



Sampling procedures for inventory of commercial volume tree species in Amazon Forest

SYLVIO P. NETTO¹, ALLAN L. PELISSARI¹, VINICIUS C. CYSNEIROS¹,
MARCELO BONAZZA² and CARLOS R. SANQUETTA¹

¹Engenharia Florestal, Universidade Federal do Paraná/UFPR, Av. Prof. Lothário Meissner, 632, Jardim Botânico, 80210-170 Curitiba, PR, Brazil

²Engenharia Florestal, Universidade Federal de Santa Catarina/UFSC, Rod. Ulysses Gaboardi, Km 3, Pessegueirinho, 89520-000 Curitibanos, SC, Brazil

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ABSTRACT

The spatial distribution of tropical tree species can affect the consistency of the estimators in commercial forest inventories, therefore, appropriate sampling procedures are required to survey species with different spatial patterns in the Amazon Forest. For this, the present study aims to evaluate the conventional sampling procedures and introduce the adaptive cluster sampling for volumetric inventories of Amazonian tree species, considering the hypotheses that the density, the spatial distribution and the zero-plots affect the consistency of the estimators, and that the adaptive cluster sampling allows to obtain more accurate volumetric estimation. We use data from a census carried out in Jamari National Forest, Brazil, where trees with diameters equal to or higher than 40 cm were measured in 1,355 plots. Species with different spatial patterns were selected and sampled with simple random sampling, systematic sampling, linear cluster sampling and adaptive cluster sampling, whereby the accuracy of the volumetric estimation and presence of zero-plots were evaluated. The sampling procedures applied to species were affected by the low density of trees and the large number of zero-plots, wherein the adaptive clusters allowed concentrating the sampling effort in plots with trees and, thus, agglutinating more representative samples to estimate the commercial volume.

Key words: Adaptive cluster sampling, spatial species distribution, volume estimation, zero-plots.

INTRODUCTION

The Amazon Forest composes the richest collection of plant species on the planet, having approximately 16 thousand tree species, where approximately 50% of the trees are concentrated in only 1.4% of species (Ter Steege et al. 2013) in a flora composed of many rare species of restricted distribution

(Hopkins 2007). For this, the spatial patterns are frequently the focus of ecological researches, due to the high diversity in tropical forests that is characterized by low density of tax (Condit et al. 2000). This knowledge is important to inventories, especially those intended for production and conservation forests.

The spatial distributions of species are fundamental for ecological modeling (Condit et al. 2000), where they reflect recruitment and mortality

Correspondence to: Allan Libanio Pelissari
E-mail: allanpelissari@gmail.com

patterns, autoecological characteristics, syndrome of dispersion and reproductive biology (Crawley 1986, Pianka 1994, Dale 1999). However, the spatial patterns can affect the consistency of the sampling procedures. If the pattern is aggregate, a sample with low number of plots can result in high or low density when the results are extrapolated to the population and, thus, appropriated sampling techniques are required (Odum and Barret 2008).

In Brazilian Amazon, conventional sampling procedures are constantly applied to estimate volumetric stock of species groups (Higuchi et al. 1982, Cavalcanti et al. 2011, Oliveira et al. 2014), where there are many rare and aggregate species responsible for high diversity that affect effectiveness of forest inventories. Nevertheless, there are no studies reported in the specialized literature about appropriate sampling techniques by specific species with different spatial patterns in the Amazon Forest.

In many studies, the adaptive cluster sampling has proved to be a versatile tool for evaluating rare and aggregate populations, because by conventional sampling you cannot estimate well their parameters (Thompson 1990, Brown and Manly 1998, Talvitie et al. 2006). Initially, the adaptive sampling was developed by Thompson (1990), which compared the results of this procedure with the conventional ones, evaluating different populations and finding satisfactory results, especially for aggregate and rare events.

In the adaptive cluster sampling, the first level is based on conventional methods as random or systematic. Through detection of a species or interesting phenomenon in the plots, the second level is started and new plots are allocated contiguously to the first ones. The second level continues until the target phenomenon is not detected and, then, the construction of the clusters is interrupted. Furthermore, its sampling structure and their estimators are presented in Thompson

(1990), Acharya et al. (2000), Talvitie et al. (2006) and Lei et al. (2012).

Considering the hypotheses that (1) the low density of trees, the spatial distribution of species and the high number of zero-plots affect the consistency of the samplings in tropical forests; and that (2) the adaptive cluster sampling allows to obtain more accurate volumetric stocks; the present study aims to evaluate the conventional sampling procedures and introduce the adaptive cluster sampling to inventory of tree species in Amazon Forest.

MATERIALS AND METHODS

STUDY AREA

A census dataset of a native forest with 1,596 ha and located in the Jamari National Forest, Rondônia State, Brazil, was used to develop this study. The Jamari National Forest occupies an area between the coordinates 09° 00' 00" S at 09° 30' 00" S and 62° 44' 05" W at 63° 16' 64" W in the Southwest Amazon Forest, where all trees with diameter at 1.3 m above the ground equal to or greater than 40 cm were measured, identified and georeferenced in an Annual Production Unit divided in 1,355 plots of 50 m × 250 m.

