



RESEARCH ARTICLE

Fish habitat associations along a longitudinal gradient in a preserved coastal Atlantic stream, Brazil

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ABSTRACT. Habitat conditions at multiple scales are one of the major factors structuring ichthyofauna. Thus, we analyzed the fish habitat associations along the headwater-mouth gradient of a coastal Atlantic stream. We categorized the sampling sites into habitat units, so that in the middle reach these categories were statistically differentiated into riffles, runs and pools. Samplings were carried out quarterly from May 2009 to February 2010 using electrofishing. Principal component analysis (PCA) indicated an environmental gradient from higher water velocity and rocky bottom to deeper and sandy areas in the headwater-mouth direction. A total of 1,495 individuals belonging to 27 species were captured, being 13, 18 and 22 from headwater, middle and mouth reaches, respectively. Shannon diversity was slightly higher in the middle reach, while beta diversity showed higher rates of addition than turnover in species along the longitudinal gradient. Fish structure, evaluated by DCA (detrended correspondence analysis) scores, showed significant differences between upper reaches and mouth reach, but the middle riffles did not differ from headwater habitats. In the middle reach, mesohabitat analysis distinguished riffles, with higher abundance of fast-water crenuchids, from pools, with a higher abundance of lentic-water characids. These results suggest that environmental differences along the stream determine the wider structural patterns. However, the middle reach amassed species from upper areas and lowlands in structured fish mesohabitat associations, possibly implying distinct local ecological interactions. These findings contribute to the assessment of stream conservation status and to recognize eventual direct impacts on fish structures along longitudinal gradients.

KEY WORDS. Abiotic gradient, diversity, ichthyofauna, mesohabitat, multiple scales.

INTRODUCTION

A major challenge in ecology of freshwater environments is understanding the functioning of river systems and identifying the mechanisms that underlie the structure of communities (Melles et al. 2012). A coherent approach to understanding these mechanisms is to compare communities along rivers, since fluvial and geomorphological differences occur longitudinally (Angermeier and Karr 1983, Ibañez et al. 2007, Silva et al. 2013). In this sense, the composition and diversity of the aquatic fauna have showed longitudinal changes, with adjustments at each local environmental conditions to its functioning along river system networks (Vannote et al. 1980, Oberdorff et al. 1993, Ferreira and Petrere Jr 2009, Suvarnaraksha et al. 2012).

Longitudinal changes in the structure and richness of ichthyofauna have been widely recorded both in temperate (Oberdorff et al. 1993, Pekárik et al. 2011) and tropical streams (Casatti 2005, Petry and Schulz 2006, Ferreira and Petrere Jr 2009, Silva et al. 2013). Several findings, summarized in Ibañez et al. (2009) have showed similar longitudinal increases of fish richness and proportional shifts of trophic guilds along rivers. Therefore, fish community structures converge to similar patterns along biogeographically distinct rivers, regardless of their phylogenetic and historical constrains (Ibañez et al. 2009). In this sense, geomorphological parameters, such as altitude, discharge, water velocity and channel depth, have more influence in predicting the increases in local fish richness/diversity and changes in fish community composition along the headwater-mouth direction,



than physicochemical water variables (Lorion et al. 2011, Suvarnaraksha et al. 2012). For example, headwater reaches with fast water and low environmental heterogeneity has shown few species that are able to survive under such conditions, while larger and more diversified downstream habitat reaches tend to support a richer and more diverse ichthyofauna (Ibañez et al. 2007, Suvarnaraksha et al. 2012). Moreover, according to local adaptations and species composition, studies have proposed that upper, middle and lower reaches of rivers are discrete zones of fish colonization (Ferreira and Petrere Jr 2009, McGarvey 2011).

However, longitudinal changes in composition and richness of fish species need a detailed assessment because their structural differences may also occur at reduced spatial scales (Rahel and Hubert 1991). At local scales, especially at moderate slopes gradients, streams are structurally complex and can be divided into mosaics of riffles, pools and runs mesohabitats (Allan 1995, Rezende et al. 2010), which are clearly delimited by different combinations of water velocity, depth and substrate composition (Angermeier and Schlosser 1989, Langeani et al. 2005). As mesohabitats have significant differences in their specific sets of environmental conditions, they play an important role in structuring the ichthyofauna (Rezende et al. 2010, Teresa and Casatti 2012), because fish communities have different strategies for foraging, breeding and obtaining refuge from predators (Matthews 1998).

Recently, efforts have been made to detect differences in functional and taxonomic diversity (Teresa and Casatti 2012), interspecific variation in habitat selection (Kano et al. 2013), guild composition (Pegg et al. 2014), and structure and complexity of food webs (Worischka et al. 2014) at mesohabitat scale. These studies have improved the knowledge on fish ecological differences among riffles, pools, and runs, and highlighted the importance of spatial mesoscale in assessing environments with contrasting conservation status (Teresa and Casatti 2012). On the other hand, since smaller spatial units are nested within broader environmental characteristics of basin network, the main challenge in understanding fish spatial structuration is assess the effects of multiple spatial scales interaction (i.e. micro and mesohabitat, stream reaches and landscape) because connections among different scales might create emergent patterns that transcend local processes (Cheek et al. 2016).

Atlantic forest is one of the world's hotspots for biodiversity conservation and, despite its threatened fauna and flora (Myers et al. 2000), coastal Atlantic streams located in this biome harbor one of the richest and ecologically most complex ichthyofaunas in the Neotropical Region (Vari and Malabarba 1998). These streams are represented by small and medium sized water bodies that run along upper cascaded channels, passing through intermediate reaches, which usually include combinations of riffles, pool and runs to slow lowlands moving channels (Rincón 1999, Oyakawa et al. 2006, Rezende et al. 2010). Coastal Atlantic streams support a high diversity of small-sized fish species with limited geographical distributions and high endemism (Esteves

and Lobón-Cerviá 2001, Barreto and Aranha 2005, Gonçalves and Braga 2012).

Upper courses of coastal Atlantic streams from southern and southeast Brazil are widely located in preserved and topographically irregular areas of Serra do Mar mountains (Oyakawa et al. 2006), making it difficult to carry out integrative spatial studies about distribution patterns of ichthyofauna (Ferreira and Petrere Jr 2009). Furthermore, most studies on fish community structure along coastal streams (Mazzoni and Lobón-Cerviá 2000, Ferreira and Petrere Jr 2009, Gonçalves and Braga 2012) have not considered the local fish-habitat associations and their influences on fish patterns in river segments. In this sense, knowledge about the fish ecology of coastal streams is still insufficient to infer accurate structuring patterns and processes, particularly at multiple scales, since fish composition and diversity are ultimately determined at both fine and coarse spatial scales (Cheek et al. 2016). Thereby, we expect that both environmental characteristics along the longitudinal gradient and local habitats induce changes on the ichthyofauna structure in a preserved Brazilian coastal Atlantic stream. We addressed the following questions: (1) Does fish structure, in density and composition, differ among headwater, middle and mouth reaches? (2) What are the relationships among local habitats and ichthyofauna along longitudinal gradient? (3) What are the patterns of fish distribution among riffle, pool, and run mesohabitats?

MATERIAL AND METHODS

The study was carried out in the Vermelho River, eastern Atlantic basin, Brazil (Fig. 1). The Vermelho River begins in the mountainous part of the Reserva Particular do Patrimônio Natural – RPPN Morro da Mina, between the coastal municipalities of Antonina and Morretes, state of Paraná, and flows into the Xaxim River, which connects the river basin network to the estuarine complex of the Paranaguá Bay. The confluence of the Vermelho and Xaxim rivers is located approximately 15 km from Antonina and 40 km from Paranaguá in a preserved area of the Serra do Mar mountains. According to the Strahler classification (Allan 1995), the Vermelho River is a first to third-order stream along its course and it is a typical stream found in the coastal Atlantic forest of southern-southeast Brazil (Oyakawa et al. 2006). The Vermelho River is about 3 km long and between 2 and 5 m wide, with a steep slope at its upper course, riffles alternating with runs and shallow pools at the intermediate course and lentic habitats in the lowland reaches. As defined by Rincón (1999), riffles are reaches of great slope with fast running waters on a rocky bed; runs are areas with relatively slow running waters, with smaller slope and no turbulence, and are usually deeper than riffles; and pools are deeper than riffles and runs, with slow flow and sandy substrate. According to the Koeppen classification (Maack 1981), the regional climate is Af (tropical rainforest, fully humid), a transitional tropical climate, with annual rainfall between 2,500 and 3,000 mm



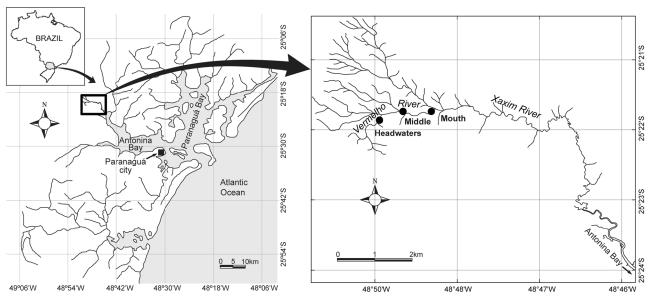


Figure 1. Location of the Vermelho River, eastern Atlantic basin, Antonina, Paraná state, Brazil. Black circles indicate the sampling reaches; arrow indicates the flow direction. Source: PEREIRA Jaime Luiz Lopes, 2015.

and mean temperature in the coldest month above 18°C. The vegetal formations of the RPPN are mangrove and 'restinga' (an herbaceous/shrubby community of coastal dunes) in lowlands and submontane Dense Ombrophilous Forest in highland areas where Vermelho River is localized (Velloso et al. 1991).

