

Molecular Genetics of Papillary Thyroid Carcinoma – Great Expectations...

ABSTRACT

Papillary thyroid carcinoma (PTC) is the most prevalent type of endocrine cancer and, in recent epidemiological surveys, one of the types of human cancer whose incidence is growing. Despite the favourable outcome and long survival rates of most patients, some tumours display an aggressive behaviour and may progress to the highly aggressive and lethal, anaplastic thyroid carcinoma. In recent years, several progresses have been made on the molecular characterization of PTC, in general, and in the genetic alterations underlying the histotype diversity of this type of cancer, in particular. This holds true regarding alterations on nuclear DNA as well as mitochondrial DNA. In this review we have summarized the most recent findings in the genetic characterization of PTC, giving a particular emphasis to the genotype-phenotype associations, the prognosis implications, and the diagnostic and therapeutic value of the newly identified genetic markers. (**Arq Bras Endocrinol Metab 2007;51/5:643-653**)

Keywords: Thyroid; Papillary thyroid carcinoma; Oncogene

RESUMO

Genética Molecular do Carcinoma Papilífero de Tireóide – Grandes Esperanças...

O carcinoma papilífero de tireóide (CPT) é o tipo mais prevalente de câncer endócrino e, em pesquisas epidemiológicas recentes, um dos tipos de câncer humano cuja incidência vêm crescendo. Apesar do prognóstico favorável e da longa taxa de sobrevivência da maioria dos pacientes, alguns tumores mostram um comportamento agressivo e podem progredir para o altamente agressivo e letal carcinoma anaplásico de tireóide. Recentemente, vários progressos foram feitos quanto à caracterização molecular do CPT, em general, e às alterações genéticas subjacentes à diversidade histológica desse tipo de câncer, em particular, particularmente com respeito às alterações dos DNAs nuclear e mitocondrial. Nesta revisão, nós resumimos os achados mais recentes da caracterização genética do CPT, dando ênfase particular às associações genótipo-fenótipo, às implicações prognósticas e ao valor diagnóstico e terapêutico dos marcadores genéticos recentemente identificados. (**Arq Bras Endocrinol Metab 2007;51/5:643-653**)

Descritores: Tireóide; Carcinoma papilífero de tireóide; Oncogenes

THYROID CANCER IS THE most prevalent type of endocrine cancer with incidence rates of 4 and 12 per 100,000 in men and women, respectively (1). Papillary thyroid cancer (PTC) represents virtually 80% of all thyroid cancers and recent epidemiological surveys indicate thyroid cancer as the type of human cancer displaying the highest growing incidence in USA [6.3 percent annual increase in the 1997–2003 period (1)]. The observation that the

revisão

VÍTOR TROVISCO
PAULA SOARES
ANA PRETO
PATRÍCIA CASTRO
VALDEMAR MÁXIMO
MANUEL SOBRINHO-SIMÕES

IPATIMUP – Institute of Molecular Pathology and Immunology of the University of Porto (VT, PS, AP, PC, VM & MS-S), Department of Pathology, Medical Faculty, University of Porto (PS, VM & MS-S), and Department of Pathology, Hospital São João (MS-S), Porto, Portugal.

Recebido em 20/02/07
Aceito em 23/02/07

incidence increase was not accompanied by a rise in deaths from the condition (0.5 per 100,000) and that nearly half the tumours are 1 centimetre or less, led some authors to suggest that better, high-tech diagnostic tests are picking up very small tumours, most of which pose no long-term threat (2). The detection of such small cancers presents a dilemma for physicians since, as stressed by Mazzaferri (3), they are not always benign. Despite the generally favourable prognosis of patients with papillary microcarcinomas, "cancer-related mortality rates may be as high as 1.0%, the rate of distant metastases as high as 2.5%, and rates of lymph node recurrence as high as 5%" (3,4). These figures, together with the fact that some cases of conventional PTC have an aggressive behaviour and may even progress to anaplastic thyroid carcinoma (ATC) (5,6), highlight the need of more powerful indicators for diagnosis and prognosis in PTC.

The essential diagnostic criteria in PTC still relies in the cytological nuclear features: large, irregular, "grooved", pale staining and with "ground glass" appearance (5,7). Whenever unequivocal nuclear features are lacking, one can use additional morphological features such as papillary architecture, stromal reaction, diffuse growth and/or the presence of psammoma bodies (8).

At present, the "*in diebus illis*" of the pathologist resides in the differential diagnosis of encapsulated lesions with some of the aforementioned nuclear characteristics and a follicular pattern of growth. This diagnostic dilemma and the aforementioned prognostic difficulties justify the great expectations that were put in the molecular characterization of PTC. In this review we will discuss whether or not they were (or will be) fulfilled.

DNA CONTENT, CYTOGENETIC AND GENETIC STABILITY OF PTC

In thyroid tumours there is no correlation between aneuploidy and malignancy and thus it cannot be used as a distinctive criterion. Aneuploidy can indeed be present in benign and malignant thyroid lesions; PTC, in particular, usually displays a diploid or near diploid DNA content (9). Despite this, an aneuploid DNA content is more frequently found in metastasising/more aggressive PTC than in conventional PTCs (10,11).

In general, PTC is cytogenetically characterized by normal karyotypes or, whenever abnormal, by simple cytogenetic alterations. Structural chromosomal alterations involve more frequently chromosomes 1, 3, 7,

and 10. Remarkably, some cases of the follicular variant of PTC (FVPTC) are characterized by chromosomal aberrations commonly found in follicular thyroid adenomas (FTA) or follicular thyroid carcinomas (FTC), such as t(2;3) and gains of chromosomes 3, 5, 7, 9, 12, 14, 17, and 20 (12). These findings suggest that there are cytogenetic changes preferentially associated with FVPTC and that FVPTC is closer to the FTC and FTA group of lesions than to classical PTC (12,13). Another signal of the genetic stability of PTC lies in the rare microsatellite instability in these carcinomas (14).

Taking the apparent genetic stability of PTC together with the frequent occurrence, in young patients, of cases of multicentric PTC, it is tempting to consider that PTC tumourigenesis reflects the end product of few carcinogenic events (8,15).

The indolent growth of PTC is reflected in the very low rates of proliferative activity, evaluated both by immunohistochemical markers (i.e. MIB-1) and by cytometric assessment of the S-phase (9,16).

RET/PTC AND NTRK1 REARRANGEMENTS

Genetic rearrangements are frequently detected in haematological and mesenchymal neoplastic diseases at variance with their rarity in carcinomas. Thyroid cancer is one of the few types of carcinoma having such genetic events, but the reason for this singularity is still unclear. The chromosomal rearrangements so far detected in PTC involve the tyrosine kinase (TK) growth factor receptors RET and NTRK1 (also known as TRKA) (17), which play a role in the regulation of growth, differentiation and programmed cell death of neurons in the peripheral and the central nervous system (18).

The rearrangements of RET (RET/PTC) and NTRK1 involve their fusion to heterologous genes (19) and result in chimeric proteins that have been extensively studied in *in vitro* models using several RET/PTC and NTRK1 (TRKT1) chimeric transcripts. Such studies have shown that the tumourigenic properties of RET/PTC and TRKT1 result from the aberrant and persistent activation of their tyrosine kinase domain (17).

RET normal expression and kinase activity is restricted to a subset of cells derived from embryonic neural crest cells (20). Consistent with this, wild-type RET is expressed at high levels in parafollicular C-cells, but its expression in follicular thyroid cells remains disputable (19).

RET gene is located in chromosome 10 and its rearrangements reflect the frequent structural cytogenetic alterations of this chromosome. RET/PTC

rearrangements can be either paracentric rearrangements, as with *H4* or *ELE1* genes (*RET/PTC1* and *RET/PTC3*, respectively), or reciprocal translocation, as the one involving *PRKAR1A* gene, encoding RI- α on 17q23 (*RET/PTC2*). *RET/PTC1*, 2, and 3 are the most frequent alterations involving *RET* proto-oncogene in PTC but at least 15 different types have been identified to date (21).