PHYTOSOCIOLOGICAL CHARACTERIZATION

The forest structure was characterized by means of phytosociological descriptors (Müller-Dombois and Ellenberg 1974) and diversity indices (Shannon and Weaver 1949, Simpson 1949, Pielou 1969). Also, the spatial patterns of species were classified through standardized Morisita's index (Morisita 1962) as random, with values between -0.5 and +0.5; uniform, characterized by values below -0.5; and aggregate, with values higher than +0.5 (Morisita 1962, Amaral et al. 2015). In addition, the Kernel density estimator (Silverman 1986) was applied to investigate the species density and their

distributions, through ArcGIS 10.3 software and Spatial Analyst package (ESRI 2016).

SAMPLING PROCEDURES

Amazon tree species were selected considering different spatial patterns and the criteria of rarity, economic and social importance. Thus, the simple random, systematic, linear cluster and adaptive cluster sampling procedures were applied with 10% of the potential sampling units of 50 m × 250 m. In simple random sampling (Figure 1a), the plots were randomly allocated to each one of the selected species populations, while the systematic sampling units in single stage were distributed with regular intervals of 250 m between them (Figure 1b).

Furthermore, linear clusters were allocated randomly in the study area, being formed by four units of 50 m × 250 m separated by plots of equal size (Figure 1c). For adaptive clusters, the first

stage consisted in a random allocation of sampling units, and those that corresponded to the inclusion criterion of at least one tree were selected for the second stage; subsequently, their neighbor plots with one or more trees were incorporated into each cluster, forming their respective networks in the final stage (Figure 1d).

The estimated volume per hectare (1 to 3) and variance of the mean (4 to 7) were calculated for simple random sampling, systematic sampling, and linear cluster sampling (Péllico Netto and Brena 1997, Husch et al. 2002). While the modified Horvitz-Thompson estimators (Thompson 1990) were used and implemented by R statistical program (R Core Team 2013) for the adaptive clusters sampling. Furthermore, the standard error (8), relative sampling error (9), and confidence interval (10) were calculated for all sampling procedures.

	Simple random and Systematic	$\bar{x} = \frac{\sum_{i=1}^n X_i}{n}$	(1)
Sample mean (\bar{x})	Linear cluster	$\bar{x} = \frac{\sum_{i=1}^n \sum_{j=1}^M X_{ij}}{nM}$	(2)
	Adaptive cluster	$\bar{x} = \frac{1}{N} \sum_{k=1}^K \frac{y_k^*}{\alpha_k}$	(3)
	Simple random	$s_{\bar{x}}^2 = \frac{s_x^2}{n} \left(\frac{N-n}{N} \right)$	(4)
Variance of the mean ($s_{\bar{x}}^2$)	Systematic	$s_{\bar{x}}^2 \cong \frac{\sum_{i=1}^{n-1} (X_i - X_{i+1})^2}{2n(n-1)} (1-f)$	(5)
	Linear cluster	$s_{\bar{x}}^2 = \frac{s_x^2}{nM} [1 + r(M-1)]$	(6)
	Adaptive cluster	$s_{\bar{x}}^2 = \frac{1}{N^2} \left[\sum_{j=1}^K \sum_{k=1}^K \frac{y_j y_k^*}{\alpha_{jk}} \left(\frac{\alpha_{jk}}{\alpha_j \alpha_k} - 1 \right) \right]$	(7)

Standard error ($s_{\bar{x}}$)

$$S_{\bar{x}} = \sqrt{S_{\bar{x}}^2} \tag{8}$$

Relative sampling error ($E\%$)

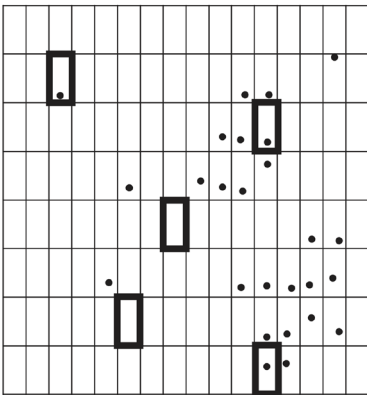
$$E\% = \pm \frac{ts_{\bar{x}}}{\bar{x}} 100 \tag{9}$$

Confidence interval the mean (CI)

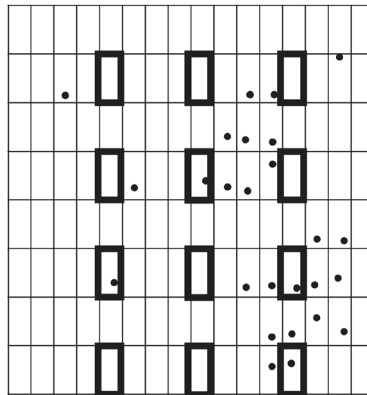
$$IC[\bar{x} - ts_{\bar{x}} \leq \mu \leq \bar{x} + ts_{\bar{x}}] = P \tag{10}$$

Where: n = number of units sampled, X_i = volume of the sample unit i , X_{ij} = volume of unit i in linear clusters j , M = number of subunits of linear clusters, N = number of potential sample units, k = number of networks of the adaptive clusters present in the sample on the first sampling level, Y_k^* = sum of observations of the net k of the adaptive clusters k , α_k = probability of initial sample belong to network k , α_{jk} = probability of initial sample to include at least one sampling unit in each network j and k , s_x^2 = sample variance of volume, $f = \frac{n}{N}$, t = value of student's t distribution, and P = associated probability of 95%.

