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Gene Flow Patterns of the *Aedes aegypti* (Diptera: Culicidae) Mosquito in Colombia: a Continental Comparison Suggests Multiple Invasion Routes and Gene Exchange

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ABSTRACT

In Colombia *Aedes aegypti* is present in 80% of the country up to 2,300 m; however, little is known of its genetic relations within a country context and, hence, within a broader context, for example, America. The aforementioned, herein, analyzed the gene flow within a context of the Americas, its directionality and genetic diversity of the mitochondrial lineages in the *A. aegypti* populations for Colombia. This called for the use of the sequences for *A. aegypti* available of the mitochondrial ND4 gene in the GenBank for Colombia and the American continent. No presence was found of nuclear mitochondrial pseudogenes (NUMTs) for Colombia. It is estimated that in Colombia the gene flow of the *A. aegypti* populations is occurring from the southeast and northeast toward the center of the country. In comparison with the mitochondrial sequences for America, the vector's haplotypes in Colombia suggest connections between the populations of mosquitoes from the south with those from the north of the continent. The gene flow model at continental scale suggests bidirectional connections between the populations from the south American scale it proposes the gene flow in all the directions with respect to the Colombian. The Colombia *A. aegypti* vector monitoring and control strategies must pay special attention to the vector's points of entry into Colombia related with Peru, Venezuela, Brazil, Mexico, and North America to avoid the entry of populations with characteristics like resistance to insecticides or vector competition.

Introduction

Aedes (Stegomyia) aegypti (Linnaeus, 1762) (Diptera: Culicidae) is the primary dengue, chikungunya, and Zika vector in the Americas (WHO, 2013, Weaver, 2014; Yakob and Walker, 2016), diseases about which there are no effective vaccines and, thereby, the most viable form to diminish its circulation is the use of chemical insecticides (Kantor, 2018). Nevertheless, excessive use of insecticides has generated globally resistance to most of the products used to control this species (WHO, 2016). *A. aegypti* originated in Africa and it is believed that its introduction to the tropical and subtropical zones of the Americas and Europe took place during the 15th and 17th centuries (Lounibos, 2002; Brown et al., 2011; Moore et al., 2013). For Colombia, it is suggested that *A. aegypti* arrived through the slave trade (OPS, 1992). In 1950, *A. aegypti* was already distributed in most of the Colombian territory; however, between 1952 and 1960 it was considered eradicated in most of the country (except for Cúcuta, Norte de Santander). By 1971, the

*Correspondence author: *E-mail*: mnavarro@ufpr.br (MA Navarro-Silva) vector had re-infested the port cities and plains of northern Colombia (Groot, 1980). In the Americas, the *A. aegypti* eradication campaign, and therefore the yellow fever, began by the Rockefeller Foundation in 1918 (Dallimore et al., 2020) and was reformulated in 1947 by PAHO, getting for 1958 to eradicate the species in much of America and South America, including Colombia and its neighboring countries, Ecuador, Peru, Brazil and Venezuela (Magalhães, 2021). However, the late vector control actions carried out in North America (initiated in 1964 and ended in 1969) led to the reinfestation of the mosquito from there to the center and south of the continent (Magalhães, 2016).

After the vector's re-infestation, three important dengue epidemics occurred in Colombia: in 1971 (450-thousand cases due to presence of dengue 2 serotype in northern Colombia); 1976 (200-thousand cases due to presence of dengue 3 serotype in central Colombia); and in 1977 (770-thousand cases of the dengue 1 serotype throughout Colombia), where special care was assigned to the central-eastern zone of the country due to re-infestation in 1978 (Groot, 1980). Thereafter, two new arboviruses, chikungunya (Martínez et al., 2015) and Zika

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(Rico-Mendoza et al., 2019), emerged in the country, causing important epidemics since their appearance. In Colombia, the species is distributed up to 2,300 m (Ruiz-López et al., 2016) representing 80% of the territory (OPS, 2011) and, with this, dengue, chikungunya, and Zika manage to circulate in 95% of Colombia (Rodríguez et al., 2003; Rodríguez and de La Hoz, 2004; Badii et al., 2007; WHO, 2013; Rico-Mendoza et al., 2019). However, in spite of efforts by the vector control programs, outbreaks of these arboviruses are still common in Colombia (Martínez et al., 2015; Guagliardo et al., 2019b).

Due to the aforementioned, understanding the patterns of structure and gene flow between the vector's populations is fundamental for the rational development of vector control programs (Urdaneta-Marquez and Failloux, 2011). For example, in South America, new alternatives are being evaluated, as is the case with the release of mosquitoes infected with *Wolbachia* (populations refractory to dengue, chikungunya, and Zika) or sterile mosquitoes (populations that once they reproduce diminish the population size) (Li and Ai, 2020; Velez et al., 2020). However, in any of both techniques, it is expected that the populations released interbreed with natural populations and these, in turn, disseminate the trait introduced into the population (Aliota et al., 2016) and undoubtedly to ensure that this happens it is necessary to know the structure and gene flow patterns where they will be released (Bosio et al., 2005).

The population structure and gene flow of the A. aegypti populations is studied through molecular markers, microsatellites, or single nucleotide polymorphisms (SNPs), which permit analyzing the populations within a contemporary context and in a finer scale (*i.e.*, a location), although also used in bigger geographic scales (*i.e.*, between countries) (Marcombe et al., 2013; Olanratmanee et al., 2013; Monteiro et al., 2014; Rašić et al., 2014). Recently, it is also possible to access the complete nuclear or mitochondrial genomic information (Pollett et al., 2020). Notwithstanding the aforementioned, using Mitochondrial DNA (mtDNA) as molecular marker has been widely applied in genetic studies of *A. aegypti* populations from different geographic points at small and large scale and in endemic regions for dengue, chikungunya, or Zika (Gonçalves et al., 2012; Seixas et al., 2013). However, its use should be cautious because nuclear mitochondrial pseudogenes (NUMTs) can be amplified, limiting its reliability in the inferences carried out due to distinct genealogies and evolutionary histories (Black and Bernhardt, 2009; Hlaing et al., 2009).

Among the mitochondrial molecular markers used to analyze the population structure and gene flow in A. aegypti, there is the mitochondrial ND4 gene, which encodes subunit 4 of the NADH dehydrogenase enzyme. The mitochondrial ND4 gene has been widely used in genetic studies of populations conducted in Brazil (Twerdochlib et al., 2012), Bolivia (Paupy et al., 2012), Peru (Yáñez et al., 2013), Venezuela (Urdaneta-Marquez and Failloux, 2011), and Mexico (Gorrochotegui-Escalante et al., 2002). For Colombia, the gene structure of the dengue vector is still poorly known. In Colombia, data available on genetic diversity were obtained by using random amplification of polymorphic DNA (RAPD) (Ocampo and Wesson, 2004; Mejía et al., 2011) and mtDNA (Caldera et al., 2013; Aguirre-Obando et al., 2015), using molecular markers of the RAPD and mtDNA type.In both cases, the results of these studies demonstrate genetic structuring in the populations. In spite of this, no work exists that analyzes and compares the mitochondrial lineages available for the vector in America with the mitochondrial lineages identified for Colombia. Consequently, this work sought to analyze within a continental context the gene flow of the mitochondrial lineages presents in the natural populations of A. aegypti of Colombia.

