

Antifouling activity of twelve demosponges from Brazil

Ribeiro, SM.^{a,b,*}, Rogers, R.^b, Rubem, AC.^b, Da Gama, BAP.^b, Muricy, G.^a and Pereira, RC.^b

^aMuseu Nacional, Universidade Federal do Rio de Janeiro, Quinta da Boa Vista s/nº, São Cristóvão, CEP 20.940-040, Rio de Janeiro, RJ, Brazil

^bDepartamento de Biologia Marinha, Universidade Federal Fluminense, CP 100.644, CEP 24.001-970, Niterói, RJ, Brazil
*e-mail: suzimir@yahoo.com.br

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Abstract

Benthic marine organisms are constantly exposed to fouling, which is harmful to most host species. Thus, the production of secondary metabolites containing antifouling properties is an important ecological advantage for sessile organisms and may also provide leading compounds for the development of antifouling paints. High antifouling potential of sponges has been demonstrated in the Indian and Pacific oceans and in the Caribbean and Mediterranean seas. Brazilian sponges remain understudied concerning antifouling activities. Only two scientific articles reported this activity in sponges of Brazil. The objective of this study was to test crude extracts of twelve species of sponges from Brazil against the attachment of the mussel *Perna perna* through laboratorial assays, and highlight promising species for future studies. The species *Petromica citrina*, *Amphimedon viridis*, *Desmapsamma anchorata*, *Chondrosia* sp., *Polymastia janeirensis*, *Tedania ignis*, *Aplysina fulva*, *Mycale angulosa*, *Hymeniacidon heliophila*, *Dysidea etheria*, *Tethya rubra*, and *Tethya maza* were frozen and freeze-dried before extraction with acetone or dichloromethane. The crude extract of four species significantly inhibited the attachment of byssus: *Tethya rubra* (p = 0.0009), *Tethya maza* (p = 0.0039), *Petromica citrina* (p = 0.0277), and *Hymeniacidon heliophila* (p = 0.00003). These species, specially, should be the target of future studies to detail the substances involved in the ability antifouling well as to define its amplitude of action.

Keywords: Porifera, Demospongiae, antifouling, *Perna perna*, Brazil.

Atividade anti-incrustante de doze demosponjas do Brasil

Resumo

Organismos bentônicos marinhos estão expostos constantemente à incrustação, que pode ser danosa para a maioria das espécies. Assim, a produção de metabólitos secundários com propriedades anti-incrustantes é uma vantagem ecológica importante para organismos sésseis e pode também orientar o estudo de substâncias para o desenvolvimento de tintas anti-incrustantes. O alto potencial anti-incrustante de esponjas tem sido demonstrado nos oceanos Índico e Pacífico, nos mares Mediterrâneo e Caribenho. Esponjas brasileiras permanecem pouco estudadas em relação à atividade anti-incrustante. Apenas dois artigos científicos registraram essa atividade em esponjas do Brasil. O objetivo desse estudo foi testar os extratos brutos de doze espécies de esponjas do Brasil contra a fixação do molusco *Perna perna* através de ensaios laboratoriais e também destacar espécies promissoras para estudos futuros. As espécies *Petromica citrina*, *Amphimedon viridis*, *Desmapsamma anchorata*, *Chondrosia* sp., *Polymastia janeirensis*, *Tedania ignis*, *Aplysina fulva*, *Mycale angulosa*, *Hymeniacidon heliophila*, *Dysidea etheria*, *Tethya rubra* e *Tethya maza* foram congeladas e liofilizadas logo após a coleta e posteriormente procedeu-se a extração com acetona ou diclorometano. O extrato bruto de quatro espécies inibiu significativamente a fixação de bissos: *Tethya rubra* (p = 0.0009), *Tethya maza* (p = 0.0039), *Petromica citrina* (p = 0.0277), e *Hymeniacidon heliophila* (p = 0.00003). Essas espécies, especialmente, devem ser priorizadas em estudos futuros para detalhamento das substâncias envolvidas na capacidade anti-incrustante, bem como para definir sua amplitude de ação.

Palavras-chave: Porifera, Demospongiae, anti-incrustação, *Perna perna*, Brazil.

1. Introduction

Benthic marine organisms are constantly affected by the settlement of larvae, propagules and microorganisms on their surface (Railkin, 2003; Harder, 2009). Sponges

often live in habitats with high level of spatial competition, such as coral reefs, but most sponge species have their surface free of fouling organisms (Rutzler, 1978; Diaz and Rutzler, 2001; Cedro et al., 2007). The presence of biofouling over the surface of a benthic organism may

increase competition by space or food supply and may also affect growth and reproduction (Jackson and Buss, 1975; Orth and van Montfrans, 1984). Secondary metabolites may prevent or reduce this type of interaction, representing an important ecological advantage for benthic marine organisms (da Gama *et al.*, 2008).