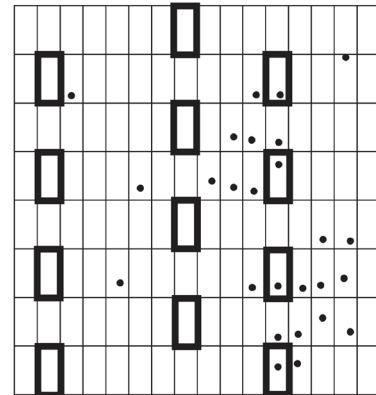
a. Simple random sampling



b. Systematic sampling

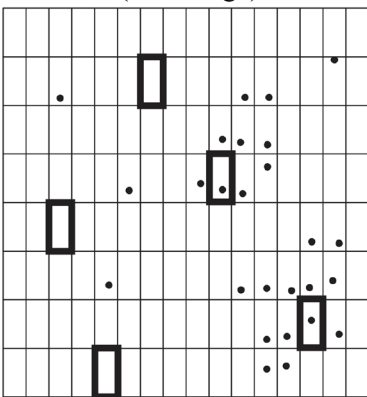


c. Linear clusters sampling

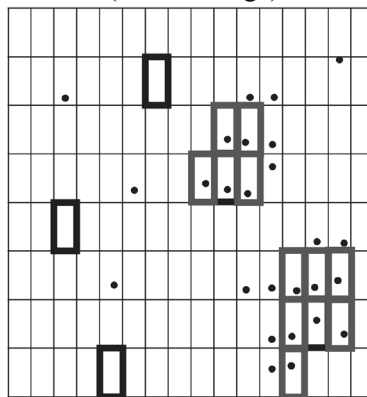


d. Adaptive cluster sampling

(First stage)



(Second stage)



(Final stage)

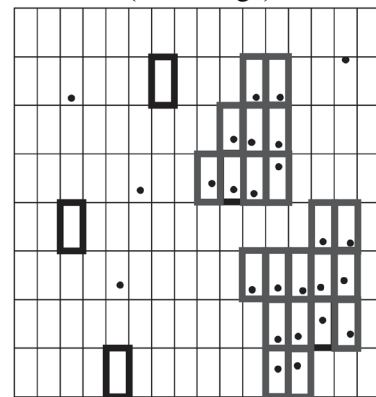


Figure 1 - Plots allocation structure in the sampling procedures.

Thereby, for each sampling procedure and species, the statistical consistency was evaluated through Z test at significance level of 0.05, using the mean census volume (μ) as the reference value. The following hypotheses were tested: null (H_0), when there is no rejection of equality between the mean volume of sample and census ($H_0: \bar{x} = \mu$); and alternative (H_1), when there is rejection of this equality ($H_1: \bar{x} \neq \mu$). Also, the relative errors were evaluated, and the zero-plots, without trees (Heinsdijk 1965), were quantified in the sampling procedures.

RESULTS

In the census, 17,557 trees were sampled of 67 species (Table I), where Fabaceae was the family with highest floristic richness (24 species), followed by Vochysiaceae with five species, and

Moraceae and Lecythidaceae with four species each. Moreover, by means of the values of diversity indices: Shannon (H') = 3.550, Pielou (J) = 0.845, and Simpson (C) = 0.038, it was evidenced a highly diverse community and with the absence of dominance of few species, representing, therefore, a balance between all taxa.

Distinct spatial distribution patterns were observed (Figure 2), where representative tree species were selected: a) *Terminalia amazonica*, a rare species, with density of only seven trees, and concentrated in a specific region (Figure 2a); (b) *Apuleia leiocarpa*, with 89 trees and concentrated in some locations (Figure 2b); c) *Cedrela fissilis*, with 81 trees dispersed in the area (Figure 2c); and d) *Bertholletia excelsa*, with high density of 904 trees (Figure 2d). In kernel maps, the density values

