

Embracing Methodological Issues in Ethnobiology and Overcoming Challenges

Ulysses Paulino Albuquerque^{1*} ⁽⁶⁾, Aníbal Silva Cantalice¹ ⁽⁶⁾, Arthur Ramalho Magalhães¹ ⁽⁶⁾, Michael A. Coe² ⁽⁶⁾, Reginaldo A. F. Gusmão¹ ⁽⁶⁾

¹Universidade Federal de Pernambuco, Departamento de Botânica, Laboratório de Ecologia e Evolução de Sistemas Socioecológicos, Recife, PE, Brazil

²Tarleton State University, Department of Biological Sciences, Stephenville, TX, United States.

*Corresponding author: upa677@hotmail.com

ABSTRACT

This article delves into the evolving landscape of ethnobiology, mainly focusing on integrating quantitative approaches and the hypothetical-deductive method. We highlight the challenges associated with adopting quantitative tools, such as the potential for oversimplification and lack of critical reflection on the principles guiding these methods. We also examine ecological diversity indices, cultural importance indices, sample sufficiency, replication in studies, and the importance of spatial-temporal context in ethnobiological research. We conclude with practical tips to enhance research's validity, reliability, and generalizability, proposing a path forward for the discipline that respects its essence while adapting to the demands of scientific evolution. While this paper broadly addresses ethnobiology, it primarily focuses on Ethnobotany literature, highlighting the field's rapid advancements and its relevance to other areas within ethnobiology.

Keywords: Ethnobotany, Quantitative Methods, Data Replicability, Sample Sufficiency, Spatial-Temporal Analysis, Cultural Importance Indices.

Introduction

For over three decades, there has been a tremendous shift in ethnobiological research from classical studies that primarily result in the helpful identification of species to human societies and their application in subsistence strategies, local livelihoods, and ethnomedicine to studies that have emphasized the development of quantitative methods, complementary to qualitative approaches, that are foundational for hypothesis testing and a greater understanding of the factors that drive humanenvironmental interactions worldwide (Prance *et al.* 1987; Turner 1988; Phillips & Gentry 1993a; b; Hoffman & Gallaher 2007; Medeiros *et al.* 2011; Gaoue *et al.* 2017; 2021; Albuquerque & Alves, 2024).

The use of quantitative methods has gained momentum and proven to be informative in ethnobiological research while providing necessary frameworks for the theoretical development of the discipline (Moerman 1979; Phillips & Gentry 1993b; Garibaldi & Turner 2004; Albuquerque & Oliveira 2007; Vandebroek & Balick 2012; Gaoue *et al.* 2017; 2021; Hart *et al.* 2017; Seyler *et al.* 2019; Bond & Gaoue

Received April 02, 2024; Accepted August 20, 2024

Editor-in-Chief: Thaís Elias Almeida; Associate Editor: Taline Silva

How to cite:

Albuquerque UP, Cantalice AS, Magalhães AR, Coe MA, Gusmão RAF. 2024. Embracing Methodological Issues in Ethnobiology and Overcoming Challenges. Acta Botanica Brasilica 38: e20240085. doi: 10.1590/1677-941X-ABB-2024-0085



2020; Coe & Gaoue 2021), a critical and honest assessment of quantitative methods themselves along with their direct links to a given research question or hypothesis that researchers aim to address is essential to their adequate and appropriate use along with their refinement as the discipline continues to evolve and methodological advancements emerge over time.

Like other emerging sciences, ethnobiology is pivotal in its evolution: the challenge of self-examination and advancement in both methods and theories. This process of self-critique is crucial, especially considering integrating quantitative approaches and using the hypotheticaldeductive method in ethnobiological research (Gaoue *et al.* 2021). Such strategies represent a notable evolution in the field, reflecting a maturation and a pursuit of greater precision in its conclusions. However, this transition to quantitative and hypothetical-deductive methods also brings challenges and implications that warrant in-depth reflection.

The increasing adoption of quantitative tools began gaining popularity, promising a new era of rigor and generalization in ethnobiological research. However, this enthusiasm for quantitative techniques was not matched by an equally necessary reflection on the principles and premises that should guide these approaches (Ferreira Júnior 2020). This lack of critical reflection might lead to an undue simplification of the complex knowledge systems that Ethnobiology aims to study or to an inappropriate application of quantitative methods that do not consider the cultural and contextual specificities of the studied groups.

Therefore, in this article, we seek to initiate a critical discussion on the challenges faced by Ethnobiology in integrating quantitative approaches and the hypotheticaldeductive method, emphasizing the need for well-defined and context-sensitive principles that can guide this approach effectively. Thus, it proposes a path for advancing Ethnobiology, respecting its essence while adapting to the demands of an ever-evolving scientific field. Although this paper addresses Ethnobiology in general terms, the vast majority of examples, references, and cases discussed pertain to Ethnobotany. Among the fields of Ethnobiology, Ethnobotany has advanced most rapidly in the discussions covered in this text. For this reason, nearly everything discussed here results from the research and advancements within Ethnobotany and, in turn, can be widely applied to other fields of Ethnobiology.

Are You Comparing? Then You Need More of the Same

Ethnobiology, throughout its history, has gone through several phases, with I (Utilitarianism) and II (Cognitive Ethnobiology) being considered more descriptive (Clément 1998; Hunn 2007). Around 1970 and 1980, phase III

(Ethnoecology) emerged to overcome the limitations of the first phases by understanding the relationships between humans and nature more broadly than just descriptive (Toledo, 1992; Hunn, 2007). Since then, ethnobiology has gone through new phases: phase IV (Indigenous Ethnobiology); phase V (Interdisciplinarity in an Era of Rapid Environmental Change); and VI (Decolonizing Institutions, Projects, and Scholarship) - that changed the object of study and scale but often maintain a descriptive bias (Hunn 2007; Wyndham et al. 2011; Nabhan et al. 2011; Wolverton 2013; McAlvay et al. 2021). The maintenance of this descriptive bias in ethnobiology may be due to a lack of understanding of the importance of replication in finding general patterns or failure to carry it out due to the higher cost of energy, time, and money for more robust research. Moreover, the strong influence of non-naturalistic sciences – social sciences – on ethnobiology may make replication less relevant or challenging to apply due to the nature of their research. For example, for non-naturalistic research groups, such as those taking a phenomenological approach, the emphasis may be more on an in-depth understanding of subjective experiences and meanings, and this difference in perspective may lead to a lower appreciation of replication in specific non-naturalistic research contexts.

Replication is important to improve the generalizability of findings across diverse contexts and populations, fostering a deeper understanding of the phenomena under study (Hurlbert 1984; Leung 2015). Here, we define generalizability as the ability to extrapolate the results of a study to broader populations than the one studied. This process advances scientific knowledge and promotes transparency and accountability within the research community, as it enables the validation of methods and data, thereby upholding ethical standards. The generalizability and reliability of a study's results are key factors that drive the implication and relevance of these studies.

