

Selenium Protective Activity Against Aflatoxin B₁ Adverse Affects on *Drosophila melanogaster*

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ABSTRACT

The aim of this work was to investigate the effects of AFB₁ and AFB₁+Se⁴⁺ on various developmental stages of *Drosophila melanogaster* were investigated. Both different concentrations of AFB₁ and Se⁴⁺ applied with AFB₁ were fed during the flies developmental period (egg, larva and pupae). When F₁ progeny of control and application groups were compared with each other, AFB₁ was found to have extending the process of metamorphosis and decreasing the total number of offsprings. But, these negative effects were inhibited with selenium treatment at different concentrations (4.0 and 8.0 ppm). These results suggested that selenium could effectively inhibit AFB₁-induced abnormalities of the developmental stages of *D.melanogaster*.

Key words: Selenium, Aflatoxin B₁, Developmental stages, Protective effect, *D. melanogaster*

INTRODUCTION

Aflatoxins are a group of naturally occurring, highly toxic mycotoxins that contain a characteristic dihydrobisfuran moiety in their molecular structures. These fungal metabolites are produced by specific strains of *Aspergillus flavus* and *Aspergillus parasiticus*, which are commonly found as contaminants of a variety of foodstuffs. The most common form of aflatoxins, aflatoxin B₁ (AFB₁), has been shown to be a potent mutagen, hepatocarcinogen and teratogen in several species of experimental animals (Wogan, 1973; Garner and Martin, 1979; Garner, 1980; Busby and Wogan, 1985).

These adverse biological effects of AFB₁ are manifested after its metabolic activation and subsequent interaction with cellular macromolecules (Wogan, 1973; Garner and Martin, 1979; Garner, 1980; Essigman et al., 1982). Several factors naturally present in foods of

common consumption have been shown to modify these critical reaction of AFB₁, i.e. its microsomal activation and interaction with DNA (Bhattacharya et al., 1984). These include vitamin, trace metals, fatty acids, flavonoids, phenolic acids and other compounds. It is expected that these substances will also counteract the adverse biological effects of AFB₁. Several studies have demonstrated that certain selenium dietary provide a protective effect against AFB₁ toxicity in several animal species. Newberne and Conner (1974) first reported that selenium supplementation up to a dietary level 1.00 ppm progressively reduced the acute toxicity of AFB₁ in rat, while greater levels enhanced the observed mortality. Studies that have been initiated to test this concept recorded that selenium and certain trace metals have exceptional ability to modify microsome mediated mutagenic activation of AFB₁ in *Salmonella typhimurium* strains TA100 and TA98 (Francis et al., 1988).

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Selenium is an essential micronutrient in the mammalian diet and deficiency of this trace element can cause a variety of severe pathological conditions (Diplock, 1981). There is increasing evidence that selenium can act as an anticarcinogen and inhibit tumor initiation and progression (Horvath and Ip, 1983; Darodo et al., 1985; Milner, 1985). Epidemiological studies have revealed an inverse relation between cancer risk and dietary intake or geographic levels of selenium content. Studies in laboratory animals suggested that selenium was also effective in inhibiting AFB₁ induced hepatocarcinogenesis (Baldwin and Parker, 1987; Yu et al., 1988; Lei et al., 1990). This protective effect was confirmed in several later studies with swine and turkeys (Burguera et al., 1983; Davila et al., 1983). In view of the demonstrated protective effect of selenium against the acute toxicity of AFB₁ and the postulated anticarcinogenic effect of selenium studies were conducted by Chen and associates to investigate the possible influence of selenium and vitamin E on certain aspects of the metabolism of AFB₁ in rats and chicks (Chen et al., 1982_a; Chen et al., 1982_b). Their results suggested that combined vitamin E-selenium deficiency enhanced aflatoxin binding to hepatic DNA and RNA in rat. However, the protective effect of selenium against AFB₁ induced teratogenic effects and influence on some development stages of *Drosophila melanogaster* has not been elucidated. The main aim of present study was to see whether selenium has any protective effect against the adverse effects of AFB₁ on *Drosophila melanogaster*.