Samples of fish and abiotic variables were all taken in third-order stream reaches of headwater, middle and mouth of Vermelho River with approximately 50 m long each (Fig. 1). The headwater reach in the upper course (25°21′54.12″S, 48°49′55.56″W, 160 m a.s.l., 47.7 m long) was characterized as a cascaded channel, with the predominance of riffles, rocky bottom and a dense riparian vegetation canopy. The middle reach at the intermediate gradient (25°21′45.12″S, 48°49′35.58″W, 80 m a.s.l., 50.2 m long) had moderate water velocity riffles and areas with runs and pools, with the predominance of sandy/rocky substrate and dense/moderate vegetation canopy. On the other hand, the mouth reach at the lower course (25°21′42.10″S, 48°49′11.08″W, 55 m a.s.l., 48 m long) was characterized by slow-moving waters, with a predominance of sandy/clayey bottom and moderate vegetation canopy (Suppl. material 1).

Each reach was subdivided into different sampling sites according to their hydrological and structural characteristics. These sampling sites were provisionally classified in mesohabitats along the headwater-mouth direction in the following order: pool, riffles, riffles and run (headwater reach); pool, run, riffles, run, riffles and pool (middle reach) and pool, riffles and run (mouth reach). However, the sampling sites classification in mesohabitats was only validated in middle reach (Fig. 2), because it presented *a posteriori* significant differences in hydrological and environmental characteristics. Environmental differences

among mesohabitats were tested with a discriminant analysis using the matrix of abiotic variables transformed as log (x+1). The Wilks' lambda statistic was used to evaluate the efficiency of environmental variables in discriminating the mesohabitats *a priori*. Values range from 1 (no discriminant power) to 0 (perfect discriminant power) (Manly 2008). The discriminant analysis assumptions were checked by the Shapiro-Wilk test for normality and Levene's test for homogeneity of variances among the groups.

Fish were collected by electrofishing using a TOYAMA® power generator (127 volts AC and 5 A) from May 2009 to February 2010. As fish species are able to move among habitats and migrate short distances seasonally (Esteves and Aranha 1999), we considered the samplings as a snapshot study (Gotelli and Ellison 2004) of existing ichthyofauna. We blocked each sampling site with 3-mm mesh seines to reduce the spatial dependence of data and prevent fish from escaping (Fig. 2), and samples were taken every three months, in order to reduce the temporal dependence of data. We performed three successive passes of electrofishing in the upstream direction, and samples included all fish captured by dip nets as well as those retained in block seines. Fish were euthanized with 10% benzocaine and after fixed in 10% formaldehyde. In the laboratory, specimens were identified and counted. Fish were collected with authorization of the Instituto Chico Mendes de Conservação da Biodiversidade (ICMBio), license number 16475-1, and the procedures for manipulation of biological material are approved by Ethics Committee on Animal Use of the Universidade Estadual de Maringá (ECAU) in accordance with protocols of ethical and methodological aspects, for the use of fish. Due to possible character introgression in



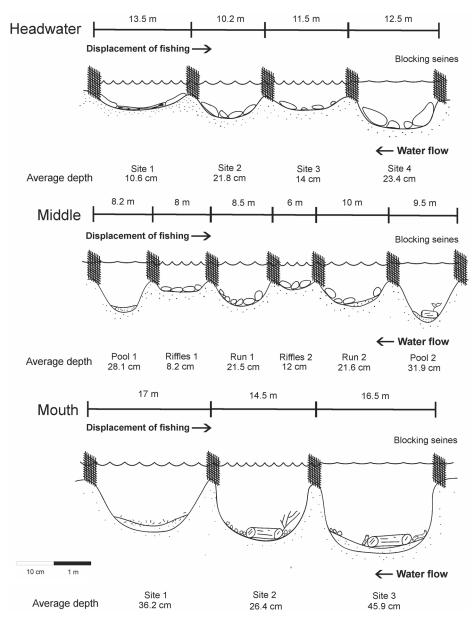


Figure 2. Fish sampling procedure. Length (above) and average depth (below) of each blocked (with seines) sampling site are showed. Middle sites were clustered into mesohabitats (see text). Displacement of fishing indicates the upstream direction of passage of electrofishing dip nets with electrodes.

the area, identification of the species belonging to *Characidium* is difficult (Paulo A. Buckup, pers. comm.), we were unable to identify two individuals of this genus with confidence, and they have been tentatively identified as *Characidium lanei* Travassos, 1967 based on photographs. Voucher specimens of 25 out of 27 species (Table 3), were deposited in the Museu de Peixes of Núcleo de Pesquisas em Limnologia, Ictiologia e Aquicultura

(Nupélia) of the Universidade Estadual de Maringá (NUP9465, NUP9524-NUP9529, NUP9541, NUP9567-NUP9572, NUP9576-NUP9595, NUP9742 and NUP19193-NUP19198), available on the SpeciesLink website (http://www.splink.org.br). For *Gymnotus pantherinus* (Steindachner, 1908) and *Hisonotus leucofrenatus* (Miranda Ribeiro, 1908) we uploaded photographs on the Fish-Base website (http://www.fishbase.org).



Water temperature (°C), dissolved oxygen (mg.l-1), pH and electrical conductivity (µS.cm⁻¹) were measured at each sampling site where fish were captured and determined as the arithmetic mean of four observations during the year. For hydraulic variables, the total length of each sampling site was measured and then three equidistant transversal sections along the site were marked. Channel width (m) was measured along each of these transversal sections, being determined as the arithmetic mean of three observations. Mean depth (cm) was estimated through five equidistant measurements along each transversal section and then determined as the arithmetic mean of 15 observations for each site. Similarly, water velocity (m.s-1), measured with a General Oceanics® mechanical flowmeter was determined from the arithmetic mean of three observations taken at the center of each transversal section at each site. The substrate bottom composition was categorized into different size classes and quantified by visual inspection. A total percentage of 100% was established for the sum of all substrate size categories at each sampling site. Size classes were based on Gordon et al. (1992): boulders (>80 mm diameter), cobbles (50-80 mm), pebbles (25-50 mm), gravel (5–25 mm), sand (<5 mm), and sand/clay (mixture of sand with silt). Organic structures, including leaf litter and tree trunks/ branches were also quantified.

We applied a principal component analysis (PCA) to the Pearson correlation matrix of environmental variables to describe the patterns of environmental variability among headwater, middle and mouth reaches. The variables were log (x+1) transformed in order to linearize the correlations. The Broken-Stick model was used for retention and interpretation of the axes, in which only axes with eigenvalues greater than those randomly generated are interpreted (Jackson 1993). Species richness, Shannon diversity and Pielou's evenness (Magurran 1988), calculated from species abundance matrix, were used as ichthyofauna descriptors. Since species richness could increase in direct proportion to the number of individuals sampled, we used the rarefaction method, with an algorithm from Krebs (1989), to compare headwater, middle and mouth reaches. Beta diversity (β_2) , calculated to evaluate the proportional addition and replacement of species along the longitudinal gradient, was obtained using Harrison index (Harrison et al. 1992). Beta diversity (β_2) measures the amount by which the regional diversity exceeds the maximum diversity obtained locally and reaches zero if adjacent sampling sites share all species, while highest β , values indicate an increase in the degree of species replacement (Petry and Schulz 2006).

To assess the volume of each sampling site, we used the following formula: Volume_{site} = Length_{site} * Mean depth_{site} * Mean channel width_{site}. Furthermore, in order to convert fish abundance data into density (i.e., individuals.m⁻³), we divided the number of individuals of each species by the respective sampling site volume. From these data, longitudinal and local patterns of ichthyofauna were described by a detrended correspondence analysis (DCA), combining the densities of each

sampling site among the stream reaches. Fish densities were log (x+1) transformed to linearize the relationships. In this analysis, only axes with eigenvalues > 0.20 were retained and interpreted (Manly 2008). DCA has been used as an alternative ordination method to CA (correspondence analysis), which eliminates the arch effect (Hill and Gauch 1980) and presumably shows more accuracy in the relationships of interest (Gotelli and Ellison 2004). Moreover, DCA preserves the distances among sites and among species observations in the multivariate space after the dimensional reduction and, therefore, allow realistic a posteriori tests of axis scores. Scores of the first significant DCA axis were subjected to a non-parametric Kruskal-Wallis analysis of variance to check for possible significant effects of the reach on ichthyofauna structure. Subsequently, a multiple comparison test was applied to determine which averages, and thus which reaches, differed from each other. Similarly, a DCA was employed to verify fish distribution patterns among mesohabitats of middle reach. Multivariate analyses, calculation of diversity indices and species rarefaction were performed in PAST 2.01 software. Drawing of scatter and line plots, discriminant and Kruskal-Wallis analyses and the tests of multiple comparisons were performed in Statistica 7.0.

RESULTS

Environmental variables

Most of the variations in environmental parameters were related to water velocity, which decreased along the stream, and to the average depth and channel width, which increased (Table 1). Boulders were the main substrate at headwater sites, while in the middle reach, heterogeneous substrate with cobbles, pebbles and gravel predominated in different mesohabitats, and at mouth reach, fine granulometric substrate with trunks and leaf litter predominated. Temperature, dissolved oxygen, pH and conductivity had low variation in headwater-mouth direction.