In this way, we might define genuine replication as the application of the same treatment (i.e., variables that determine the variable response, such as ethnic group, IDH-Human Development Index, temperature, urbanization on knowledge of local people; Figure 1) to multiple and independent sample units, while pseudoreplication occurs when incorrectly assumed independence of a sample unit (Hurlbert, 1984; Lazic, 2016). Sample independence is when the observations or elements in our sample are not dependent on each other, meaning other observations or elements do not influence them. However, the presence of pseudoreplication has been the main obstacle to the quality of ethnobiological studies, and there is often confusion in the definition between genuine replication and pseudoreplication (Giday et al. 2003; Asase et al. 2005). An example of pseudoreplication in ethnobiological studies is when individual people are used as replicates in a study, comparing the treatment in villages where the sampling or experimental unit is the village. However, the analysis

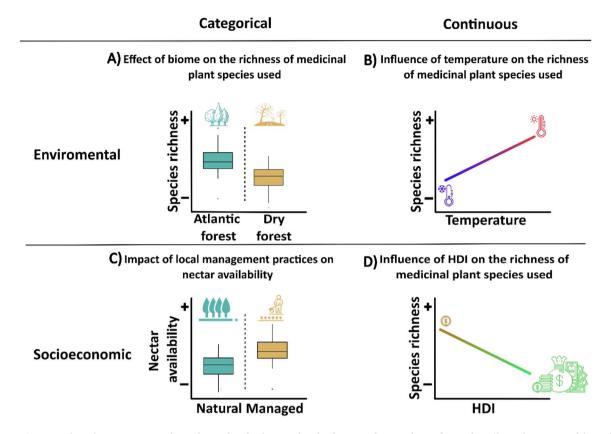


Figure 1. Examples of treatments evaluated on ethnobiology. Ethnobiology studies might evaluate the effect of categorical (A and C) or Continuous treatment (B and D) in the variables of interest, represented on the "Y" axis. The treatments are independent variables, represented on the "X" axis, and could be environmental (A and B) or socioeconomic (C and D). HDI - Human Development Index.

is performed using the individual as the unit of analysis. When using individuals from a community as a sampling unit, results are obtained about the specific community, consequently not allowing generalization to other villages with different socioeconomic and environmental conditions. Therefore, the results cannot be generalized to other villages; they only have implications for the village studied. In this way, the most appropriate way would be to sample people in different villages and analyze the villages with sampling units, allowing greater generalization of the result (Figure 1).

In studies of different areas, including ethnobiology, there are three common causes of pseudoreplication (Schank & Koehnle 2009; Davies & Gray 2015; Gaoue *et al.* 2021):

(I) Inappropriate use of individual people as replicates is the most frequent cause of pseudoreplication in ethnobiology research. Studies that compare phenomena at the community, ethnic, or village level tend to inappropriate use of individual subjects as replications. For example, sampling several people from a single village of an ethnic group and discussing the level of ethnicity is a common error because it disregards the socioeconomic and environmental differences within this ethnic group. This practice gives rise to the false determination that the subjects are independent replicates;

(II) Inadequate consideration of spatial autocorrelation-Insufficient attention is often dedicated to spatial autocorrelation in the data. Samples taken close to one another may not be independent, for example, studies that evaluate the distribution of knowledge about fauna and flora in distinct local communities. Thus, it is expected that geographically closer communities will have more remarkable species similarity than distant communities, which, by not taking into account the spatial autocorrelation between these communities, can result in pseudoreplication and lead to an overestimation of the degree of freedom as well as an inflation of the Type I error rate in statistical tests (reject the null hypothesis when it is true - find significance when there is none);

(III) Temporal pseudoreplication- This happens when data are not collected independently between experimental units due to changes in timing, which may lead to pseudoreplication. For example, data on the local knowledge of communities are obtained in a specific year, and after a period (e.g., five years), the same community is resampled. These data may not be temporally independent; if the resampling period is short, there may be an influence from previous knowledge, and even with long periods between samplings, it is impossible to be sure of this independence due to the transmission of knowledge. Furthermore, statistical tools are recommended to reduce this bias (Generalized mixed models with the sample unit like a random effect - see more details Gaoue *et al.* 2021).

Thus, it is important to evaluate each data point's independence thoroughly; consequently, genuine replication in ethnobiological studies might guarantee the validity of statistical analyses and study conclusions. The use of genuine replication in ethnobiological studies has several advantages, including increased statistical power, improved generalizability, and higher reliability of results (Lazic *et al.* 2018). On the other hand, there are also some disadvantages to using genuine replication, such as increased cost, time, and complexity of the studies (Lazic *et al.* 2018). Overall, the advantages of genuine replication in ethnobiological studies outweigh the disadvantages. Thus, it is recommended that future ethnobiology studies add genuine replications and appropriate analyses to mitigate the problems associated with replication.

The Number of Participants Matters, But So Does Who They Are

Sample size sufficiency is a central criterion in any research requiring data collection, widely discussed across various fields (Ramsey & Hewitt 2005; Martínez-Mesa *et al.* 2014; Schooling & Jones 2014; Olsen & Orr 2016; Walters 2021; Silva *et al.* 2022). Among ethnobiologists, there seems to be a belief that the choice of the minimum number of individuals in a study does not need to be supported by any protocol or statistical inference, often due to the logistical challenges of field data collection.

This approach often leads to 'black box' scenarios, where information about the criteria used to determine the study's participants is limited or completely missing. This lack of clarity ultimately compromises the validity and representativeness of the results, opportunities for meta-analyses and generalizable conclusions across study areas. Traditional or Local Ecological Knowledge, being heterogeneous and subject to variations (Berkes *et al.* 2000; Fernández-Llamazares *et al.* 2015; Mata *et al.* 2023; Santos *et al.* 2023), underscores the need for a sample that captures this diversity for a more comprehensive understanding.

The sample size is intrinsically linked to the studied population and available information, such as population size. This information determines whether sampling should be probabilistic or non-probabilistic (Gil 2008; Hibberts *et al.* 2012; Albuquerque *et al.* 2014). In an ideal scenario, probabilistic sampling is (a) random, where every individual has an equal chance of being chosen; (b) numerically representative, with a minimum number of individuals selected based on criteria like confidence level and sampling error; (c) stratified by groups, where the proportion among groups (e.g., men and women; age groups, etc.) is similar in both the sample and the studied population (Gil 2008; Hibberts *et al.* 2012; Albuquerque *et al.* 2014).

In probabilistic samples, we can estimate the sample size using formulas $n_0 = \frac{1}{E_0^2}$ and $n = \frac{N \cdot n_0}{N + n_0}$, where \mathbf{n}_0 represents the initial sample size estimate; \mathbf{E}_0 is the tolerable sampling error, commonly used in ethnobiological studies as 0.05, or 5%; N is the total population size. These formulas are used in simple random sampling (Barbetta 2012; Albuquerque et al. 2014). The following formula is used for stratified sampling: $n = \frac{(Z\alpha/2)^2 p \cdot q}{e^2}$, where **n** is the sample size; **Z**_a is the critical value corresponding to the desired confidence level; **p** is the population proportion of individuals belonging to a specific category; **q** is the proportion of individuals not belonging to the specific category (1-p); and e is the margin of error (Barbetta 2012; Albuquerque et al. 2014). Several software programs, such as G*Power (Faul et al. 2007) or R (R Development Core Team 2023), using packages like pwr (Champely et al. 2018), can be used to determine sample sizes. Online platforms also allow for sample size calculation - SurveyMonkey (https://www.surveymonkey.com/mp/ sample-size-calculator/).

Complex cultural data about the human community such as the number of hunters, midwives, local experts, etc., often makes it difficult to determine an accurate minimum number of participants. Researchers may opt for specific methodologies, like the snowball method (Albuquerque et al. 2014), to access these individuals. This approach should be combined with statistical methods, demonstrating the representativeness of the information obtained to the reader. Peroni et al. (2014) recommended using species accumulation curves (Colwell & Coddington 1994; Gotelli & Colwell 2001) to indicate the likelihood of new information emerging as more interviews are conducted. This method is often observed in studies involving hunters (Whiting et al. 2011; Oliveira et al. 2020). Various software like EstimateS (Colwell 2013) or R (R Development Core Team 2023), using packages like vegan (Oksanen et al. 2019), can be employed for these analyses.

A probabilistic approach is recommended when feasible, as it provides a more robust statistical basis for generalizations. Furthermore, the inclusion or exclusion criteria for participants play a crucial role in determining the sample size. Transparent disclosure of these criteria enhances the study's replicability and allows for evaluating the consistency of results in different contexts.