Somatic rearrangements of the *RET* proto-oncogene have been detected in 3–60% of sporadic PTC (19,21). The prevalence of *RET/PTC* in PTC varies significantly in different studies, probably reflecting the different methodologies and the geographic sampling, as well as the histological composition of the series (see below). In most series dealing with sporadic PTC *RET/PTC1* is the most common type, comprising up to 60–70% of the rearrangements, whereas *RET/PTC3* accounts for 20–30% (16,19,22,23). At variance with this, in Chernobyl-related thyroid cancers, *RET/PTC3* rearrangements are the most frequent (19,24–26), at least for the “first wave” of cancers arising in this setting, since *RET/PTC1* seems to predominate in cases with a longer latency period [for a revision see (27)]. Other rare types of *RET/PTC* rearrangements appear to be mainly associated with radiation exposure (19).

The existence of a precursor lesion of PTC was a longstanding question and the detection of *RET/PTC* rearrangements in inflammatory lesions of the thyroid has recently fuelled this issue (28). Using highly sensitive methods (real-time PCR), some authors described *RET/PTC* rearrangements in up to 95% of cases of Hashimoto’s thyroiditis (HT) (29,30). However, other groups were unable to reproduce these data using a similar methodology (28,31). It remains to be clarified if the use of very sensitive methods highlights “spurious” genetic alterations with low biological potential, or if HT is a real “pre-neoplastic” entity as it has been suggested by some authors (29,30).

Oncogenic rearrangements of *NTRK1* gene are also found in PTC. The *NTRK1* gene, localized in chromosome 1 (again, one of the chromosomes with more frequent structural alterations in PTC), codes for the high-affinity nerve growth factor (NGF) receptor, and its activation has been reported to elicit the activation of the RAF-MEK-ERK pathway (32). *NTRK1* rearrangements are rare, usually found in less than 10% of cases of sporadic PTC (33–35). In 2004, Frattini and co-workers (36) studied *NTRK1* gene rearrangement by RT-PCR and found the expression of *NTRK1* TK domain, which is suggestive of oncogenic rearrangement in three out of 55 PTC cases (5.5%).

NTRK1 cell signalling is modulated by the presence of p75 (NTR) (32). In contrast to *NTRK1*, p75 (NTR) is able to bind all neurotrophins but lacks intrinsic tyrosine kinase activity (18). Although initially described as a low affinity receptor, p75 has the same affinity for NGF as does *NTRK1*, and when co-expressed with *NTRK1* enhances its ability to bind and to respond to neurotrophins. p75 also modulates/enhances the specificity of other TRKs for their preferred ligands (18).

The detection of *NTRK1* rearrangements in PTC and the observation that the thyroid targeted expression of the rearranged *NTRK1* chimeric protein (TRKT1) in transgenic mice leads to the development of PTC (37), support the involvement of *NTRK1* and of p75 (NTR) in the etiopathogenesis of PTC. Recently we have observed neoexpression of p75, particularly in conventional PTC (38). We also shown that the cellular localization of p75 appears to be related to the presence of the *BRAF*^{V600E} mutation; the biological significance of this finding remains to be clarified (38).

BRAF MUTATIONS

BRAF is one of the three members of the conserved RAF family of serine/threonine kinases – ARAF, *BRAF*, and CRAF – which are critical effectors of the canonical MAPK pathway RAF-MEK-ERK. This pathway is critical in the transduction of signals by growth factors, hormones and cytokines, being involved in the regulation of cell proliferation, differentiation and apoptosis (39,40).

RAF genes have been described as proto-oncogenes because RAFs are the immediate downstream effectors of RAS oncoproteins and because their initial description was its oncogenic viral form *v-raf* (41). Yet the ultimate evidence was the finding of activating mutations in *BRAF* gene in a wide panel of human cancers, most prominently in cutaneous melanoma (63–66%) (42,43) but also in serous ovarian carcinoma (33–40%) (42,44) and colorectal carcinoma (11–20%) (45,46). Of notice, *BRAF* mutations were mostly non coexistent with *RAS* mutations and the great majority of the mutations (~90%) were of a single type: the 1799T-A transition, leading to the substitution of a valine by a glutamic acid at the position 600 (V600E) (42).

Within thyroid neoplasias, *BRAF* mutations (V600E) were frequently detected in sporadic PTC (29–83%) (47–50), ranking as the major genetic alteration of this type of human cancer. *BRAF* mutations are

almost always exclusive to the relatively rare *RAS* genes mutations and also to *RET* (*RET/PTC*) and *NTRK1* rearrangements, altogether accounting for about 70% of PTC cases (33,36,47,48,51). The mutually exclusive oncogenic activation of *RET/PTC*, *RAS* and *BRAF* in PTC supports the existence of a linear oncogenic signalling pathway involving *RET/PTC*-*RAS*-*BRAF*-*MEK*-*ERK* in these tumours, a concept further reinforced by functional experiments *in vitro* (52,53).

Following the detection of *RET/PTC* rearrangements in HT, *BRAF* mutations were also screened in these lesions (54-56). *BRAF* mutations were solely detected by Kim and co-workers (56) and only in cases in which the HT areas coexisted with *BRAF*-mutated PTC areas. Hence, as the hypothesis of contamination with PTC-positive cells cannot be excluded, these studies do not demonstrate the pre-neoplastic role of HT.

BRAF mutations were found to be rare in childhood PTC in the post-Chernobyl setting (57-60). As in the sporadic cases, *BRAF* gene alterations do not coexist with the highly frequent *RET/PTC* rearrangements (58,59). It has been suggested that the ionizing radiation contributes to the occurrence of PTC primarily by the induction of double-strand DNA breaks and their subsequent illegitimate recombination, thus leading to *RET/PTC* (and *NTRK1*) rearrangements (57,61). High prevalences of *RET/PTC* rearrangements and reciprocal low prevalences of *BRAF* mutations are also hallmarks of sporadic childhood and young PTC (16,54,58,60,62-64). Taking these facts together with the above discussed *RET/PTC* 'waves' in post-Chernobyl PTC (first *RET/PTC3*, and then *RET/PTC3*) (27,65), it is possible that a third wave of *BRAF*-mutated PTC cases in the Chernobyl setting, thus recapitulating the sequence observed in sporadic PTC.

RAS MUTATIONS

Three human *RAS* genes – *NRAS*, *HRAS*, and *KRAS* – are frequently involved in human tumorigenesis (about 15% of all human tumours harbour mutations in these genes) (66). The *RAS* mutations found in tumours typically occur in codons 12, 13 or 61 of any of the three genes and produce constitutively active *RAS* proteins. *RAS* proteins are key intracellular signal transducers that can activate several downstream pathways, namely the classical *RAF*-*MEK*-*ERK* pathway (40).

RAS genes mutations are particularly prevalent in FTA and FTC and, less frequently, in PTC (67). Their prevalence in PTC varies widely, depending on the studied series (0–16%) (67-70). The mutations

associated to PTC (and to thyroid lesions in general) predominantly involve codons 61 of *NRAS* and, to a less extent, of *HRAS* (13,67,71,72) (see below the genotype-phenotype correlations with regard to *BRAF* and *RAS* mutations).

PAX8/PPAR γ REARRANGEMENTS

Translocations involving chromosomes 2 and 3 and LOH in chromosome 3 have been detected in FTC cases (73), suggesting the putative existence of a tumour suppressor gene at 3p25 locus. Kroll and co-workers (74) showed that the translocation t(2;3) (q13;p25) results in the fusion of the DNA-binding domains of the thyroid transcription factor *PAX8* (2q13) to the A to F domains of peroxisome proliferator-activated receptor γ (*PPAR γ 1*)(3p25). The tumourigenic effect of this event appears to be due to the loss of proper *PAX8* and *PPAR γ* transcriptional function in the rearranged *PAX8/PPAR γ* form, as well as of the remnant normal *PAX8* and *PPAR γ* proteins due to a dominant-negative effect (74,75).

In the initial description, *PAX8/PPAR γ* mRNA and protein were detected in FTC but not in FTA, PTC, or multinodular hyperplasias, and it was thus advanced as a marker of FTC (74). However, the genetic alteration was later on also described in a number of FTA cases (76-78). This ruled out the possibility of considering *PAX8/PPAR γ* rearrangements as a molecular indicator of malignancy.

