ISSN 0104-6632 Printed in Brazil

Brazilian Journal of Chemical Engineering

Vol. 20, No. 04, pp. 363 - 374, October - December 2003

THE EFFECTS OF GEOMETRY AND OPERATIONAL CONDITIONS ON GAS HOLDUP, LIQUID CIRCULATION AND MASS TRANSFER IN AN AIRLIFT REACTOR

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(Received: May 29, 2002; Accepted: May 28, 2003)

Abstract - In airlift reactors transport phenomena are achieved by pneumatic agitation and circulation occurs in a defined cyclic pattern through a loop. In the present work, the effect of geometrical relations on gas holdup and liquid velocity, and consequently on the gas-liquid mass transfer coefficient, was studied in a 6-liter airlift bioreactor with $A_D/A_R = 0.63$; A_D , downcomer cross-sectional area, and A_R , riser cross-sectional area. Measurements of the volumetric oxygen transfer coefficient ($k_L a$) were taken in a water-air system using a modified sulfite oxidation method. Different conditions were examined by varying parameters such as superficial air velocity in the riser (U_{GR}), bottom clearance (d_1) and top clearance (d_2). It was observed from the experimental results that d_1 and d_2 have a remarkable effect on $k_L a$ values. The effect is due to their influence on gas holdup and liquid velocity, consequently affecting $k_L a$. Superficial air velocity in the riser (U_{GR}) ranged from 0.0126 to 0.0440 m.s⁻¹ and $k_L a$ varied between 40 to 250 h⁻¹, whereas gas holdup (ϵ) reached values up to 0.2. The volumetric oxygen transfer coefficient ($k_L a$), gas holdup in the riser (ϵ_R) and downcomer (ϵ_D) and superficial liquid velocity in the riser (U_{LR}) for all the geometrical relations were successfully correlated with dimensionless numbers, namely, the Sherwood number (Sh) and the Froude number (Fr) as well as with geometrical relations such as the bottom space ratio ($E_L a$) and top sp

Keywords: airlift reactor, oxygen transfer, gas holdup, hydrodynamics, $k_L a$.

INTRODUCTION

Non conventional bioreactors are currently the focus of studies in many biochemical engineering laboratories, since the success of an industrial fermentation process depends heavily on the efficiency of the reactor. Particularly oxygen transfer and the corresponding energy input necessary to meet the oxygen demand strongly affects production costs. Airlift reactors are agitated pneumatically and circulation takes place in a defined cyclic pattern

through a loop, which divides the reactor into two zones: a flow-upward and a flow-downward zone. The gas-sparged zone or the riser, has higher gas holdup than the relatively gas-free zone, the downcomer, where the flow is downward (Chisti, 1989).

The possibility of meet the aeration and agitation requirements during the fermentation process with low energy input led to a growing interest in using airlift bioreactors in industrial processes. The simplicity of their design and construction, better defined flow patterns, low power input, low shear fields, good

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mixing and extended aseptic operation, made possible by the absence of stirrer shafts, seals and bearings, are important advantages of airlift reactors in fermentation applications (Chisti and Moo-Young, 1987; Chisti, 1989; Moresi, 1981).

In the concentric-tube airlift bioreactor, some geometrical parameters affect gas holdup, liquid circulation, mixing time and the volumetric oxygen transfer coefficient. Extensive study of reactor hydrodynamics and reactor geometry enhances the importance of the geometrical parameters in the design and scale-up of concentric-tube airlift bioreactors (Gavrilescu and Tudose, 1998a, b and c).

Particularly in recent work, the performance of airlift reactors has been studied in terms of oxygen transfer capability for further utilization in fed-batch cultures (Gallindez-Mayer et al., 2001) and in cultivations with a pellet-type growing microorganism (Freitas and Teixeira, 2001). The latter authors mention that results in the literature for mass transfer in airlift reactors vary widely and are contradictory, since the reactors and experimental procedures are all different. On the other hand, bench scale (1 to 10 liters) reactors are the most practical for development of new bioprocesses, particularly those dealing with valuable metabolites and utilizing somewhat expensive components. This is the case in the production of biologically active substances by filamentous microorganisms or hybridoma cells, which are very sensitive to shear stress, while demanding na appropriate supply of oxygen. So far, little work has focused on the hydrodynamics and mass transfer performance of small-scale airlift reactors in view of establishing operational conditions for batch, fed-batch and even continuous cultivation of cells.