MATERIALS AND METHODS

The flies used in experiments were Oregon-R wild type (w.t.) strain of *Drosophila melanogaster* Meigen (Diptera: Drosophilidae). This stock had been maintained for many years in culture vials and was, therefore, highly inbred with little genetic variation. The experimental medium was a yeast-agar-sugar medium (Standard *Drosophila* Medium:SDM). Flies were grown and aged in culture bottles containing SDM. All experiments were carried out at 25°C and 40-60% relative humidity.

The tested substance, crystalline aflatoxin B₁ (AFB₁, Acros Organics, No:227340100, New Jersey, USA), was dissolved in a 10% solution of dimethyl sulfoxide (DMSO, Sigma-Aldrich Laborchemicalies GmbH). A sodium selenite solution (4.0 and 8.0 ppm) was prepared by adding of Na₂SeO₃ (Sigma Chemical Co., St. Louis, MO) to distilled water. In our experiments, parental generations of *D.melanogaster* were treated with various concentrations of both AFB₁ (0.2, 0.5 and 0.8 ppm AFB₁/ ml SDM) and sodium selenite (4.0 and 8.0 ppm Se⁴⁺/ ml SDM). To test the effects of the AFB₁ and AFB₁+Se⁴⁺ on the growth of *D.melanogaster*, this species was cultivated in 9 different media, shown in Table 1. All the females used in the experiments were virgins. The flies with the same age were used for experiments and seven pairs were mated. Then, the developmental stages were followed daily. Offsprings were counted everyday from the first day of eclosion and phenotypic abnormalities of F₁ individuals examined under microscope were observed. Statistical analysis of data was done using Duncan's one-way range test.

Table 1 - The composition of different media used for this study

Composition				
Medium	SDM (ml)	DMSO (ml/100ml)	AFB ₁ (ppm)	Selenium (ppm)
Control (C)	100	—	—	—
I (C+DMSO)	100	10	—	—
II	100	—	0.2	—
III	100	—	0.5	—
IV	100	—	0.8	—
V	100	—	0.2	4.0
VI	100	—	0.2	8.0
VII	100	—	0.5	4.0
VIII	100	—	0.5	8.0
IX	100	—	0.8	4.0
X	100	—	0.8	8.0

RESULTS AND DISCUSSION

The effects of the AFB₁ and AFB₁+Se⁴⁺ on the development stages of *D. melanogaster* are shown in Table 2. In the laboratory at 25±1°C, the life cycle (egg-adult) was 9 days (Uysal and Bahçeci, 1995; Uysal et al., 2002). In control (C), C+DMSO (Medium I) and all experimental groups laid eggs were observed at the second day of

mating. In control and Medium I, first adult was emerged from the pupa at 9th days of mating. But in the presence of 0.2, 0.5 and 0.8 ppm of AFB₁ (Medium II, III and IV), various development stages were retarded. At these concentrations of AFB₁, first offsprings of F₁ progeny were seen at 10th, 11th and 13th days, respectively (Table 2).

Table 2 - Occurrence of egg- adult developmental stages after application of different doses of AFB₁ and AFB₁+ Se⁴⁺

Developmental stages	Days of stages at control and applications (ppm)										
	C	I	II	V	VI	III	VII	VIII	IV	IX	X
Mating	1	1	1	1	1	1	1	1	1	1	1
Egg	2	2	2	2	2	2	2	2	2	2	2
1 th instar larvae	3	3	4	3	3	4	3	3	5	3	3
2 nd instar larvae	4	4	5	4	4	6	4	4	7	4	4
3 rd instar larvae	5	5	6	5	5	7	5	5	9	5	5
Prepupa	6	6	7	6	6	8	6	6	10	6	6
Pupa	7	7	8	7	7	10	7	7	11	7	7
Adult	9	9	10	9	9	11	9	9	13	9	9

On the other hand, one of the most interesting results obtained was that the developmental stages of *D. melanogaster* were observed at the same days with control (C) with shown that Se⁴⁺ applications (Medium. V-X). Table 2, both 4.0 and 8.0 ppm Se applications prevented the negative effects of AFB₁ on developmental stages and these stages were completed within their normal developmental phases (9 days).