Recently, we reported for the first time the *PAX8/PPAR γ* fusion gene in a relatively high percentage of cases of FVPTC (37.5%) (13) (see below the genotype-phenotype meaning of this finding).

So far it is not known if *PAX8/PPAR γ* rearrangements are prevalent events in poorly differentiated thyroid carcinoma (PDTC) and ATC and, hence, if they may be used as progression markers of follicular lesions (FTA, FTC and FVPTC) towards more aggressive forms of thyroid neoplasias. It remains also to be confirmed the advanced association of *PAX8/PPAR γ* rearrangements with vascular invasiveness in FTC (77).

ALTERATIONS IN MITOCHONDRIAL GENES IN PTC

We agree with the new WHO classification of thyroid tumours in which oncocytic (Hürthle cell) tumours are considered as variants of their non-oncocytic counterparts (e.g. oncocytic variant of PTC and oncocytic variant of FTC) instead of constituting a category by itself (7,8).

The mitochondrial DNA (mtDNA) is small (16,569bp) and encodes 13 essential components of the cellular energy-production apparatus, being absolutely vital for life.

The high copy number of mtDNA and the cytoplasmic location of the mitochondria contribute to its high mutation rate, about 10 to 20 times higher than that of the nuclear DNA (nDNA) [for a thorough review see (79)].

Alterations of mtDNA have been demonstrated in various types of human cancer including thyroid tumours, and include large deletions, missense mutations, frameshift mutations and small deletions/insertions, but the role of mtDNA somatic mutations in tumourigenesis has not been yet fully understood (80-82).

Yeh and co-workers (83) identified three somatic mtDNA mutations in PTC in a series of 21 thyroid tumours with different histotypes and suggested that somatic mtDNA alterations may be involved in thyroid tumourigenesis. Despite their interesting results, the authors only studied a small series of tumours with an overrepresentation of PTC. These results have been, in part, confirmed in a study by our group in a much larger series of thyroid tumour (84). We found that FTC and PTC carried a significantly higher prevalence of non-silent point mutations in complex I genes than FTA (84). We have also found large deletions of mtDNA in all types of tumours, with a striking prominence for Hürthle cell tumours (up to 16%) of mtDNA common deletion independently of the histological variant (84). The same held true to sequence variants of *ATPase 6* gene which were significantly more prevalent in patients with Hürthle cell tumours than in patients with non-Hürthle cell neoplasms (84).

Mutations and sequence variants in mtDNA complex I genes appear to be more frequent in malignant than in benign thyroid tumours, suggesting a role in tumour progression of these alterations. The mtDNA common deletion and *ATPase 6* variants are more frequent in Hürthle cell tumours than in non-Hürthle cell tumours, appearing to be involved with Hürthle cell transformation rather than with tumourigenesis (84). With regard to mtDNA alterations, PTCs do not differ from FTCs, i.e. apparently no specific mtDNA alterations are associated with the papillary histotype (84).

Alterations in *GRIM-19* gene, a nuclear gene encoding a complex I mitochondrial protein, have been recently found in Hürthle cell tumours (85). *GRIM-19* is one of several proteins associated with retinoid-interferon-induced mortality (*GRIM*); it is considered as a cell death regulator that promotes apoptosis, a negative regulator of cell growth, and it is also involved in mito-

chondrial metabolism (86). *GRIM-19* mutations were detected in three cases of Hürthle cell variant of PTC and in one case of Hürthle cell variant of FTC, whereas no mutations were detected in any non-Hürthle cell carcinoma (85). The detection of *RET/PTC1* in one of the cases of Hürthle cell variant of PTC with a *GRIM-19* mutation suggests that *GRIM-19* mutations may play a role together with the oncogenic activation in tumour development. Although a larger series of benign and malignant thyroid tumours with and without Hürthle cell features needs to be studied, it seems that *GRIM-19* alterations, like mtDNA alterations, are associated with Hürthle cell features and not with specific histotypes. This finding suggests that *GRIM-19* mutations may serve as a predisposing alteration for the occurrence of tumours with cell oxyphilia. Other alterations, such as *RET/PTC* rearrangements or *BRAF* mutations, appear to be necessary in Hürthle cell tumours, like in any non-Hürthle cell variant of PTC, for the cancer development (48,87).

PHENOTYPE-GENOTYPE RELATIONSHIP IN THYROID NEOPLASIA

PTC represent a heterogeneous group of tumours, comprising several histological variants that share the peculiar and diagnostic nuclear features of PTC (5,7,88). Despite their common classification as 'PTC', convincing molecular evidence has appeared in the last years that support the previous histological distinction of some PTC variants. The most frequent *RET/PTC* rearrangement, *RET/PTC1*, is related to PTC cases displaying the conventional histotype (16,22,23,64), whereas *RET/PTC3* is associated with the solid variant of PTC (19,24,25). *RET/PTC3* was also reported to occur frequently in cases of the tall cell variant of PTC (89).

Regarding *BRAF* mutations, the most frequent *BRAF^{V600E}* mutant form is almost exclusively detected in PTC cases with a papillary or mixed papillary/follicular architecture (irrespectively of the variant being conventional, tall cell, oxyphilic or microcarcinoma) (13,36,54,64,90-92), whereas the less frequent and less reported *BRAF^{K601E}* form (~7%) is exclusively detected in cases of FVPTC (54,91).

RAS mutations are also closely associated with the tumour's histotype, being rarely detected in conventional PTC (0-16%) and frequently detected in FVPTC (25-100%) (13,64,71,72).

Finally, the relatively high frequency of *PAX8/PPAR γ* rearrangements in FVPTC, in conjunction with the aforementioned data on *RET*, *BRAF*

and *RAS* alterations, reinforce the assumption that some FVPTC cases share some of the molecular features of follicular tumours (FTA and FTC), constituting a sort of intermediate category between conventional PTC and FTC (13).

There is a particular type of thyroid tumour that usually occurs in the context of Familial Adenomatous Polyposis (FAP) and that is considered by most authors as a variant of PTC – the so-called cribriform-morular variant of PTC (93,94). The molecular link between FAP and the cribriform morular variant of PTC is the *APC* gene, since germline and/or somatic alterations have been detected in FAP-associated, as well as in sporadic thyroid carcinomas with a cribriform-morular appearance (95,96). Moreover, the *APC* deleterious mutations related to these tumours are associated with the 5' region of exon 15 and are unfrequent in the considered hotspot mutation region of *APC* (97). The search for germline *APC* mutations in apparently sporadic cases of the cribriform-morular variant of PTC is a must; this approach is of clinical relevance, as these cases may represent an occult FAP condition (93,94).

A last point to stress another type of genotype-phenotype correlation: Hürthle cell tumours display similar mitochondrial alterations (proteins encoded by mtDNA and/or nDNA) regardless of the specific thyroid tumour histotype (PTC or FTC or even FTA) (see above). In this setting one is referring to a mitochondria-rich phenotype rather than to the oncogenic-related tumour phenotype. It remains to be clarified whether or not it will be advantageous to treat patients with Hürthle cell carcinomas targeting both the specific oncogenes involved in the tumour or the mitochondrial abnormalities.

DIAGNOSTIC AND PROGNOSTIC USEFULNESS OF THE SEVERAL PTC-ASSOCIATED GENETIC ALTERATIONS

FNA is the best tool for the early diagnosis of thyroid lesions. As the PTC diagnosis relies on the typical nuclear features, cases with less than typical PTC nuclei in FNA samples, as it frequently occurs in FVPTC, may be misdiagnosed as FTA or as follicular tumour.

The diagnosis of FTC cannot be usually made by FNA since it depends on the observation of capsular or vascular invasion (5,7). All these limitations led to the assumption that good ancillary tools would be very useful for the FNA diagnosis of difficult thyroid tumours.

PAX8-PPAR γ rearrangements and *RAS* genes mutations are quite prevalent in cases of FVPTC

(13,64,71). Yet, such genetic events are similarly frequent in the lesions we would like to distinguish from PTC – e.g. FTC and, more importantly, FTA (13,67,74,78,98). Thus, the identification of *PAX8-PPAR γ* rearrangements and/or *RAS* mutations in FNA samples does not provide useful diagnostic improvements in difficult, concrete, follicular patterned thyroid tumours.