Good Theories, Good Methods

Ethnobiology is situated at the confluence of various disciplines, encompassing the complexity of human interactions with the natural environment (Delgado *et al.* 2023). This field of study consistently faces eminent challenges that demand sophisticated theoretical evolution and substantial methodological refinement. This necessity is not merely pragmatic but also epistemological, reflecting the multiplicity and diversity of human-environmental phenomena that Ethnobiology seeks to understand. Added to this are ethical, social, and philosophical challenges of a world in need of transitioning towards environmental, social, and racial justice (Albuquerque *et al.* 2024; Hanazaki 2024) along with a balanced coexistence between humans and the environment.

The relevance of a solid theoretical foundation in Ethnobiology is indisputable, serving as a guiding axis for research, interpretation of human-environmental phenomena, and application in real-world contexts amidst worldwide socio-economic inequalities driving access to natural resources. Robust theories enable the articulation of testable hypotheses and the generation of explanatory models that reflect the intricate relationships between humans and the environment (Gaoue *et al.* 2021). In this context, theory is not static but dynamic and evolutionary, mirroring the continuous interplay between empirical discoveries and conceptual reflections.

Although intrinsically subjective, qualitative research is indispensable for understanding the cultural, social, and symbolic dimensions of human-environmental phenomena of interest to ethnobiologists. Theories in qualitative research play a crucial role as they do in quantitative research, offering deep insights into human perceptions, values, and beliefs, revealing aspects often hidden in strictly quantitative analyses. Unfortunately, there seems to be little advancement in qualitative and quantitative analyses to test robust theories, as if theory and practice could not mutually nourish each other to enhance our understanding of the relationship between humans and nature and how this understanding can help tackle the challenges different peoples face, for example, the social-ecological resilience of people in the face climate and land-use change (see Delgado et al., 2023). Furthermore, to evaluate good theories effectively and to understand how different factors (e.g., socioeconomic, environmental, cultural, etc.) influence the interrelation between humans and nature, we need to adopt good methods with higher reliability.

Beyond these considerations, the incorrect or misguided use of qualitative or quantitative methods also affect the quality of ethnobiological research. In this article, we present some aspects, within the exclusive realm of quantitative research, that can advance the quality of the science we produce. Thus, we explore the integration of quantitative approaches and the hypothetical-deductive method in Ethnobiology, emphasizing challenges like oversimplification and a lack of critical reflection. We examine ecological and cultural diversity indices, sample sufficiency, replication, and the importance of spatial-temporal context in research offering practical suggestions to enhance validity, reliability, and generalizability in Ethnobiological research, aiming to preserve its essence while adapting to scientific evolution.

Ecological Indices: Use Only as Prescribed

In Ecology, several diversity indexes (e.g., Shannon, Simpson, Jaccard, and Morisita) have been used to summarize complex and multidimensional information about ecosystems, however aggregating so much information into a single index has proven problematic in several ways (Magurran 1988; Barrantes & Sandoval 2009). For example, the Shannon index summarizes the communities' characteristics, like species richness, abundance, and evenness, on a single value that becomes more difficult to understand if changes in the value of the index represent variation in which of these characteristics. However, diversity indexes have been commonly used in other areas, such as ethnobiology used mainly to understand the pattern of natural resources used by humans (Begossi 1996; Peroni et al. 2014). Consequently, ethnobiology has faced some of the same methodological challenges as ecology. The most common problems with using indexes are that they are often not comparable, do not represent reality well, and poorly summarize complex data from the study system.

For instance, when examining a medicinal plant system within a community following a disturbance, relying solely on diversity indices may not yield a comprehensive understanding of the changes that have taken place. Simply observing a shift in the Shannon index from 2.5 to 2 does not provide sufficient insight into the nature of these changes, particularly when compared to assessments of species richness and abundance. A decrease of 0.5 in the Shannon index does not accurately reflect a 20% loss in species compared to the pre-disturbance period. It fails to capture the proportional loss or gain of species, abundance, and evenness, which are crucial for understanding the dynamics of the system. This is because a decrease of 0.5 does not represent the loss of 20% of species in relation to the pre-disturbance period, that is, the decrease is not proportional. As a result, several ethnobiology studies tend to present the index values with reasonable interpretations due to the limited information provided by the indices, like a single number (Green & Chapman 2011; Daly et al. 2018). In this way, the use of single variables (i.e., species richness, abundance, and identity of species) often provides better information about the studied system than the index values.



Quantitative ethnobiological studies often incorporate diversity ecological indexes, like the Shannon and Simpson index which has been indiscriminately and mistakenly used in prior research. Ethnobiology studies need to make clear the limitations of a given method employed and the context in which they are used. The inappropriate use of indices can result in shallow and non-generalizable interpretations when presenting only the value of the index (Barrantes & Sandoval 2009; Green & Chapman 2011). For example, it is likely incorrect to say that the Shannon index is high when a value is close to 4 because values for this index tend to vary from 0 to infinity (Segnon & Achigan-Dako 2014). Thus, four is relatively low when compared to higher values like 10, 20, and 50. This shows that a study area has a specific Shannon index value that is context dependent, or comparing between study areas does not always mean that these results are generalizable to other sites or biological/ human communities.

Another example is the interpretation of the dissimilarity indices, which may be shallow and limited in their predictions. Studies using these indices present the results of whether the communities are not similar, but detecting and affirming these differences with the indices alone is impossible. For example, when evaluating the community of plants used in two local communities, it might be observed dissimilarity Bray-Curtis index next to 1, representing that both communities used are distinct, but is not possible to detect if this difference is due to changes in species richness - number of species - or identity of used plants. Consequently, studies commonly present the index values and state that the index is limited (Toledo *et al.* 2007). Therefore, the question arises: why continue using these indices?

Therefore, understanding the limitations of the indices and seeking alternatives are essential for the appropriate use of the indices and for advancing the understanding of the complexity of ecosystems. The main challenge in ecology and ethnobiology tends to follow the same path: finding alternative paths to indexes that allow us to summarize the complexity of ecosystems without losing explanatory power. Recent studies that evaluate the efficiency of the diversity index in Ecology have shown that these indexes are insensitive to species differences and abundance (Izsák & Papp 2000; Ricotta & Avena 2003; Lamb et al. 2009). Moreover, the diversity indexes do not provide good information about environmental changes, like climate and land-use changes (Thiebaut et al. 2002; Lamb et al. 2009). The following are the principal objections to the diversity indices used in ecological and ethnobiological research (Barrantes & Sandoval 2009):

(I) Failure to recognize intraspecific differences within populations, such as age classes or sexes, results in loss of information. For example, in ethnobiological research, these differences might influence knowledge strategies, social hierarchies, foraging behaviors, and responses to the environment;

(II) most indexes are highly sensitive to sample size because they depend on the number of observed species. This sensitivity is due to nearly all diversity indices incorporating the number of species (S) as a fundamental component in their calculations, making them particularly sensitive to changes in sample size that are correlated with the number of species;

(II) the indexes ignored the heterogeneous sampling units. Thus, the lack of care can lead to using variables with different sampling units, for example, using species abundance and vegetation cover simultaneously and as if they were the same unit of measurement. The indices did not detect these differences;

(IV) It allows only limited insights into communities and evolutionary patterns. The use of indexes is only to describe, and it is not comparable because assigning a probability value to an index is impossible.

Alternatively, to remedy some of these challenges, adopting multivariate analysis and model-based approaches can allow a greater understanding of the drivers of humanenvironmental interactions in a given study system more adequately. For example, understanding the effect of environmental change on the use of medicinal plants by local people can be assessed more adequately by using explicit variables, like plant species richness and abundance, rather than diversity indexes of used plants (Barrantes & Sandoval, 2009). Explicit variables allow a mechanistic and clear comprehension of communities' responses to environmental changes. Furthermore, we recommended that future studies in ethnobiology continue to consider the methodological advances in ecology and ethnobiology while adopting new model-based approaches, and the use of explicit variables (e.g., the use of piecewise and hierarchical models allowing multiple dependent and independent variables simultaneously - see more details Gaoue et al. 2021).