In the present work the effects of geometrical factors upon gas holdup and liquid velocity, and

consequently on the mixing and mass transfer coefficient ($k_L a$), was studied in a concentric-tube airlift bioreactor with six-liter working volume for further utilization in bioprocess development. Mixing time and the volumetric oxygen transfer coefficient ($k_L a$) were measured under different conditions by varying the superficial air velocity in the riser (U_{GR}), bottom clearance (d_1) and top clearance (d_2) in a water-air system.

MATERIALS AND METHODS

Equipment

A scheme of the concentric-tube airlift reactor used in this work with dimensions given in Table 1 is shown in Figure 1. Riser diameter, D_R , is an equivalent diameter, calculated as $D_R = (4/\Pi(A_T - A_D))^{1/2}$ (m²), where A_T is the total superficial area and A_D , the downcomer superficial area (m²). It was made of glass, with the bottom and top plates made of rigid nylon. Air sparger and other pipes were made of stainless steel. The distance from the reactor base to the draft tube (d₁) and the distance from the top of the draft tube to the liquid level (d₂) were changed within the range of 0.025 to 0.045 m and of 0 to 0.040 m, respectively.

For determination of $k_L a$, experiments were carried out with aqueous sulfite solution and air as the gaseous phase. Air was sparged through a 0.114 m ring, with 35 holes with a diameter of 0.0007 m and a superficial air velocity in the riser (U_{GR}) set within the range of 0.0126 to 0.0440 m.s⁻¹. Airflow rates were measured with a mass flowmeter from Cole Parmer (series 33116-42). All experimental runs were carried out at atmospheric pressure and a temperature of $28^{\circ}C$.

Table 1: Geometrical characteristics of the airlift reactor

Dimension – Symbol	Value (m)
Riser Diameter - D _R	0.100
Downcomer Diameter - D _D	0.080
Bottom Clearance - d ₁	0.025; 0.035; 0.045
Top Clearance - d ₂	0.000; 0.020; 0.040
Inner Tube Height - H _D	0.37
Pressure Measurement Distance - d	0.13
External Tube Height - H _R	0.60

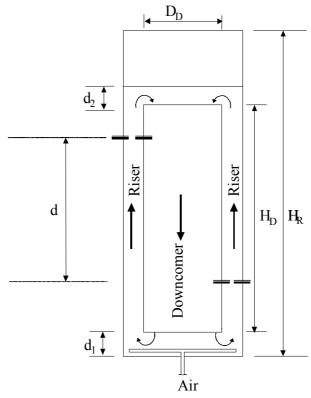


Figure 1: Schematic view of the concentric-tube airlift reactor

Gas Holdup

The total gas holdup (ϵ_T) was determined by the volume expansion method, while for the riser (ϵ_R) and downcomer (ϵ_D) zones, the manometer method was used. Chisti (1989) describes both methods in details. An electronic pressure meter model SC 990 from T&S – Equipamentos Eletrônicos, S. Carlos SP (Brazil) was utilized to measure the pressure drop between the riser and downcomer for each set of experiments. The distance between the two data acquisition points (d) was 0.13 m (Figure 1). The average of three recorded values was used for holdup calculation according to the following equation:

$$\varepsilon_{i} = \frac{\rho_{L}}{\rho_{L} - \rho_{G}} - \frac{\Delta P}{(\rho_{L} - \rho_{G})g \ d}$$
 (1)

where subscript "i" stands for D (downcomer) or R (riser).

Characterization of the Liquid Phase

Acid solution was used as a tracer to evaluate mixing time (t_m) , circulation time (t_C) and linear or interstitial liquid velocity (V_L) by analysis of the pH

forcing function response data. Mixing time (t_m) was defined as the time required to reach 95% of the final pH value after addition of the tracer. Initially, the airlift reactor was filled with 6 liters of tap water and the air was turned on up to the point of maximum flow rate to saturate the water with oxygen. After 5 minutes, aeration was set to the desired value and pH was adjusted with HCl 2N solution to 6.0 (\pm 0.1). Then the tracer, a 10 mL citrate buffer 1M (pH 3.4), was injected into the reactor exactly in the center of the liquid level, at the top of the downcomer zone. Meanwhile, a chart recorder was switched on to monitor the pH in the bioreactor via a New Brunswick (NBS, Edison NJ, USA) pH meter/controller with an Ingold probe. From the time interval between two tracer peaks, circulation time (t_C) was measured.