Similar findings have been observed in previous studies also. For example, Lalor et al., (1976) found that growth on AFB₁ media of *D. melanogaster* caused significant increases in egg-to- adult developmental time. Larval and pupal toxic effects caused by AFB₁ have also been demonstrated by various authors in *Musca domestica* (Beard and Walton, 1971) and *D. melanogaster* (Kirk et al., 1971). Besides, according to Chinnici et al., (1979), second and third larvae of strain A-9 showed significant mortality rates when grown at 0.88 ppm AFB₁. At the 0.88 ppm concentration, both A-9 and A-11 strains showed significant mortality rates for first instar larvae, but the A-9 larvae died at higher rates than the A-11 larvae. Similar results were also obtained after the application of mycotoxins such as rubratoxin B, patulin and diacetoxyscirpenol (Reis, 1975).

The total number of offsprings (for F₁ progeny) of aflatoxin B₁ treated *D. melanogaster* were

affected by all application groups (Medium II, III and IV) of toxin (Table 3). The maximum inhibition effect was produced with higher toxin concentration (0.8 ppm/ ml, Medium IV). As seen in Table 3 too, the difference between the number of the offsprings of control (2305) and control+DMSO (Medium I, 2218) was statistically unimportant (P> 0.05). But, as 2305 F₁ individuals were in control group, the number of offsprings decreased in the application groups (Medium II, III and IV), depending on the dosage increase. Statistical analysis showed that this decreasing was significant (P< 0.01).

In our unpublished study, besides the above mentioned dosages of AFB₁, three different dosages have also been examined (1.1, 1.4 and 1.7 ppm/ ml), but, no offsprings were obtained. Furthermore, metamorphosis stopped in 1st, 2nd and 3rd instar larvae according to the increasing concentrations and was not completed.

For many researchers, the most important reason of the decrease of offsprings in *D. melanogaster* exposed AFB₁ was the decrease of fertility, both in male and female (Matsumura and Knight, 1967; Chinnici et al., 1976). Similar effects have also been observed in different animal groups such as mice, rat and mouse (Bashandy et al., 1994; Nair and Verma, 2000; Verma and Nair, 2001).

Table 3 - The effect of different concentrations of selenium together with AFB₁ on the total number of F₁ individuals and formation of malformed individuals (%)

Medium	F ₁ progeny	Malformed individuals	%
C	2305	9	0.39
I	2218	7	0.32
II	1120**	73	65.1
V	1623*	52	32.04
VI	1761*	20	11.36
III	924**	75	81.1
VII	1536*	50	32.55
VIII	1602*	16	0.99
IV	39**	3	76.92
IX	1062*	36	33.8
X	1130*	10	0.88

Duncan's one-way range test: * P<0.05 ; ** P<0.01

Besides, the extreme sensitivity of larvae and pupae and also their mortality in high concentrations (1.4 ppm) were one of the most important reasons of the decrease in the number of offsprings (Kirk et al., 1971; Llwellyn and Chinnici, 1978).

Furthermore, when F₁ individuals were examined as phenotypic, malformed individuals were also observed and malformation was concentrated mainly on wing, leg and thorax and neither the formation of extremities nor their lackness have been found. While the rate of the malformed individuals in the control was 0.39%, they changed between 65.17-81.16% in Medium II, III and IV, respectively. This ratio of change was statistically important (P<0.01). Depending on the increase of concentration in Medium II, III and IV, both the number of individuals decreased and malformation increased (Table 3).