Materials and methods

To estimate the structure, gene flow, and potential directionality of the gene flow of the vector populations in Colombia with respect to the rest of the Americas, the work used sequences of the mitochondrial ND4 gene published for Colombia (Caldera et al., 2013; Aguirre-Obando et al., 2015) and America (Gonçalves et al., 2012). The publications for Colombia were obtained from a prior search in Scopus and Google Scholar by using the following criteria "*Aedes aegypti*" followed by DNA "Colombia", AND "genetic diversity", OR "gene flow", "gene structure" and "mtDNA". It considered all the databases and years. The origin (country and location) of the sequences, frequency, numbering and access numbers to GenBank are shown in Table 1.

Sequences referring to the American continent correspond to five countries: Mexico-North America, Colombia, Brazil, Venezuela, and Peru and were taken from Gonçalves et al., (2012). The sequences analyzed and compiled by Gonçalves et al. (2012) for America have as main characteristic the lack of NUMT. The sequences obtained were aligned by using the MAFT software, version 7, to detect haplotypes (H) present in them (Katoh and Standley, 2013), and these were numbered based on their frequency, thus, the most frequent was H1, followed by H2 and so on. The H found in Colombia followed the same numbering system, adding "Col" at the end (*i.e.*, Hn-Col). In addition, to reduce the error caused by the presence of NUMT in the sequences published for Colombia, two analyses were performed: 1. A search was made for heterozygous sites in the sequences and additional stop codons and, 2. The H were compared with the NUMT list verified by (Hlaing et al., 2009), Hlaing et al. (2009), and Black and Bernhardt (2009). If any NUMT was found, it was removed from the analysis and reported. Bearing in mind the H listed in Table 1, two haplotype networks were constructed: one for the departments of Colombia, Colombian scale, and another for the American continent (including Colombian H; continental scale). Both were constructed through the R software, version 3.5.1, using the Pegas package (Paradis et al., 2020) and the haplonet function based on the parsimony probability calculation (Templeton et al., 1992). The networks were processed graphically in Inkscape (www.inkscape.org).

Estimation of the nucleotide diversity (π), haplotypic diversity (Hd), and the D Tajima neutrality tests were estimated by location and country by using the Pegas and Strata packages in the R software.

With the H for Colombia and the Americas, the jModelTest - version 2.1.1 – was used to search for the best evolutionary model, selected through the Akaike information criterion (AIC) (Darriba et al., 2012). Thereafter, the model selected was implemented in Beast2 software (Bouckaert et al., 2014) to obtain the phylogenetic tree through Bayesian inference using the following parameters: use of a strict clock model, a Bayesian horizon coalescing model, executed with 10-million generations for the Markov Chain Monte Carlo (MCMC) with sampling every 1,000 generations and pre-burn in 10-thousand chains.

To infer the directionality of the gene flow among the A. aegypti populations of Colombia at continental and South American scales, two migration scenarios were tested by using a Bayesian coalescing approach in Migrate-N 4.21 (Beerli, 2004). The first scenario, at continental scale, took as comparison nodes the populations grouped for Mexico-North America, Venezuela, Peru, Brazil (Manaus, Porto Velho - Rio Branco, Belem, Boa Vista, and South-eastern Brazil), and Colombia (Sucre and Quindio). This examined six hypotheses of gene-flow models: panmixia hypothesis, total migration, two staggered stages (stepping-stone 1 and stepping-stone 2), gene flow from the north to the south or migration hypothesis from the south to the north (Fig. 1a). The second scenario, at South American scale, took as comparison nodes populations grouped for Peru, Venezuela, and south-eastern Brazil. For populations located in the Brazilian Amazon (Manaus, Porto Velho - Rio Branco, Belem, and Boa Vista) and Colombia (departament of Sucre and Quindio) these were used as independent nodes (Fig. 1b). This scenario tested five hypotheses of gene-flow models: total-migration hypothesis, two staggered scenarios (stepping-stone 1 and stepping-stone 2), gene flow from the north of South

Table 1

Hap -	Continent			Brazilian						Colombia		CanDani	Citation
	VZ	Peru	M-NA	MAO	BrAM	BV	PV-RB	SEBr	BL	Sucre	Quindio	Genbalik	Citation
H1	94	36	330	1	19	0	16	11	1	71	10	EU650409.1	Lima Júnior and Scarpassa (2009)
H2	99	0	9	22	30	7	0	2	1	38	0	EU650415.1	Lima Júnior and Scarpassa (2009)
H3	6	0	8	0	3	0	1	3	2	0	0	AY906852.1	Paduan and Ribolla (2008)
H4	10	0	11	1	6	0	4	0	1	0	0	EU650407.1	Lima Júnior and Scarpassa (2009)
H5	0	0	0	0	0	0	0	1	0	0	0	JQ303018	Gonçalves et al. (2012)
H6	85	0	0	0	1	0	0	0	1	0	0	EU650413.1	Lima Júnior and Scarpassa (2009)
H7	331	0	1	0	1	1	0	1	0	0	0	DQ176835.2	Bracco et al. (2007)
H8	1	0	0	0	0	0	0	2	0	0	0	EU446272.1	Burugu et al. (2008)
H9	0	0	0	0	0	0	0	1	0	0	0	AY906840.1	Paduan and Ribolla (2008)
H10	0	15	355	0	9	2	1	47	6	0	0	EU650414.1	Lima Júnior and Scarpassa (2009)
H11	0	0	0	0	1	0	0	3	1	0	0	EU650412.1	Lima Júnior and Scarpassa (2009)
H12	0	0	55	1	1	0	0	3	0	1	0	DQ176840.2	Bracco et al. (2007)
H13	0	0	26	0	0	0	0	3	0	0	0	DQ176831.2	Bracco et al. (2007)
H14	0	0	283	0	0	0	0	0	0	0	0	AF334858.1	Gorrochotegui-Escalante et al. (2002)
H15	0	0	55	0	0	0	0	0	0	0	0	AF334850.1	Gorrochotegui-Escalante et al. (2002)
H16	0	0	214	0	0	0	0	0	0	0	0	AF334851.1	Gorrochotegui-Escalante et al. (2002)
H17	0	0	93	0	0	0	0	0	0	0	0	AF334852.1	Gorrochotegui-Escalante et al. (2002)
H18	0	0	139	0	0	0	0	0	0	0	0	AF334854.1	Gorrochotegui-Escalante et al. (2002)
H19	0	0	130	0	0	0	0	0	0	0	0	AF334864.1	Gorrochotegui-Escalante et al. (2002)
H20	0	0	154	0	0	0	0	0	0	0	0	AF334859.1	Gorrochotegui-Escalante et al. (2002)
H21	0	0	57	0	0	0	0	0	0	0	0	AF334865.1	Gorrochotegui-Escalante et al. (2002)
H22	0	0	12	0	0	0	0	0	0	0	0	AF334848.1	Gorrochotegui-Escalante et al. (2002)
H23	0	0	17	0	0	0	0	0	0	0	0	AF334849.1	Gorrochotegui-Escalante et al. (2002)
H24	0	0	18	0	0	0	0	0	0	0	0	AF334857.1	Gorrochotegui-Escalante et al. (2002)
H25	0	0	0	1	1	0	0	0	0	0	0	EU650416.1	Gorrochotegui-Escalante et al. (2002)
H26	0	0	3	0	0	0	0	0	0	0	0	AF334861.1	Gorrochotegui-Escalante et al. (2002)
H27	0	0	4	0	0	0	0	0	0	0	0	AF334847.1	Gorrochotegui-Escalante et al. (2002)
H28	0	0	9	0	0	0	0	0	0	0	0	AF334862.1	Gorrochotegui-Escalante et al. (2002)
H29	0	0	0	1	1	0	0	0	0	0	0	EU650417.1	Lima Júnior and Scarpassa (2009)
H30	0	0	0	0	0	0	0	0	0	31	20	KF241756	Aguirre-Obando et al. (2015)
H31	0	0	0	0	0	0	0	0	0	1	0	JN896656	Caldera et al. (2013)
H32	0	0	0	0	0	0	0	0	0	1	11	KF241755	Aguirre-Obando et al. (2015)
H33	0	0	0	0	0	0	0	0	0	1	0	JN896678	Caldera et al. (2013)
H34	0	0	0	0	0	0	0	0	0	1	0	JN896686	Caldera et al. (2013)
H35	0	0	0	0	1	1	0	0	0	0	0	JQ303017	Gonçalves et al. (2012)
Total	626	51	1983	27	74	11	22	77	13	145	41		

BrAM (Brazilian Amazon), SEBr: Southeastern Brazil; PE: Peru; VZ: Venezuela; M-NA: Mexico - North America; MAO: Manaus; BL: Belém; BV: Boa Vista; PV - RB: Rio Branco - Porto Velho.