Besides the question of the ecological role of antifouling molecules of marine organisms, they are a promising tool for the development of commercial antifouling paints (Hellio *et al.*, 2009). Biofouling on artificial structures causes financial losses of over \$6.5 billion dollars per year due to reduction in navigation efficiency of commercial ships caused by the increased friction by the irregular shape of the fouling organism (Holmes, 1970; Houghton, 1978; da Gama *et al.*, 2003; Bhadury and Wright, 2004). Other important effect of biofouling on ship hulls is the dispersion of invasive species that may cause serious damage to native communities (Gollasch, 2002). The problem of marine fouling for navigation is so old that ancient civilizations such as the phoenicians (700 B.C.) used waxes, tar, asphalt and pitch on ship wood to avoid these organisms (Callow, 1990; Almeida *et al.*, 2007). The first antifouling paints arose in the mid 19th century using a strong toxic substance dispersed in a polymeric binder (Almeida *et al.*, 2007). Around 1950, the development of organometallic paints (with tin, arsenic, mercury and others) preceded tributyltin-based antifouling paints, which became famous due to their efficiency and to their toxic effects on marine organisms and environment (Ruiz *et al.*, 1995; Evans *et al.*, 2000; Fernandez *et al.*, 2005). Thus, several countries control the use of industrial antifouling products and the use of TBT was banned by the International Maritime Organization since January 2008 (IMO, 2002). Some alternative antifouling paints have been used to substitute organotin-based paints, but they may also alter the aquatic environment (Armstrong *et al.*, 2000; Karlsson and Eklund, 2004; Löschau and Krätke, 2005). Antifouling molecules derived from marine organisms are promising leads for new commercial paints because these metabolites already exist in the sea and are biodegradable (Rittschof, 2001; Bhadury and Wright, 2004).

Several studies showed the antifouling potential of sponge secondary metabolites in the Indian and Pacific oceans, Caribbean and Mediterranean seas (Sera *et al.*, 1999a; Kubanek *et al.*, 2002; Hellio *et al.*, 2005; Limna Mol *et al.*, 2009). The antifouling property of marine benthic invertebrates from the Brazilian coast are still poorly known; to this date, only a single species of gorgonian, *Phyllogorgia dilatata* (Esper) and three of sponges were investigated: *Geodia corticostylifera* Hajdu *et al.*, *Mycale microsigmatosa* Arndt and *Aplysina fulva* (Pallas). The extract of *G. corticostylifera* had strong antifouling activity, preventing the establishment of the bivalve *Perna perna* in laboratorial tests (Clavico *et al.*, 2006), while extracts of *M. microsigmatosa* and *P. dilatata* selectively inhibited the establishment of barnacles and *A. fulva* did not showed antifouling effects in field experiments (Pereira *et al.*, 2002). As there are more

than 300 species of marine sponges in the Brazilian coast, this area of knowledge is clearly little explored in Brazil and there is great potential for the discovery of new antifouling compounds. The aim of this study is perform a screening for antifouling activity of twelve abundant sponge species from NE and SE Brazil, and point out promising species for further and more detailed investigations.

2. Materials and Methods

2.1. Collection, maintenance of organisms and extraction

Twelve species of sponges belonging to five orders were collected through free or SCUBA diving from tide zone to 20 m depth (Rio de Janeiro and Bahia States) from 2006 to 2008 (Table 1).

Juveniles of *Perna perna*, a common mussel in the Brazilian coast, were collected in December 2008 from the rocky coastal area at Itaipu beach (Niterói City, Rio de Janeiro State, Brazil, 23°00'34" S-44°26'10" W). They were kept in a 230 L recirculating aquarium at constant temperature (20 °C), salinity (35) and aeration to be further used in the assays.

After collection, sponge species were refrigerated through ice packages until arrive to laboratory and be frozen. Lyophilized sponges were extracted with acetone, except to *Petromica citrina*, which was extracted with dichloromethane. Sponges were extracted two times, first during 24 hours before be filtrated, and then extracted for 72 hours. The resulting extract from each filtration were added to previous filtered extract, to obtain the final extract.

2.2. Antifouling assays

Juveniles of *Perna perna* were carefully cleaned and separated from each other. The selection was done by considering active exposition of foot and crawling, showing that the mussels exhibit an exploratory behavior of the substrate.

The method described in da Gama *et al.* (2003) was followed to measure the antifouling activity against *Perna perna*. Water-resistant filter papers (control) were cut in circles of 9 cm in diameter and soaked with dichloromethane. A second set of 9 cm filter papers (treatment) were cut in a chessboard pattern (1.5 cm² squares). Treatment filters were soaked in sponge extracts diluted in dichloromethane, receiving the same amount of extract or, in case of the control, soaked only with solvent and allowed to air dry. Control and treatment filters were placed in sterile polystyrene Petri dishes (9 cm diameter x 1 cm height, Figure 1). Natural concentration of crude extracts were calculated and used for each sponge species (see Table 1).