The accurate detection of both *RET/PTC* rearrangements and *BRAF* gene mutations in thyroid FNA samples has proved useful in certain situations (99-104). The detection of *RET/PTC* rearrangements or of *BRAF* mutations (V600E or K601E) will lead to an unequivocal diagnosis of PTC and will therefore allow a better clinical approach in such cases (105). The strong association of *BRAF*^{V600E} mutation and, to a lesser extent, *RET/PTC1*, to PTC cases displaying typical papillary or mixed papillary-follicular architecture limits, however, their screening usefulness since those are the easiest cases to diagnose by FNA.

PTCs rarely cause the death of the patients (106,107) as they usually respond very well to the surgical removal of the tumour followed by radioactive iodine treatment. Yet, some cases have an aggressive behaviour and some may progress to the highly aggressive and lethal, ATC (5,6). The identification of the PTCs with guarded prognosis would have major clinical relevance and that is the reason why so many groups have been trying to find genetic markers with clinical significance for the outcome of the patients.

Most studies published so far have focused on the clinical significance of *RET/PTC* rearrangements and, particularly, of mutant *BRAF* forms. The presence of *RET/PTC* rearrangements have been associated in some studies with lymph node metastatization (34,62,64,70) while other groups did not find this association (16,108,109). Of the several clinicopathological parameters analysed, *RET/PTC* positive cases have only been convincingly associated with younger age at diagnosis (16,34,62-64). *RET/PTC3* seems to be associated with tumours with a guarded prognosis but larger series with longer follow-up are needed to clarify this issue (5,7).

Also controversial has been the putative association of *BRAF* mutations with poor prognosis parameters in PTC. The majority of the groups advanced that PTCs bearing these mutations are prone to a more aggressive clinical behaviour (64,90,110-112). The disclosure of *BRAF* mutations in PDTC and ATC, particularly in cases apparently derived from pre-existent PTC, provided additional support to their prognostic meaning (90,113-115).

Some studies have described a statistically significant association of *BRAF* mutations to older age at diagnosis (54,64,90), male gender (112,116), extrathyroid extension (64,90,110), lymph node metastases (110), distant metastases (111), higher tumour staging (64,90,110,111), and recurrence (110,112). Other groups did not find any of the aforementioned associations (33,51,92,117,118). Moreover, given the strict relationship of *BRAF* mutations to PTCs with predominant papillary architecture, the aforementioned relationship of *BRAF* mutations to poor prognosis indicators may be biased by the constitution of each series with regard to the histological types of PTC. For instance, the tall cell variant of PTC, accepted by some groups as being more aggressive than conventional PTC (119-121), has particularly high frequency of *BRAF* mutations (36,64,90). The alleged histological bias is well shown in the study by Xing and co-workers (110), in which the associations of *BRAF* mutations to extrathyroid extension, lymph node metastasis and high tumour staging were lost upon the stratification of the series by histotype.

Finally, it remains to be verified the implications of *PAX8/PPAR γ* in the behaviour of the tumours. We have found a significant association between *PAX8/PPAR γ* rearrangements and multifocality and vascular invasiveness in FVPTC, suggesting that the rearrangement confers a higher invasive potential (13). A strong association with vascular invasiveness was described in FTC harbouring these genetic alterations (77). Besides the association with invasiveness, it remains to be demonstrated if cases of FTC or FVPTC positive for *PAX8/PPAR γ* are more prone to give rise to blood born/haematological metastases (13).

NEW THERAPEUTIC APPROACHES TARGETING GENETIC ALTERATIONS IN PTC

The treatment of patients with PTC has usually an excellent outcome, leading to a much lower risk for death or disease recurrence than other human malignancies. Initially PTC is treated by surgery (total or partial thyroidectomy), followed by radioiodine (^{131}I) treatment (122). Radioiodine therapy is an extremely effective treatment in well-differentiated thyroid carcinomas (FTC and PTC), being, at present, the only effective treatment in patients with distant metastases (122).

Although the majority of tumours respond well to radioiodine therapy, there are PTCs that are resistant to this classical therapy, namely those that are inoperable and have lost radioiodine avidity. Until

recently, there were few clinical trials available for patients with iodine nonresponsive thyroid cancers. The molecular alterations that have been identified in PTC and thought to be important in oncogenesis, namely *RET/PTC* rearrangement and *BRAF^{V600E}* mutation raised the possibility of targeting these molecules as a potential therapeutic approach.

Several inhibitors targeting different kinases are available. BAY 43-9006, a bi-aryl urea (also known as Sorafenib), first designed as a RAF1 kinase inhibitor (123), inhibits *in vitro* both wild-type and V600E-mutant BRAF (124). In addition, BAY 43-9006 has significant activity against VEGFR2 and 3, PDGFR β , Flt-3, c-Kit and RET. BAY 43-9006 has been approved by the Food and Drug Administration (FDA) for therapy of renal cell carcinoma despite the absence of *BRAF* mutations in this subset of tumours, and it is under evaluation for melanoma and thyroid cancer treatment, the two human malignancies which harbour the highest percentage of *BRAF* mutations. With regard to thyroid tumours, it was observed that BAY 43-9006 inhibits growth of thyroid carcinoma derived cell lines (125).

Two potent RAF kinase inhibitors, NVP-AAL881-NX and NVP-LBT613-AG-8, have been tested *in vitro* and *in vivo* against a panel of thyroid cancer cell lines, and both drugs were found to inhibit proliferation and to induce cell death (126).

Recently, new data concerning the requirement of the Hsp90 chaperone for the stability of BRAF^{V600E} rendered the possibility of targeting BRAF^{V600E} by the use of Hsp90 inhibitors, such as 17-AAG, but further studies are needed (127,128).

Small molecules of various chemical classes have been reported to inhibit RET (two pyrazolopyrimidines – PP1 and PP2 –, a 2-indolinone – RPI-1 – and two indolocarbazole derivatives – CEP-701 and CEP-751); for the moment, ZD6474 is the only one in an advanced phase of clinical trials (129).

Loss of radioiodine uptake is related to low or absent Sodium Iodine Symporter (NIS) protein, and this observation led to the attempt of therapeutically inducing an increased expression of NIS. It has been shown in clinical trials that retinoic acid analogues act by increasing NIS expression and promoting higher ^{131}I uptake (122).

PAX8/PPAR γ rearrangement have been found in a subset of FVPTC (13), PPAR γ agonists, as rosiglitazone, are already in phase II clinical trials in patients with thyroglobulin-positive and radioiodine-negative differentiated thyroid cancer in an attempt to induce radioiodine uptake (130). Other PPAR γ agonists, like

troglistazone and the new, high affinity, RS5444, have been shown to inhibit *in vitro* the proliferation in thyroid carcinoma derived cell lines (131,132).

Summing up, the available compounds targeting different molecules that are thought to be involved in PTC carcinogenesis represent promising drugs that are being evaluated in the treatment of patients with thyroid cancer. These drugs vary in their specificity and have been tested with relative success in preclinical studies and in early clinical trials (133). The challenge now is to progress in the validation of the actual targets of the drugs and in the exploitation of the possibility of using combined therapy in patients with progressive thyroid cancer.

ACKNOWLEDGEMENTS

We would like to thank the colleagues of our group for their support and collaboration. Some of the work herein cited was supported by the Portuguese Science and Technology Foundation (FCT) through a PhD grant to Vítor Trovisco (SFRH/BD/13055/2003), Post-doc grants to Ana Preto (SFRH/BPD/14882/2003) and Patrícia Castro (SFRH/BPD/26553/2006), and a project funding (POCTI project POCTI/NSE/48171/2002).

We would like to apologize to the authors whose works were not cited by space constrains or unintended omission.