America to the south and gene flow from the south of South America to the north (Fig. 1b). In both scenarios, the Bayesian search strategy was conducted by using the following parameters: 2×10^5 generations, number of replicates = 4, burn = 1000, using a static heating scheme with four temperatures (temperatures: 1.0, 1.5, 3.0, and 100,000). The population sizes were assumed similar (that is, θ estimated). For DNA sequences, Migrate-N assumes an F84 model of nucleotide substitution (Kishino and Hasegawa, 1989; Felsenstein and Churchill, 1996), with or without variable substitution rates with gamma distribution between sites. The best test model was selected in each scenario evaluated according with the Log Bayes factor (LBF) based on the marginal accuracy of the Bezier approximation score generated for the models from each scenario.

Results

In all, for the *A. aegypti* populations from the Americas in the GenBank, the work obtained 2,996 sequences from the ND4 gene that after aligning and cutting had a length of 395 pb. These are distributed in the following manner: Mexico - North America (64.58%), Venezuela

(20.37%), Peru (1.66%), Brazil (7.32%; South - Eastern Brazil (2.50%), Brazilian Amazon (2.40%), Manaus (0.87%), Rio Blanco- Porto Velho (0.72%), Belém (0.45%), Boa Vista (0.35%)), and Colombia (6.05%). For Colombia, sequences were obtained from the departments of Sucre (4.72% distributed in the municipalities of: Sincelejo (2,03%), Guaranda (1.33%), Corazal (0.73%) and Sampues (0.70%)) and Quindio ((1.33%) distributed in the municipalities of: Armenia (0.40%), Montenegro (0.33%), Quimbaya (0.33%) and Barcelona (0.33%)).

Table 1 displays the distribution and H number of the ND4 gene for *A. aegypti* in the Americas, including Colombia. The study observed 35 H without presence of NUMTs, of which eight are in the Colombian populations. In general, H1 is the most frequent (19.20%) and is distributed in all the countries. The haplotype network at continental level suggests that the Colombian populations from the departments of Sucre and Quindio are connected with populations of mosquitoes from Peru, Venezuela, southeastern Brazil, the Brazilian Amazon, and Mexico-North America (Fig. 2a). In turn, the haplotype network at Colombian level suggests that H1-Col is the most frequent and ancestral H, located in the departments of Quindio and Sucre (Fig. 2b).



Figure 1 Geographic location of the *A. aegypti* populations included in this study and gene-flow models evaluated. In both graphics, circles indicate the populations and arrows represent the gene flow between populations. A) Scale at American continent level (N = 2,996 specimens from six locations: Mexico – North America (M-NA), Venezuela (VZ), Peru (PE), Brazilian Amazon (BrAm), southeastern Brazil (SEBr), and Colombia (CO). B) Scale at South America level (N = 1,083 specimens from six locations: Venezuela, Peru, Brazilian Amazon (5 locations; Brazilian Amazon (BrAM), Manaus (MAO), Belém (BL). Boa Vista (BV), Rio Branco (RB), Porto Velho (PV)), Southeastern Brazil (SEBr) and Colombia (2 locations; Sucre (S), Quindio (Q)).

Table 2 shows results of haplotypic diversity (Hd), nucleotide diversity (π), and neutrality test by countries and by locations. In general, the Hd from the Americas is 0.606 ± 0.14 (mean ± SD), while by countries the greater diversity occurred in Mexico-North America (0.731 ± 0.001), followed by Venezuela (0.655 ± 0.000), Colombia (0.648 ± 0.141), Brazil (0.595 ± 0.010), and Peru (0.423 ± 0.002). The π at Americas level was 0.008, while by countries it varied between 0.002 (Mexico-North America) and 0.017 (Brazilian Amazon). The Tajima D neutrality test (D) at Americas level indicates that most of the *A. aegypti* populations are in constant expansion, showing negative values in most cases (P < 0.05).

The migration scenario at continental scale suggests the hypothesis of the *stepping-stone* 1 gene-flow model with the highest subsequent probability (Model probability; 0.999, Log; -914.836 and LBF; 189.900) (Table 3), suggesting a connection of the bidirectional gene flow between the populations from Mexico-North America with the Colombian populations and connection between the populations from South America (Peru, Venezuela, Brazilian Amazon and southeastern Brazil) with the Colombian populations. A trampoline connection is shown between Mexico-North America with mosquito populations from southeastern Brazil (and from this population to the rest of the populations from South America) (Fig. 3a).

The migration scenario at South American spatial scale suggests that the hypothesis of the *full-migration* model had the highest subsequent probability (Model probability; 0.999, Log; -725.010, LBF; 0.073) (Table 3), indicating that gene flow among the populations from South America with respect to the Colombian populations (Sucre and Quindio) occurs in all the directionalities (Fig. 3b).

The phylogenetic analysis indicates that the 35 American H are grouped into two clades, I and II (Fig. 4). Clade I groups six of the eight H observed in the populations of Colombia (H1, H2, H12, H30, H31, H32, H33, and H34), seven H exclusive for Mexico-North America (H15, H16, H17, H19, H22, H23, H26) and four H widely disseminated in the Americas (H12, H4, H3, and H1). In turn, clade II contains haplotypes distributed in populations from Brazil, Venezuela, Mexico-North America, Peru, and two Colombian H. The terminals of the phylogenetic tree, represented and differentiated with blue, red, and green, indicate the H endemic for Mexico-North America, Colombia, and Brazil, respectively.

Discussion

The mitochondrial haplotypes detected for Colombia come from vector populations from the departments of Quindio (1,845 km², Latitude: 4.533872, Longitude: -75.676974) and Sucre (10,670 km², Latitude: 4.525659, Longitude:75.648716). The department of Quindio is located in the central-eastern zone of Colombia (Andean region)



Figure 2. Haplotype network for the mitochondrial ND4 gene from natural *A. aegypti* populations from: **A.** American continent, including sequences for Colombia; **B.** Colombia. In both haplotype networks, the size of each circle is proportional with the haplotypic frequency. The number next to each circle indicates the haplotype number.

Table 2

Results of genetic diversity and neutrality tests for Aedes aegypti populations from America.

