2)	(5.3)	(10.6)	(2.6)	(42.0)	(100.0)

n: number of samples analized.

final step of treatment reached levels of 99.6 and 100%, respectively. In this respect, it should be pointed out that the Governador Island sewage plant was working at reduced capacity throughout 1990 due to repairs (Table IV), what may explain

the lower rate of *Campylobacter* elimination obtained at this plant compared to the Penha plant (Table V).

Another interesting aspect was the detection of a high percentage of untyped strains at all sewage

treatment plants (Tables III, IV, V). A possible explanation for this fact is the hypothesis of environmental effects or even selective pressure on the microorganisms (Arimi et al. 1988, Höller 1988, Jacob et al. 1990, Jones et al. 1990b). In general, survival of campylobacters in sewage may be limited by the high organic load (Höller & Schomakers-Revaka 1994).

The loss of viability during laboratory identification is considered to be due to the sudden change in environmental conditions, and exposure to high or low temperatures, oxygen or toxic substances that may quickly result in transition to a non-cultivable stage, with a sudden change to coccoid forms (Rollins & Colwell 1986). In addition, in agreement with data reported by most investigators, we also observed the presence of a large number of viable but not cultivable *Campylobacter* cells (Rollins & Colwell 1986, Carter et al. 1987, Höller 1988, Humphrey 1989, Pokorny 1990, Jacob & Stelzer 1992, Höller & Schomakers-Revaka 1994).

Finally, analysis of species and biotype distribution at the sewage plants showed the predominance of *C. jejuni* biotypes I and II, in agreement with data reported by others (Taylor et al. 1983, Carter et al. 1987, Arimi et al. 1988, Jones et al. 1990c, Stampi et al. 1992). This is explained by the fact that *C. jejuni* is the species most frequently detected in man and most frequently responsible for enteritis caused by campylobacters, probably reflecting its best adaptation and diffusion among human beings.

REFERENCES

- Arimi SM, Fricker CR, Park RWA 1988. Occurrence of thermophilic campylobacters in sewage and their removal by treatment process. *Epidem Inf* 101: 279-286.
- Betaieb M, Jones K 1990. Thermophilic campylobacters in two treatment plants in Libya. *Lett Appl Microbiol* 11: 93-95.
- Carter AM, Pacha RE, Clark GW, Williams EA 1987. Seasonal occurrence of *Campylobacter* spp. in surface waters and their correlation with standard indicator bacteria. *Appl Environ Microbiol* 53: 523-526.
- Filgueiras ALL, Hofer E 1989. Ocorrência de Campylobacter termofílico em diferentes pontos de uma estação de tratamento de esgotos na cidade do Rio de Janeiro, RJ. Rev Microbiol (São Paulo) 20: 303-308.
- George HA, Hoffman PS, Smibert RM, Krieg NR 1978. Improved media for growth and aerotolerance of *C. fetus. J Clin Microbiol* 8: 36-41.
- Hofer E 1974. Métodos utilizados para o isolamento e identificação de Vibrio cholerae. Informe Patologia Clínica, Rio de Janeiro, 1: 5-18.
- Höller C 1988. Long-term study of occurrence, distribution and reduction of *Campylobacter* sp. in the sewage system and wastewater treatment plant of a big town. *Wat Sci Tech* 20: 529-531.

- Höller C, Schomakers-Revaka U 1994. A note: comparison of different homogenization procedures for detecting *Campylobacter* spp. in sewage sludge. *J Appl Bacteriol* 77: 591-596.
- Humphrey TJ 1989. An appraisal of the efficacy of preenrichment for the isolation of *Campylobacter jejuni* from water and food. *J Appl Bacteriol* 66: 119-126.
- Jacob J, Stelzer W 1992. Comparison of two media for the isolation of thermophilic *Campylobacter* from waste waters of different quality. *Zentralbl Mikrobiol* 147: 41-44.
- Jacob J, Bindemann U, Stelzer W 1990. Identification of *Campylobacter jejuni* and *Campylobacter coli* from wastewater by SDS-disc-page of whole cell protein. *Zbl Hyg 190*: 357-364.
- Jones K, Betaieb M, Telford DR 1990a. Thermophilic campylobacters in surface waters around Lancaster, UK: negative correlation with *Campy-lobacter* infections in the community. *J Appl Bacteriol* 69: 758-764.
- Jones K, Betaieb M, Telford, DR 1990b. Correlation between environmental monitoring of thermophilic campylobacters in sewage effluent and the incidence of *Campylobacter* infection in the community. *J Appl Bacteriol* 69: 235-240.
- Jones K, Betaieb M, Telford DR 1990c. Seasonal variation of thermophilic campylobacters in sewage sludge. *J Appl Bacteriol* 69: 185-189.
- Ketley JM 1997. Pathogenesis of enteric infection by *Campylobacter. Microbiology* 143: 5-21.
- Koenraad PMFJ, Hazeleger WC, van der Laan T, Beumer RR, Rombouts FM 1994. Survey of Campylobacter spp. in sewage plants in The Netherlands. Food Microbiol 11: 65-73.
- Lior H 1984. New extended biotyping scheme for Campylobacter jejuni, Campylobacter coli and "Campylobacter laridis". J Clin Microbiol 20: 636-640.
- Pokorny J 1990. To the problem of *C. jejuni* detectability in water. *J Hyg Epidemiol Microbiol Immun* 34: 57-61.
- Rollins DM, Colwell RR 1986. Viable but nonculturable stage of *Campylobacter jejuni* and its role in survival in the natural aquatic environment. *Appl Environ Microbiol* 52: 531-538.
- Smibert RM 1974. Campylobacter, p. 207-211. In RE Buchanan & NE Gibbons (eds), *Bergey's Manual of Determinative Bacteriology*, 8th ed., The Williams & Wilkins Co., Baltimore.
- Smibert RM 1984. Campylobacter, p. 111-118. In NR Krieg & JG Holt (eds), Bergey's Manual of Systematic Bacteriology, 8th ed., vol. 1, The Williams & Wilkins Co., Baltimore.
- Stampi S, Varoli O, de Luca G, Zanetti F 1992. Occurrence, removal and seasonal variation of "thermophilic" campylobacters in a sewage treatment plant in Italy. *Zbl Hyg 191*: 199-210.
- Stelzer W, Jacob J, Schulze E 1991. Environmental aspects of *Campylobacter* infections. *Zentralbl Mikrobiol* 146: 3-15.
- Taylor DN, McDermott KT, Little JR, Wells JG, Blaser MJ 1983. Campylobacter enteritis from untreated water in the Rocky mountains. Ann Int Med 99: 38-40.