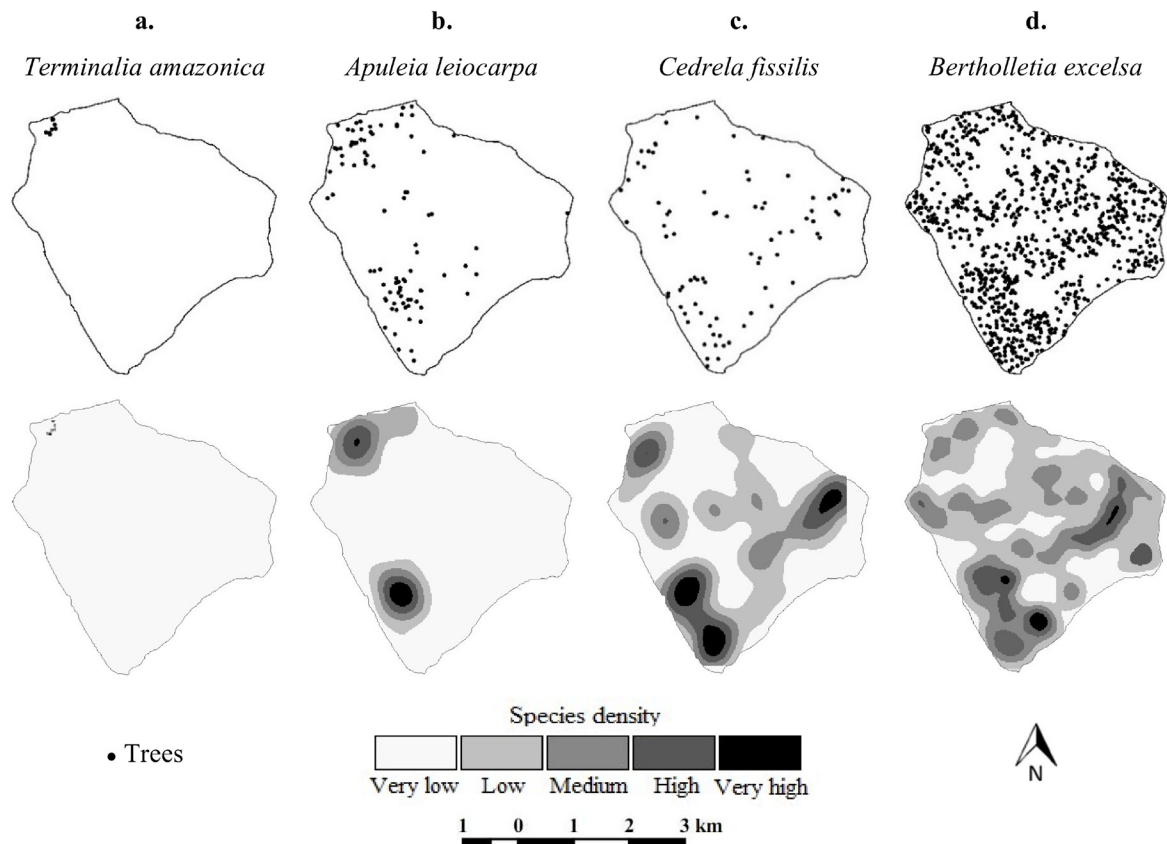


Figure 2 - Spatial distribution of the commercial tree species in Jamari National Forest, Brazilian Amazon.

TABLE I
Phytosociological descriptors of the commercial tree species in Jamari National Forest, Brazilian Amazon.