In previous studies, similar results have also been obtained. For example, teratogenic effects of AFB₁ were observed on fetuses of hamster, mice, rat and cow by Aleksandrowicz and Smyk (1973). These teratogenic effects were also found as follows: decreases in body size (Chinnici et al., 1976) and wing length (Lalor et al., 1976), formation of tumor on different body parts (Sidorov et al., 2001), hepatocarcinogen effects at vertebrates (Pier, 1981). These findings are in accordance with our results. According to our data, the most important reason for the formation of phenotypic abnormalities was delaying of metamorphosis, because AFB₁ caused some faults during transcription of developmental genes and defects

in homeotic genes which affected the final condition of imaginal discs (Wallace et al., 1991). It has also been reported that AFB₁ is metabolically activated by the microsomal mixed-function monooxygenase system to the 8, 9-epoxide, which readily binds to the nucleophilic sites in DNA to form DNA adducts (Stark, 1986). AFB₁ preferentially attacks guanine residues in DNA and the major adduct for is 8,9- dihydro-9-hydroxy-(N7-guanyl) AFB₁, which accounts for over 90% of the total adducts (Essigmann et al., 1982). The formation of AFB₁- DNA adducts in the target cell gives rise to promutagenic sites in DNA.

The addition of different concentrations of Se⁴⁺ to aflatoxin containing media (Medium V-X), was found to be interesting for the increase of F₁ individuals and the decrease of the rate of malformation. The individuals number belonging to the F₁ was 1120 and the rate of malformed individuals was 65.17 % in the medium containing 0.2 ppm AFB₁ (Medium II). In the media containing 4.0 and 8.0 ppm Se⁴⁺ (Medium V and VI), F₁ individuals were 1623 and 1761, respectively. Furthermore, the number of malformation decreased from 65.17% to 11.36%. Similarly, while the number of F₁ individuals was 924 in the medium of 0.5 ppm AFB₁ (Medium III), this number was 1536 and 1602 in Se⁴⁺ added media (Medium VII and VIII). The rate of malformed individuals dropped from 81.16 to 0.99%. The antagonistic effect of Se⁴⁺ was clearly observed in 0.8 ppm AFB₁ (Medium IV, Table 3).