Country	Localidad	Number of sequences		Neutrality test			
Country	LUCAIIUAU	Number of sequences	Number of haplotypes	Hd	π(%)	Tajimas' D	
Colombia	Sucre	145	8	0,650 ± 0,000	0,007	-2,604	*
	Quindio	41	3	0,646 ± 0,002	0,006	-2,208	*
Brazil	Brazilian Amazon ^a	74	12	0,755 ± 0,001	0,017	0,361	
	Manaus	27	6	0,341 ± 0,013	0,007	-3,151	*
	Rio blanco-Porto Velho	22	4	0,454 ± 0,013	0,003	-2,512	*
	Belem	14	7	0,813 ± 0,008	0,016	-2,147	*
	Boa Vista	11	4	0,600 ± 0,023	0,005	-4,396	
	Southeast of Brazil	77	11	0,606 ± 0,003	0,003	-2,504	*
Mex - Nort America		1984	22	0,731 ± 0,001	0,002	-0,475	
Peru		51	2	0,423 ± 0,002	0,013	2,669	*
Venezuela		626	7	$0,655 \pm 0,000$	0,012	-1,577	

^a Manaus, Rio Branco - Porto Velho, Belem, Boa Vista are part of the Brazilian Amazon; * significant difference (p < 0.05).

and is divided into 12 municipalities, with Armenia (1,573 m) as its capital city. In turn, the department of Sucre is located to the north of Colombia (Caribbean region) and comprises 26 municipalities, with Sincelejo (213 m) as the capital city. Both departments, Quindio and Sucre, are separated by 520 km and are interconnected through the Pan-American highway (a roadway system that joins almost all the countries of the western hemisphere in the American continent, with

the exception of the Darién Gap in Colombia (Holmes, 1963) and the Colombian roadway system (INVIAS, 2014).

Two mitochondrial lineages were found in the populations evaluated, a pattern evidenced in prior studies (Bosio et al., 2005; Gonçalves et al., 2012; Aguirre-Obando et al., 2015; Monsalve et al., 2021). The haplotypes present in populations from Colombia and Mexico-North America and the bidirectional gene flow between these populations, suggested in

Logarithmic marginal likelihood values, logarithmic Bayes factor, and probability models of gene-flow models on continental and South American scales.

Stage	Model	Log(mL)	LBF	Model probability
America	Full migration	-914,8366	189,90038	0,0000
	Nort-Sur	-972,5316	247,5954	0,0000
	Sur-Nort	-921,2833	196,3471	0,0000
	Panmitico	-948,5178	223,5816	0,0000
	St1	-887,6193	162,6831	1,0000
	St2	-966,4143	241,4781	0,0000
South América	Nort-Sur	-829,6100	104,6738	0,0000
	Sur-Nort	-819,7800	94,8438	0,0000
	Full migration	-725,0100	0,0738	1,0000
	St1	-799,5600	74,6238	0,0000
	St2	-826,6500	101,7138	0,0000



Figure 3. Representation of the gene-flow model with greater subsequent probability in the scenarios of the Americas and South America. **A.** Scale at American continent level (N = 2,996 specimens from six locations: Mexico–North America, Venezuela, Peru, Brazilian Amazon, southeastern Brazil, and Colombia. Within the continent context, the *Stepping Stone 1* model shows through the green vectors the way the gene flow occurs among the different populations evaluated. **B.** Scale at South America level; the *Full-migration* model indicates migration among all the populations, hence, plotting of the potential migration routes through passive land transport (Pan-American highway) and riverine paths (principal rivers).

our *stepping-stone – 1* model, could be supported by the presence of maritime routes between both countries, given that these have been associated with the global propagation of *A. aegypti*, bearing in mind population genetics studies conducted in maritime ports in Asia and Brazil (Paduan and Ribolla, 2008; Hlaing et al., 2010).

Among the mechanisms that could explain gene flow between the mosquito populations from Colombia toward Mexico-North America, there is passive migration due to human activity (Gorrochotegui-Escalante et al., 2002; Aguirre-Obando et al., 2015), specifically, the essential role of maritime ports in disseminating mosquitoes of the genus Aedes (Futami et al., 2015; Kollars, 2017). For example, in Colombia, the Port of Buenaventura on the Pacific is the main cargo port in the country, responsible for 50% of imports in Colombia (Pérez-Valbuena, 2007); it is located 165 km from the Department of Quindio. The Port of Barranquilla on the Atlantic, the second-most important in Colombia, has a direct maritime flow with the Gulf of Mexico and is located 730 km from the department of Quindio and 415 km from the department of Sucre (Otero, 2012). There is qualitative and quantitative evidence that recognizes the fundamental role of passive transportation of maritime flow and maritime ports in the dispersion and dissemination of immature mosquito forms (Chadee, 1984; Sprenger, 1987; Halstead, 2000; Fonzi et al., 2015). For example, immature forms transported by used tires (Medlock et al., 2012) and "Wet Footed" plants, such as

Lucky Bamboo (Demeulemeester et al., 2014) have been observed. Historically, in the Americas, the transportation of tires used by sea, and subsequently by land, has favored the dispersion of Aedes mosquitoes (Sprenger, 1987), where it has been suggested that this was one of the main sources of entrance of mosquitoes in the reinfestation of A aegypti from the United States to South America(Halstead, 2000). In Colombia, for 2021, for example, a total 4,653 tons of used tires from Mexico and USA (1.429 t), Brazil (2.397 t), Venezuela (25 t), Peru (141 t) and China (212 t) (ITC, 2022). The high plasticity of A. aegypti to the use of containers to deposit eggs and their ability to enter the temporary diapaus process allow them to survive to the evaporation process (draining) and long trips in small water bodies formed in small temporary ponds in the containers (Dallimore et al., 2020). Hence, it is suggested that the entry of individuals of this vector into the country from Mexico-North America to Colombia takes place through passive maritime transport. Moreover, additional genetic and population studies are suggested of the vector aimed at the Colombian ports to verify our hypothesis.

The connection of the Colombian populations with those from South America (Peru, Venezuela, and Brazil (= Brazilian Amazon + Southeastern Brazil) may be explained by two factors: riverine connectivity and land transport. Riverine connectivity has evidenced that it plays an important role in passive transport of *A. aegypti* populations between



Figure 4. Phylogeny constructed through Bayesian inference estimated from the 35H found of the ND4 gene for the *A. aegypti* populations in the American continent. The blue horizontal bars above the branches reflect the 95% CI for the branch supports. The color bars (blue, green, and red) on the tree terminals indicate which haplotypes are exclusive for a specific population. The dotted lines on the right side of the tree and numbers I or II indicate to what clade each of the terminals belong. H1-Col (Colombia (Sucre and Quindio), Venezuela, Peru, M-NA, Brazil (MA-O, RBPV, SEBr, BE-L)), H4 (Venezuela, M-NA, Brazil (MA-O, RBPV, SEBr, BE-L)), H3 (Venezuela, Peru, M-NA, Brazil (RBPV, SEBr, BE-L)), H3 (M-NA, Brazil (SEBr)), H8 (Venezuela, Brazil (SEBr)).

Peru and the Brazilian Amazon and is supported by observation of pupae on ships and barges between these locations (Guagliardo et al., 2014, 2015, 2019a). For land transport, along the American continent is the Pan-American highway that connects and facilitates land transport among most countries in the Americas, except for a 130-km stretch in Panama, Central America. In this sense, the Pan-American highway could facilitate passive migration between populations of vector mosquitoes from Colombia to the other countries in South America, given that it is recognized that roadway connectivity (Hlaing et al., 2010; Gonçalves et al., 2012; Maffey et al., 2020), tire cargo transport (Moore and Mitchell, 1997; Bennett et al., 2019), and flow of private vehicles (Eritja et al., 2017) influence directly on the passive migration of mosquitoes from the genus Aedes and its associated diseases (Mahabir et al., 2012). In any scenario, whether terrestrial or riverine, both are associated globally with passive transport of immature or adult forms of the dengue vector (Morrison et al., 2006; Gonçalves et al., 2012). A mathematical model proposed by our research group, incorporating land cargo transport for A. aegypti and the Mayaro virus, suggests that the presence of roadways with greater flow of cargo influence on the dissemination of the vector at broad geographic scales and, consequently, contribute with the epidemiological spread of arbovirus (Valencia-Marín et al., 2020).