The first PCA axis retained for interpretation (eigenvalue = 5.2) explained 35% of total variance and primarily distinguished the reaches (Fig. 3). With more negative scores, all sites from the headwater reach and riffles from the middle reach showed higher current velocity and rocky bottoms, while at the positive end, habitats from the mouth reach were deeper and had wider channels, with sand and sand/clay substrates, and also higher quantity of trunks/branches and leaf litter. Runs and pools from middle reach were located at the intermediate environmental gradient of PCA.

Mesohabitats determined a priori

The discriminant analysis distinguished the groups of riffle, pool and run mesohabitats predefined according to hydraulics and structural characteristics from middle reach (Wilks' lambda = 0.0017, F = 10.96, p < 0.001). Average depth and cobble substrate percentage were the variables that significantly influenced the separation of groups (Table 2). Riffles had



Table 1. Mean ± standard deviation of abiotic variables and main substrate bottom for sampling sites in headwater, middle and mouth reaches of the Vermelho River, Paraná state, Brazil. Temp. = temperature, DO = dissolved oxygen, Cond. = conductivity. Deviations represent environmental variation between May 2009 and February 2010. * Sites at middle reach were classified as riffles, runs and pools.

	Sites	Water vel. (m.s ⁻¹)	Depth (cm)	Channel width (m)	Temp. (°C)	DO (mg.l ⁻¹)	рН	Cond. (µS/cm ⁻¹)	Boulders (%)
Middle* Headwater	Site 1	7.0 ± 2.1	10.6 ± 2.9	3.1 ± 0.9	20.0 ± 2.4	7.7 ± 1.2	7.0 ± 0.4	45.1 ± 4.9	55 ± 8.7
	Site 2	4.4 ± 2.2	21.8 ± 4.8	4.1 ± 1.0	20.1 ± 2.5	7.8 ± 0.9	7.2 ± 0.4	41.6 ± 2.6	$36,3 \pm 6.5$
	Site 3	6.6 ± 1.8	14 ± 5.9	2.9 ± 1.0	20.1 ± 2.3	7.4 ± 1.4	7.3 ± 0.3	42.1 ± 2.3	$37,5 \pm 8.3$
	Site 4	5.5 ± 1.8	23.4 ± 8.3	2.6 ± 1.0	20.1 ± 2.3	7.9 ± 1.5	7.4 ± 0.3	41.8 ± 1.2	75 ± 5
	Pool1	1.7 ± 1.8	28.1 ± 5.1	2.7 ± 0.4	20.6 ± 2.4	8.0 ± 0.7	6.5 ± 0.5	40.7 ± 1.6	1.3 ± 2.2
	Riffles1	6.4 ± 1.8	8.2 ± 2.2	2.6 ± 0.3	20.5 ± 2.3	7.8 ± 0.5	7.0 ± 0.6	39.3 ± 4.2	1.3 ± 2.2
	Runs1	2.0 ± 1.5	21.5 ± 8.2	2.8 ± 0.4	20.6 ± 2.2	7.1 ± 0.4	7.1 ± 0.4	39.4 ± 4.0	0 ± 0
	Riffles2	6.5 ± 3.1	12.0 ± 5.2	2.0 ± 0.8	20.7 ± 2.4	7.5 ± 0.7	7.2 ± 0.3	39.4 ± 4.8	0 ± 0
	Runs2	1.2 ± 1.5	21.6 ± 4.7	3.7 ± 0.2	20.6 ± 2.3	7.3 ± 0.7	7.2 ± 0.4	39.3 ± 4.3	0 ± 0
Mouth	Pool2	0.7 ± 0.6	31.9 ± 6.4	3.5 ± 0.3	20.7 ± 2.3	7.6 ± 1.0	7.2 ± 0.4	39.3 ± 4.4	0 ± 0
	Site 1	2.1 ± 1.0	35.2 ± 20.0	3.9 ± 0.8	20.4 ± 3.3	7.5 ± 1.1	6.1 ± 0.2	38.8 ± 4.4	0 ± 0
	Site 2	4.2 ± 1.4	26.4 ± 22.7	4.2 ± 1.0	20.5 ± 3.3	7.3 ± 1.0	6.7 ± 0.6	36.6 ± 5.2	0 ± 0
	Site 3	1.4 ± 1.5	45.9 ± 29.7	5.4 ± 1.1	20.5 ± 3.3	7.6 ± 0.9	6.7 ± 0.5	39.9 ± 1.7	0 ± 0

	Sites	Boulders (%)	Cobbles (%)	Pebbles (%)	Gravel (%)	Sand (%)	Sand/Clay (%)	Trunk/branch (%)	Leaf litter (%)
Headwater	Site 1	55 ± 8.7	30 ± 0	8.8 ± 4.1	6.3 ± 5.4	0 ± 0	0 ± 0	0 ± 0	0 ± 0
	Site 2	$36,3 \pm 6.5$	35 ± 6.1	15 ± 5	13.8 ± 8.1	0 ± 0	0 ± 0	0 ± 0	0 ± 0
	Site 3	$37,5 \pm 8.3$	33.8 ± 10.8	6.3 ± 2.1	22.5 ± 14.4	0 ± 0	0 ± 0	0 ± 0	0 ± 0
	Site 4	75 ± 5	17.5 ± 4.3	1.3 ± 2.1	6.3 ± 5.4	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Middle*	Pool1	1.3 ± 2.2	3.8 ± 2.2	3.8 ± 2.2	57.5 ± 17.9	2.5 ± 4.3	23.8 ± 12.9	0 ± 0	7.5 ± 13
	Riffles1	1.3 ± 2.2	26.3 ± 13.9	67.5 ± 10.9	2.5 ± 2.5	0 ± 0	0 ± 0	0 ± 0	2.5 ± 2.5
	Runs1	0 ± 0	28.8 ± 11.3	35 ± 5	23.8 ± 9.6	0 ± 0	2.5 ± 4.3	5 ± 3.5	5 ± 3.5
	Riffles2	0 ± 0	72.5 ± 4.3	18.8 ± 4.1	1.3 ± 2.2	0 ± 0	0 ± 0	0 ± 0	7.5 ± 5.6
	Runs2	0 ± 0	27.5 ± 8.3	12.5 ± 4.3	46.3 ± 4.1	1.3 ± 2.2	0 ± 0	5 ± 6.1	7.5 ± 5.6
Mouth	Pool2	0 ± 0	16.3 ± 8.2	8.8 ± 2.2	57.5 ± 4.3	0 ± 0	0 ± 0	6.3 ± 4.1	11.3 ± 7.4
	Site 1	0 ± 0	0 ± 0	0 ± 0	2.5 ± 4.3	28.8 ± 18.2	31.3 ± 20.1	17.5 ± 2.5	20 ± 6.1
	Site 2	0 ± 0	0 ± 0	15 ± 15	35 ± 8.7	12.5 ± 8.3	2.5 ± 4.3	28.5 ± 7.4	6.3 ± 4.1
Σ	Site 3	0 ± 0	0 ± 0	13.8 ± 6.5	5 ± 0	1.3 ± 2.1	26.3 ± 6.5	25 ± 3.5	28.8 ± 5.4

higher percentage of cobbles, while pools were deeper and had the bottom covered mostly by leaf litter, and runs presented intermediate abiotic conditions between riffles and pool.

Richness and diversity

A total of 1,495 fish individuals were captured, belonging to 27 species, 12 families and six orders. Siluriformes contributed with 14 (51.9%), Characiformes with seven (25.9%), and the other orders had only one or two species (Table 3).

Fish species richness increased along the stream, with 13, 18 and 22 species in headwater, middle and mouth reaches, respectively (Table 3). Rarefaction analysis showed that the expected richness was also higher at the mouth reach, although an asymptote was not attained in its rarefaction curve (Fig. 4). Species composition also differed among reaches: from 27

species, only nine, particularly the characids, *Deuterodon langei* Travassos, 1957, *Mimagoniates microlepis* (Steindachner, 1876), *Characidium lanei* Travassos, 1967 and *Characidium pterostictum* Gomes, 1947 occurred throughout the stream.

In the headwater reach, *C. lanei*, *D. langei* and *Rhamdioglanis frenatus* Ihering, 1907 were the most abundant, while *Ituglanis proops* (Miranda Ribeiro, 1908) was rare and exclusive. In the middle reach, in addition to *C. lanei* and *D. langei*, *M. microlepis* also predominated. *Rivulus luelingi* Seegers, 1984, with only one individual, was exclusive. At the mouth reach, abundances were higher for *M. microlepis*, *Pseudotothyris obtusa* (Miranda Ribeiro, 1911) and *C. lanei*, while *Astyanax* aff. *ribeirae* Eigenmann, 1911, *Hoplias malabaricus* (Bloch, 1794) and *Gymnotus* spp. had only one individual each (Table 3).

Shannon diversity and evenness had their highest absolute values in the middle reach (Table 3). Beta diversity



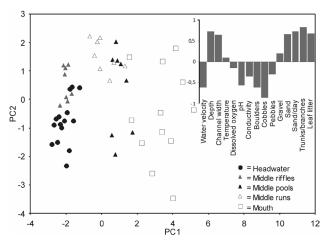


Figure 3. Scatter plot of sampling sites and mesohabitats in the Vermelho River, Paraná state, Brazil, along the environmental gradient produced by the first two PCA axes (PC1 and PC2). Inner figure shows Pearson correlations of variables with PC1.

Table 2. Results of discriminant analysis with environmental variables that contributed to separation of groups of mesohabitats (i.e., riffles, pools and runs) in the middle reach of the Vermelho River. Bold values are significant (p<0.05).