The reactor has two diametrically opposed 45° upward-inclined side ports located at a point corresponding to one third of the distance from the bottom to the top of the inner tube. The Ingold pH probe was inserted into one of them and the Mettler-Toledo, InPro 6000 model DO (dissolved oxygen) probe, into the other one. The probes were inserted in such a way that only the tip was in contact with the fluid in the riser zone.

The liquid linear velocity in the riser section (V_{LR}) can be estimated from circulation time (t_C) by the following procedure. Average liquid linear velocity (\overline{V}_L) is calculated from circulation time (t_C) given by equation (2):

$$\overline{V}_{L} = \frac{2 H_{D} + d_{1} + d_{2}}{t_{C}}$$
 (2)

Circulation time (t_C) is the sum of the time which elapsed in the riser (t_R) and that which elapsed in the downcomer (t_D) :

$$t_{C} = t_{R} + t_{D} \tag{3}$$

and t_R and t_D are defined as

$$t_{R} = \frac{H_{D} + d_{1}/2 + d_{2}/2}{V_{LR}}$$
 (4)

$$t_{\rm D} = \frac{H_{\rm D} + d_1/2 + d_2/2}{V_{\rm LD}}$$
 (5)

Furthermore, the continuity criterion (U_{LR} $A_R = U_{LD}$ A_D) can be used to show the existence of the following relationship between the liquid linear velocities in the riser (V_{LR}) and the downcomer (V_{LD}) (Chisti, 1989):

$$V_{LR} (1 - \varepsilon_R) A_R = V_{LD} (1 - \varepsilon_D) A_D$$
 (6)

By substituting equations (3), (4) and (5) into equation (2), we obtain

$$\overline{V}_{L} = \frac{2 V_{LR} V_{LD}}{V_{LR} + V_{LD}}$$
 (7)

Thus, by measuring circulation time (t_C) , the liquid linear velocities in the riser (V_{LR}) and in the downcomer (V_{LD}) can be calculated as follows:

$$V_{LR} = \frac{H_D + d_1 + d_2}{t_C} \left(1 + \frac{A_D (1 - \varepsilon_D)}{A_R (1 - \varepsilon_R)} \right)$$
 (8)

$$V_{LD} = \frac{H_D + d_1 + d_2}{t_C} \left(1 + \frac{A_R (1 - \varepsilon_R)}{A_D (1 - \varepsilon_D)} \right)$$
(9)

Measurement of the Volumetric Oxygen Transfer Coefficient

The volumetric oxygen transfer coefficient (k_La) was determined by a modified sulfite method (Vilaça et

al., 2000). Initially, distilled water was added to the reactor. Air was turned on and the airflow rate was adjusted to the desired value. Enough copper sulfate crystal catalyst to give 0.7 g.L⁻¹ was added and dissolved. Enough sodium sulfite crystals to give 8.83 g.L⁻¹ was added to the reactor and the recorder, coupled to a dissolved oxygen analyzer (New Brunswick Scientific Co., Edison NJ, USA), was turned on. The DO analyzer was linked to the DO probe placed in the riser, as described above.

The reaction between sodium sulfite and oxygen in the liquid phase is given by the stoichiometric equation

$$Na_2SO_3 + 1/2 O_2 \xrightarrow{CuSO_4} Na_2SO_4$$
 (10)

The amount of sodium sulfite added was enough to maintain the dissolved oxygen concentration around zero during a long time period, increasing the accuracy of the method. The time (Δt) necessary to consume all the sodium sulfite was determined by the rise in dissolved oxygen level. The time interval varied from around 30 minutes to 3 hours. The dissolved oxygen, DO, rapidly fell to zero (in a few seconds), and this fact was easily detected since the slope of the DO curve is very steep, abruptly leveling off at zero or a near-zero level. Also, the rise in the DO curve is steep and can be easily observed. The error should be around the range of a few seconds to 30 minutes, at the most. Actually this method is utilized by New Brunswick Inc. to evaluate the oxygen transfer capabilities of their fermentors. This method (modified sulfite oxidation) was utilized to have a standard reference for comparison with commercial fermentors. Under these conditions, the reaction rate is much higher than diffusion through the gas-liquid interface and the process is diffusioncontrolled. Therefore the volumetric oxygen transfer rate, N_V (molO₂.L⁻¹.s⁻¹), can be determined from the stoichiometry of Equation (10), leading to Equation (11).

$$N_{V} = \frac{0.5 \text{ n}}{V \Delta t} \tag{11}$$

where n is the number of moles of Na₂SO₃ consumed in the reaction and V is the total liquid volume in the reactor.