REFERENCES

1. Ries LAG, Harkins D, Krapcho M, Mariotto A, Miller BA, Feuer EJ, et al. (eds). **SEER Cancer Statistics Review, 1975-2003**. National Cancer Institute, 2006. Based on November 2005 SEER data submission, posted to the SEER web site, 2006.
2. Davies L, Welch HG. Increasing incidence of thyroid cancer in the United States, 1973-2002. **JAMA** 2006;295:2164-7.
3. Mazzaferri EL. Managing small thyroid cancers. **JAMA** 2006;295:2179-82.
4. Chow SM, Law SC, Chan JK, Au SK, Yau S, Lau WH. Papillary microcarcinoma of the thyroid – prognostic significance of lymph node metastasis and multifocality. **Cancer** 2003;98:31-40.
5. Rosai J, Carcangiu ML, DeLellis RA. **Tumours of the thyroid gland**. Washington: Armed Force Institute of Pathology, 1992.
6. Wynford-Thomas D. Origin and progression of thyroid epithelial tumours: cellular and molecular mechanisms. **Horm Res** 1997;47:145-57.
7. DeLellis RA LR, Heitz PU, Eng C (ed). **World Health Organization Classification of Tumours. Pathology and Genetics of Tumours of Endocrine Glands**. Lyon: IARC Press, 2004.
8. Fonseca E, Soares P, Cardoso-Oliveira M, Sobrinho-Simões M. Diagnostic criteria in well-differentiated thyroid carcinomas. **Endocr Pathol** 2006;17:109-17.
9. Johannessen JV, Sobrinho-Simões M, Tangen KO, Lindmo T. A flow cytometric deoxyribonucleic acid analysis of papillary thyroid carcinoma. **Lab Invest** 1981;45:336-41.
10. Backdahl M, Cohn K, Auer G, Forsslund G, Granberg PO, Lundell G, et al. Comparison of nuclear DNA content in primary and metastatic papillary thyroid carcinoma. **Cancer Res** 1985;45:2890-4.
11. Johannessen JV, Sobrinho-Simões M, Lindmo T, Tangen KO, Kaalhus O, Brennhovd IO. Anomalous papillary carcinoma of the thyroid. **Cancer** 1983;51:1462-7.
12. Roque L, Nunes VM, Ribeiro C, Martins C, Soares J. Karyotypic characterization of papillary thyroid carcinomas. **Cancer** 2001;92:2529-38.
13. Castro P, Rebocho AP, Soares RJ, Magalhães J, Roque L, Trovisco V, et al. PAX8-PPAR γ rearrangement is frequently detected in the follicular variant of papillary thyroid carcinoma. **J Clin Endocrinol Metab** 2006;91:213-20.
14. Soares P, dos Santos NR, Seruca R, Lothe RA, Sobrinho-Simões M. Benign and malignant thyroid lesions show instability at microsatellite loci. **Eur J Cancer** 1997;33:293-6.
15. Sobrinho-Simões M, Preto A, Rocha AS, Castro P, Maximo V, Fonseca E, et al. Molecular pathology of well-differentiated thyroid carcinomas. **Virchows Arch** 2005;447:787-93.
16. Soares P, Fonseca E, Wynford-Thomas D, Sobrinho-Simões M. Sporadic ret-rearranged papillary carcinoma of the thyroid: a subset of slow growing, less aggressive thyroid neoplasms? **J Pathol** 1998;185:71-8.
17. Tallini G. Molecular pathobiology of thyroid neoplasms. **Endocr Pathol** 2002;13:271-88.
18. Teng KK, Hempstead BL. Neurotrophins and their receptors: signaling trios in complex biological systems. **Cell Mol Life Sci** 2004;61:35-48.
19. Nikiforov YE. RET/PTC rearrangement in thyroid tumors. **Endocr Pathol** 2002;13:3-16.
20. Schuchardt A, D'Agati V, Larsson-Blomberg L, Costantini F, Pachnis V. Defects in the kidney and enteric nervous system of mice lacking the tyrosine kinase receptor **Ret**. **Nature** 1994;367:380-3.
21. Kondo T, Ezzat S, Asa SL. Pathogenetic mechanisms in thyroid follicular-cell neoplasia. **Nat Rev Cancer** 2006;6:292-306.
22. Sugg SL, Ezzat S, Rosen IB, Freeman JL, Asa SL. Distinct multiple RET/PTC gene rearrangements in multifocal papillary thyroid neoplasia. **J Clin Endocrinol Metab** 1998;83:4116-22.
23. Tallini G, Santoro M, Helie M, Carlomagno F, Salvatore G, Chiappetta G, et al. RET/PTC oncogene activation defines a subset of papillary thyroid carcinomas lacking evidence of progression to poorly differentiated or undifferentiated tumor phenotypes. **Clin Cancer Res** 1998;4:287-94.
24. Thomas GA, Bunnell H, Cook HA, Williams ED, Nerovnya A, Cherstvoy ED, et al. High prevalence of RET/PTC rearrangements in Ukrainian and Belarussian post-Chernobyl thyroid papillary carcinomas: a strong correlation between RET/PTC3 and the solid-follicular variant. **J Clin Endocrinol Metab** 1999;84:4232-8.
25. Nikiforov YE, Rowland JM, Bove KE, Monforte-Munoz H, Fagin JA. Distinct pattern of ret oncogene rearrangements in morphological variants of radiation-induced and sporadic thyroid papillary carcinomas in children. **Cancer Res** 1997;57:1690-4.
26. Motomura T, Nikiforov YE, Namba H, Ashizawa K, Nagataki S, Yamashita S, et al. ret rearrangements in Japanese pediatric and adult papillary thyroid cancers. **Thyroid** 1998;8:485-9.
27. Williams D. Cancer after nuclear fallout: lessons from the Chernobyl accident. **Nat Rev Cancer** 2002;2:543-9.
28. Nikiforov YE. RET/PTC Rearrangement – a link between Hashimoto's thyroiditis and thyroid cancer... or not. **J Clin Endocrinol Metab** 2006;91:2040-2.
29. Rhoden KJ, Unger K, Salvatore G, Yilmaz Y, Vovk V, Chiappetta G, et al. RET/papillary thyroid cancer rearrangement in nonneoplastic thyrocytes: follicular cells of Hashimoto's thyroiditis share low-level recombination events with a subset of papillary carcinoma. **J Clin Endocrinol Metab** 2006;91:2414-23.
30. Sheils OM, O'Eary JJ, Uhlmann V, Lattich K, Sweeney EC. ret/PTC-1 activation in Hashimoto thyroiditis. **Int J Surg Pathol** 2000;8:185-9.