Treatment filter paper was placed in the upper face (squared) and control in the lower face (entire) to offer the same area of treatment and control for byssus fixation. The dishes were completely filled with filtered natural sea water and three mussels ranging from 1.5 to

Table 1 - Species studied and their localities of collection. RJ, Rio de Janeiro and BA, Bahia. *Calculated on the dry weight of sponge.

Species/Author	Family/order	Locality/state	Natural concentration* of crude extract (%)
<i>Amphimedon viridis</i> Duchassaing & Michelotti, 1864	Niphatidae/Haplosclerida	Angra dos Reis, RJ	1.57
<i>Aplysina fulva</i> (Pallas, 1766)	Aplysinidae/Verongida	Arraial do Cabo, RJ	1.62
<i>Chondrosia</i> sp.	Chondrillidae/Hadromerida	Angra dos Reis, RJ	2.13
<i>Desmapsamma anchorata</i> (Carter, 1882)	Myxiliidae/Poecilosclerida	Angra dos Reis, RJ	1.60
<i>Dysidea etheria</i> (de Laubenfels, 1936)	Dysideidae/Dictyoceratida	Angra dos Reis, RJ	2.00
<i>Hymeniacion heliophila</i> (Parker, 1910)	Halichondriidae/Halichondrida	Niterói, RJ	2.05
<i>Mycale angulosa</i> (Duchassaing & Michelotti, 1864)	Mycalidae/Poecilosclerida	Angra dos Reis, RJ	1.95
<i>Petromica citrina</i> Muricy et al., 2001	Halichondriidae/Halichondrida	Rio de Janeiro, RJ	2.06
<i>Polymastia janeirensis</i> (Boury-Esnault, 1973)	Polymastiidae/Hadromerida	Arraial do Cabo, RJ	1.59
<i>Tedania ignis</i> (Duchassaing & Michelotti, 1864)	Tedaniidae/Poecilosclerida	Angra dos Reis, RJ	1.83
<i>Tethya maza</i> Selenka, 1879	Tethyidae/Hadromerida	Paraty, RJ	2.68
<i>Tethya rubra</i> Ribeiro & Muricy, 2004	Tethyidae/Hadromerida	Salvador, BA	2.29

2.5 cm length were added (Figure 1). We also used an independent blank control in which both filter papers were embedded only with dichloromethane. For each assay, ten replicates were performed.

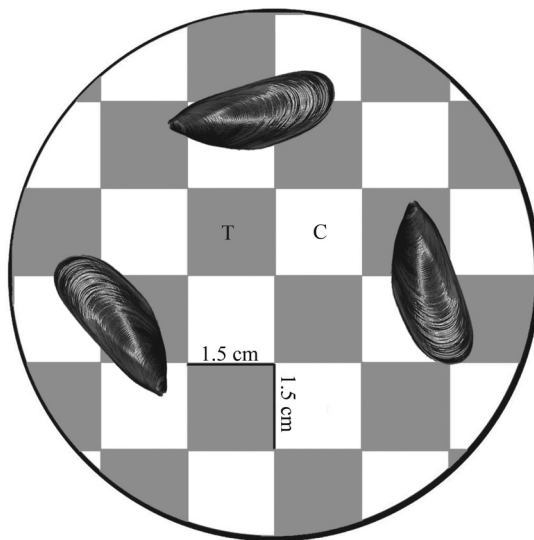


Figure 1 - Schematic draw of experimental unit containing treatment (T, gray squares) and control (C, white squares) filter papers and three juveniles of *Perna perna* used in assays.

After 24 hours of experiment, all the records of attachment were quantified. Mussels were placed in plastic mesh bags tagged according to treatment and suspended in an aquarium, and checked for possible mortality due to exposure to the compounds tested, after 24 hours.

2.3. Statistical analysis

One-way analyses of variance (ANOVA) was used to compare the number of byssal thread attached among treatments and Dunnetts *post hoc* test was employed to detect the differences between treatments and independent controls. *T* test were used to detected differences in

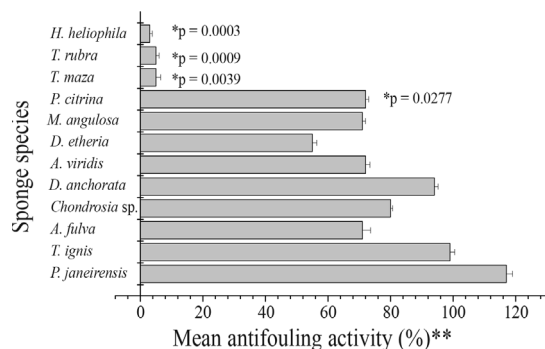


Figure 2 - Mean antifouling activity of sponge crude extracts. **Value in percentage of control with standard error indicated by bar.

attachment between dependent control and treatment only for *Petromica citrina* assays (see Discussion). As assumptions of normality and homogeneity of variances were not met even after data transformations, Kruskal-Wallis non-parametric tests were run (Zar, 1999). Differences were considered significant when $p < 0.05$ ($\alpha = 5\%$).