In summary, we recommend employing multivariate analysis and model-based approaches instead of relying solely on diversity indices for a more comprehensive understanding of the drivers of human-environmental interactions. Clearly report the specific variables used in your studies, such as species richness and abundance, to provide a more detailed and accurate representation of the study system. Additionally, acknowledge and discuss the limitations of the indices used and the specific contexts in which they are applicable to avoid misinterpretation and overgeneralization.

Ethnobiological Indices Do Not Necessarily Make Your Work More Attractive or Robust

The use of cultural importance indices to quantify the level of importance of a given plant species to a particular cultural group has become widespread, often with aims to understand better medicinal plant use and selection, species use, economic, and ethnotaxonomic values, and aid in defining conservation priorities (Turner 1988; Stoffle et al. 1990; Phillips & Gentry 1993a; Bennett & Prance 2000; Pieroni 2001; Garibaldi & Turner 2004; Reyes-García et al. 2006; Silva et al. 2006; Hoffman & Gallaher, 2007; Tardío & Pardo-De-Santayana 2008; Thomas et al. 2009; Franco et al. 2014; Tudela-Talavera et al. 2015; Mateo-Martín et al. 2023). While studies seeking to quantify the cultural importance of plant taxa often aim to do so by defining a particular set of unique variables that comprise a given index proposed to provide a replicable and objective measure of species cultural values, significant limitations to these approaches hinder the ability of these indices to provide a generalizable methodology that can be confidently applied to ensure appropriate and adequate estimates of cultural importance over time, various study systems and geographic ranges. First, there lacks a clear, concise, and operational definition of species' cultural importance in ethnobiology beyond the general statement, "the importance of the role that a plant species plays in a given culture." This brings into question, "What specific roles does a species have to fulfill within a given society to be culturally important, and to what extent?"

In a foundational study, Turner (1988) asked fundamental questions on how species cultural importance could be documented and compared in a meaningful, systematic, and quantifiable way while building on the work of her mentors (Berlin et al. 1973; Hunn 1982) and highlighting the complexities of such an undertaking. However, only some ethnobiological studies have continued to pose these essential questions on what specific factors or criteria adequately define species' cultural importance while systematically testing predictions to reach a consensus among researchers and a generalizable conclusion. Instead, numerous studies arbitrarily propose a given quantitative index assuming it estimates specy's cultural importance with little to no theoretical support for the choice or inclusion of specific variables and discussion on why they are adequate to provide a rigorous, generalizable, and unbiased estimate. Second, most studies aiming to quantify cultural importance define variables of a given index primarily from the researcher's perspective and need to incorporate adequate metrics of cultural importance defined from the participant's emic perspective. It has been suggested that adequate evaluations of cultural importance should be done by Indigenous peoples and local communities themselves (Turner 1988). As such, there are inherent biases driven by the researcher's values included in estimates of species' cultural importance from the etic perspective regardless of whether a variable for participant consensus is included as a metric (see Silva et al. 2006) along with variables defined, ranked, or estimated from the researchers themselves (Tardío & Pardo-De-Santayana 2008; Thomas et al. 2009). This topic has been discussed in the literature (Coe & Gaoue 2020) and warrants further research and honest discussions to determine if the quantification of species' cultural importance can be adequately estimated from the emic or etic perspectives. Third, there has been a noticeable trend for ethnobiologists to propose a novel index based on either current study objectives or aims to improve upon existing indices that have been criticized for limitations (e.g., for lacking a given variable or for incorporating researcher subjectivity) (Stoffle et al. 1990; Pieroni 2001; Reyes-García et al. 2006; Tardío & Pardo-De-Santayana 2008; Franco et al. 2014; Tudela-Talavera et al. 2015).

While innovation has been an integral force at the forefront of methodological advancements developed from and built upon the entire body of ethnobiological research, it is important for researchers to clearly understand and consider the assumptions of a given index concerning appropriate methods employed for study questions being addressed or hypotheses being tested (Hoffman & Gallaher 2007). In a thorough examination of 12 indices used to predict species' cultural importance and identify cultural keystone species, Coe and Gaoue (2020) found that most indices were correlated, meaning that most importance indices contain similar variables suggesting that the species' cultural values aim to quantify were repetitive and most often not unique. Given that these indices were also shown to perform poorly in predicting species cultural values (Coe & Gaoue 2020) within the cultural keystone species framework (Garibaldi & Turner 2004), it is important to consider their appropriate use to avoid unreliable and ungeneralizable data interpretation (Medeiros et al. 2011).

As the emphasis on quantification and hypothesis testing in ethnobiology continues, it is important to ask if ethnobiologists need an index to quantify species' cultural importance. As noted in the seminal paper by Turner (1988), who provided the first quantifiable index to estimate species' cultural importance, and asked a knowledgeable elder participant directly, "Which plants are important?" she replied, "I would pick them all – they are all important."

In summary, we recommend establishing clear and operational definitions for cultural importance and the specific roles that species fulfill within a society. It is important to ensure that cultural importance indices include metrics defined from the participants' perspectives to reduce researcher bias (see Sousa *et al.* 2024). Additionally, critically evaluate the need for new indices and their theoretical support to ensure they provide meaningful and generalizable estimates.



Where Are We? When Are We?

Most ethnobiological studies are conducted at local scales, aiming to address issues related to the knowledge or use of natural resources. However, these studies often neglect spatial and temporal information, resulting in biases that impede the findings' understanding, replication, and generalization.

Spatial biases are linked to the absence of specific spatial information about the study location. Often, studies provide generic descriptions of the locality, such as "The study was conducted in São Desidério, Bahia, Brazil" with minor variations including the name of the community "The study was conducted in the Licuri community, in the municipality [...]" or the central coordinate of the municipality "The study [...], 12° 21' 8"S, 44° 59' 3"W." The issue with this information is that it does not precisely provide the exact location of the community; this is the specific geographical position. In this example, the community could be located anywhere within the ~15,200 km² of the locality.

The sociocultural, environmental, and political context in which the investigated communities are embedded plays an equally crucial role in constructing a comprehensive spatial overview for the reader. For example, in the context of ethnobotanical research aimed at understanding the use of medicinal species, the presence of information such as the availability of modern health services (Saynes-Vásquez *et al.* 2016; Weckmüller *et al.* 2019), the dynamics of different land use patterns (Kunwar *et al.* 2015; Arjona-García *et al.* 2021), and cultural context (Kidane *et al.* 2014) are elements that often explain the dynamics of medicinal plant knowledge. Without this information, the reader might accept the presented results without fully understanding the scenario in which the community is embedded.

Among the spatial biases discussed, we can classify the community's location as essential in all studies. Other information is relevant in specific contexts and should be adjusted according to the objectives of each research, emphasizing the socio-environmental, cultural, and political contexts in which Indigenous Peoples and Local Communities are situated, as these elements can influence the interpretation of results.

Temporal biases, in turn, are linked to the lack of temporal information during the study. The presence of information indicating the period in which the data were collected, such as "Data was collected between April/2023 and July/2023" becomes crucial. This precision becomes even more critical when considering the dynamic nature of traditional or local ecological knowledge over time. For example, Sousa *et al.* (2022) observed significant changes in the most popular plants in a rural community in Northeast Brazil over two years.

Furthermore, studies with clear spatial and temporal information provide a broader range of data, enabling

greater robustness and eligibility in systematic reviews and meta-analyses. For example, Santoro et al. (2017) tested whether the pharmacopoeias of African populations - broad spatial scales - incorporated more medicinal species as the incidence of malaria increased, which was not corroborated. However, due to the use of temporal data - articles with different publication years - it was possible to understand that the stability of pharmacopoeias may be related to public interventions aimed at controlling malaria, where pharmacopoeias do not need to increase the number of species. In this sense, when conducting ethnobiological studies, it is essential to provide clear spatial and temporal information to ensure a more complete and contextualized understanding of the studied communities that seek to understand the relationship between human beings and biodiversity. By addressing these aspects, researchers can build a more solid scientific foundation, promoting significant advances in understanding and addressing ethnobiological issues.