The volumetric oxygen transfer coefficient ($k_L a$) can be determined by Equation (12).

$$k_{L}a = \frac{N_{V}}{C^* - C} \tag{12}$$

where k_L is the mass transfer coefficient, a is the

transfer area per unit volume, C^* is the saturatedoxygen concentration and C is the dissolved oxygen concentration; during Δt , C = 0.

For each measure, the reactor content was replaced with distilled water and the above procedure was repeated.

RESULTS AND DISCUSSION

A set of experiments was conducted using three values of d_1 (0.025, 0.035 and 0.045 m), three values of d_2 (0, 0.020 and 0.040 m) and six values of U_{GR} (0.0126-0.0440 m.s⁻¹). The ratio between downcomer and riser areas was $A_D/A_R = 0.63$. Figure 2 shows the results obtained for the gas holdup in the riser (ϵ_R) and in the downcomer (ϵ_D) zones. It was observed that the gas holdup in both zones increases with the decrease in bottom clearance (d_1). This fact was also observed by Gravilescu and Tudose (1998a) while working with three large concentric-tube airlift reactors of 70, 2,500 and 5,200 liters.

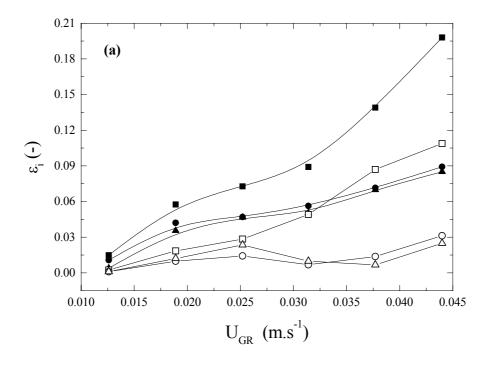
Higher ϵ_i values were observed for $d_2=0$, particularly for larger values of U_{GR} . Regardless of d_1 values, for $d_2 \geq 0.020$ m, ϵ_R and ϵ_D values were very similar. However, when $d_2=0$, ϵ_R and ϵ_D are very large, particularly for higher U_{GR} . Working with d_2 from 0 to 0.28 m (draft-tube height = 1.68 to 8.26 m) Gravilescu and Tudose (1998a) also observed this effect. Probably, above a certain critical value of d_2 the drag of bubbles is hindered, resulting in faster

bubble disengagement and lower gas holdup. Mention should be made that when $d_2=0$, measured with $U_{GR}=0$, the volume of liquid is smaller. Therefore for a given airflow rate, ϵ_R and ϵ_D are higher. The importance of studying the $d_2=0$ condition lies in the fact that this equipment will also be utilized as a single and repeated fed-batch reactor. The first step will be cultivation with minimum volume as a batch. This step can last for several hours to a few days. After this step, feeding with concentrated medium will take place.

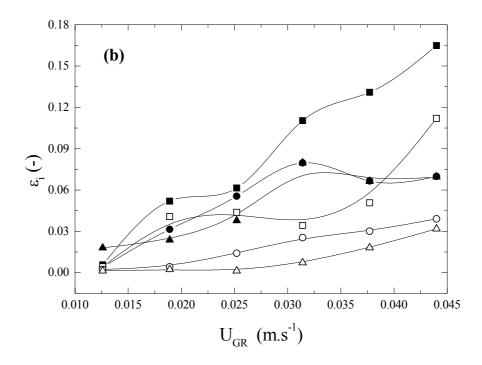
Similarly to the riser (ϵ_R) and downcomer (ϵ_D) gas holdup, the superficial liquid velocity in the riser (U_{LR}) depends on the geometrical characteristics. In the riser, U_{LR} depends on the superficial air velocity in the raiser (U_{GR}) and on the reactor geometrical characteristics, as shown in Figure 3.

Figure 4 shows the effect of superficial air velocity in the raiser (U_{GR}) on mixing time (t_m) . It can be observed that the larger the top clearance (d_2) , the higher is the mixing time (t_m) . Similarly, when d_1 is increased, mixing time decreases for constant d_2 .