Existing genetic connections among the vector's populations in southeastern Brazil and the Brazilian Amazon would be based on two

facts: the first, on the history of the vector's re-infestation in Brazil after the 1955 eradication campaign, which is presumed to have started in southeastern Brazil, reaching the central-western region and regions in the northeast by mid-1980 and, finally, reaching all the other Brazilian states in 1998 (Figueiredo, 2003); the second is based on the existence of the highways BR364 (communicating São Paulo-Acre) and BR153 (Rio Grande do Sul-Belém) in Brazil, which connect the north and south regions in Brazil (Gonçalves et al., 2012). Existing connections among *A. aegypti* populations between Venezuela and the northern Brazilian Amazon (Boa Vista) are based on the presence of dengue serotypes in Boa Vista, previously only registered in Venezuela (Figueiredo, 2003; Lourenço-de-Oliveira et al., 2004; Codeço et al., 2009).

In a national scale, the gene flow of *A. aegypti* populations in Colombia, according to that found in the migration hypothesis at South American scale occurs in panmictic manner (Fig. 3b). This hypothesis is supported by recent studies with microsatellites for three Colombian populations that evidence panmictic behavior (Monsalve et al., 2021). Consequently, the vector's dispersion toward the center of Colombia could occur through land or river communication routes. Land communication would be taking place through the principal roadways in Colombia; among them, the Pan-American highway, which communicates the central-eastern region with the north of the country. It is estimated that annually close to 9.5-million t (Pérez-Valbuena, 2007) are transported from Valle del

Cauca (mainly from Buenaventura) to the four principal capital cities in Colombia (Bogotá, Cali, Medellín, and Barranquilla). Most of this cargo goes through highways in the department of Quindio, which permit roadway communication with Bogotá, Medellín, and Barranquilla.

In turn, riverine transport, principally through the Magdalena and Cauca rivers, contribute with the passive dispersion of the vector from the central-eastern region to the north of Colombia and vice versa. The Magdalena River originates on the border between the departments of Cauca and Huila and the Cauca River in the department of Cauca; both departments located in the central-western region of Colombia. The Magdalena and Cauca rivers form the first and second riverine artery in Colombia, respectively. The Magdalena River, whose principal affluent is the Cauca River, empties into the Caribbean Sea at 7.5 km from Barranquilla. The Magdalena River occupies 24% of the Colombian continental territory, where 18 departments are located and where 80% of the population inhabits and 85% of the Colombian gross domestic product (GDP) is produced (Ministerio de Ambiente, Vivienda y Desarrollo Territorial, 2010).

However, more studies are needed in different parts of Colombia to corroborate the hypotheses exposed in this document.

Conclusions

Strategies for *A. aegypti* monitoring and vector control must pay close attention to the vector's points of entry to Colombia related with Peru, Venezuela, Brazil, Mexico, and North America, to avoid the entry (and exit) of new populations with characteristics, like resistance to insecticides or vector competition to arboviruses circulating in Colombia or to new emerging arboviruses. Our results show the importance and implication of the different mechanisms and migration routes of the vector in two spatial scales, continental and subcontinental, which can be considered to implement and plan vector monitoring and control programs.

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Conflicts of Interest

The authors declare having no conflict of interests.

References

Aguirre-Obando, O., Dalla Bonna, A., Luna, D., Navarro-Silva, M. A., 2015. Insecticide resistance and genetic variability in natural populations of *Aedes* (*Stegomyia*) aegypti (Diptera: Culicidae) from Colombia. Zoologia 32 (1), 14-22. http://dx.doi.org/10.1590/ S1984-46702015000100003.

- Aliota, M. T., Peinado, S. A., Velez, I. D., Osorio, J. E., 2016. The wMel strain of Wolbachia reduces transmission of Zika virus by *Aedes aegypti*. Sci. Rep. 6 (1), 28792. http://dx.doi.org/10.1038/srep28792.
- Badii, M., Landeros, J., Cerna, E., Abreu, J., 2007. Ecología e historia del dengue en las Américas. Daena Int. J. Good Conscience 2, 248-273.
- Beerli, P. 2004. Migrate Documentation. School of Computational Science and Department of Biological Science, Florida State University, Tallahassee, FL. Available in: http://popgen.sc.fsu.edu/ oldversions/0.x/0.7.1/migdoc.pdf (accessed 5 August 2021).
- Bennett, K. L., Gómez Martínez, C., Almanza, A., Rovira, J. R., McMillan, W. O., Enriquez, V., Barraza, E., Diaz, M., Sanchez-Galan, J. E., Whiteman, A., Gittens, R. A., Loaiza, J. R., 2019. High infestation of invasive *Aedes* mosquitoes in used tires along the local transport network of Panama. Parasit. Vectors 12 (1), 264. http://dx.doi.org/10.1186/ s13071-019-3522-8.
- Black 4th, I. V., Bernhardt, S. A., 2009. Abundant nuclear copies of mitochondrial origin (NUMTs) in the *Aedes aegypti* genome. Insect Mol. Biol. 18 (6), 705-713. http://dx.doi.org/10.1111/j.1365-2583.2009.00925.x.
- Bosio, C. F., Harrington, L. C., Norris, D., Scott, T. W., Jones, J. W., Sithiprasasna, R., 2005. Genetic structure of *Aedes aegypti* populations in Thailand using mitochondrial DNA. Am. J. Trop. Med. Hyg. 72 (4), 434-442. http://dx.doi.org/10.4269/ajtmh.2005.72.434.
- Bouckaert, R., Heled, J., Kühnert, D., Vaughan, T., Wu, C.-H., Xie, D., Suchard, M. A., Rambaut, A., Drummond, A. J., 2014. BEAST 2: a software platform for Bayesian evolutionary analysis. PLOS Comput. Biol. 10 (4), e1003537. http://dx.doi.org/10.1371/journal.pcbi.1003537.
- Bracco, J., Capurro, M. L., Lourenço-de-Oliveira, R., Sallum, M., 2007. Genetic variability of *Aedes aegypti in* the Americas using a mitochondrial gene: evidence of multiple introductions. Mem. Inst. Oswaldo Cruz 102 (5), 573-580. http://dx.doi.org/10.1590/ S0074-02762007005000062.
- Brown, J. E., McBride, C. S., Johnson, P., Ritchie, S., Paupy, C., Bossin, H., Lutomiah, J., Fernandez-Salas, I., Ponlawat, A., Cornel, A. J., Black 4th, W. C., Gorrochotegui-Escalante, N., Urdaneta-Marquez, L., Sylla, M., Slotman, M., Murray, K. O., Walker, C., Powell, J. R., 2011. Worldwide patterns of genetic differentiation imply multiple 'domestications' of *Aedes aegypti*, a major vector of human diseases. Proc. Biol. Sci. 278 (1717), 2446-2454. http://dx.doi.org/10.1098/rspb.2010.2469.
- Burugu, M., Sang, R., Kamau, L., 2008. Genetic Structure of *Aedes aegypti* Populations in Coastal and Inland Kenya Using Mitochondrial DNA. Kenya Medical Research Institute, Kenya.
- Caldera, S., Jaramillo, S., Cochero, S., Pérez-Doria, A., Bejarano, E., 2013. Diferencias genéticas entre poblaciones de *Aedes aegypti* de municipios del norte de Colombia, con baja y alta incidencia de dengue. Biomedica 33, 89-98.
- Chadee, D. D., 1984. *Aedes aegypti* aboard boats at Port-of-Spain, Trinidad, West Indies (1972-82). Mosq. News 44, 1-3.
- Codeço, C. T., Honório, N. A., Ríos-Velásquez, C. M., Santos, M., Mattos, I. V., Luz, S. B., Reis, I. C., Cunha, G. B., Rosa-Freitas, M. G., Tsouris, P., Castro, M. G., Hayd, R. L. N., Luitgards-Moura, J. F., 2009. Seasonal dynamics of *Aedes aegypti*(Diptera: Culicidae) in the northernmost state of Brazil: a likely port-of-entry for dengue virus 4. Mem. Inst. Oswaldo Cruz 104 (4), 614-620. http://dx.doi.org/10.1590/S0074-02762009000400014.
- Dallimore, T., Goodson, D., Batke, S., Strode, C., 2020. A potential global surveillance tool for effective, low-cost sampling of invasive *Aedes* mosquito eggs from tyres using adhesive tape. Parasit. Vectors 13 (1), 1-16. http://dx.doi.org/10.1186/s13071-020-3939-0.
- Darriba, D., Taboada, G. L., Doallo, R., Posada, D., 2012. jModelTest 2: more models, new heuristics and parallel computing. Nat. Methods 9 (8), 772. http://dx.doi.org/10.1038/nmeth.2109.