Species	Family	N	D%	F%	Do%	VI%	Morisita's index	
<i>Dinizia excelsa</i> Ducke	Fabaceae	986	5.62	4.67	13.11	7.80	0.50	Aggregate
<i>Peltogyne paniculata</i> Benth.	Fabaceae	1,598	9.10	6.62	5.89	7.20	0.50	Aggregate
<i>Astronium lecointei</i> Ducke	Anacardiaceae	1,296	7.38	5.84	6.35	6.53	0.50	Aggregate
<i>Bertholletia excelsa</i> Humb. & Bonpl.	Lecythidaceae	904	5.15	4.89	8.93	6.32	0.50	Aggregate
<i>Cariniana micrantha</i> Ducke	Lecythidaceae	676	3.85	3.86	6.90	4.87	0.50	Aggregate
<i>Huberodendron swietenoides</i> (Gleason) Ducke	Malvaceae	828	4.72	4.03	4.81	4.52	0.50	Aggregate
Tachigali sp.	Fabaceae	807	4.60	3.94	3.28	3.94	0.50	Aggregate
<i>Couratari stellata</i> A. C. Sm.	Lecythidaceae	581	3.31	3.51	3.91	3.58	0.50	Aggregate
<i>Copaifera multijuga</i> Hayne	Fabaceae	690	3.93	3.83	2.38	3.38	0.50	Aggregate
<i>Clarisia racemosa</i> Ruiz & Pav.	Moraceae	636	3.62	3.42	2.40	3.15	0.50	Aggregate
<i>Aspidosperma</i> sp.	Apocynaceae	514	2.93	3.07	3.10	3.03	0.50	Aggregate
<i>Protium robustum</i> (Swart) D.M. Porter	Burseraceae	658	3.75	3.01	2.12	2.96	0.50	Aggregate
<i>Hymenolobium heterocarpum</i> Ducke	Fabaceae	456	2.60	2.89	3.14	2.87	0.33	Random
<i>Pouteria guianensis</i> Aubl.	Sapotaceae	565	3.22	3.17	2.04	2.81	0.50	Aggregate
<i>Dipteryx odorata</i> (Aubl.) Willd.	Fabaceae	405	2.31	2.64	2.03	2.33	0.41	Random
<i>Caryocar glabrum</i> Pers.	Caryocaraceae	359	2.04	2.44	1.96	2.15	-0.17	Random
<i>Goupia glabra</i> Aubl.	Goupiaceae	343	1.95	2.19	2.11	2.09	0.50	Aggregate
<i>Erismia bicolor</i> Ducke	Vochysiaceae	336	1.91	2.20	1.74	1.95	0.50	Aggregate
<i>Qualea paraensis</i> Ducke	Vochysiaceae	354	2.02	2.23	1.39	1.88	0.50	Aggregate
<i>Allantoma decandra</i> (Ducke)	Lecythidaceae	292	1.66	1.96	1.78	1.80	0.39	Random
<i>Vataireopsis speciosa</i> Ducke	Fabaceae	302	1.72	2.05	1.23	1.67	0.15	Random
<i>Brosimum rubescens</i> Taub.	Moraceae	287	1.63	1.92	1.33	1.63	0.50	Aggregate
<i>Hymenaea palustris</i> Ducke	Fabaceae	261	1.49	1.75	1.13	1.45	0.50	Aggregate
<i>Brosimum</i> sp.	Moraceae	230	1.31	1.59	1.11	1.33	0.20	Random
<i>Erismia fuscum</i> Ducke	Vochysiaceae	235	1.34	1.52	1.09	1.32	0.50	Aggregate
<i>Vatairea guianensis</i> Aubl.	Fabaceae	235	1.34	1.56	1.00	1.30	0.50	Aggregate
<i>Cedrelinga cateniformis</i> (Ducke) Ducke	Fabaceae	166	0.95	1.03	1.77	1.25	0.50	Aggregate
<i>Iryanthera paradoxa</i> (Schwacke) Warb.	Myristicaceae	223	1.27	1.53	0.86	1.22	0.34	Random
<i>Handroanthus incanus</i> (A.H. Gentry) S. O. Grose	Bignoniaceae	168	0.96	1.20	0.98	1.05	0.05	Random
<i>Caryocar villosum</i> (Aubl.) Pers.	Caryocaraceae	148	0.84	1.05	1.04	0.98	0.26	Random
<i>Minquartia guianensis</i> Aubl.	Olacaceae	183	1.04	1.27	0.58	0.96	0.48	Random
<i>Parkia pendula</i> (Willd.) Benth. ex Walp.	Fabaceae	130	0.74	0.94	1.09	0.92	0.18	Random
<i>Enterolobium schomburgkii</i> (Benth.) Benth.	Fabaceae	149	0.85	1.07	0.74	0.89	0.43	Random
<i>Simarouba amara</i> Aubl.	Simaroubaceae	162	0.92	1.10	0.54	0.85	0.50	Aggregate
<i>Mezilaurus synandra</i> (Mez) Kosterm.	Lauraceae	146	0.83	1.00	0.61	0.82	0.50	Aggregate
<i>Handroanthus impetiginosus</i> (Mart. ex DC.) Mattos	Bignoniaceae	108	0.62	0.74	0.73	0.70	0.50	Aggregate
<i>Bowdichia nitida</i> Spruce ex Benth.	Fabaceae	109	0.62	0.81	0.42	0.62	-0.17	Random

TABLE I (continuation)