	Axis 1	Axis 2	Wilks' Lambda	Partial Lambda	F	р
Depth	-2.187	-0.059	0.0052	0.321	7.39	0.019
Water velocity	-0.369	-0.240	0.0017	0.983	0.06	0.942
Cobbles	1.647	2.591	0.0053	0.314	7.65	0.017
Pebbles	0.417	1.142	0.0028	0.593	2.40	0.161
Leaf litter	-0.144	-2.953	0.0032	0.526	3.15	0.106
Sand	1.604	0.828	0.0035	0.474	3.88	0.074
Gravel	-0.220	1.042	0.0028	0.597	2.36	0.165
Sand/clay	-0.384	1.619	0.0023	0.730	1.29	0.333
рН	1.577	1.277	0.0034	0.488	3.67	0.081
Dissolved oxygen	0.119	-1.352	0.0025	0.673	1.70	0.251
Trunks/branches	-1.192	-0.646	0.0028	0.598	2.36	0.165
Temperature	-1.506	0.622	0.0027	0.622	2.13	0.190
Conductivity	-2.440	2.088	0.0028	0.595	2.38	0.163
Boulders	0.725	-0.355	0.0022	0.761	1.10	0.385
Channel width	-1.410	0.520	0.0022	0.772	1.03	0.405
Eigenvalues	89.2	5.6				
%	94	100				

indicated greater addition than replacement of fish species along the stream. The index values were low when comparing headwater with middle (β_2 = 0.23) and middle with mouth reaches (β_2 = 0.30). Poeciliid, gymnotid and callichthyid species were the main additions, while *Ancistrus multispinnis* (Regan, 1912) and *R. frenatus* were the main replacements downstream (Table 3).

Table 3. Number of individuals per fish species collected and ichthyofauna descriptors in headwater, middle and mouth reaches in the Vermelho River, Paraná state, Brazil. In bold, predominant species.