- Demeulemeester, J., Deblauwe, I., de Witte, J. C., Jansen, F., Hendy, A., Madder, M., 2014. First interception of *Aedes* (*Stegomyia*) *albopictus* in Lucky bamboo shipments in Belgium. J. Eur. Mosq. Control Assoc. 32, 14-16.
- Eritja, R., Palmer, J. R. B., Roiz, D., Sanpera-Calbet, I., Bartumeus, F., 2017. Direct evidence of adult *Aedes albopictus* dispersal by car. Sci. Rep. 7 (1), 1-15. http://dx.doi.org/10.1038/s41598-017-12652-5.
- Felsenstein, J., Churchill, G. A., 1996. A Hidden Markov Model approach to variation among sites in rate of evolution. Mol. Biol. Evol. 13 (1), 93-104. http://dx.doi.org/10.1093/oxfordjournals.molbev.a025575.
- Figueiredo, L. T. M., 2003. Dengue in Brazil: Past, Present and Future Perspective. WHO Regional Office for South-East Asia, New Delhi.
- Fonzi, E., Higa, Y., Bertuso, A. G., Futami, K., Minakawa, N., 2015. Humanmediated marine dispersal influences the population structure of *Aedes aegypti* in the Philippine Archipelago. PLoS Negl. Trop. Dis. 9 (6), e0003829. http://dx.doi.org/10.1371/journal.pntd.0003829.
- Futami, K., Valderrama, A., Baldi, M., Minakawa, N., Marin Rodriguez, R., Chaves, L. F., 2015. New and common haplotypes shape genetic diversity in Asian tiger mosquito populations from Costa Rica and Panamá. J. Econ. Entomol. 108 (2), 761-768. http://dx.doi. org/10.1093/jee/tou028.
- Gonçalves, A., Cunha, I., Santos, W., Luz, S., Ribolla, P., Abad-Franch, F., 2012. Gene flow networks among American *Aedes aegypti* populations. Evol. Appl. 5 (7), 664-676. http://dx.doi.org/10.1111/j.1752-4571.2012.00244.x.
- Gorrochotegui-Escalante, N., Gomez-Machorro, C., Lozano-Fuentes, S., Fernandez-Salas, I., Munoz, M. L., Farfan-Ale, J. A., Garcia-Rejon, J., Beaty, B. J., Black 4th, W. C., 2002. Breeding structure of *Aedes aegypti* populations in Mexico varies by region. Am. J. Trop. Med. Hyg. 66 (2), 213-222. http://dx.doi.org/10.4269/ajtmh.2002.66.213.
- Groot, H., 1980. The reinvasion of Colombia by *Aedes aegypti*: aspects to remember. Am. J. Trop. Med. Hyg. 29 (3), 330-338. http://dx.doi. org/10.4269/ajtmh.1980.29.330.
- Guagliardo, S. A. J., Ardila Roldan, S. C., Santacoloma, L., Luna, C., Cordovez Alvarez, J. M., Rojas Gacha, J. D., Mansur, M., Levine, R. S., Lenhart, A., Oviedo, P. F., 2019b. Enhanced vector surveillance to control arbovirus epidemics in Colombia. Rev. Panam. Salud Publica 43, 1. http://dx.doi.org/10.26633/RPSP.2019.50.
- Guagliardo, S. A. J., Lee, Y., Pierce, A. A., Wong, J., Chu, Y. Y., Morrison, A. C., Astete, H., Brosi, B., Vazquez-Prokopec, G., Scott, T. W., Kitron, U., Stoddard, S. T., 2019a. The genetic structure of *Aedes aegypti* populations is driven by boat traffic in the Peruvian Amazon. PLoS Negl. Trop. Dis. 13 (9), e0007552. http://dx.doi.org/10.1371/journal. pntd.0007552.
- Guagliardo, S. A., Barboza, J. L., Morrison, A. C., Astete, H., Vazquez-Prokopec, G., Kitron, U., 2014. Patterns of geographic expansion of *Aedes aegypti* in the Peruvian Amazon. PLoS Negl. Trop. Dis. 8 (8), e3033. http://dx.doi.org/10.1371/journal.pntd.0003033.
- Guagliardo, S. A., Morrison, A. C., Luis Barboza, J., Wesson, D. M., Ponnusamy, L., Astete, H., Vazquez-Prokopec, G., Kitron, U., 2015. Evidence for *Aedes aegypti*(Diptera: Culicidae) oviposition on boats in the Peruvian Amazon. J. Med. Entomol. 52 (4), 726-729. http:// dx.doi.org/10.1093/jme/tjv048.
- Halstead, S. B., 2000. Successes and failures in dengue control-global experience. Dengue Bull. 24, 20-24.
- Hlaing, T., Tun-Lin, W., Somboon, P., Socheat, D., Setha, T., Min, S., Chang, M. S., Walton, C., 2009. Mitochondrial pseudogenes in the nuclear genome of *Aedes aegypti* mosquitoes: implications for past and future population genetic studies. BMC Genet. 10 (1), 11. http:// dx.doi.org/10.1186/1471-2156-10-11.
- Hlaing, T., Tun-Lin, W., Somboon, P., Socheat, D., Setha, T., Min, S., Thaung, S., Anyaele, O., de Silva, B., Chang, M. S., Prakash, A., Linton, Y., Walton,

C., 2010. Spatial genetic structure of *Aedes aegypti* mosquitoes in mainland Southeast Asia. Evol. Appl. 3 (4), 319-339. http://dx.doi. org/10.1111/j.1752-4571.2009.00113.x.