Species	Family	N	D%	F%	Do%	VI%	Morisita's index	
<i>Apuleia leiocarpa</i> (Vogel) J.F.Macbr.	Fabaceae	89	0.51	0.59	0.65	0.58	0.50	Aggregate
<i>Martiodendron elatum</i> (Ducke) Gleason	Fabaceae	97	0.55	0.71	0.43	0.56	0.21	Random
<i>Diploptropis rodriguesii</i> H.C. Lima	Fabaceae	100	0.57	0.74	0.33	0.54	0.05	Random
<i>Manilkara elata</i> (Allemão ex Miq.) Monach.	Sapotaceae	91	0.52	0.70	0.35	0.52	-0.45	Random
<i>Cedrela fissilis</i> Vell.	Meliaceae	81	0.46	0.61	0.33	0.47	-0.06	Random
<i>Peltogyne venosa</i> (Vahl) Benth.	Fabaceae	77	0.44	0.54	0.35	0.44	0.50	Aggregate
<i>Laetia procera</i> (Poepp.) Eichler	Salicaceae	76	0.43	0.57	0.27	0.42	-0.02	Random
<i>Virola</i> sp.	Myristicaceae	67	0.38	0.50	0.19	0.36	0.07	Random
<i>Dipteryx alata</i> Vogel	Fabaceae	57	0.32	0.43	0.22	0.33	-0.04	Random
<i>Cordia goeldiana</i> Huber	Boraginaceae	50	0.28	0.36	0.20	0.28	0.50	Aggregate
<i>Hevea guianensis</i> Aubl.	Euphorbiaceae	45	0.26	0.31	0.14	0.23	0.50	Aggregate
<i>Jacaranda copaia</i> (Aubl.) D.Don	Bignoniaceae	39	0.22	0.30	0.14	0.22	-0.19	Random
<i>Qualea</i> sp.	Vochysiaceae	39	0.22	0.27	0.14	0.21	0.50	Aggregate
<i>Parkia multijuga</i> Benth.	Fabaceae	25	0.14	0.18	0.13	0.15	0.50	Aggregate
<i>Bagassa guianensis</i> Aubl.	Moraceae	24	0.14	0.17	0.10	0.14	0.50	Aggregate
<i>Osteophloeum platyspermum</i> (Spruce ex A. DC.) Warb.	Myristicaceae	15	0.09	0.10	0.07	0.09	0.51	Aggregate
<i>Zollernia paraensis</i> Huber	Fabaceae	15	0.09	0.11	0.05	0.08	-0.07	Random
<i>Pouteria eugeniifolia</i> (Pierre) Baehni	Sapotaceae	11	0.06	0.07	0.05	0.06	0.51	Aggregate
<i>Terminalia amazonica</i> (J.F.Gmel) Exell.	Combretaceae	7	0.04	0.03	0.04	0.04	0.59	Aggregate
<i>Hymenolobium modestum</i> Ducke	Fabaceae	5	0.03	0.04	0.04	0.03	-0.02	Random
<i>Aspidosperma sandwithianum</i> Markgr.	Apocynaceae	6	0.03	0.05	0.02	0.03	-0.03	Random
<i>Coccoloba latifolia</i> Lam.	Polygonaceae	3	0.02	0.02	0.03	0.02	-0.01	Random
<i>Anacardium parviflorum</i> Ducke	Anacardiaceae	3	0.02	0.02	0.02	0.02	-0.01	Random
<i>Vochysia</i> sp.	Vochysiaceae	3	0.02	0.02	0.02	0.02	-0.01	Random
<i>Trattinnickia rhoifolia</i> Willd*	Burseraceae	1	0.01	0.01	0.03	0.02	-	-
<i>Lueheopsis rosea</i> (Ducke) Burret*	Malvaceae	1	0.01	0.01	0.01	0.01	-	-
<i>Parkia</i> sp.*	Fabaceae	1	0.01	0.01	0.01	0.01	-	-
<i>Hymenaea intermedia</i> Ducke*	Fabaceae	1	0.01	0.01	0.00	0.01	-	-
<i>Aspidosperma</i> sp.*	Apocynaceae	1	0.01	0.01	0.00	0.01	-	-
<i>Inga edulis</i> Mart.*	Fabaceae	1	0.01	0.01	0.00	0.01	-	-
Total		14,666	100	100	100	100		

Where: N = number of trees, D% = density, F% = frequency, Do% = dominance, VI% = value of importance, * = it was not possible to calculate the Morisita's Index for these species, because occurred only one tree in the population, and highlighted lines = indicate de species selected in this study.

were higher in locations with trees and decreased with increasing distance from these points.

When the Morisita's index was applied (Table I), the random spatial distribution was observed for *Cedrela fissilis*, while the aggregate pattern was identified for *Apuleia leiocarpa* and *Bertholletia excelsa*, as well as for *Terminalia amazonica* that showed the highest level of aggregation and one of the greatest levels of rarity with density ($D\%$) equal to 0.04%. Considering all species, 44.26% were classified as random, 55.73% as aggregate, and none with uniform distribution.

By means of the Z test at significance level of 0.05 (Table II), the null hypothesis was rejected only for mean volume (\bar{x}) in systematic sampling applied for *Apuleia leiocarpa*. However, statistical differences in association to the census (μ) were not found between the estimates by others sampling procedures, resulting in consistent confidence intervals (CI) for the means of studied species.

High relative errors ($E\%$) were observed for the selected species (Table II), especially for *Terminalia amazonica*, for which the application of adaptive cluster sampling increases the accuracy, as well as for *Bertholletia excelsa* and *Cedrela fissilis*. However, linear clusters were not effective for *Terminalia amazonica*, but they were superior than other procedures applied to *Apuleia leiocarpa*.

Regarding the percentage of zero-plots observed in each sampling procedure (Figure 3), lower values were observed when using adaptive cluster sampling, mainly to *Bertholletia excelsa* (Figure 3d). Moreover, the absence of trees per sampling unit resulted in zero-plots frequencies near to 100% for *Terminalia amazonica* (Figure 3a), *Apuleia leiocarpa* (Figure 3b) and *Cedrela fissilis* (Figure 3c).

DISCUSSION

According to the selected species with distinct spatial distribution patterns (Figure 2), *Apuleia*

leiocarpa and *Cedrela fissilis* have wood with high commercial value (Carvalho 2003) and wide geographical distribution in the Amazon territory (Pennington 1981, Oliveira Filho and Fontes 2000). *Terminalia amazonica* is commonly cultivated in Central America due to its fast growth (Piotto et al. 2003, Moya et al. 2009), while *Bertholletia excelsa* is the most relevant Amazonian species for non-timber purposes (Thomas et al. 2014), whose nuts are important for sustaining the Amazonian rural communities (Salomão 2009).