The Quantity of Ingredients is as Important as the Ingredients Themselves

Researchers frequently overlook important information like the mean or median value of their samples, the spread of the data (standard deviation), and whether any unusual data points - outliers - were excluded (Zuur et al. 2010; Gerstner *et al.* 2017). Highlighting the means or medians of samples can provide insights into the central tendency of the dataset. At the same time, the standard deviation shows how spread out the data points are from the average. For example, after interviewing individuals from a community, we calculated that, on average, each person used seven medicinal plants. While researchers often stop after pointing out the mean of a sample in the results section, they can offer more pertinent details. For instance, the average number of cited species was 7, with a standard deviation of ±3.5 ranging from 3 to 15 species. This gives a general idea of the data obtained and gives readers a clearer idea of the sample discussed. Neglecting to report such details hampers the study's reproducibility and impedes transparency in understanding the primary results.

Another common but lesser problem facing data transparency is the need for more information about outliers, whether the author removed them from the samples or kept them (Zuur *et al.* 2010). An outlier is a data point representing an observation that differs significantly from other observations. It can happen for many reasons, such as measurement errors or conditions that differ from norm (Kelley & Preacher 2012). Sometimes, an outlier can be a genuine great variation in the studied phenomenon. For instance, encountering an elderly individual who is a specialist or generally well-versed in medicinal herbs during interviews could introduce outliers compared to the sampled population. These outliers can skew averages and affect statistical evaluations, particularly those relying on mean-based analyses like linear models. Genuine outliers are often kept in the sample; however, genuine outliers are often difficult to tell apart from measurement errors. Whether it be the decision of the ethnobiologist, it is a transparent and reproducible practice to inform why the decision better fits the research goals. Therefore, providing clarity on handling outliers ensures the integrity and accuracy of the research findings.

Similarly, when testing a hypothesis, it is common for researchers to prioritize p-value reports over the effect sizes in analysis and hypothesis testing (Popovic et al., 2024). P-values indicate the probability of observing the data or more extreme ones, assuming the null hypothesis is true (Kelley & Preacher 2012; Gerstner et al. 2017). A small p-value, typically less than 0.05, suggests that the observed data is unlikely to have occurred by chance alone (under the null hypothesis), providing strong evidence supporting the alternative hypothesis and representing the phenomenon being tested. The effect sizes, in turn, represent the magnitude of the relationship between dependent variables (those that are measured, such as knowledge of medicinal species) and independent variables (those that are used to explain the knowledge, such as age or gender) (Kelley & Preacher 2012; Rosenberg et al. 2013). In systematic reviews and meta-analyses, effect sizes are standardized for comparison across different studies, even when study designs differ (Rosenberg et al. 2013; O'Dea *et al.* 2021). Interpreting p-values and effect sizes helps the researcher validate the studied phenomenon and its magnitude.

Moreover, these values provide insight into the strength of a particular phenomenon, including coefficients in a linear model, differences between means (common in ethnobiology studies), correlation coefficients, and more (Kelley & Preacher 2012; O'Dea et al. 2021). For example, two distinct studies investigate whether there are differences in the knowledge of medicinal species among populations living in urban and rural contexts. In both studies, the authors were limited to reporting the following statement: - "Our study demonstrated that, compared to urban populations, rural populations know more medicinal species (p<0.05)". However, upon observing the effect, we noticed that, on average, one population knew 5.5 more plants than the other, whereas in the other study, the average difference was only 1.5 plants. This information facilitates the reader's interpretation of the study's main message and the search for explanations for such discrepancies.

Additionally, consider a linear regression example: when testing a hypothesis on whether the diversity of medicinal plants responds to an environmental gradient (e.g., urbanization, altitude, proximity to main roads) and obtaining a p-value of 0.04 and an effect size of 0.19 (in this case, represented by a linear coefficient β) strongly suggests that the environmental gradient correlates with the response variable – in this case, the diversity of species declared by individuals. In linear regression, the linear coefficient is the value that multiplies the predictor variable (consider $Y = \beta x + a$, the linear coefficient would be β) and gives an idea of the magnitude of how much independent variables (x) affect the dependent variables (Y). In our example, this can be interpreted as every one-unit change in the predictor variable (urbanization, altitude, proximity to main roads) corresponds to a 0.19 change in the number of species. Therefore, solely reporting p-values may be insufficient for fully interpreting results (Popovic *et al.* 2024) and should be accompanied by effect sizes that are crucial for theoretical understanding and conveying the magnitude of a statistically significant phenomenon.

Similarly, graphs depicting various approaches, such as barplots, boxplots, and scatter plots, often need more detail, mirroring sample deficiencies (Gerstner *et al.* 2017). For instance, barplots and boxplots are frequently presented without indicating means or medians, maximum and minimum values, and even without explicitly specifying the measures depicted on the axes (e.g., the number of species, biomass (g), temperature (°C), etc.; see Table 1) and their respective units. Moreover, whether the unit of measure has undergone any transformation, such as log transformations, remains unclear. This presents a significant challenge as it not only diminishes the reproducibility of the study but also impedes the comprehension of the results, making it difficult to grasp the main ideas of the study (Gerstner *et al.* 2017).

Furthermore, improving reporting mean or median values, addressing outliers, and providing standard deviation and effect sizes are important for replicability and help validate the research conducted by ethnobiologists. However, considering the limited word count and space typically available in research articles, it is advisable to include these details in supplementary materials and make the article's metadata available for readers and potential researchers conducting reviews. Thus, adopting these practices enhances the probability of the research being included in a systematic review or meta-analysis, thereby increasing its impact (Gerstner *et al.* 2017; Popovic *et al.* 2024). Consequently, even a small additional effort in handling challenging data can yield significant rewards.

Future Directions

As we look ahead in Ethnobiology, several key directions emerge for future research. Firstly, developing integrated methodologies that seamlessly blend quantitative tools with the rich, qualitative roots of Ethnobiology is paramount. This approach would maintain the depth of traditional knowledge and enhance it with the precision of quantitative analysis.

| Thematic | Common Problems | How to Solve Them |
|---|---|---|
| Mean or median, standard deviation, and maximum and minimum values | Lack of details in reporting sample characteristics | Provide sample characteristics, including key statistics and range. |
| | "[] the average number of medicinal plant species known in the community is seven species." | "[] the number of known medicinal species ranges from 3 to 15 (7 mean ± 3.5 SD species)." |
| Effect sizes | Prioritization of p-value reporting over effect sizes | Emphasize the importance of reporting effect sizes alongside p-values for a comprehensive understanding of relationships between variables. |
| | "[] urbanized and rural communities differed significantly (P<0.05)." | "[] Rural areas had an average of 3.2 more species than urban areas (P=0.032), indicating a significant difference between the two." |
| Graphical communication | Lack of detail and clarity in graph representation | Enhance graph clarity by including essential details like standard deviation and axis labels complete with units (e.g., °C, mm, etc.), ensuring transparency in data representation. The legends should encompass adequate information to ensure the comprehension of the graph independently from associated textual explanations. |
| | + Medicinal plant diversity - rural urban | Medicinal plant diversity (Community mean) rural urban |

Table 1. A Synthesis of Principal Thematic Problems and Solutions in Data Reporting and Analysis

Secondly, critically evaluating the tools and indices currently in use is essential. This involves thoughtful examination of ecological and ethnobiological indices and exploring alternatives that can adequately estimate the intricacies of ecosystems and cultural importance without oversimplification.

Thirdly, the issue of sample representation and replication in studies needs to be addressed with greater rigor. Establishing robust sampling strategies that accurately reflect the diversity within communities and ensuring replication will enhance the reliability and applicability of research findings.