- Holmes, J., 1963. The Pan American Highway. J. Geog. 62 (4), 145-152. http://dx.doi.org/10.1080/00221346308982184.
- Instituto Nacional de Vias INVIAS, 2014. Galeria de mapas del Instituto Nacional de Vias (INVIAS) de Colombia. Available in: http://www. icde.org.co/web/guest/galeriam_invias (accessed 9 January 2019).
- International Trade Centre ITC, 2022. Trade Map: 4012 Retreaded or Used Tires "Pneumatic Rims". Available in: https://intracen.org/ (accessed 26 June 2022).
- Kantor, I. N., 2018. Dengue, zika, chikungunya y el desarrollo de vacunas. Rev. Med. 78 (1), 23-28.
- Katoh, K., Standley, D. M., 2013. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. Mol. Biol. Evol. 30 (4), 772-780. http://dx.doi.org/10.1093/molbev/mst010.
- Kishino, H., Hasegawa, M., 1989. Evaluation of the maximum likelihood estimate of the evolutionary tree topologies from DNA sequence data, and the branching order in Hominoidea. J. Mol. Evol. 29 (2), 170-179. http://dx.doi.org/10.1007/BF02100115.
- Kollars, T. M., 2017. Assessing likely invasion sites of Zika virus-infected mosquitoes in civilian and naval maritime ports in Florida. Res. Rep. Trop. Med. 8, 1-6. http://dx.doi.org/10.2147/RRTM.S123456.
- Li, J., Ai, S., 2020. Impulsive releases of sterile mosquitoes and interactive dynamics with time delay. J. Biol. Dyn. 14(1), 289-307. http://dx.doi. org/10.1080/17513758.2020.1748239.
- Lima Júnior, R., Scarpassa, V., 2009. Evidence of two lineages of the dengue vector *Aedes aegypti* in the Brazilian Amazon, based on mitochondrial DNA ND4 gene sequences. Genet. Mol. Biol. 32 (2), 414-422. http://dx.doi.org/10.1590/S1415-47572009005000036.
- Lounibos, L. P., 2002. Invasions by insect vectors of human disease. Annu. Rev. Entomol. 47 (1), 233-266. http://dx.doi.org/10.1146/ annurev.ento.47.091201.145206.
- Lourenço-de-Oliveira, R., Vazeille, M., de Filippis, A. M. B., Failloux, A. B., 2004. *Aedes aegypti* in Brazil: genetically differentiated populations with high susceptibility to dengue and yellow fever viruses. T R. Soc Trop Med H 98 (1), 43-54. http://dx.doi.org/10.1016/S0035-9203(03)00006-3.
- Maffey, L., Garzón, M. J., Confalonieri, V., Chanampa, M. M., Hasson, E., Schweigmann, N., 2020. Genome-wide screening of *Aedes aegypti* (Culicidae: Diptera) populations from northwestern Argentina: active and passive dispersal shape genetic structure. J. Med. Entomol. 57 (6), 1930-1941. http://dx.doi.org/10.1093/jme/tjaa125.
- Magalhães, R. C. S., 2016. A erradicação do *Aedes aegypti*: febre amarela, Fred Soper e saúde pública nas Américas (1918-1968). Editora FIOCRUZ, Rio de Janeiro. https://doi.org/10.7476/9788575414798.
- Magalhães, R. C. S., 2021. As Américas contra o mosquito: a Campanha Continental para a Erradicação do *Aedes aegypti* e a cooperação interamericana em saúde (1947-1968). Contraponto 10, 438.
- Mahabir, R. S., Severson, D. W., Chadee, D. D., 2012. Impact of road networks on the distribution of dengue fever cases in Trinidad, West Indies. Acta Trop. 123 (3), 178-183. http://dx.doi.org/10.1016/j. actatropica.2012.05.001.
- Marcombe, S., Paris, M., Paupy, C., Bringuier, C., Yebakima, A., Chandre, F., David, J.-P., Corbel, V., Despres, L., 2013. Insecticide-Driven Patterns of Genetic Variation in the Dengue Vector *Aedes aegypti* in Martinique Island. PLoS One 8 (10), e77857. http://dx.doi.org/10.1371/journal. pone.0077857.
- Martínez, M., Estévez, A., Quijada, H., Walteros, D., Tolosa, N., Paredes, A., 2015. IQEN (Informe Quincenal Epidemiológico Nacional). Bogotá. Available in: https://www.ins.gov.co/buscador-eventos/IQEN/ IQEN%20vol%2020%202015%20num%205.pdf (accessed 6 June 2018).

- Medlock, J. M., Hansford, K. M., Schaffner, F., Versteirt, V., Hendrickx, G., Zeller, H., van Bortel, W., 2012. A review of the invasive mosquitoes in Europe: Ecology, public health risks, and control options. Vector Borne Zoonotic Dis. 12 (6), 435-447. http://dx.doi.org/10.1089/ vbz.2011.0814.
- Mejía, G., Mora, G., Ramos, E., Mestre, R., Mazanett, E., Malambo, D., Gomez, D., 2011. Identificación genética de sub-poblaciones de *Aedes aegypti* en Cartagena de Indias-Colombia. Rev. Cinc. Biomed. 1, 5. https://doi.org/doi.org/10.7705/biomedica.v38i0.3728.
- Ministerio de Ambiente, Vivienda y Desarrollo Territorial, 2010. Política nacional para la gestión integral del recurso hídrico. Imprenta Nacional de Colombia, Bogotá.
- Monsalve, Y., Triana-Chávez, O., Gómez-Palacio, A., 2021. Population structure and ancestry prediction of *Aedes aegypti*(Diptera: Culicidae) supports a single African origin of Colombian populations. Mem. Inst. Oswaldo Cruz 116, e200441. http://dx.doi.org/10.1590/0074-02760200441.
- Monteiro, F., Shama, R., Martins, A., Gloria-Soria, A., Brown, J., Powell, J., 2014. Genetic diversity of Brazilian *Aedes aegypti*: patterns following an Eradication Program. PLoS Negl. Trop. Dis. 8 (12), e3460. http:// dx.doi.org/10.1371/journal.pntd.0003460.
- Moore, C. G., Mitchell, C., 1997. *Aedes albopictus* in the United States: ten-year presence and public health implications. Emerg. Infect. Dis. 3 (3), 329-334. http://dx.doi.org/10.3201/eid0303.970309.
- Moore, M., Sylla, M., Goss, L., Burugu, M. W., Sang, R., Kamau, L. W., Kenya, E. U., Bosio, C., Munoz, M. L., Sharakova, M., Black, W. C., 2013. Dual African Origins of Global *Aedes aegyptis.*l. populations Revealed by Mitochondrial DNA. PLoS Negl. Trop. Dis. 7 (4), e2175. http://dx.doi.org/10.1371/journal.pntd.0002175.
- Morrison, A. C., Sihuincha, M., Stancil, J. D., Zamora, E., Astete, H., Olson, J. G., Vidal-Ore, C., Scott, T. W., 2006. *Aedes aegypti* (Diptera: Culicidae) production from non-residential sites in the Amazonian city of Iquitos, Peru. Ann. Trop. Med. Parasitol. 100 (sup1), 73-86. http://dx.doi.org/10.1179/136485906X105534.
- Ocampo, C., Wesson, D., 2004. Population dynamics of *Aedes aegypti* from a dengue hyperendemic urban setting in Colombia. Am. J. Trop. Med. Hyg. 71 (4), 506-513. http://dx.doi.org/10.4269/ajtmh.2004.71.506.
- Olanratmanee, P., Kittayapong, P., Chansang, C., Hoffmann, A. A., Weeks, A. R., Endersby, N. M., 2013. Population genetic structure of *Aedes* (*Stegomyia*) *aegypti* (L.) at a micro-spatial scale in Thailand: implications for a dengue suppression strategy. PLoS Negl. Trop. Dis. 7 (1), e1913. http://dx.doi.org/10.1371/journal.pntd.0001913.
- Organización Panamericana de la Salud OPS, 1992. El dengue y la fiebre hemorrágica del dengue en las Américas: una visión general del problema. Bol. Epidemiologico. 13, 9-10.
- Organización Panamericana de la Salud OPS, 2011. Guía de vigilancia entomológica y control del dengue. Colombia. Available in: http:// new.paho.org/col/index.php?option=com_docman&task=doc_ download&gid=1215&Itemid= (accessed 5 August 2021).
- Otero, A., 2012. El puerto de Barranquilla: retos y recomendaciones. Rev. Econ. Caribe 10, 126-159.
- Paduan, K. D. S., Ribolla, P. E. M., 2008. Mitochondrial DNA polymorphism and heteroplasmy in populations of *Aedes aegypti* in Brazil. J. Med. Entomol. 45 (1), 59-67. http://dx.doi.org/10.1093/jmedent/45.1.59.
- Paradis, E., Kamvar, Z.N., Jombart, T., Brian, K., Frederic, M., 2020. Pegas: Population and Evolutionary Genetics Analysis System. R Foundation for Statistical Computing, Vienna.
- Paupy, C., le Goff, G., Brengues, C., Guerra, M., Revollo, J., Barja Simon, Z., Hervé, J.-P., Fontenille, D., 2012. Genetic structure and phylogeography of *Aedes aegypti*, the dengue and yellow-fever mosquito vector in Bolivia. Infect. Genet. Evol. 12 (6), 1260-1269. http://dx.doi. org/10.1016/j.meegid.2012.04.012.