The Morisita's index equal to 0.59 for *Terminalia amazonica* was the largest among the species (Table I), while the aggregate distribution (0.50) found for *Apuleia leiocarpa* and *Bertholletia excelsa* is common among tropical species due to soil conditions and syndrome of species dispersion (Condit et al. 2000, Plotkin et al. 2000, Seidler and Plotkin 2006). The random distribution (-0.06) for *Cedrela fissilis* is less frequent, because it implies in more homogeneity of environment or in less specificity of the habitat (Matteucci and Colma 1982).

In this context, to compare the influence of environmental factors in the distribution of Amazonian tree species, Barroso et al. (2011) concluded that the soil attributes affect the abundance of species, although present a weak correlation with species occurrence. However, *Apuleia leiocarpa* (Figure 2b) and *Cedrela fissilis* (Figure 2c) holds high commercial value and, thus, the historical factors of the exploitation can reveal a strong influence on their current spatial distributions (Tassin and Riviere 2003), since the study area is inserted into a region of intense timber exploitation.

Bertholletia excelsa was observed in groups that ranged in size and number of trees (Figure 2d), and associated to other large trees on not flooded lands, where the natural clearings and the dispersion of its fruits by animals expand the population (Salomão 2009, Thomas et al. 2014).

TABLE II
Commercial volume estimation and percentage of zero-plots by sampling procedures in Jamari National Forest, Brazilian Amazon.

Sampling procedure	μ	\bar{x}	$E\%$	CI	$z-p\%$
	$(m^3 ha^{-1})$			$(m^3 ha^{-1})$	
<i>Terminalia amazonica</i>					
Simple random	0.017	0.051 ^{ns}	187.6%	$0.000 \leq \mu \leq 0.148$	99.3%
Systematic		0.035 ^{ns}	189.1%	$0.000 \leq \mu \leq 0.101$	99.2%
Linear cluster		-	-	-	100%
Adaptive cluster		0.083 ^{ns}	116.0%	$0.000 \leq \mu \leq 0.178$	97.8%
<i>Apuleia leiocarpa</i>					
Simple random	0.439	0.255 ^{ns}	82.8%	$0.044 \leq \mu \leq 0.467$	95.6%
Systematic		0.095*	146.9%	$0.000 \leq \mu \leq 0.234$	98.4%
Linear cluster		0.463 ^{ns}	69.1%	$0.143 \leq \mu \leq 0.783$	93.4%
Adaptive cluster		0.278 ^{ns}	74.8%	$0.070 \leq \mu \leq 0.486$	83.6%
<i>Cedrela fissilis</i>					
Simple random	0.265	0.267 ^{ns}	66.1%	$0.091 \leq \mu \leq 0.443$	94.1%
Systematic		0.213 ^{ns}	68.7%	$0.067 \leq \mu \leq 0.360$	94.5%
Linear cluster		0.277 ^{ns}	73.6%	$0.073 \leq \mu \leq 0.480$	94.1%
Adaptive cluster		0.288 ^{ns}	64.9%	$0.101 \leq \mu \leq 0.475$	92.1%
<i>Bertholletia excelsa</i>					
Simple random	3.704	3.821 ^{ns}	20.3%	$3.043 \leq \mu \leq 4.598$	51.5%
Systematic		3.980 ^{ns}	22.2%	$3.097 \leq \mu \leq 4.863$	50.4%
Linear cluster		3.216 ^{ns}	25.3%	$2.404 \leq \mu \leq 4.028$	59.6%
Adaptive cluster		3.746 ^{ns}	20.2%	$2.990 \leq \mu \leq 4.502$	14.2%

Where: μ = mean volume (census), \bar{x} = mean sample volume, $E\%$ = relative sampling error, CI = confidence interval, $z-p\%$ = percentage of zero-plots, ns = no rejection of equality between the mean volume of sample and census, and * = rejection of the equality by Z test at significance level of 0.05.

In addition to these factors, archaeological and historical evidences suggest the influence of the man, through collecting and cultivation, on the spatial distribution of the species (Peres et al. 2003, Thomas et al. 2014).

In inventories carried out in the Amazon Forest, the random, systematic and cluster sampling procedures are the most commonly applied (Higuchi 1987, Ubialli et al. 2009, Cavalcanti et al. 2011, Queiroz et al. 2011, Andrade et al. 2015). However, in approaches by species, these procedures have not shown satisfactory performance in the estimates of the present study (Table II), possibly due to low density of trees that affect the composition of representative samples,

especially for *Terminalia amazonica* and, thus, confirming the first hypothesis of this study.

The spatial patterns (Figure 2) affected in the effectiveness of the sampling procedures (Table II), endorsing the first hypothesis, whose spatial behaviors are concomitant effects of several mechanisms, as dispersion, predation, pathogenic disease, tolerance, germination and competition (Myser and Malahay 2012). On the other hand, the abiotic factors, such as topography and lighting, also influencing on the dispersal of species (Svenning 1999), turn the comprehension of aspects that determine the distribution pattern of tropical species more complex (Table II).