31. Nikiforova MN, Caudill CM, Biddinger P, Nikiforov YE. Prevalence of RET/PTC rearrangements in Hashimoto's thyroiditis and papillary thyroid carcinomas. **Int J Surg Pathol** **2002**;10:15-22.
32. Miller FD, Kaplan DR. Neurotrophin signalling pathways regulating neuronal apoptosis. **Cell Mol Life Sci** **2001**;58:1045-53.
33. Liu RT, Chen YJ, Chou FF, Li CL, Wu WL, Tsai PC, et al. No correlation between BRAF^{V600E} mutation and clinicopathological features of papillary thyroid carcinomas in Taiwan. **Clin Endocrinol (Oxf)** **2005**;63:461-6.
34. Musholt TJ, Musholt PB, Khaladj N, Schulz D, Scheumann GF, Klempnauer J. Prognostic significance of RET and NTRK1 rearrangements in sporadic papillary thyroid carcinoma. **Surgery** **2000**;128:984-93.
35. Kuo CS, Lin CY, Hsu CW, Lee CH, Lin HD. Low frequency of rearrangement of TRK protooncogene in Chinese thyroid tumors. **Endocrine** **2000**;13:341-4.
36. Frattini M, Ferrario C, Bressan P, Balestra D, De Cecco L, Mondellini P, et al. Alternative mutations of BRAF, RET and NTRK1 are associated with similar but distinct gene expression patterns in papillary thyroid cancer. **Oncogene** **2004**;23:7436-40.
37. Russell JP, Powell DJ, Cunnane M, Greco A, Portella G, Santoro M, et al. The TRK-T1 fusion protein induces neoplastic transformation of thyroid epithelium. **Oncogene** **2000**;19:5729-35.
38. Rocha AS, Risberg B, Magalhães J, Trovisco V, de Castro IV, Lazarovici P, et al. The p75 neurotrophin receptor is widely expressed in conventional papillary thyroid carcinoma. **Hum Pathol** **2006**;37:562-8.
39. Tanoue T, Nishida E. Molecular recognitions in the MAP kinase cascades. **Cell Signal** **2003**;15:455-62.
40. Peyssonnaud C, Eychene A. The Raf/MEK/ERK pathway: new concepts of activation. **Biol Cell** **2001**;93:53-62.
41. Rapp UR, Goldsborough MD, Mark GE, Bonner TI, Groffen J, Reynolds FH, Jr., et al. Structure and biological activity of v-raf, a unique oncogene transduced by a retrovirus. **Proc Natl Acad Sci U S A** **1983**;80:4218-22.
42. Davies H, Bignell GR, Cox C, Stephens P, Edkins S, Clegg S, et al. Mutations of the BRAF gene in human cancer. **Nature** **2002**;417:949-54.
43. Brose MS, Volpe P, Feldman M, Kumar M, Rishi I, Gerrero R, et al. BRAF and RAS mutations in human lung cancer and melanoma. **Cancer Res** **2002**;62:6997-7000.
44. Singer G, Oldt R, III, Cohen Y, Wang BG, Sidransky D, Kurman RJ, et al. Mutations in BRAF and KRAS characterize the development of low-grade ovarian serous carcinoma. **J Natl Cancer Inst** **2003**;95:484-6.
45. Rajagopalan H, Bardelli A, Lengauer C, Kinzler KW, Vogelstein B, Velculescu VE. Tumorigenesis: RAF/RAS oncogenes and mismatch-repair status. **Nature** **2002**;418:934.
46. Wang L, Cunningham JM, Winters JL, Guenther JC, French AJ, Boardman LA, et al. BRAF mutations in colon cancer are not likely attributable to defective DNA mismatch repair. **Cancer Res** **2003**;63:5209-12.
47. Kimura ET, Nikiforova MN, Zhu Z, Knauf JA, Nikiforov YE, Fagin JA. High Prevalence of BRAF mutations in thyroid cancer: genetic evidence for constitutive activation of the RET/PTC-RAS-BRAF signaling pathway in papillary thyroid carcinoma. **Cancer Res** **2003**;63:1454-7.
48. Soares P, Trovisco V, Rocha AS, Lima J, Castro P, Preto A, et al. BRAF mutations and RET/PTC rearrangements are alternative events in the etiopathogenesis of PTC. **Oncogene** **2003**;22:4578-80.
49. Cohen Y, Xing M, Mambo E, Guo Z, Wu G, Trink B, et al. BRAF mutation in papillary thyroid carcinoma. **JNCI Cancer Spectrum** **2003**;95:625-7.
50. Kim KH, Kang DW, Kim SH, Seong IO, Kang DY. Mutations of the BRAF gene in papillary thyroid carcinoma in a Korean population. **Yonsei Med J** **2004**;45:818-21.
51. Puxeddu E, Moretti S, Elisei R, Romei C, Pascucci R, Martinelli M, et al. BRAF^{V599E} mutation is the leading genetic event in adult sporadic papillary thyroid carcinomas. **J Clin Endocrinol Metab** **2004**;89:2414-20.
52. Melillo RM, Castellone MD, Guarino V, De FV, Cirafici AM, Salvatore G, et al. The RET/PTC-RAS-BRAF linear signaling cascade mediates the motile and mitogenic phenotype of thyroid cancer cells. **J Clin Invest** **2005**;115:1068-81.
53. Mitsutake N, Miyagishi M, Mitsutake S, Akeno N, Mesa JC, Knauf JA, et al. BRAF mediates RET/PTC-induced MAPK activation in thyroid cells: functional support for requirement of the RET/PTC-RAS-BRAF pathway in papillary thyroid carcinogenesis. **Endocrinology** **2005**.
54. Trovisco V, Soares P, Preto A, de Castro IV, Lima J, Castro P, et al. Type and prevalence of BRAF mutations are closely associated with papillary thyroid carcinoma histotype and patients' age but not with tumour aggressiveness. **Virchows Arch** **2005**;446:589-95.
55. Smyth P, Finn S, Cahill S, O'Regan E, Flavin R, O'Leary JJ, et al. ret/PTC and BRAF act as distinct molecular, time-dependent triggers in a sporadic Irish cohort of papillary thyroid carcinoma. **Int J Surg Pathol** **2005**;13:1-8.
56. Kim KH, Suh KS, Kang DW, Kang DY. Mutations of the BRAF gene in papillary thyroid carcinoma and in Hashimoto's thyroiditis. **Pathol Int** **2005**;55:540-5.
57. Nikiforova MN, Ciampi R, Salvatore G, Santoro M, Gandhi M, Knauf JA, et al. Low prevalence of BRAF mutations in radiation-induced thyroid tumors in contrast to sporadic papillary carcinomas. **Cancer Lett** **2004**;209:1-6.
58. Lima J, Trovisco V, Soares P, Maximo V, Magalhães J, Salvatore G, et al. BRAF mutations are not a major event in post-cholesterol childhood thyroid carcinomas. **J Clin Endocrinol Metab** **2004**;89:4267-71.
59. Kumagai A, Namba H, Saenko VA, Ashizawa K, Ohtsuru A, Ito M, et al. Low frequency of BRAF^{T1796A} mutations in childhood thyroid carcinomas. **J Clin Endocrinol Metab** **2004**;89:4280-4.
60. Powell N, Jeremiah S, Morishita M, Dudley E, Bethel J, Bogdanova T, et al. Frequency of BRAF T1796A mutation in papillary thyroid carcinoma relates to age of patient at diagnosis and not to radiation exposure. **J Pathol** **2005**;205:558-64.
61. Nikiforova MN, Stringer JR, Blough R, Medvedovic M, Fagin JA, Nikiforov YE. Proximity of chromosomal loci that participate in radiation-induced rearrangements in human cells. **Science** **2000**;290:138-41.
62. Sugg SL, Zheng L, Rosen IB, Freeman JL, Ezzat S, Asa SL. ret/PTC-1, -2, and -3 oncogene rearrangements in human thyroid carcinomas: implications for metastatic potential? **J Clin Endocrinol Metab** **1996**;81:3360-5.
63. Bongarzone I, Fugazzola L, Vigneri P, Mariani L, Mondellini P, Pacini F, et al. Age-related activation of the tyrosine kinase receptor protooncogenes RET and NTRK1 in papillary thyroid carcinoma. **J Clin Endocrinol Metab** **1996**;81:2006-9.
64. Adeniran AJ, Zhu Z, Gandhi M, Steward DL, Fidler JP, Giordano TJ, et al. Correlation between genetic alterations and microscopic features, clinical manifestations, and prognostic characteristics of thyroid papillary carcinomas. **Am J Surg Pathol** **2006**;30:216-22.
65. Williams D, Baverstock K. Chernobyl and the future: too soon for a final diagnosis. **Nature** **2006**;440:993-4.
66. Bos JL. ras oncogenes in human cancer: a review. **Cancer Res** **1989**;49:4682-9.
67. Vasko V, Ferrand M, Di Cristofaro J, Carayon P, Henry JF, de Micco C. Specific pattern of RAS oncogene mutations in follicular thyroid tumors. **J Clin Endocrinol Metab** **2003**;88:2745-52.
68. Nikiforova MN, Lynch RA, Biddinger PW, Alexander EK, Dorn GW, Tallini G, et al. RAS point mutations and PAX8-PPAR γ rearrangement in thyroid tumors: evidence for distinct molecular pathways in thyroid follicular carcinoma. **J Clin Endocrinol Metab** **2003**;88:2318-26.
69. Lazzereschi D, Mincione G, Coppa A, Ranieri A, Turco A, Baccheschi G, et al. Oncogenes and antioncogenes involved in human thyroid carcinogenesis. **J Exp Clin Cancer Res** **1997**;16:325-32.
70. Sugg SL, Ezzat S, Zheng L, Freeman JL, Rosen IB, Asa SL. Oncogene profile of papillary thyroid carcinoma. **Surgery** **1999**;125:46-52.