Species					
Characidae Deuterodon langei Travassos, 1957 NUP9541 105 155 33 Hollandichthys multifasciatus (Eigenmann & Norris, 1900) Mimagoniates microlepis (Steindachner, 1876) Astyanax aff. ribeirae Eigenmann, 1911 NUP9526 10 73 202 10 Characidium lanei Travassos, 1967 NUP9526 117 111 38 Characidium lanei Travassos, 1967 NUP9525 117 111 38 Characidium petrostictum Gomes, 1947 NUP9588 29 22 30 Erythrynidae Hoplias malabaricus (Bloch, 1794) NUP19193 -	Species		Headwater	Middle	Mouth
Deuterodon langei Travassos, 1957 Hollandichthys multifiasciatus (Eigenmann & NuP9529 4 36 13 36 13 NuP9529 4 36 13 36 13 NuP9529 4 36 13 36 13 36 33 34 36 33 36 33 36 33 36 33 36 33 36 33 34 36 33 34 35 34 35 34 35 34 35 34 35 34 35 34 35 34 35 36 36 36 36 36 36 36	Characiformes	зресппенз			
Hollandichthys multifasciatus (Eigenmann & NUP9529	Characidae				
& Norris, 1900) NUP9529 4 36 13 Mimagoniates microlepis (Steindachner, 1876) NUP9526 10 73 202 Astyanax aff. ribeirae Eigenmann, 1911 NUP9742 - 2 1 Crenuchidae Characidium larei Travassos, 1967 NUP9525 117 111 38 Characidium pterostictum Gomes, 1947 NUP9588 29 22 30 Erythrynidae Hoplias malabaricus (Bloch, 1794) NUP19193 - - 1 Eyprinodontiformes Poeciliidae Phaloloceros harpagos Lucinda, 2008 NUP9527 - 27 6 Rivulidae Rivulis luelingi Seegers, 1984 NUP19198 - 1 - Gymnotiformes Gymnotitos paritherinus (Steindachner, 1908) NUP19194 - 1 - Ferciformes Gobiidae Awaous tajasica (Lichtenstein, 1822) NUP19195 - 4 1 Siluriformes Callichthydae Scleromystax barbatus (Quoy & Gaimard, 1822) NUP9570 - 16 21 Schizol	Deuterodon langei Travassos, 1957	NUP9541	105	155	33
& Norris, 1900) Mimagoniates microlepis (Steindachner, 1876) NUP9526 10 73 202 Astyanax aff. ribeirae Eigenmann, 1911 NUP9742 - 2 1 Crenuchidae Characidium lanei Travassos, 1967 NUP9525 117 111 38 Characidium pterostictum Gomes, 1947 NUP9588 29 22 30 Erythrynidae Hoplics malabaricus (Bloch, 1794) NUP19193 - - 1 Cyprinodontiformes Poecilidae Phalloceros harpagos Lucinda, 2008 NUP9527 - 27 6 Rivulidae Rivulus luelingi Seegers, 1984 NUP19198 - 1 - Gymnotidae Gymnotius pantherinus (Steindachner, 1908) - - 4 1 Gymnotus pantherinus (Steindachner, 1908) - - 4 1 Gymnotus carapo Linnaeus, 1758 NUP19194 - - 1 Perciformes Callichthydae - - 4 1 Golidae Ancistrus multispinnis (Regan, 1912) NUP9570 <t< td=""><td>Hollandichthys multifasciatus (Eigenmann</td><td>NII IDQ5 2Q</td><td>4</td><td>36</td><td>12</td></t<>	Hollandichthys multifasciatus (Eigenmann	NII IDQ5 2Q	4	36	12
1876 NUP9526 10 73 202	& Norris, 1900)	NUP9329	4	30	13
1876 Astyonax aff. ribeirae Eigenmann, 1911 NUP9742 - 2 1	Mimagoniates microlepis (Steindachner,	NII IP9526	10	73	202
Crenuchidae Characidium lanei Travassos, 1967 Characidium pterostictum Gomes, 1947 Erythrynidae Hoplias malabaricus (Bloch, 1794) NUP19193 1 Cyprinodontiformes Poeciliidae Phalloceros harpagos Lucinda, 2008 Rivulidae Rivulus luelingi Seegers, 1984 NUP19198 - 1 - Cymnotiformes Gymnotidae Gymnotidae Gymnotidae Gymnotidae Gymnotidae Gymnotidae Gymnotidae Gymnotidae Gymnotus pantherinus (Steindachner, 1908) Gymnotus carapo Linnaeus, 1758 NUP19194 1 Perciformes Gobilidae Awaous tajasica (Lichtenstein, 1822) NUP19195 4 Siluriformes Callichthydae Scleromystax barbatus (Quoy & Gaimard, 1824) Loricariidae Ancistrus multispinnis (Regan, 1912) Schizolecis guntheri (Miranda Ribeiro, 1918) Schizolecis gs. 2 NUP10903 Kronichthys cf. lacerta (Nichols, 1919) Pseudotothyris obtusa (Miranda Ribeiro, 1908) Kronichthys cf. lacerta (Nichols, 1919) Rineloricaria sp. Heptapteridae Acentronichthys leptos Eigenmann & Eigenmann, 1889 Rhamdioglanis frenatus Ihering, 1907 NUP13613 NUP19524 A 35 Alizoleci squelen (Quoy & Gaimard, 1925 Rhamdia quelen (Quoy & Gaimard, 1925 Rhamdia quelen (Quoy & Gaimard, 1824) Trichomycteridae Ituglanis proops (Miranda Ribeiro, 1908) NUP1997 Synbranchiformes	1876)	14017520	10	,,	202
Characidium lanei Travassos, 1967 NUP9525 117 111 38 Characidium pterostictum Gomes, 1947 NUP9588 29 22 30 Erythrynidae Hoplias malabaricus (Bloch, 1794) NUP19193 - - 1 Cyprinodontiformes Poeciliidae Phalloceros harpagos Lucinda, 2008 NUP9527 - 27 6 Rivullidae Rivullidae NUP19198 - 1 - Gymnotiformes Gymnotids Luclingi Seegers, 1984 NUP19198 - 1 - Gymnotids and Silvingi Seegers, 1984 NUP19198 - 1 - Gymnotids and Silvinginis Seegers, 1984 NUP19198 - 1 - Gymnotids and Silvinginis Seegers, 1984 NUP19198 - 1 - Gymnotids carapo Linnaeus, 1758 NUP19194 - - 1 Perciformes Cobicidae NUP19195 - - 4 1 Siluriformes Callichthydae Scaleromystax barbatus (Quoy & Gaimard, 1822) NUP19195 -		NUP9742	-	2	1
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Perciformes Gobiidae **Awaous tajasica** (Lichtenstein, 1822)** NUP19195** 4 **Siluriformes Callichthydae **Scleromystax barbatus** (Quoy & Gaimard, 1824)** Loricariidae **Ancistrus multispinnis** (Regan, 1912)** NUP13626** 12 9 **Schizolecis guntheri** (Miranda Ribeiro, 1918)** NUP19465** 7 70 5 **Schizolecis sp. 1 NUP10903** 32 **Schizolecis sp. 2 NUP10904** 8 41 **Hisonotus leucofrenatus** (Miranda Ribeiro, 1908)** NUP10900** 3 **Fronichthys cf. lacerta** (Nichols, 1919)** NUP10900** 3 **Heptapteridae** NuP10902** 59 **Heptapteridae** NuP10908** - 12 12 **Heptapteridae** NuP10908** - 12 12 **Heptapteridae** NuP10908** - 12 12 **Heptapteridae** NuP10909**	•				_
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Schizolecis guntheri (Miranda Ribeiro, 1918) NUP9465 7 70 5 Schizolecis sp. 1 NUP10903 - - 32 Schizolecis sp. 2 NUP10904 8 41 - Hisonotus leucofrenatus (Miranda Ribeiro, 1908) - - - 3 Kronichthys cf. lacerta (Nichols, 1919) NUP10900 - - 11 Pseudotothyris obtusa (Miranda Ribeiro, 1911) NUP10902 - - 59 1911) Rineloricaria sp. NUP10898 - 12 12 Heptapteridae Acentronichthys leptos Eigenmann & Eigenmann, 1889 NUP9592 19 17 2 Rhamdioglanis frenatus Ihering, 1907 NUP13614 32 9 - Pimelodella pappenheimi Ahl, 1925 NUP13613 - - 6 Rhamdia quelen (Quoy & Gaimard, 1824) NUP9524 4 35 4 Trichomycteridae Ituglanis proops (Miranda Ribeiro, 1908) NUP19197 2 - - - Synbranchiformes	Loricariidae				
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1918 Schizolecis sp. 1	Schizolecis guntheri (Miranda Ribeiro,	NUP9465	7	70	5
Schizolecis sp. 2 NUP10904 8 41 - Hisonotus leucofrenatus (Miranda Ribeiro, 1908) - - - 3 Kronichthys cf. lacerta (Nichols, 1919) NUP10900 - - 11 Pseudotothyris obtusa (Miranda Ribeiro, 1911) NUP10902 - - 59 1911) NUP10898 - 12 12 Heptapteridae - - 12 12 Heptapteridae - NUP9592 19 17 2 Eigenmann, 1889 Rhamdioglanis frenatus Ihering, 1907 NUP13614 32 9 - Pimelodella pappenheimi Ahl, 1925 NUP13613 - - 6 Rhamdia quelen (Quoy & Gaimard, 1824) NUP9524 4 35 4 Trichomycteridae Ituglanis proops (Miranda Ribeiro, 1908) NUP19197 2 - - Synbranchiformes	,		•		
Hisonotus leucofrenatus (Miranda Ribeiro, 1908) - - - 3 Kronichthys cf. lacerta (Nichols, 1919) NUP10900 - - 11 Pseudotothyris obtusa (Miranda Ribeiro, 1911) NUP10902 - - 59 Heptapterida petro Eigenmann & Eigenmann, 1889 NUP10898 - 12 12 Heptapteridae NUP9592 19 17 2 2 Rhamdioglanis frenatus Ihering, 1907 NUP13614 32 9 - Pimelodella pappenheimi Ahl, 1925 NUP13613 - - 6 Rhamdia quelen (Quoy & Gaimard, 1824) NUP9524 4 35 4 Trichomycteridae Ituglanis proops (Miranda Ribeiro, 1908) NUP19197 2 - - Synbranchiformes	•		-	-	32
1908) Kronichthys cf. lacerta (Nichols, 1919) NUP10900 11 Pseudotothyris obtusa (Miranda Ribeiro, 1911) Rineloricaria sp. NUP10898 - 12 12 Heptapteridae Acentronichthys leptos Eigenmann & Eigenmann, 1889 Rhamdioglanis frenatus Ihering, 1907 NUP13614 32 9 - Pimelodella pappenheimi Ahl, 1925 NUP13613 6 Rhamdia quelen (Quoy & Gaimard, 1824) Trichomycteridae Ituglanis proops (Miranda Ribeiro, 1908) NUP19197 2 - Synbranchiformes	·	NUP10904	8	41	-
Kronichthys cf. lacerta (Nichols, 1919) NUP10900 - - 11 Pseudotothyris obtusa (Miranda Ribeiro, 1911) NUP10902 - - 59 Heptapteridae NUP10898 - 12 12 Heptapteridae NUP10898 - 12 12 Acentronichthys leptos Eigenmann & Eigenmann, 1889 NUP9592 19 17 2 Eigenmann, 1889 NUP13614 32 9 - Pimelodella pappenheimi Ahl, 1925 NUP13613 - - 6 Rhamdia quelen (Quoy & Gaimard, 1824) NUP9524 4 35 4 Trichomycteridae Ituglanis proops (Miranda Ribeiro, 1908) NUP19197 2 - - Synbranchiformes		_	_	-	3
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1911) Rineloricaria sp. Heptapteridae Acentronichthys leptos Eigenmann & Eigenmann, 1889 Rhamdioglanis frenatus Ihering, 1907 Pimelodella pappenheimi Ahl, 1925 Rhamdia quelen (Quoy & Gaimard, 1824) Trichomycteridae Ituglanis proops (Miranda Ribeiro, 1908) NUP19197 A Company Company NUP19197 Rhamdia quelen (Quoy & Gaimard, NUP19197 NUP19197 NUP19197 NUP19197 Rhamdia Quelen (Quoy & Gaimard, NUP19197 NUP19197 NUP19197 NUP19197 NUP19197 NUP19197 NUP19197 NUP19197		NUP10900	-	-	- 11
Rineloricaria sp. NUP10898 – 12 12 Heptapteridae Acentronichthys leptos Eigenmann & Eigenmann, 1889 NUP9592 19 17 2 Eigenmann, 1889 NUP13614 32 9 – Pimelodella pappenheimi Ahl, 1925 NUP13613 – – 6 Rhamdia quelen (Quoy & Gaimard, 1824) NUP9524 4 35 4 Trichomycteridae Ituglanis proops (Miranda Ribeiro, 1908) NUP19197 2 – – Synbranchiformes		NUP10902	-	-	59
Heptapteridae Acentronichthys leptos Eigenmann & NUP9592 19 17 2 Eigenmann, 1889 Rhamdioglanis frenatus Ihering, 1907 NUP13614 32 9 - Pimelodella pappenheimi Ahl, 1925 NUP13613 6 Rhamdia quelen (Quoy & Gaimard, 1824) Trichomycteridae Ituglanis proops (Miranda Ribeiro, 1908) NUP19197 2 - Synbranchiformes	,	NII IP10898	_	12	12
Acentronichthys leptos Eigenmann & NUP9592 19 17 2 Eigenmann, 1889 Rhamdioglanis frenatus Ihering, 1907 NUP13614 32 9 - Pimelodella pappenheimi Ahl, 1925 NUP13613 6 Rhamdia quelen (Quoy & Gaimard, 1824) Trichomycteridae Ituglanis proops (Miranda Ribeiro, 1908) NUP19197 2 Synbranchiformes	•	1101 10070		12	12
Eigenmann, 1889 Rhamdioglanis frenatus Ihering, 1907 NUP13614 32 9 - Pimelodella pappenheimi Ahl, 1925 NUP13613 6 Rhamdia quelen (Quoy & Gaimard, 1824) Trichomycteridae Ituglanis proops (Miranda Ribeiro, 1908) NUP19197 2 Synbranchiformes					
Pimelodella pappenheimi Ahl, 1925 NUP13613 6 Rhamdia quelen (Quoy & Gaimard, 1824) Trichomycteridae Ituglanis proops (Miranda Ribeiro, 1908) NUP19197 2 Synbranchiformes	Eigenmann, 1889	NUP9592	19	17	2
Rhamdia quelen (Quoy & Gaimard, NUP9524 4 35 4 1824) Trichomycteridae Ituglanis proops (Miranda Ribeiro, 1908) NUP19197 2 Synbranchiformes		NUP13614	32	9	-
1824) Trichomycteridae Ituglanis proops (Miranda Ribeiro, 1908) NUP19197 2 – – Synbranchiformes	Pimelodella pappenheimi Ahl, 1925	NUP13613	_	-	6
1824) Trichomycteridae Ituglanis proops (Miranda Ribeiro, 1908) NUP19197 2 – – Synbranchiformes	Rhamdia quelen (Quoy & Gaimard,	NII IDQ524	4	3.5	4
Ituglanis proops (Miranda Ribeiro, 1908) NUP19197 2 – – Synbranchiformes	1824)	1101 3324	4	33	4
Synbranchiformes	Trichomycteridae				
		NUP19197	2	-	-
Synbranchidae					
Combranchus marmoratus Plach 1705 NUIDOSC7 4 5 13	-	NII IDOS CZ	4	F	12
Synbranchus marmoratus Bloch, 1795 NUP9567 4 5 12 Richness 13 18 22		/מכנייטויו			
Abundance 353 645 497					
Shannon diversity 1.87 2.35 2.17					
Evenness 0.73 0.81 0.70	-				

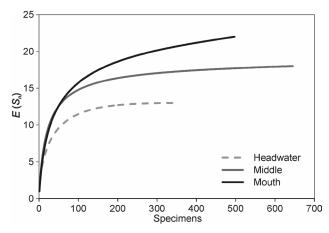


Figure 4. Species rarefaction curves for reaches in the Vermelho River, Paraná state, Brazil. $E(S_n)$ denotes the expected number of species according to abundance.

Longitudinal and local fish community structure

The first DCA axis retained for interpretation ($\lambda 1 = 0.50$) classified the ichthyofauna structure into headwater, middle and mouth reaches and explained 21.3% of total variance (Fig. 5). Kronichthys cf. lacerta (Nichols, 1919), Awaous tajasica (Lichtenstein, 1822) and Schizolecis sp. 1 were positively correlated with DCA1, while A. multispinnis, Acentronichthys leptos Eigenmann, 1889 and R. frenatus were negatively correlated. For DCA2, I. proops and Gymnotus carapo Linnaeus, 1758 were positively correlated, while Kronichthys cf. lacerta and R. luelingi were negatively correlated. For the DCA1 scores, there was a significant difference in fish structure only between the upper reaches (headwater, middle) and the mouth reach (Kruskal-Wallis test = 31.66, p < 0.05; multiple comparisons p < 0.05). Headwater and middle reaches did not differ because of the similarity in fish structures between middle riffles and headwater sites. These habitats shared many lotic species, such as A. multispinnis, A. leptos and Characidium spp. Middle pools showed a greater similarity in regard to fish structure with lentic sites of the mouth reach, sharing exclusive species such as Scleromystax barbatus (Quoy & Gaimard, 1824) and Phalloceros harpagos Lucinda, 2008 in addition to a great number of individuals of M. microlepis.