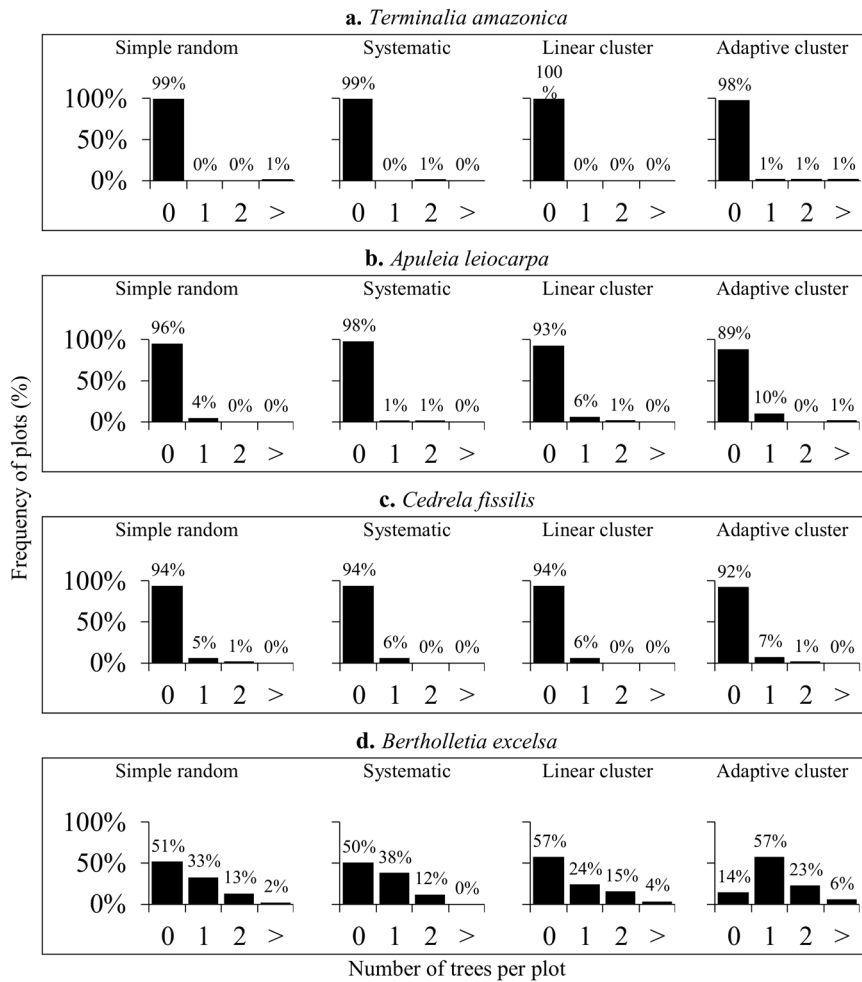


Figure 3 - Frequency of plots with zero, one, two or more than two trees for each species and sampling procedure in Jamari National Forest, Brazilian Amazon.

In addition, the high presence of zero-plots (Figure 3) interferes on the volumetric estimates, resulting in increase of the sampling error, which proves the first hypothesis (Table II). Therefore, the adaptive cluster sampling allowed to concentrate the sampling effort on non-zero-plots of the selected species. This enables us to maximize the efficiency of forest inventories and compose more representative samples for estimating commercial volume.

When applying adaptive cluster sampling, Talvitie et al. (2006) and Lei et al. (2012) observed that this procedure results in higher efficiency, when compared to conventional sampling procedures,

highlighting its importance for surveying rare and aggregate populations. However, few studies have considered the problems of forest sampling with adaptive clusters (Roesch Jr 1993, Acharya et al. 2000), since the sub-sampling of rare species results in a considerable underestimation of the biodiversity (Hopkins 2007).

As an alternative to fixed area plots, sampling methods with probability proportional to size could reduce or eliminate the presence of zero-plots in the composition of samples. However, Prodan's points and Strand's lines have showed operationally impracticable, due to the area size and the difficulty to include trees of rare species in the sampling units.

Thus, the results confirm the second hypothesis, that the adaptive cluster sampling reduces the zero-plots to estimate the commercial volume (Table II), and is a quite appropriated procedure for sampling rare populations of many kinds (Thompson 1990, Roesch Jr 1993, Brown 2003, Talvitie et al. 2006, Soares et al. 2009, Lei et al. 2012).

The estimator's effectiveness of the sampling procedures is directly related to spatial patterns, levels of aggregation and species density, where the adaptive clusters enable to concentrate the sampling effort to plots with occurrence of trees, reducing the percentage of zero-plots and maximizing the accuracy of commercial volume estimates in Amazon Forest inventories. However, the sampling procedures applied independently to species could result in unsatisfactory statistical performance in the volumetric estimations, due to the low density of individuals and to the high number of zero-plots.

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