3. Results and Discussion

From twelve sponge crude extracts, eleven were able to reduce mussel attachment, although only four in a statistically significant manner: *Tethya rubra*, *T. maza*, *H. heliophila* and *P. citrina* ($p = 0.0009$, 0.0039 , 0.00003 and 0.0277 , respectively - Figure 2).

In all cases, the crude extract were nontoxic, since 24 h after test no mollusc was found dead.

Several metabolites from marine sponges were recorded by possessing antifouling properties against macro and/or microorganisms (Willemsen, 1994; Hellio *et al.*, 2009), which reinforce the high potential of sponges to produce bioactive compounds.

Since most crude extracts tested in the present study were obtained with acetone (an organic solvent with polar affinities), we consider the dissolution of metabolites, from treatment filter paper, through the stagnant sea water in the Petri dish. These metabolites may act directly on the mussel inhibiting or stimulating byssus emission. All extracts tested were able to reduce mussel fixation, except for *P. janeirensis*.

Hymeniacidon heliophila was tested previously for antifouling activity, through *in situ* experiments, while *Tethya* spp. and *P. citrina* were tested here for the first time. The extract of *H. heliophila* was highly antifoulant in the present study (in which only 23 byssus were fixed, in total, against 132 in the blank controls), but did not deter fouling in North Carolina, U.S.A., in experiments *in situ* (Henrikson and Pawlik, 1998). The effectiveness of *H. heliophila* crude extract from Rio de Janeiro should represent an important ecological advantage, since this sponge occurs in the same zone of *P. perna* and is usually found living over them. Moreover, antipredation activity against fishes, sea urchins and hermit crabs had been demonstrated in extracts from *H. heliophila* (Ribeiro *et al.*, 2010). On the other hand, field and laboratory experiments differ in several aspects, making hard direct comparisons. We must consider that in laboratory assays still water was used in contrast larvae to constant natural flow, and a single fouling organism in contrast with the large array of larve available in water column (da Gama *et al.*, 2003). Although laboratory tests are very useful and reliable, the ecological significance is questionable and requires field testing to better interpretation of results (da Gama *et al.*, 2003; Bakus *et al.*, 1985). Different results found in the present study and Henrikson and Pawlik (1998) could be due to chemical differences between Rio de Janeiro and North Carolina specimens, or due to limitation concerning the method applied. It is an interesting point to be hereafter investigated.

From the four active species, three have sterols as majority components (pers. obs. in *T. rubra*, *T. maza* and *H. heliophila*). Despite being common and abundant in sponges, sterols were found to be effective inhibiting mussel and barnacle settlement and/or had lethal effect to larvae ascidian (Tsukamoto *et al.*, 1997; Tomono *et al.*, 1998; Sera *et al.*, 1999a, b; Qiu *et al.*, 2008). In addition to the sterols, other compounds as terpenoids, alkaloids, saponins, and fatty-acids, were indicated as active against barnacle larvae or mollusc settlement (Goto *et al.*, 1992; Fusetani, 2004; Roper *et al.*, 2009). It is necessary investigate sterols, as well as minority components of *T. maza*, *T. rubra*, *P. citrina* and *H. heliophila* to identify the substance acting in *P. perna* attachment.

Although, the crude extract of *A. viridis*, *P. janeirensis* and *T. ignis* did not show activity against the settlement of mussels, the high antimicrobial property recorded from these species might be functional by inhibiting the attachment of marine microbiota in the early stages of fouling process (Kelman *et al.*, 2001; Muricy *et al.*, 1993). Experiments in natural conditions are good indicators by permitting several larvae in contact with extracts. Despite various experiments with larvae and macroinvertebrates that reveal the inhibitory capacity of sponges against the settlement, an *in situ* experiment found that just one out of 26 sponge species inhibited larval recruitment, suggesting that competent larvae use chemical characteristics of the sponges as cues for appropriate settlement sites (Bingham and Young, 1991).

We recommend the sponges *T. maza*, *T. rubra*, *H. heliophila* and *P. citrina* for future studies. Otherwise, the model organism used (bacteria, invertebrate larvae, or mussel) may have different levels of sensitivity by the chemical components. Thus, antifouling activity can be also detected using other organisms, and additional tests would be necessary to verify how effective these extracts are concerning antifouling process.

The existence of antifouling activity in these sponges seems to reflect the evolutionary history of each species, once it is not related to a pattern of taxonomic affinity or geographical distribution.

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