In the middle reach, fish richness and abundance were higher in pools and runs (Table 4). *Phalloceros harpagos, Gymnotus pantherinus* (Steindachner, 1908), *S. barbatus, Rhamdia quelen* (Quoy & Gaimard, 1824) and *Synbranchus marmoratus* Bloch, 1795 were exclusive to pools and runs; *Acentronichthys leptos* to riffles and runs; *Astyanax* aff. *ribeirae* to pools and *R. luelingi* to riffles. Other species occurred throughout all mesohabitats. However, we observed a clear pattern, where lentic habitat fishes (i.e. *D. langei, M. microlepis* and *Hollandichthys multifasciatus*

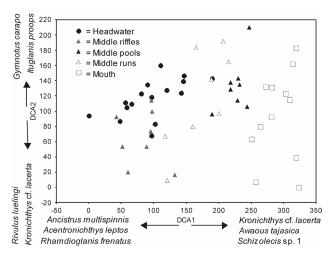


Figure 5. Ordination of the sampling sites through a detrended correspondence analysis (DCA) applied to the density matrix of fish species in the Vermelho River, Paraná state, Brazil.

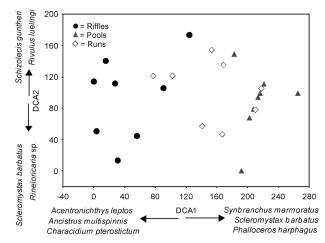


Figure 6. Ordination of sampling mesohabitats in the middle reach through a detrended correspondence analysis (DCA) applied to density matrix of fish species in the Vermelho River, Paraná state, Brazil.

(Eigenmann & Norris, 1900)), were more abundant in pools, while fast-current dwelling (i.e. *Characidium* spp., *A. leptos* and *A. multispinnis*), were more abundant in riffles. Runs tended to present intermediate abundance for most of the species. For mesohabitat fish structure, the first DCA axis retained ($\lambda 1 = 0.49$), explained 33% of total variance, segregating pools (positive scores) from riffles (less positive scores) (Fig. 6). *Synbranchus marmoratus*, *S. barbatus* and *P. harpagos* were positively correlated with DCA1, while *A. leptos*, *A. multispinnis* and *C. pterostictum* were negatively correlated.



Table 4. Number of individuals per fish species collected and richness in riffle, pool and run mesohabitats of middle reach in the Vermelho River, Paraná state, Brazil. In bold, predominant species.

Species	Riffles	Pools	Runs
Characiformes			
Deuterodon langei Travassos, 1957	7	103	45
Hollandichthys multifasciatus (Eigenmann & Norris, 1900)	1	27	8
Mimagoniates microlepis (Steindachner, 1876)	1	62	10
Astyanax aff. ribeirae Eigenmann, 1911	-	2	-
Characidium lanei Travassos, 1967	64	17	30
Characidium pterostictum Gomes, 1947	12	2	8
Cyprinodontiformes			
Phalloceros harpagos Lucinda, 2008	-	22	5
Rivulus luelingi Seegers, 1984	1	-	-
Gymnotiformes			
Gymnotus pantherinus (Steindachner, 1908)	-	1	3
Siluriformes			
Scleromystax barbatus (Quoy & Gaimard, 1824)	-	11	5
Ancistrus multispinnis (Regan, 1912)	5	1	3
Schizolecis guntheri (Miranda Ribeiro, 1918)	25	18	27
Schizolecis sp. 2	10	16	15
Rineloricaria sp.	1	5	6
Acentronichthys leptos Eigenmann & Eigenmann, 1889	13	-	4
Rhamdioglanis frenatus Ihering, 1907	3	3	3
Rhamdia quelen (Quoy & Gaimard, 1824)	_	30	5
Synbranchiformes			
Synbranchus marmoratus Bloch, 1795	-	4	1
Richness	12	16	16
Abundance	143	324	178

DISCUSSION

Based on the literature, between 20 and 50 fish species are estimated to occur in coastal Atlantic streams of Paraná state (Fogaça et al. 2003, Barreto and Aranha 2005, Abilhoa and Bastos 2009, Guimarães et al. 2010). Channels located on steep slopes of the Serra do Mar mountains and geographical isolation due the subdivision of eastern Atlantic basin in several sub basins, have conferred to fish smaller sizes and higher degree of endemism (Vari and Malabarba 1998, Menezes et al. 2007). Here, the Vermelho River presented similar fish richness (27 species) and composition similarities to 30 species recorded by Barreto and Aranha (2005) in a stream on the northern coast of Paraná, which is similar in length. As expected to fish communities in coastal Atlantic streams, we sampled fish with small-sized bodies and species such as *D. langei*, *M. microlepis* and *H. multifasciatus* which are not endemic but typically occur in coastal basins (Menezes et al. 2007).

We also registered three species that are probably new. *Rineloricaria* sp. corresponds to *Rineloricaria* sp. 1 from Abilhoa and Bastos (2009), with three dark-brown bands on dorsal region of the body between dorsal and caudal fins. *Schizolecis guntheri*

presents a large dark stripe along the sides of the body and a small light area within a blotch on the base of the caudal fin (Abilhoa and Bastos 2009). However, Schizolecis sp. 1 and Schizolecis sp. 2 differed because both do not present a continuous dark stripe. The first has a large blotch above the pectoral fin and another one in the base of the caudal fin, while the latter has similar large blotches, but with irregular dots and longitudinal small bars along the body. We also initially identified two specimens as a distinct species of Characidium. However, this genus includes probable cases of interspecific hybridization (Pansonato-Alves et al. 2014), generating individuals with intermediate morphologies. In the study region, C. lanei and C. pterostictum live in sympatry, which increases the probability of hybridization between this two species (Paulo A. Buckup, pers. comm.). After a review of photographs, the two specimens were tentatively identified as C. lanei because of the higher color similarity (Abilhoa and Bastos 2009). Reanalysis of the data, after this reidentification, presented slight, but not significant differences in outputs, which did not change the interpretation of ecological patterns.

The richness and species rarefaction increased in headwater-mouth direction, while Shannon diversity was higher in the middle reach. Similar results were reported by Casatti (2005), who argued that the increase in richness and in diversity is likely related to higher structural complexity downstream. The middle reach of the Vermelho River showed typical characteristics of a transitional environment between the headwater reach (rocky bottom and high water velocity) and mouth reach (sandy substrate and low water velocity). Thus, the sampling sites of middle reach, which were scattered at the intermediate abiotic gradient of the PCA, presented higher environmental variability, which provides a more diversified meso- and microhabitats, allowing a more diversified fish fauna to inhabiting this reach.

The Vermelho River had higher rates of species addition than replacement along its course, corroborating results of Petry and Schulz (2006), who reported higher addition over replacement in the Sinos River, southern Brazil. Higher additions occur in rivers with few abrupt geomorphological transitions, enabling a gradual adjustment in fish composition to different environmental conditions downstream (Rahel and Hubert 1991). For the Vermelho River, which shows a pronounced slope between headwater and mouth reaches, the most frequent species additions likely related to increased volume and availability of different habitats downstream and, also, to the ingression of estuarine species, such as A. tajasica, in the mouth reach. However, the replacements of A. multispinnis and R. frenatus, which did not occur in the mouth reach, are likely associated with the longitudinal decrease of water velocity and changes from a rocky substrate to sandy bottom downstream. Ancistrus multispinnis inhabits moderate to fast currents dwelling on boulders and cobbles (Barreto and Aranha 2005) and presents larger ventral sucker mouth than other loricariids, which allows it to adhere to the substratum (Breda et al. 2005). A congeneric of R. frenatus, Rhamdioglanis transfaciatus Miranda Ribeiro, 1908 with



similar elongated body shape, inhabits upper fast-water reaches (Guimarães et al. 2010), indicating the capacities of these species of swimming among boulders and cobbles.

The DCA and statistical tests showed that fish community structure differed only between upper and mouth reaches. Fast-water sites from headwater reach did not differ from middle riffles because these sites share similar composition and density of fish. On the other hand, the middle pool ichthyofauna was more similar to lentic habitats of the mouth reach. According to Frissell et al. (1986), due to the hierarchical nature of river systems, processes at large scales will ultimately influence fish structures at smaller spatial scales. However, the similarity in fish fauna between fast- and slow-moving water habitats in different reaches of the Vermelho River might reflect the low influence of large-scale processes, such as human activities, in its surrounds. In turn, different reaches could be offering similar conditions to the input of allochthonous matter and filtering mechanisms to reduce sedimentation along the entire stream, maintaining similarity in some of the fish-local habitats associations at a larger scale. Additionally, we observed higher environmental similarity, such as rocky substrate and fast waters between headwater and middle riffles habitats, which is an indicative of similar fish structures in function of their habitat (Taylor 2000, Langeani et al. 2005).