71. Zhu Z, Gandhi M, Nikiforova MN, Fischer AH, Nikiforov YE. Molecular profile and clinical-pathologic features of the follicular variant of papillary thyroid carcinoma. An unusually high prevalence of ras mutations. **Am J Clin Pathol** 2003;120:71-7.
72. Di Cristofaro J, Marcy M, Vasko V, Sebag F, Fakhry N, Wynford-Thomas D, et al. Molecular genetic study comparing follicular variant versus classic papillary thyroid carcinomas: association of N-ras mutation in codon 61 with follicular variant. **Hum Pathol** 2006;37:824-30.
73. Roque L, Castedo S, Gomes P, Soares P, Clode A, Soares J. Cytogenetic findings in 18 follicular thyroid adenomas. **Cancer Genet Cytogenet** 1993;67:1-6.
74. Kroll TG, Sarraf P, Pecciarini L, Chen CJ, Mueller E, Spiegelman BM, et al. PAX8-PPAR γ 1 fusion oncogene in human thyroid carcinoma [corrected]. **Science** 2000;289:1357-60.
75. Au AY, McBride C, Wilhelm KG, Jr., Koenig RJ, Speller B, Cheung L, et al. PAX8-peroxisome proliferator-activated receptor gamma (PPAR γ) disrupts normal PAX8 or PPAR γ transcriptional function and stimulates follicular thyroid cell growth. **Endocrinology** 2006;147:367-76.
76. Cheung L, Messina M, Gill A, Clarkson A, Learoyd D, Delbridge L, et al. Detection of the PAX8-PPAR γ fusion oncogene in both follicular thyroid carcinomas and adenomas. **J Clin Endocrinol Metab** 2003;88:354-7.
77. Nikiforova MN, Biddinger PW, Caudill CM, Kroll TG, Nikiforov YE. PAX8-PPAR γ rearrangement in thyroid tumors: RT-PCR and immunohistochemical analyses. **Am J Surg Pathol** 2002;26:1016-23.
78. Marques AR, Espadinha C, Catarino AL, Moniz S, Pereira T, Sobrinho LG, et al. Expression of PAX8-PPAR γ 1 rearrangements in both follicular thyroid carcinomas and adenomas. **J Clin Endocrinol Metab** 2002;87:3947-52.
79. Wallace DC. Diseases of the mitochondrial DNA. **Annu Rev Biochem** 1992;61:1175-212.
80. Polyak K, Li Y, Zhu H, Lengauer C, Willson JK, Markowitz SD, et al. Somatic mutations of the mitochondrial genome in human colorectal tumours. **Nat Genet** 1998;20:291-3.
81. Richard SM, Bailliet G, Paez GL, Bianchi MS, Peltomaki P, Bianchi NO. Nuclear and mitochondrial genome instability in human breast cancer. **Cancer Res** 2000;60:4231-7.
82. Habano W, Sugai T, Nakamura SI, Uesugi N, Yoshida T, Sasou S. Microsatellite instability and mutation of mitochondrial and nuclear DNA in gastric carcinoma. **Gastroenterology** 2000;118:835-41.
83. Yeh JJ, Lunetta KL, van Orsouw NJ, Moore FD, Jr., Mutter GL, Vijg J, et al. Somatic mitochondrial DNA (mtDNA) mutations in papillary thyroid carcinomas and differential mtDNA sequence variants in cases with thyroid tumours. **Oncogene** 2000;19:2060-6.
84. Maximo V, Soares P, Lima J, Cameselle-Teijeiro J, Sobrinho-Simões M. Mitochondrial DNA somatic mutations (point mutations and large deletions) and mitochondrial DNA variants in human thyroid pathology: a study with emphasis on Hurthle cell tumours. **Am J Pathol** 2002;160:1857-65.
85. Maximo V, Botelho T, Capela J, Soares P, Lima J, Taveira A, et al. Somatic and germline mutation in GRIM-19, a dual function gene involved in mitochondrial metabolism and cell death, is linked to mitochondrion-rich (Hurthle cell) tumours of the thyroid. **Br J Cancer** 2005;92:1892-8.
86. Angell JE, Lindner DJ, Shapiro PS, Hofmann ER, Kalvakolanu DV. Identification of GRIM-19, a novel cell death-regulatory gene induced by the interferon-beta and retinoic acid combination, using a genetic approach. **J Biol Chem** 2000;275:33416-26.
87. Grieco M, Santoro M, Berlingieri MT, Melillo RM, Donghi R, Bongarzone I, et al. PTC is a novel rearranged form of the ret proto-oncogene and is frequently detected in vivo in human thyroid papillary carcinomas. **Cell** 1990;60:557-63.
88. Vickery AL, Jr., Carcangiu ML, Johannessen JV, Sobrinho-Simões M. Papillary carcinoma. **Semin Diagn Pathol** 1985;2:90-100.
89. Basolo F, Giannini R, Monaco C, Melillo RM, Carlomagno F, Pancrazi M, et al. Potent mitogenicity of the RET/PTC3 oncogene correlates with its prevalence in tall-cell variant of papillary thyroid carcinoma. **Am J Pathol** 2002;160:247-54.
90. Nikiforova MN, Kimura ET, Gandhi M, Biddinger PW, Knauf JA, Basolo F, et al. BRAF mutations in thyroid tumors are restricted to papillary carcinomas and anaplastic or poorly differentiated carcinomas arising from papillary carcinomas. **J Clin Endocrinol Metab** 2003;88:5399-404.
91. Trovisco V, Vieira de Castro I, Soares P, Maximo V, Silva P, Magalhães J, et al. BRAF mutations are associated with some histological types of papillary thyroid carcinoma. **J Pathol** 2004;202:247-51.
92. Fugazzola L, Mannavola D, Cirello V, Vannucchi G, Muzza M, Vicentini L, et al. BRAF mutations in an Italian cohort of thyroid cancers. **Clin Endocrinol (Oxf)** 2004;61:239-43.
93. Harach HR, Williams GT, Williams ED. Familial adenomatous polyposis associated thyroid carcinoma: a distinct type of follicular cell neoplasm. **Histopathology** 1994;25:549-61.
94. Cameselle-Teijeiro J, Chan JK. Cribriform-morular variant of papillary carcinoma: a distinctive variant representing the sporadic counterpart of familial adenomatous polyposis-associated thyroid carcinoma? **Mod Pathol** 1999;12:400-11.
95. Cameselle-Teijeiro J, Ruiz-Ponte C, Loidi L, Suarez-Penaranda J, Baltar J, Sobrinho-Simões M. Somatic but not germline mutation of the APC gene in a case of cribriform-morular variant of papillary thyroid carcinoma. **Am J Clin Pathol** 2001;115:486-93.
96. Iwama T, Konishi M, Iijima T, Yoshinaga K, Tominaga T, Koike M, et al. Somatic mutation of the APC gene in thyroid carcinoma associated with familial adenomatous polyposis. **Jpn J Cancer Res** 1999;90:372-6.
97. Cetta F, Montalto G, Gori M, Curia MC, Cama A, Olschwang S. Germline mutations of the APC gene in patients with familial adenomatous polyposis-associated thyroid carcinoma: results from a European cooperative study. **J Clin Endocrinol Metab** 2000;85:286-92.
98. Nakabashi CC, Guimarães GS, Michaluart P, Jr., Ward LS, Cerutti JM, Maciel RM. The expression of PAX8-PPAR γ rearrangements is not specific to follicular thyroid carcinoma. **Clin Endocrinol (Oxf)** 2004;61:280-2.
99. Salvatore G, Giannini R, Faviana P, Caleo A, Migliaccio I, Fagin JA, et al. Analysis of BRAF point mutation and RET/PTC rearrangement refines the fine-needle aspiration diagnosis of papillary thyroid carcinoma. **J Clin Endocrinol Metab** 2004;89:5175-80.
100. Cheung CC, Carydis B, Ezzat S, Bedard YC, Asa SL. Analysis of ret/PTC gene rearrangements refines the fine needle aspiration diagnosis of thyroid cancer. **J Clin Endocrinol Metab** 2001;86:2187-90.
101. Xing M, Tufano RP, Tufano AP, Basaria S, Ewertz M, Rosenbaum E, et al. Detection of BRAF mutation on fine needle aspiration biopsy specimens: a new diagnostic tool for papillary thyroid cancer. **J Clin Endocrinol Metab** 2004; 89:2867-72.
102. Hayashida N, Namba H, Kumagai A, Hayashi T, Ohtsuru A, Ito M, et al. A rapid and simple detection method for the BRAF(T1796A) mutation in fine-needle aspirated thyroid carcinoma cells. **Thyroid** 2004;14:910-5.
103. Domingues R, Mendonça E, Sobrinho L, Bugalho MJ. Searching for RET/PTC rearrangements and BRAF V599E mutation in thyroid aspirates might contribute to establish a preoperative diagnosis of papillary thyroid carcinoma. **Cytopathology** 2005; 16: 27-31.
104. Cohen Y, Rosenbaum E, Clark DP, Zeiger MA, Umbricht CB, Tufano RP, et al. Mutational analysis of BRAF in fine needle aspiration biopsies of the thyroid: a potential application for the preoperative assessment of thyroid nodules. **Clin Cancer Res** 2004;10:2761-5.
105. Mojica WD, Khoury T. Presence of the BRAF^{V600E} point mutation in morphologically benign appearing thyroid inclusions of cervical lymph nodes. **Endocr Pathol** 2006;17:183-9.
106. Jemal A, Murray T, Ward E, Samuels A, Tiwari RC, Ghafoor A, et al. Cancer statistics, 2005. **CA Cancer J Clin** 2005;55:10-30.
107. Sawka AM, Thephamongkhon K, Brouwers M, Thabane L, Browman G, Gerstein HC. Clinical review 170: A systematic review and metaanalysis of the effectiveness of radioactive iodine remnant ablation for well-differentiated thyroid cancer. **J Clin Endocrinol Metab** 2004;89:3668-76.