Moreover, greater attention should be given to ethnobiological studies' spatial-temporal context. Detailed reporting of geographical and temporal data is crucial for a comprehensive understanding of ecological knowledge and practices' dynamic nature.

Additionally, enhancing data transparency and reporting standards is vital for the field's progress. This includes

providing detailed information on sample characteristics, the handling of outliers, and effect sizes, which would improve the reproducibility and validity of research findings.

Lastly, embracing new technologies and analytical tools to understand and visualize complex ethnobiological data will be a step forward in this evolving field. By addressing these future directions, Ethnobiology can continue to grow as a discipline that respects its qualitative heritage and thrives on methodological innovation and scientific rigor.

Acknowledgments

RAFG and ARM thank the National Council for Scientific and Technological Development (CNPQ) for their financial support, provided through n° 381253/2024-4 and n° 380461/2024-2, respectively. ASC extends sincere appreciation to the Coordination for the Improvement of Higher Education Personnel (CAPES), code n° 001.

Authors' Contribution

Ulysses Albuquerque: Conceptualization; All authors: Writing Original draft, Writing- Reviewing and Editing,

Conflicts of Interest

The authors have no conflicts of interest to declare.

References

Albuquerque UP, Alves RRN. 2024. Integrating depth and rigor in ethnobiological and ethnomedical research. Journal of Ethnobiology and Ethnomedicine 20: 6. doi: 10.1186/s13002-023-00643-y

Albuquerque UP, Maroyi A, Ladio AH *et al.* 2024. Advancing ethnobiology for the ecological transition and a more inclusive and just world: A comprehensive framework for the next 20 years. Journal of Ethnobiology and Ethnomedicine 20: 18. doi: 10.1186/s13002-024-00661-4

Albuquerque UP, Lucena RFP, Lins Neto EMF. 2014. Selection of Research Participants. In: Albuquerque UP, Lucena RFP, Cunha LVFC (orgs.). Methods and Techniques in Ethnobiology and Ethnoecology. New York, Springer Protocols Handbooks. p. 1-14. doi: 10.1007/978-1-4614-8636-7_1 p. 255-276

Albuquerque UP, Oliveira RF. 2007. Is the use-impact on native caatinga species in Brazil reduced by the high species richness of medicinal plants? Journal of Ethnopharmacology 113: 156-170. doi: 10.1016/j. jep.2007.05.025

Arjona-García C, Blancas J, Beltrán-Rodríguez L *et al.* 2021. How does urbanization affect perceptions and traditional knowledge of medicinal plants? Journal of Ethnobiology and Ethnomedicine 17: 1-26. doi: 10.1186/s13002-021-00473-w

Asase A, Oteng-Yeboah AA, Odamtten GT, Simmonds MSJ. 2005. Ethnobotanical study of some Ghanaian anti-malarial plants. Journal of Ethnopharmacology 99: 273-279. doi: 10.1016/j.jep.2005.02.020

Barbetta PA. 2012. Técnicas de Amostragem. In: Barbetta PA. Estatística aplicada às Ciências Sociais. 8nd. edn. Florianópolis, Editora da UFSC. p. 41-64.

Barrantes G, Sandoval L. 2009. Conceptual and statistical problems associated with the use of diversity indices in ecology. Revista De Biologia Tropical 57: 451-460. doi: 10.15517/rbt.v57i3.5467

Begossi A. 1996. Use of ecological methods in ethnobiology: Diversity indices. Economic Botany 50: 280-289. doi:10.1007/BF02907333

Bennett BC, Prance GT. 2000. Introduced Plants in the Indigenous Pharmacopoeia of Northern South America. Economic Botany 54: 90-102. doi: 10.1007/BF02866603

Berkes F, Colding J, Folke C. 2000. Rediscovery of traditional ecological knowledge as adaptive management. Ecological Applications 10: 1251-1262. doi: 10.1890/1051-0761(2000)010[1251:ROTEKA]2.0.CO;2

Berlin B, Breedlove DE, Laughlin RM, Raven PH. 1973. Cultural significance and lexical retention in Tzeltal-Tzotzil ethnobotany. In: Edmonson MS (eds.). Meaning in Mayan Languages: Ethnolinguistic Studies. Texas, Mouton. p. 143-164. doi: 10.1515/9783110869675.143

Bond MO, Gaoue OG. 2020. Prestige and homophily predict network structure for social learning of medicinal plant knowledge. PLoS ONE 15: 1-27. doi: 10.1371/journal.pone.0239345

Champely S, Ekstrom C, Dalgaard P *et al.* 2018. Power analysis functions along the lines of Cohen (1988). https://cran.r-project.org/web/packages/. 14 november 2022.

Clément D. 1998. The Historical Foundations of Ethnobiology (1860-1899). Journal of Ethnobiology 18: 161-187. Coe MA, Gaoue OG. 2020. Most cultural importance indices do not predict species cultural keystone status. Human Ecology 48: 721-732. doi: 10.1007/ s10745-020-00192-y

Coe MA, Gaoue OG. 2021. Phylogeny explains why less therapeutically redundant plant species are not necessarily facing greater use pressure. People and Nature 3: 770-781. doi: 10.1002/pan3.10216

Colwell RK, Coddington JA. 1994. Estimating terrestrial biodiversity through extrapolation. Philosophical Transactions of the Royal Society of London 345: 101-118. doi: 10.1098/rstb.1994.0091

Colwell RK. 2013. EstimateS: Statistical estimation of species richness and shared species from samples. Version 9 and earlier. https://www.robertkcolwell.org/pages/1407-estimates. 01 april 2024.

Daly AJ, Baetens JM, Baets B. 2018. Ecological Diversity: Measuring the Unmeasurable. Mathematics 6: 119. doi:10.3390/math6070119

Davies GM, Gray A. 2015. Don't let spurious accusations of pseudoreplication limit our ability to learn from natural experiments (and other messy kinds of ecological monitoring). Ecology and Evolution 5: 5295-5304. doi: 10.1002/ece3.1782

Delgado AN, Ludwig D, El-Hani C. 2023. Pluralist ethnobiology: between philosophical reflection and transdisciplinary action. Journal of Ethnobiology 43: 191-197. doi: 10.1177/02780771231194774

Faul F, Erdfelder E, Lang AG, Buchner A. 2007. G*Power 3: A Flexible Statistical Power Analysis Program for the Social, Behavioral, and Biomedical Sciences. Behavior Research Methods 39: 175-191. doi:10.3758/ BF03193146

Fernández-Llamazares Á, Díaz-Reviriego I, Luz AC, Cabeza M, Pyhälä A, Reyes-García V. 2015. Rapid ecosystem change challenges the adaptive capacity of Local Environmental Knowledge. Global Environmental Change 31: 272-284. doi: 10.1016/j.gloenvcha.2015.02.001

Ferreira Júnior WS. 2020. Reflections on the theoretical advance in ethnobiology: Are we pointing to the wrong direction? Ethnobiology and Conservation 9. doi: 10.15451/ec2020-05-9.20-1-8

Franco FM, Ghani BAA, Hidayati S. 2014. Terras (Eusideroxylon zwageri Teijsm. & Binn.), a Cultural Keystone Species of the Berawan People of Sarawak, Malaysia. Social Sciences & Humanities 22: 891-902.