The experimental values of the volumetric oxygen transfer coefficient ($k_L a$), obtained for six different values of superficial air velocity in the riser (U_{GR}) and four different pairs of d_1 and d_2 , are given in Figure 5. Results show that $k_L a$ was higher for both $d_1 = 0.045$ m and $d_2 = 0.040$ m. This behavior is directly associated with the smaller values of mixing time (t_m), which led to higher liquid circulation velocity under this geometrical relationship.



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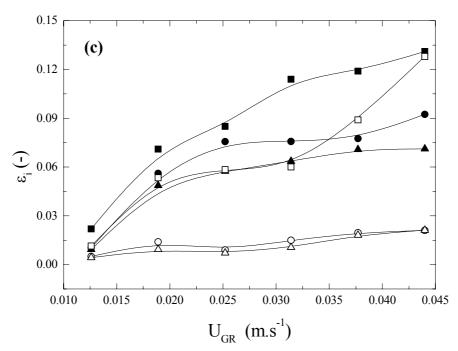


Figure 2: Gas holdup in the riser (ϵ_R) and downcomer (ϵ_D) zones of the concentric-tube airlift reactor as a function of superficial air velocity (U_{GR}) . a) $d_1 = 0.025$ m, b) $d_1 = 0.035$ m and c) $d_1 = 0.045$ m. ϵ_R : (\bullet - $d_2 = 0.000$ m), (ℓ - $d_2 = 0.020$ m), (ℓ - $d_2 = 0.040$ m), ϵ_D : (ℓ - ℓ -

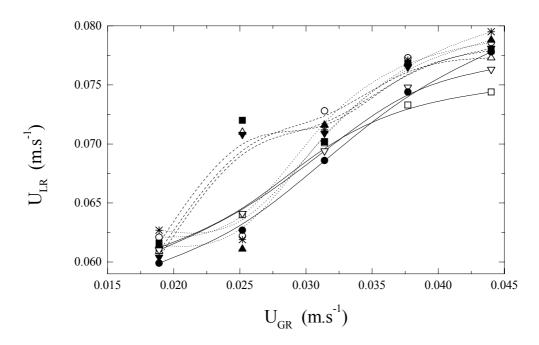


Figure 3: Liquid superficial velocity in the riser zone (U_{LR}) as a function of superficial air velocity (U_{GR}): —□—: $d_1 = 0.025$ m and $d_2 = 0.000$ m, "o": $d_1 = 0.025$ m and $d_2 = 0.020$ m, —△—: $d_1 = 0.025$ m and $d_2 = 0.040$ m, —0.035 m and $d_2 = 0.035$ m and $d_2 = 0.035$ m and $d_2 = 0.035$ m and $d_2 = 0.020$ m, —0.035 m and $d_2 = 0.035$ m and $d_2 = 0.045$ m

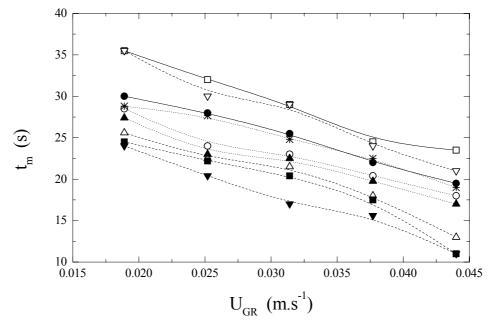


Figure 4: Reactor mixing time (t_m) as a function of superficial air velocity (U_{GR}) . $\neg \neg \neg$: $d_1 = 0.025$ m and $d_2 = 0.000$ m, $\neg \neg \neg$: $d_1 = 0.025$ m and $d_2 = 0.020$ m, $- \neg \neg \neg$: $d_1 = 0.025$ m and $d_2 = 0.040$ m, $\neg \neg \neg \neg$: $d_1 = 0.035$ m and $d_2 = 0.000$ m, $\neg \neg \neg \neg$: $d_1 = 0.035$ m and $d_2 = 0.040$ m, $- \ell \neg$: $d_1 = 0.045$ m and $d_2 = 0.040$ m, $- \neg \neg \neg$: $d_1 = 0.045$ m and $d_2 = 0.040$ m, $- \neg \neg \neg$: $d_1 = 0.045$ m and $d_2 = 0.040$ m