- Pérez-Valbuena, G.J., 2007. Historia, geografía y puerto como determinantes de la situación social de Buenaventura. Centro de Estudios Económicos Regionales, Cartagena. (Documentos de Trabajo Sobre Economía Regional y Urbana, 91).
- Pollett, S., Fauver, J. R., Maljkovic Berry, I., Melendrez, M., Morrison, A., Gillis, L. D., Johansson, M. A., Jarman, R. G., Grubaugh, N., 2020. Genomic epidemiology as a public health tool to combat mosquitoborne virus outbreaks. Int. J. Infect. Dis. 221 (Suppl.3), S308-S318. http://dx.doi.org/10.1093/infdis/jiz302.
- Rašić, G., Filipović, I., Weeks, A., Hoffmann, A., 2014. Genome-wide SNPs lead to strong signals of geographic structure and relatedness patterns in the major arbovirus vector, *Aedes aegypti*. BMC Genomics 15 (1), 275. http://dx.doi.org/10.1186/1471-2164-15-275.
- Rico-Mendoza, A., Porras-Ramírez, A., Chang, A., Encinales, L., Lynch, R., 2019. Co-circulation of dengue, chikungunya, and Zika viruses in Colombia from 2008 to 2018. Rev. Panam. Salud Publica 43, e49. http://dx.doi.org/10.26633/RPSP.2019.49.
- Rodríguez, G., de La Hoz, R., 2004. Dengue and dengue and vector behaviour in Cáqueza, Colombia. Rev. Salud Publica 7, 1-15.
- Rodríguez, G., Velandia, M., Boshell, J., 2003. Fiebre amarilla, la enfermedad y su control. Instituto Nacional de Salud, Bogotá.
- Ruiz-López, F., González-Mazo, A., Vélez-Mira, A., Gómez, G. F., Zuleta, L., Uribe, S., Vélez-Bernal, I. D., 2016. Presencia de *Aedes* (*Stegomyia*) *aegypti* (Linnaeus, 1762) y su infección natural con el virus del dengue en alturas no registradas para Colombia. Biomedica 36 (2), 303-308. http://dx.doi.org/10.7705/biomedica.v36i2.3301.
- Seixas, G., Salgueiro, P., Silva, A. C., Campos, M., Spenassatto, C., Reyes-Lugo, M., Novo, M. T., Ribolla, P. E. M., Pinto, J. P. S. S., Sousa, C. A., 2013. *Aedes aegypti* on Madeira Island (Portugal): genetic variation of a recently introduced dengue vector. Mem. Inst. Oswaldo Cruz 108 (Suppl.1), 3-10. http://dx.doi.org/10.1590/0074-0276130386.
- Sprenger, P. R. D., 1987. The used tire trade: a mechanism for the worldwide dispersal of container breeding mosquitoes. J. Am. Mosq. Control Assoc. 3, 2904963.
- Templeton, A. R., Crandall, K. A., Sing, C. F., 1992. A cladistic analysis of phenotypic associations with haplotypes inferred from restriction endonuclease mapping and DNA sequence data. III. Cladogram estimation. Genetics 132 (2), 619-633. http://dx.doi.org/10.1093/ genetics/132.2.619.
- Twerdochlib, A. L., Bonna, A. C. D., Leite, S. S., Chitolina, R. F., Westphal, B., Navarro-Silva, M. A., 2012. Genetic variability of a population of *Aedes aegypti* from Paraná, Brazil, using the mitochondrial ND4 gene. Rev. Bras. Entomol. 56 (2), 249-256. http://dx.doi.org/10.1590/ S0085-56262012005000030.
- Urdaneta-Marquez, L., Failloux, A.-B., 2011. Population genetic structure of *Aedes aegypti*, the principal vector of dengue viruses. Int. J. Infect. Dis. 11, 253-261. https://doi.org/doi.org/10.1016/j.meegid.2010.11.020.
- Valencia-Marín, B. S., Gandica, I. D., Aguirre-Obando, O. A., 2020. The Mayaro virus and its potential epidemiological consequences in Colombia: an exploratory biomathematics analysis. Parasit. Vectors 13 (1), 508. http://dx.doi.org/10.1186/s13071-020-04354-1.
- Velez, I. D., Santacruz, E., Kutcher, S. C., Duque, S. L., Uribe, A., Barajas, J., Gonzalez, S., Patino, A. C., Zuluaga, L., Martínez, L., Muñoz, E., Mejia, M. C., Arbelaez, M. P., Pulido, H., Jewell, N. P., Dufault, S. M., O'Neill, S. L., Simmons, C. P., Anders, K. L., Tanamas, S. K., 2020. The impact of city-wide deployment of Wolbachia-carrying mosquitoes on arboviral disease incidence in Medellín and Bello, Colombia: study protocol for an interrupted time-series analysis and a test-negative design study. F1000 Res. 8, 1327. http://dx.doi.org/10.12688/f1000research.19858.2.
- Weaver, S., 2014. Arrival of Chikungunya virus in the new Word. Prospects for spread and impact on public health. PLoS Negl. Trop. Dis. 8 (6), e2921. http://dx.doi.org/10.1371/journal.pntd.0002921.

- World Health Organization WHO, 2013. Sustaining the Drive to Overcome the Global Impact of Neglected Tropical Diseases. Ginebra. Available in: http://apps.who.int/iris/bitstream/10665/77950/1/9789241564540_ eng.pdf (accessed 5 August 2021).
- World Health Organization WHO, 2016. Test Procedures for Insecticide Resistance Monitoring in Malaria Vector Mosquitoes, 2nd ed. Ginebra. Available in: https://apps.who. int/iris/bitstream/handle/10665/250677/9789241511575-eng.

pdf;jsessionid=032D674946ED0785F4522A191C5D0E5B?sequence=1 (accessed 5 August 2021).

- Yakob, L., Walker, T., 2016. Zika virus outbreak in the Americas: the need for novel mosquito control methods. Lancet Glob. Health 4 (3), e148-e149. http://dx.doi.org/10.1016/S2214-109X(16)00048-6.
- Yáñez, P., Mamani, E., Valle, J., García, M. P., León, W., Villaseca, P., Torres, D., Cabezas, C., 2013. Variabilidad genética del *Aedes aegypti* determinada mediante el análisis del gen mitocondrial ND4 en once áreas endémicas para dengue en el Perú. Rev. Peru. Med. Exp. Salud Publica 30, 246-250.