Gaoue OG, Coe MA, Bond M, Hart G, Seyler, BC, McMillen H. 2017. Theories and Major Hypotheses in Ethnobotany. Economic Botany 71: 269-287. doi: 10.1007/s12231-017-9389-8

Gaoue OG, Moutouama JK, Coe MA *et al.* 2021, Methodological advances for hypothesis-driven ethnobiology. Biological Reviews 96: 2281-2303. doi: 10.1111/brv.12752

Garibaldi A, Turner N. 2004. Cultural keystone species: Implications for ecological conservation and restoration. Ecology and Society 9: art1. doi: 10.5751/ES-00669-090301

Gerstner K, Moreno-Mateos D, Gurevitch J *et al.* 2017. Will your paper be used in a meta-analysis? Make the reach of your research broader and longer lasting. Methods in Ecology and Evolution 8: 777-784. doi: 10.1111/2041-210X.12758

Giday M, Asfaw Z, Elmqvist T, Woldu Z. 2003. An ethnobotanical study of medicinal plants used by the Zay people in Ethiopia. Journal of Ethnopharmacology 85: 43-52. doi: 10.1016/S0378-8741(02)00359-8

Gil AC. 2008. Amostragem na Pesquisa Social. In: Gil AC. Métodos e Técnicas de Pesquisa Social. 6nd. edn. São Paulo, Atlas. p. 89-99

Gotelli NJ, Colwell RK. 2001. Quantifying biodiversity: Procedures and pitfalls in the measurement and comparison of species richness. Ecology Letters 4: 379-391. doi: 10.1046/j.1461-0248.2001.00230.x

Green R, Chapman PM. 2011. The problem with indices. Marine Pollution Bulletin. 62: 1377-1380. doi:10.1016/j.marpolbul.2011.02.016

Hanazaki N. 2024. Local and traditional knowledge systems, resistance, and socioenvironmental justice. Journal of Ethnobiology and Ethnomedicine 20: 5. doi: 10.1186/s13002-023-00641-0

Hart G, Gaoue OG, De La Torre *et al.* 2017. Availability, diversification and versatility explain human selection of introduced plants in Ecuadorian traditional medicine. PLoS ONE 12: 1-16. doi: doi.org/10.1371/journal. pone.0184369

Hibberts M, Johnson RB, Hudson K. 2012. Common survey sampling techniques. In: Gideon L (org.). Handbook of Survey Methodology for the Social Sciences. Springer, New York. p. 53-74. doi: 10.1007/978-1-4614-3876-2_5

Hoffman B, Gallaher T. 2007. Importance indices in ethnobotany. Ethnobotany Research and Applications 5: 201-218. doi: 10.17348/ era.5.0.201-218

Hunn E. 1982. The utilitarian factor in folk biological classification. American Anthropologist 84: 830-847. doi: 10.1525/aa.1982.84.4.02a00070

Hunn E. 2007. Ethnobiology in Four Phases. Journal of Ethnobiology 27: 1-10. doi: 10.2993/0278-0771(2007)27[1:EIFP]2.0.CO;2

Hurlbert SH. 1984. Pseudoreplication and the Design of Ecological Field Experiments. Ecological Monographs 54: 187-211. doi:10.2307/1942661

Izsák J, Papp L. 2000. A link between ecological diversity indices and measures of biodiversity. Ecological Modeling 130: 151-156. doi:10.1016/S0304-3800(00)00203-9

Kelley K, Preacher KJ. 2012. On effect size. Psychological methods 17: 137. doi: 10.1037/a0028086

Kidane B, Van Andel T, Van Der Maesen LJG, Asfaw Z. 2014. Use and management of traditional medicinal plants by Maale and Ari ethnic communities in southern Ethiopia. Journal of Ethnobiology and Ethnomedicine 10: 46. doi: 10.1186/1746-4269-10-46

Kunwar RM, Acharya RP, Chowdhary CL, Bussmann RW. 2015. Medicinal plant dynamics in indigenous medicines in farwest Nepal. Journal of Ethnopharmacology 163: 210-219. doi: 10.1016/j.jep.2015.01.035

Lamb EG, Bayne E, Holloway G *et al.* 2009. Indices for monitoring biodiversity change: Are some more effective than others? Ecological Indicators 9: 432-444. doi:10.1016/j.ecolind.2008.06.001

Lazic SE. 2016. Experimental design for laboratory biologists: Maximising information and improving reproducibility. United Kingdom, Cambridge University Press.

Lazic SE, Clarke-Williams CJ, Munafo MR. 2018. What exactly is 'N' in cell culture and animal experiments? PLoS Biology 16: e2005282. doi:10.1371/journal.pbio.2005282

Leung L. 2015. Validity, reliability, and generalizability in qualitative research. Journal of Family Medicine and Primary Care 4: 324-327. doi: 10.4103/2249-4863.161306

Magurran AE. 1988. Ecological diversity and its measurement. Princeton, Princeton University.

Martínez-Mesa J, González-Chica DA, Bastos JL, Bonamigo RR, Duquia RP. 2014. Sample size: How many participants do I need in my research? Anais Brasileiros de Dermatologia 89: 609-615. doi: 10.1590/abd1806-4841.20143705

Mata PT, Oliveira ARS, Arnan X, Reyes-García V, Silva TC. 2023. Teenagers' ecological knowledge about dry forests in Northeastern Brazil: Theoretical and practical implications in ethnobiology. SN Social Sciences 3: 58. doi: 10.1007/s43545-023-00636-4

Mateo-Martín J, Benítez G, Gras A *et al.* 2023. Cultural importance, availability and conservation status of Spanish wild medicinal plants: Implications for sustainability. People and Nature 5: 1512-1525. doi: doi. org/10.1002/pan3.10511

MCalvay AC, Armstrong CG, Baker J *et al.* 2021. Ethnobiology Phase VI: Decolonizing Institutions, Projects, and Scholarship. Journal of Ethnobiology 41: 170-191.

Medeiros MFT, Silva OS, Albuquerque UP. 2011. Quantification in ethnobotanical research: An overview of indices used from 1995 to 2009. Sitientibus Série Ciências Biológicas 11: 211-230.

Moerman DE. 1979. Symbols and selectivity: A statistical analysis of Native American medical Ethnobotany. Journal of Ethnopharmacology 1: 111-119. doi: doi.org/10.1016/0378-8741(79)90002-3

Nabhan GP, Wyndham F, Lepofsky D. 2011. Ethnobiology for a Diverse World Ethnobiology Emerging from a Time of Crisis. Journal of Ethnobiology 31: 172-175.

O'Dea RE, Lagisz M, Jennions MD *et al.* 2021. Preferred reporting items for systematic reviews and meta-analyses in ecology and evolutionary biology: A PRISMA extension. Biological Reviews 96: 1695-1722. doi: doi.org/10.1111/brv.12721

Oksanen J, Blanchet FG, Friendly M *et al.* 2019. Package 'vegan'. Community ecology package. https://cran.r-project.org/web/packages/vegan/index. html. 28 august 2024.

Oliveira WSL, Borges AKM, Lopes FS, Vasconcellos A, Alves RRN. 2020. Illegal trade of songbirds: An analysis of the activity in an area of northeast Brazil. Journal of Ethnobiology and Ethnomedicine 16: 16. doi: 10.1186/ s13002-020-00365-5

Olsen RB, Orr LL. 2016. On the "where" of social experiments: Selecting more representative samples to inform policy. New Directions for Evaluation 2016: 61-71. doi: 10.1002/ev.20207

Peroni N, Araujo HFP, Hanazaki N. 2014. Ecological Methods in Ethnobotanical and Ethnobiological Research: Using Diversity Measurements and Richness Estimators. In: Albuquerque UP, Lucena, RFP, Cunha LVFC, Alves, RRN (eds.). Methods and Techniques in Ethnobiology and Ethnoecology. New York, Springer Protocols Handbooks. doi: 10.1007/978-1-4614-8636-7_25

Phillips O, Gentry AH. 1993a. The useful plants of Tambopata, Peru: I. Statistical hypotheses tests with a new quantitative technique. Economic Botany 47: 15-32. doi: 10.1007/BF02862203.

Phillips O, Gentry AH. 1993b. The useful plants of Tambopata, Peru: II. Additional hypothesis testing in quantitative ethnobotany. Economic Botany 47: 33-43. doi: doi.org/10.1007/BF02862204

Pieroni A. 2001. Evaluation of the cultural significance of wild food botanicals traditionally consumed in Northwestern Tuscany, Italy. Journal of Ethnobiology 21: 89-104.