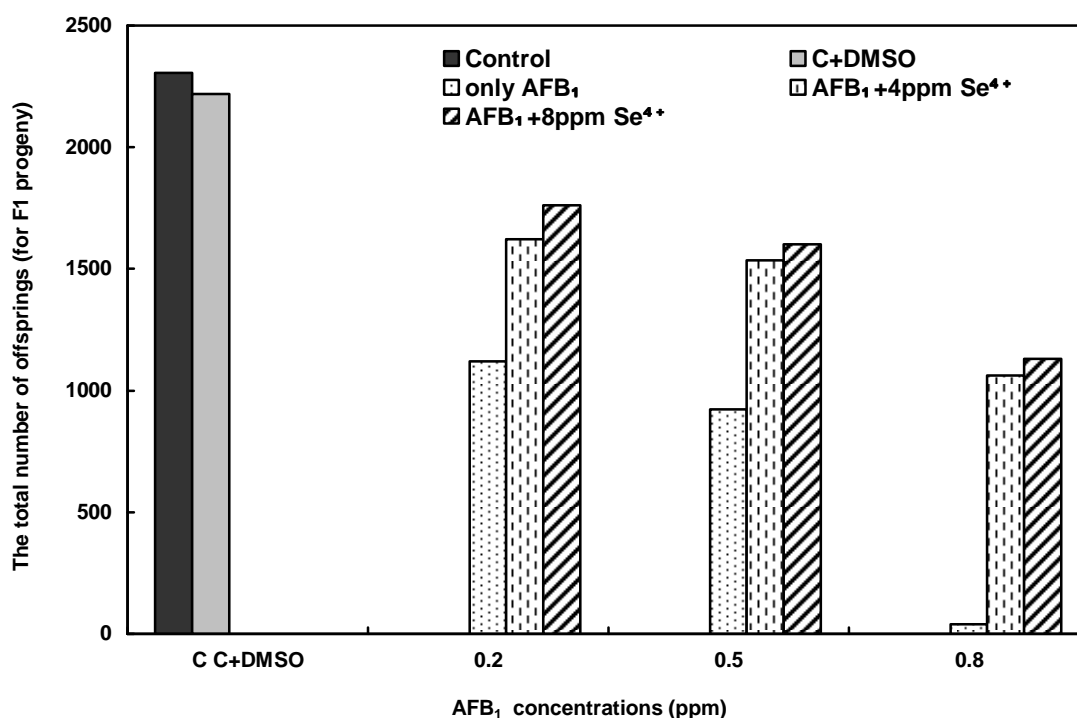


Figure 1 - Comparison of the total number of offsprings obtained from *D.melanogaster* treated with different concentrations of AFB₁ and AFB₁+Se⁴⁺.

As only 39 individuals were counted at the 0.8 ppm AFB₁, the individuals number with the increasing Se⁴⁺ concentrations was found to be important. This was statistically important ($P < 0.05$), (Fig. 1).

Although protective effects of selenium on teratogenic disorders has not been identified, some studies demonstrated that it has protective effect against the acute toxicity of AFB₁ and carcinogenesis in animal species (Chen et al., 1982a). Carcinogen-induced cellular oxidative damage and its role in the cytotoxicity and carcinogenesis have attracted much attention (Imlay and Linn, 1988; Farber et al., 1990). Oxidative damage usually refers to the impairment of the function of cellular components, e.g., enzymes, nucleic acids, membranes and proteins, by reactive oxygen species such as superoxide radicals (O_2^-), hydroxyl free radical (OH), and hydrogen peroxide (H_2O_2). Oxidative damage causes cellular damage, in particular ageing and some irreversible pathologic processes such as cancer (Gracy et al., 1999). Therefore, it could be possible that AFB₁ also contributed to the genotoxic damage. The identification and function of free radicals have increased the interest in

substances with antioxidant properties such as selenium, vitamin A, β karotene etc. Selenium has antimutagenic and anticarcinogenic activity. Previous studies on the protective effects of antioxidants such as selenium and vitamin A against the cytotoxicity and genotoxicity of AFB₁ mostly focused on the metabolism and detoxification of AFB₁ or the formation of AFB₁-DNA adduct (Chen et al., 1982b; Mandel et al., 1987; Decoudu et al., 1992). It was also suggested that the protective effect of selenium could be mediated through a cellular mechanism related to glutathione detoxification pathways (Shen et al., 1994). Glutathione has been shown to play an important role in the detoxification of AFB₁ through conjugating reactions catalyzed by glutathione S transferase. Selenium is closely involved in metabolic pathways of glutathione, being an essential element in the selenoprotein glutathione peroxidase (GPX). Because selenium is a component part of glutathione peroxidase, it is possible that selenium neutralizes free radicals present in cell following the hydrogen peroxide treatment that induces DNA damage (Bronzetti et al., 2001).

Our results have also showed that selenium has a protective effect on the development disorders of *D. melanogaster*. It could be concluded that more investigations should be carried on in order to understand whether or not the mechanism of the effect of selenium on *D. melanogaster* is as shown in the present experiments.

RESUMO

O objetivo deste trabalho foi investigar os efeitos da AFB₁ e AFB₁ + Se⁴ em vários estágios de desenvolvimento da *Drosophila melanogaster*. Ambos diferentes concentrações da AFB₁ e Se⁴⁺ aplicado com AFB₁ foram alimentados durante a fase de desenvolvimento da mosca (ovo, larva e pulpa). Quando a progenese F₁ do controle e aplicações foram comparadas com outros grupos, AFB₁ ampliou o processo de metamórfose e na redução do número total de ovos. Porém esses efeitos negativos foram inibidos com o tratamento com selênio em diferentes concentrações (4.0 e 8.0 ppm). Esses resultados sugerem que o selênio pode efetivamente inibir AFB₁ que induz anomalias nos estágios do desenvolvimento da *Drosophila melanogaster*

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