Similarly, Cheek et al. (2016) observed that landscape- and stream reach-scale variables had a low degree of explanation for the variation of the fish structure in a minimally disturbed stream when compared to other mesoscale variables. These findings open the discussion about the strength that finer spatial scales might have on ichthyofauna structure in preserved systems, since environmental homogenization in impacted streams (Olden et al. 2004) could obscure the local fish patterns included in broader scales. For Vermelho River, which is a preserved stream, the middle reach has higher local environmental heterogeneity when compared with the headwater and mouth reaches, creating some combinations of fish-mesohabitat associations. At middle reach, the mesohabitat scale seems to transcend longitudinal processes, not completely differentiating its fish structure from the others reaches, but mixing fish composition from upper and lower reaches in the structuring of local mesohabitats. Therefore, results might be interpreted as an alternative to mesohabitat-specific fish distributions among riffles, runs and pools according to fish adaptations. In this sense, it is expected that from middle reach to upstream or to downstream, the fish-habitat associations are more homogeneous in function of the greater longitudinal scale influence.

Analyzing only longitudinal scale, similar patterns in fish distribution among river segments were identified by Ferreira and Petrere Jr (2009), who established altitudinal zones with a predominance of different fish families in streams of the Atlantic coast, São Paulo. At reach scale, fish communities in the Vermelho River seem to reflect adjustments in the specific composition according to the abiotic gradient. Therefore, the stream might be

characterized by a headwater reach with a poorer ichthyofauna, with species such as *A. multispinnis* and *Characidium* spp., which are adapted to high water velocity; middle reach with the highest diversity, fish-mesohabitat structuring and an increase in abundance of water column dwelling species, such as characids and poeciliids; and a mouth reach, within boundaries of the coastal plain, with slow water velocity and deeper environments, that favor colonization by characids such as *M. microlepis*, loricariids of lentic habitats, and eventually estuarine species.

Focusing only on the mesohabitat scale, several authors recognize that within fish community, some species preferably inhabit riffles or runs while others inhabit pools (Taylor 2000, Langeani et al. 2005). Likewise, Rezende et al. (2010) identified and established fish species as indicators of mesohabitats according to their relative densities in riffles and pools in a coastal stream. In the Vermelho River, although headwater and mouth reaches are the most environmentally homogeneous habitats, the middle reach was geomorphologically structured into riffles, pools and runs. In riffles, which are hydrologically more dynamic mesohabitats, the ichthyofauna probably directs large amounts of energy to maintain their position against the current (Matthews and Styron Jr 1981). Energy-saving strategies could explain the high relationship of some species, such as A. multispinnis, and Characidium spp., with the middle riffles, since these species have depressed and fusiform bodies, minimizing the energy cost of swimming in fast waters (Breda et al. 2005). Barreto and Aranha (2005) observed A. multispinnis and C. pterostictum in moderate to torrential currents, the latter maintaining themselves by standing on their pectoral and pelvic fins in protected areas behind cobbles, possibly to reduce energy expenditure.

On the other hand, pools have been considered as mesohabitats with the highest habitat availabilities and environmental complexity, which may support a greater number of species (Taylor 2000, Langeani et al. 2005). We also reported higher richness and habitat preferences for *P. harpagos*, *S. barbatus* and *S. marmoratus* in pools. Similar to our findings, *P. harpagos* was defined as an indicator of pools by Rezende et al. (2010), while *S. barbatus* occurs in areas with slow currents on sandy substrate, and *S. marmoratus* occurs in holes of clay margins (Barreto and Aranha 2005). In addition, pools presented higher amounts of trunks, branches, leaf litter and were deeper, that favors the colonization by characids with laterally compressed body, such as *D. langei* and *M. microlepis*. Since pools were more environmentally structured, they might also represent a refuge from predators (Matthews 1998).

Different from riffles and pools, totally segregated from one another, fish community structure of runs was slightly similar in composition with other mesohabitats. However, runs harbor a high number of lentic species such as *D. langei*, *M. microlepis* and *H. multifasciatus* in relation to riffles, and an intermediate abundance of fast-water fish, such as *Characidium* spp. Similar to our study, where the ichthyofauna was not completely distinct in composition, but mainly in proportional abundance, Rezende



et al. (2010) found no indicator species for runs. As there are no boundaries between mesohabitats to prevent fish movement, species may be using runs to forage or as transient areas to access their preferential sites. In this sense, these runs would act as ecotones between riffle and pool mesohabitats (Jowett 1993).

In short, our results showed differences in composition and structure of ichthyofauna in the Vermelho River, related both to reach and mesohabitat scales. These results reveal differences in distribution, abundance and ecological requirements of species and suggest that environmental differences along stream determine the wider structural patterns of ichthyofauna. However, combining reach and habitat spatial scales, some patterns in fish similarities showed that the middle reach is a transitional heterogeneous area that congregates species from the headwater and mouth reaches, but is locally structured in fish-mesohabitat associations. Additionally, these local patterns revealed distinct fish structures between riffles and pools, which likely imply distinct local ecological interactions. Our findings reflect the preserved conservation status of the Vermelho River, contributing to identifying possible direct impacts on fish fauna along longitudinal gradients in other Atlantic coastal streams.

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LITERATURE CITED

- Abilhoa V, Bastos LP (2009) Fish, Cubatão river basin, Atlantic Rainforest stream, Paraná, Brazil. Check List 5: 8–18. https://doi.org/10.15560/5.1.8
- Allan JD (1995) Stream ecology: structure and function of running waters. Chapman and Hall, London, 388 pp. https://doi.org/10.1007/978-94-011-0729-7
- Angermeier PL, Karr JR (1983) Fish communities along environmental gradients in a system of tropical streams. Environmental Biology of Fish 9: 117–135. https://doi.org/10.1007/BF00690857

- Angermeier PL, Schlosser IJ (1989) Species-area relationships for stream fishes. Ecology 70: 1450–1642. https://doi.org/10.2307/1938204
- Barreto AP, Aranha JMR (2005) Assembleia de peixes de um riacho de Floresta Atlântica: composição e distribuição espacial (Guaraqueçaba, Paraná, Brasil). Acta Scientiarum Biological Sciences 27: 153–160. https://doi.org/10.4025/actascibiolsci. v27i2.1326
- Breda L, Oliveira EF, Goulart E (2005) Ecomorfologia de locomoção de peixes com enfoque para espécies neotropicais. Acta Scientiarum Biological Sciences 27: 371–381. https://doi.org/10.4025/actascibiolsci.v27i4.1271
- Casatti L (2005) Fish assemblage structure in a first order stream, southeastern Brazil: longitudinal distribution, seasonality, and microhabitat diversity. Biota Neotropica 5: 75–83. https://doi.org/10.1590/S1676-06032005000100009
- Cheek BD, Grabowski TB, Bean PT, Groeschel JR, Magnelia SJ (2016) Evaluating habitat associations of a fish assemblage at multiple spatial scales in a minimally disturbed stream using low-cost remote sensing. Aquatic Conservation: Marine and Freshwater Ecosystems 26: 20–34. https://doi.org/10.1002/agc.2569
- Esteves KE, Aranha JMR (1999) Ecologia trófica de peixes de riachos. In: Caramaschi EP, Mazzoni R, Peres-Neto PR (Eds) Ecologia de peixes de riachos. PPGE-UFRJ, Série Oecologia Brasiliensis, Rio de Janeiro, 157–182. https://doi.org/10.4257/oeco.1999.0601.05
- Esteves KE, Lobón-Cerviá J (2001) Composition and trophic structure of a clear water Atlantic rainforest stream in southeastern Brazil. Environmental Biology of Fish 62: 429–440. https://doi.org/10.1023/A:1012249313341
- Ferreira FC, Petrere Jr M (2009) The fish zonation of the Itanhaém river basin in the Atlantic Forest of southeast Brazil. Hydrobiologia 363: 11–34. https://doi.org/10.1007/s10750-009-9932-4
- Fogaça FNO, Aranha JMR, Ester MLP (2003) Ictiofauna do rio do Quebra (Antonina, PR, Brasil): Ocupação espacial e hábito alimentar. Interciencia 28: 168–173.
- Frissell CA, Liss WJ, Warren CE, Hurley MD (1986) A hierarchical framework for stream habitat classification: viewing streams in a watershed context. Environmental Management 10: 199–214. https://doi.org/10.1007/BF01867358
- Gonçalves CS, Braga FMS (2012) Changes in ichthyofauna composition along a gradient from clearwaters to blackwaters in coastal streams of Atlantic forest (southeastern Brazil) in relation to environmental variables. Neotropical Ichthyology 10: 675–684. https://doi.org/10.1590/S1679-62252012000300022
- Gordon ND, McMahon TA, Finlayson BL (1992) Stream hydrology: an introduction for ecologists. John Wiley and Sons, Chichester, 526 pp.
- Gotelli NJ, Ellison AM (2004) A primer of ecological statistics. Sinauer Associates, Sunderland, 614 pp.
- Guimarães ATB, Menezes MS, Peret AC (2010) Composição da ictiofauna em função da fisiografia de um riacho costeiro de