Popovic G, Mason TJ, Drobniak SM *et al.* 2024. Four principles for improved statistical ecology. Methods in Ecology and Evolution 15: 266-281. doi: 10.1111/2041-210X.14270

Prance GT, Balee W, Boom BM, Carneiro RL. 1987. Quantitative Ethnobotany and the Case for Conservation in Amazonia. Conservation Biology 1: 296-310. doi: 10.1111/j.1523-1739.1987.tb00050.x

R Development Core Team. 2023. R a language and environment for statistical computing. Vienna, Austria, R Foundation for Statistical Computing.

Ramsey CA, Hewitt AD. 2005. A methodology for assessing sample representativeness. Environmental Forensics 6: 71-75. doi: 10.1080/15275920590913877

Reyes-García V, Huanca T, Vadez V, Leonard W, Wilkie D. 2006. Cultural, practical, and economic value of wild plants: A quantitative study in the Bolivian Amazon. Economic Botany 60: 62-74. doi: 10.1663/0013-0001(2006)60[62:CPAEVO]2.0.CO;2

Ricotta C, Avena G. 2003. On the relationship between Pielou's evenness and landscape dominance within the context of Hill's diversity profiles. Ecological Indicators 2: 361-365. doi: 10.1016/S1470-160X(03)00005-0

Rosenberg MS, Rothstein HR, Gurevitch J. 2013. Effect sizes: Conventional choices and calculations. In: Koricheva J, Gurevitch J, Mengersen K (eds.). Handbook of meta-analysis in ecology and evolution. Princeton, Princeton University Press. p. 61-71. doi: 10.23943/ princeton/9780691137285.003.0006

Santoro FR, Santos GC, Ferreira Júnior WS *et al.* 2017. Testing an ethnobiological evolutionary hypothesis on plant-based remedies to

treat malaria in Africa. Evolutionary Biology 44: 216-226. doi: 10.1007/s11692-016-9400-9

Santos FIS, Medeiros MJL, Morais RF, Lopes CGR. 2023. Sociodemographic factors influencing environmental perception and local ecological knowledge in forests of different ages in northeastern Brazil. Ethnobotany Research and Applications 26: 1-13. doi: 10.32859/era.26.72.1-13

Saynes-Vásquez A, Vibrans H, Vergara-Silva F, Caballero J. 2016. Intracultural differences in local botanical knowledge and knowledge loss among the Mexican isthmus Zapotecs. PLoS ONE 11: e0151693. doi: 10.1371/journal.pone.0151693

Schank JC, Koehnle TJ. 2009. Pseudoreplication is a pseudoproblem. Journal of Comparative Psychology 123: 421-433. doi: 10.1037/a0013579

Schooling CM, Jones HE. 2014. Is representativeness the right question? International Journal of Epidemiology 43: 631-632. doi: 10.1093/ije/ dyt264

Segnon AC, Aachigan-Dako EG. 2014. Comparative analysis of diversity and utilization of edible plants in arid and semi-arid areas in Benin. Journal of Ethnobiology and Ethnomedicine 10: 80. doi: 10.1186/1746-4269-10-80

Seyler BC, Gaoue OG, Tang Y, Duffy DC. 2019. Understanding knowledge threatened by declining wild orchid populations in an urbanizing China (Sichuan). Environmental Conservation 46: 318-325. doi: 10.1017/S0376892919000171

Silva JG, Caetano RA, Silva RRV, Medeiros PM. 2022. Sampling Bias in Ethnobotanical Studies on Medicinal Plants Conducted in Local Markets. Journal of Ethnobiology 42: 20-30. doi: 10.2993/0278-0771-42.1.20

Silva VA, Andrade LHC, Albuquerque UP. 2006. Revising the Cultural Significance Index : The Case of the Fulni-ô in Northeastern Brazil. Field Methods 18: 98-108. doi: doi.org/10.1177/1525822X05278025

Sousa DCP, Ferreira Júnior WS, Albuquerque UP. 2022. Short-term temporal analysis and children's knowledge of the composition of important medicinal plants: The structural core hypothesis. Journal of Ethnobiology and Ethnomedicine 18: 51. doi: 10.1186/s13002-022-00548-2

Sousa R, Cantalice AS, Santos, FIR, Silva TC, Albuquerque UP. 2024. Contributions to the Identification of Cultural Keystone Species from an Emic Perspective: A Case Study from Northeast Brazil. Economic Botany 78: 182-196. doi: 10.1007/s12231-024-09603-3

Stoffle RW, Halmo DB, Evans MJ, Olmsted JE. 1990. Calculating the cultural significance of American Indian plants: Paiute and Shoshone ethnobotany at Yucca Mountain, Nevada. American Anthropologist 92: 416-432. doi: 10.1525/aa.1990.92.2.02a00100

Tardío J, Pardo-De-Santayana M. 2008. Cultural importance indices: A comparative analysis based on the useful wild plants of southern Cantabria

(northern Spain). Economic Botany 62: 24-39. doi: 10.1007/s12231-007-9004-5

Thiebaut G, Guérold F, Muller S. 2002. Are trophic and diversity indices based on macrophyte communities pertinent tools to monitor water quality? Water Research 36: 3602-3610. doi:10.1016/S0043-1354(02)00052-0

Thomas E, Vandebroek I, Sanca S, Damme PV. 2009. Cultural significance of medicinal plant families and species among Quechua farmers in Apillapampa, Bolivia. Journal of Ethnopharmacology 122: 60-67. doi: 10.1016/j.jep.2008.11.021

Toledo BA, Colantonio S, Galetto L. 2007. Knowledge and Use of Edible and Medicinal Plants in Two Populations from the Chaco Forest, CÓRdoba Province, Argentina. Journal of Ethnobiology 27: 218-232. doi: 10.2993/0278-0771_2007_27_218_kauoea_2.0.co_2

Tudela-Talavera P, Torre-Cuadros MA, Native Community of Vencedor. 2015. Cultural Importance and Use of Medicinal Plants in the Shipibo-Conibo Native Community of Vencedor (Loreto) Peru. Ethnobotany Research and Applications 14: 533-548. doi: 10.17348/era.14.0.533-548

Turner NJ. 1988. The Importance of a Rose : Evaluating the Cultural Significance of Plants in Thompson and Lillooet Interior Salish. American Anthropologist 90: 272-290. doi: 10.1525/aa.1988.90.2.02a00020

Vandebroek I, Balick MJ. 2012. Globalization and loss of plant knowledge: Challenging the paradigm. PLoS One 7: e37643. doi: 10.1371/journal. pone.0037643

Walters WH. 2021. Survey design, sampling, and significance testing: Key issues. The Journal of Academic Librarianship 47: 102344. doi: 10.1016/j. acalib.2021.102344

Weckmüller H, Barriocanal C, Maneja R, Boada M. 2019. Factors affecting traditional medicinal plant knowledge of the Waorani, Ecuador. Sustainability 11: 4460. doi: 10.3390/su11164460

Whiting MJ, Williams VL, Hibbitts TJ. 2011. Animals traded for traditional medicine at the Faraday market in South Africa: Species diversity and conservation implications. Journal of Zoology 284: 84-96. doi: 10.1111/j.1469-7998.2010.00784.x

Wolverton S. 2013. Ethnobiology 5: Interdisciplinarity in an Era of Rapid Environmental Change. Ethnobiology Letters 4: 21-25. doi:10.14237/ ebl.4.2013.11

Wyndham FS, Lepofsky D, Tiffany S. 2011. Taking Stock in Ethnobiology: Where Do We Come From? What Are We? Where Are We Going? Journal of Ethnobiology 31: 110-127. doi: 10.2993/02780771-31.1.10

Zuur AF, Ieno EN, Elphick CS. 2010. A protocol for data exploration to avoid common statistical problems. Methods in ecology and evolution 1: 3-14. doi: 10.1111/j.2041-210X.2009.00001.x