108. Basolo F, Molinaro E, Agate L, Pinchera A, Pollina L, Chiappetta G, et al. RET protein expression has no prognostic impact on the long-term outcome of papillary thyroid carcinoma. **Eur J Endocrinol** **2001**;145:599-604.
109. Puxeddu E, Moretti S, Giannico A, Martinelli M, Marino C, Avenia N, et al. Ret/PTC activation does not influence clinical and pathological features of adult papillary thyroid carcinomas. **Eur J Endocrinol** **2003**;148:505-13.
110. Xing M, Westra WH, Tufano RP, Cohen Y, Rosenbaum E, Rhoden KJ, et al. BRAF mutation predicts a poorer clinical prognosis for papillary thyroid cancer. **J Clin Endocrinol Metab** **2005**;90:6373-9.
111. Namba H, Nakashima M, Hayashi T, Hayashida N, Maeda S, Rogounovitch TI, et al. Clinical implication of hot spot BRAF mutation, V599E, in papillary thyroid cancers. **J Clin Endocrinol Metab** **2003**;88:4393-7.
112. Kim TY, Kim WB, Rhee YS, Song JY, Kim JM, Gong G, et al. The BRAF mutation is useful for prediction of clinical recurrence in low-risk patients with conventional papillary thyroid carcinoma. **Clin Endocrinol (Oxf)** **2006**;65:364-8.
113. Quiros RM, Ding HG, Gattuso P, Prinz RA, Xu X. Evidence that one subset of anaplastic thyroid carcinomas are derived from papillary carcinomas due to BRAF and p53 mutations. **Cancer** **2005**;103:2261-8.
114. Soares P, Trovisco V, Rocha AS, Feijão T, Rebocho AP, Fonseca E, et al. BRAF mutations typical of papillary thyroid carcinoma are more frequently detected in undifferentiated than in insular and insular-like poorly differentiated carcinomas. **Virchows Arch** **2004**;444:572-6.
115. Knauf JA, Ma X, Smith EP, Zhang L, Mitsutake N, Liao XH, et al. Targeted expression of BRAF^{V600E} in thyroid cells of transgenic mice results in papillary thyroid cancers that undergo dedifferentiation. **Cancer Res** **2005**;65:4238-45.
116. Xu X, Quiros RM, Gattuso P, Ain KB, Prinz RA. High prevalence of BRAF gene mutation in papillary thyroid carcinomas and thyroid tumor cell lines. **Cancer Res** **2003**;63:4561-7.
117. Sedliarou I, Saenko V, Lantsov D, Rogounovitch T, Namba H, Abrosimov A, et al. The BRAF^{T1796A} transversion is a prevalent mutational event in human thyroid microcarcinoma. **Int J Oncol** **2004**;25:1729-35.
118. Kim TY, Kim WB, Song JY, Rhee YS, Gong G, Cho YM, et al. The BRAF mutation is not associated with poor prognostic factors in Korean patients with conventional papillary thyroid microcarcinoma. **Clin Endocrinol (Oxf)** **2005**;63:588-93.
119. Akslen LA, LiVolsi VA. Prognostic significance of histologic grading compared with subclassification of papillary thyroid carcinoma. **Cancer** **2000**;88:1902-8.
120. DeLellis RA. Pathology and genetics of thyroid carcinoma. **J Surg Oncol** **2006**;94:662-9.
121. Michels JJ, Jacques M, Henry-Amar M, Bardet S. Prevalence and prognostic significance of tall cell variant of papillary thyroid carcinoma. **Hum Pathol** **2007**;38:212-9.
122. Baudin E, Schlumberger M. New therapeutic approaches for metastatic thyroid carcinoma. **Lancet Oncol** **2007**;8:148-56.
123. Lyons JF, Wilhelm S, Hibner B, Bollag G. Discovery of a novel Raf kinase inhibitor. **Endocr Relat Cancer** **2001**;8:219-25.
124. Wan PT, Garnett MJ, Roe SM, Lee S, Niculescu-Duvaz D, Good VM, et al. Mechanism of activation of the RAF-ERK signaling pathway by oncogenic mutations of B-RAF. **Cell** **2004**;116:855-67.
125. Salvatore G, De Falco V, Salerno P, Nappi TC, Pepe S, Troncone G, et al. BRAF is a therapeutic target in aggressive thyroid carcinoma. **Clin Cancer Res** **2006**;12:1623-9.
126. Ouyang B, Knauf JA, Smith EP, Zhang L, Ramsey T, Yusuff N, et al. Inhibitors of Raf kinase activity block growth of thyroid cancer cells with RET/PTC or BRAF mutations in vitro and in vivo. **Clin Cancer Res** **2006**;12:1785-93.
127. da Rocha Dias S, Friedlos F, Light Y, Springer C, Workman P, Marais R. Activated B-RAF is an Hsp90 client protein that is targeted by the anticancer drug 17-allylamino-17-demethoxygeldanamycin. **Cancer Res** **2005**;65:10686-91.
128. Grbovic OM, Basso AD, Sawai A, Ye Q, Friedlander P, Solit D, et al. V600E B-Raf requires the Hsp90 chaperone for stability and is degraded in response to Hsp90 inhibitors. **Proc Natl Acad Sci U S A** **2006**;103:57-62.
129. Carlomagno F, Anaganti S, Guida T, Salvatore G, Troncone G, Wilhelm SM, et al. BAY 43-9006 inhibition of oncogenic RET mutants. **J Natl Cancer Inst** **2006**;98:326-34.
130. Kebebew E, Peng M, Reiff E, Treseler P, Woeber KA, Clark OH, et al. A phase II trial of rosiglitazone in patients with thyroglobulin-positive and radioiodine-negative differentiated thyroid cancer. **Surgery** **2006**;140:960-6; discussion 6-7.
131. Park JW, Zarnegar R, Kanauchi H, Wong MG, Hyun WC, Ginzinger DG, et al. Troglitazone, the peroxisome proliferator-activated receptor-gamma agonist, induces antiproliferation and redifferentiation in human thyroid cancer cell lines. **Thyroid** **2005**;15:222-31.
132. Copland JA, Marlow LA, Kurakata S, Fujiwara K, Wong AK, Kreinest PA, et al. Novel high-affinity PPAR γ agonist alone and in combination with paclitaxel inhibits human anaplastic thyroid carcinoma tumor growth via p21WAF1/CIP1. **Oncogene** **2006**;25:2304-17.
133. Espinosa AV, Porchia L, Ringel MD. Targeting BRAF in thyroid cancer. **Br J Cancer** **2007**;96:16-20.

Address for correspondence:

Manuel Sobrinho-Simões
 IPATIMUP
 Rua Dr. Roberto Frias s/n
 4200-465 Porto, Portugal
 Fax: (351) 22557-0799
 E-mail: ssimoese@ipatimup.pt