- Floresta Atlântica Brasil. Biota Neotropica 10: 57–65. https://doi.org/10.1590/S1676-06032010000200006
- Harrison S, Ross SJ, Lawton JH (1992) Beta diversity on geographic gradients in Britain. Journal Animal Ecology 61: 151–158. https://doi.org/10.2307/5518
- Hill MO, Gauch Jr HG (1980) Detrended correspondence analysis: an improved ordination technique. Vegetatio 42: 47–58. https://doi.org/10.1007/BF00048870
- Ibañez C, Belliard J, Hughes RM, Irz P, Kamdem-Toham A, Lamouroux N, Tedesco PA, Oberdorff T (2009) Convergence of temperate and tropical stream fish assemblages. Ecography 32: 658–670. https://doi.org/10.1111/j.16000587.2008.05591.x
- Ibañez C, Oberdorff T, Teugels G, Mamononekene V, Lavoué S, Fermon Y, Paugy P, Toham AK (2007) Fish assemblages structure and function along environmental gradients in rivers of Gabon (Africa). Ecology of Freshwater Fish 16: 315–334. https://doi.org/10.1111/j.1600-0633.2006.00222.x
- Jackson DA (1993) Stopping rules in principal components analysis: a comparison of heuristical and statistical approaches. Ecology 74: 2204–2214. https://doi.org/10.2307/1939574
- Jowett IG (1993) A method for objectively identifying pool, run, and riffle habitats from physical measurements. New Zealand Journal of Marine and Freshwater Research 27: 241–248. https://doi.org/10.1080/00288330.1993.9516563
- Kano Y, Miyazaki Y, Tomiyama Y, Mitsuyuki C, Nishida S, Rashid ZA (2013) Linking Mesohabitat Selection and Ecological Traits of a Fish Assemblage in a Small Tropical Stream (Tinggi River, Pahang Basin) of the Malay Peninsula. Zoological Science 30: 178–184. https://doi.org/10.2108/zsj.30.178
- Krebs CJ (1989) Ecological Methodology. Harper and Row, New York, 654 pp.
- Langeani F, Casatti L, Gameiro HS, Bellucco-do-Carmo A, Rossa-Feres DC (2005) Riffle and pool fish communities in a large stream of southeastern Brazil. Neotropical Ichthyology 3: 305–311. https://doi.org/10.1590/S167962252005000200009
- Lorion CM, Kennedy BP, Braatne JH (2011) Altitudinal gradients in stream fish diversity and the prevalence of diadromy in Sixaola River basin, Costa Rica. Environmental Biology of Fishes 91: 487–499. doi:10.1007/s10641-011-9810-6
- Maack R (1981) Geografia física do estado do Paraná. Editora José Olympio, Rio de Janeiro, 450 pp.
- Magurran AE (1988) Ecological Diversity and its Measurements. Croom Helm Limited, London, 179 pp. https://doi.org/10.1007/978-94-015-7358-0
- Manly BJF (2008) Métodos estatísticos multivariados: uma introdução. Bookman, Porto Alegre, 3rd ed., 230 pp.
- Matthews WJ (1998) Patterns in freshwater fish ecology. Chapman and Hall, New York, 756 pp. https://doi.org/10.1007/978-1-4615-4066-3
- Matthews WJ, Styron Jr JT (1981) Tolerance of headwater vs. mainstream fishes for abrupt physicochemical changes. The American Midland Naturalist 105: 149–158. https://doi.org/10.2307/2425020

- Mazzoni R, Lobón-Cerviá J (2000) Longitudinal structure, density and production rates of a Neotropical stream fish assemblage: the river Ubatiba in the Serra do Mar, southeast Brazil. Ecography 23: 588–602. https://doi.org/10.1111/j.1600-0587.2000.tb00178.x
- McGarvey DJ (2011) Quantifying ichthyofaunal zonation and species richness along a 2800-km reach of the Rio Chama and Rio Grande (USA). Ecology of Freshwater Fish 20: 231–242. https://doi.org/10.1111/j.1600-0633.2011.00485.x
- Melles SJ, Jones NE, Schmidt B (2012) Review of theoretical developments in stream ecology and their influence on stream classification and conservation planning. Freshwater Biology 57: 415–434. https://doi.org/10.1111/j. 1365-2427.2011.02716.x
- Menezes NA, Wzmeitan SH, Oyakawa OT, Lima FCT, Castro RMC, Wzmeitan MJ (2007) Peixes de água doce da Mata Atlântica: lista preliminar das espécies e comentários sobre conservação de peixes de água doce neotropicais. Museu de Zoologia, Universidade de São Paulo, São Paulo, 408 pp.
- Myers N, Mittermeier RA, Mittermeier CG, Fonseca GA, Kent J (2000) Biodiversity hotspots for conservation priorities. Nature 403: 853–858. https://doi.org/10.1038/35002501
- Oberdorff T, Guilbert E, Luchetta JC (1993) Patterns of fish species richness in the Seine River basin, France. Hydrobiologia 259: 157–167. https://doi.org/10.1007/BF00006595
- Olden JD, Leroy Poff N, Douglas MR, Douglas ME, Fausch D (2004) Ecological and evolutionary consequences of biotic homogenization. Trends in Ecology and Evolution 19: 18–24. https://doi.org/10.1016/j.tree.2003.09.010
- Oyakawa OT, Akama A, Mautari KC, Nolasco JC (2006) Peixes de riachos da Mata Atlântica nas unidades de conservação do vale do rio Ribeira de Iguape no estado de São Paulo. Editora Neotrópica, São Paulo, 201 pp.
- Pansonato-Alves JC, Serrano EA, Utsunomia R, Camacho JPM, Silva GJC, Vicari MR, Artoni RF, Oliveira C, Foresti F (2014) Single origin of sex chromosomes and multiple origins of B chromosomes in fish genus *Characidium*. PLOSone 9: e107169. doi. org/10.1371/journal.pone.0107169
- Pegg BMA, Behmer AT, Parasaweicz P, Rogers JN (2014) Application of mesohabitat fish use information to identify guilds for lotic systems. Applied Ichthyology 30: 1065–1068. https://doi.org/10.1111/jai.12502
- Pekárik L, Švátora M, Černý J, Kosco J (2011) Longitudinal structure of fish assemblages in a minimally disrupted stream. Biologia 66: 886–892. https://doi.org/10.2478/s11756-011-0097-z
- Petry AC, Schulz UH (2006) Longitudinal changes and indicator species of the fish fauna in the subtropical Sinos River, Brazil. Journal of Fish Biology 69: 272–290. https://doi.org/10.1111/j. 1095-8649.2006.01110.x
- Rahel FJ, Hubert WA (1991) Fish assemblages and habitat gradients in a rocky mountain great plains stream: biotic zonation and additive patterns of community change. Transactions of the American Fisheries Society 120: 319–332. https://doi.org/10.1577/1548-8659(1991)120<0319:FAAHGI>2.3.CO;2



Rezende CF, Moraes M, Manna LR, Leitão RP, Caramaschi EP, Mazzoni R (2010) Mesohabitat indicator species in a coastal stream of the Atlantic rainforest, Rio de Janeiro-Brazil. Revista de Biologia Tropical 58: 1479–1487.

Rincón PA (1999) Uso do micro-hábitat em peixes de riachos: métodos e perspectivas. In: Caramaschi EP, Mazzoni R, Peres-Neto PR (Eds) Ecologia de peixes de Riachos. PPGE-UFRJ, Série Oecologia Brasiliensis, Rio de Janeiro, 23–90. https://doi.org/10.4257/oeco.1999.0601.02

Silva JFM, Raio CB, Bernardino DFS, Bennemann ST (2013) Longitudinal patterns of fish assemblages in mountain streams from tropical forest biome. Biota Neotropica 13(3): 64–73. https://doi.org/10.1590/S1676-06032013000300008

Suvarnaraksha A, Lek S, Lek-Ang S, Jutagate T (2012) Fish diversity and assemblage patterns along the longitudinal gradient of a tropical river in the Indo-Burma hotspot region (Ping-Wang River Basin, Thailand). Hydrobiologia 694: 153–169. https://doi.org/10.1007/s10750-012-1139-4

Taylor CM (2000) A large-scale comparative analysis of riffle and pool fish communities in an upland stream system. Environmental Biology of Fish 58: 89–95. https://doi. org/10.1023/A:1007677718275

Teresa FB, Casatti L (2012) Influence of forest cover and mesohabitat types on functional and taxonomic diversity of fish communities in Neotropical lowland streams. Ecology of Freshwater Fish 21: 433–442. https://doi.org/10.1111/j.1600-0633.2012.00562.x

Vannote RL, Minshall GW, Cummins KW, Sedell JR, Cushing CE (1980) The river continuum concept. Canadian Journal Fisheries and Aquatic Sciences 37: 130–137. https://doi.org/10.1139/f80-017

Velloso HP, Rangel-Filho ALR, Lima JCA (1991) Classificação da vegetação brasileira adaptada a um sistema universal. IBGE/ Departamento de Recursos Naturais e Estudos Ambientais, Rio de Janeiro, 123 pp.

Vari RP, Malabarba LR (1998) Neotropical ichthyology: an overview. In: Malabarba LR, Reis RE, Vari RP, Lucena ZMS, Lucena CAS (Eds) Phylogeny and Classification of Neotropical Fishes. Edipucrs, Porto Alegre, 1–11.

Worischka S, Hellmann C, Berendonk TU, Winkelmann C (2014) Fish predation can induce mesohabitat-specific differences in food web structures in small stream ecosystems. Aquatic Ecology 48: 367–378. https://doi.org/10.1007/s10452-014-9490-3

Supplementary material 1

Figure S1. Hydrological and structural characteristics of the sampling reaches of the Vermelho River, state of Paraná, Brazil

Authors: Luciano Lazzarini Wolff, Norma Segatti Hahn Data type: specimens data

Explanation note: (A) Cascaded channel in the headwater; (B, C and D) riffles, pool and runs mesohabitats, respectively, in the middle; (E-F) slow-moving waters habitats in the mouth

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