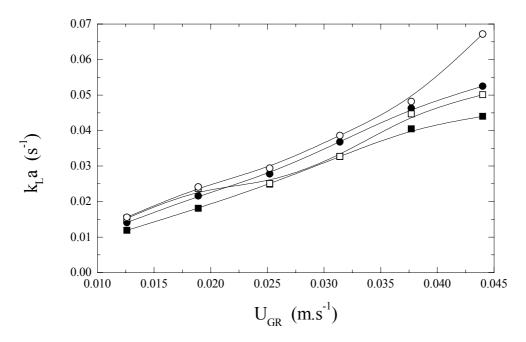


Figure 5: Volumetric oxygen transfer coefficient ($k_L a$) as a function of superficial air velocity (U_{GR}). \bullet : $d_1 = 0.025$ m and $d_2 = 0.00$ m, ℓ : $d_1 = 0.025$ m and $d_2 = 0.04$ m, \circ : $d_1 = 0.045$ m and $d_2 = 0.00$ m, o: $d_1 = 0.045$ m and $d_2 = 0.040$ m

From the results, it can be observed that the gas holdup (ϵ_i) , the superficial liquid velocity in the riser (U_{LR}) and the volumetric oxygen transfer coefficient $(k_L a)$ were clearly affected by the geometrical characteristics of the bioreactor, making possible the application of correlations between these variables and the geometrical factors, as proposed by Gavrilescu and Tudose (1998a, b and c). The correlations obtained from the experimental results for each gas holdup are as follows:

$$\epsilon_T = 1.32 \text{ Fr}^{0.77} \text{ B}^{0.39} \text{ T}^{0.08}, r^2 = 0.95$$
 (13)

$$\epsilon_R = 11.97$$
 Fr^{1.31} B^{0.42} T^{-2.44}, $r^2 = 0.80$ (14)

$$\varepsilon_{\rm D} = 13.52 \quad \text{Fr}^{1.46} \ \text{B}^{0.72} \ \text{T}^{-5.21}, \ \ r^2 = 0.71$$
 (15)

where

$$B = \frac{d_1}{D_R} \tag{16}$$

$$T = \frac{d_2}{D_R} + 1 \tag{17}$$

$$Fr = \frac{U_{GR}}{\sqrt{g D_R}}$$
 (18)

The main variable affecting partial gas holdup is top clearance, d₂ in variable T. Gavrilescu and Tudose (1998a) observed that the Froude number (Fr) was the variable that most affected gas holdup in any bioreactor zone. However, in the present work, this was not observed, probably due to the fact that the range of superficial gas velocities in the riser (U_{GR}) , from 0.0126 to 0.0440 m.s⁻¹, was too narrow. Gavrilescu and Tudose (1998a) worked in the range of U_{GR} between 0.01 and 0.11 m.s⁻¹. Figure 6 contains the experimental values of gas holdup and values calculated from equations (13), (14) and (15). The dispersion of data for ε_R and ε_D is due to oscillations in the digital electronic pressure meter readings used for calculation of holdup. The full range of the "electronic pressure meter" (pressure transducer) is 200 cm H₂O and accuracy is 0.25% of the full range; therefore ϵ_R and ϵ_D error is around 23%, taking into account the oscillations of the digital dial during measurement. This fact is reflected in the low correlation coefficients found, as shown in Figures 6b and 6c.

Also, the following relationship between the experimental values of superficial liquid velocity in the riser (U_{LR}) and the geometrical factors was obtained:

$$U_{LR} = 0.178 \text{ Fr}^{0.297} \text{ B}^{-0.004} \text{ T}^{0.095}, r^2 = 0.91 (19)$$

Figure 7 contains the values predicted by equation (19), compared with the experimental values for the riser superficial liquid velocity (U_{LR}).

Regarding k_La , it can be observed in Figure 5 that this variable was also affected by geometrical parameters d_1 and d_2 . In addition, it was possible to apply the relationship proposed by Gavrilescu and Tudose (1998c), taking into account parameters B, T and Fr as follows:

Sh =
$$7.16 \cdot 10^6 \text{ Fr}^{1.121} \text{ B}^{0.201} \text{ T}^{0.410}, \quad r^2 = 0.98 \quad (20)$$

where

$$Sh = \frac{k_L a D_R^2}{D_L}$$
 (21)

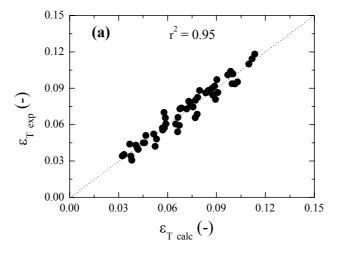
and

$$D_L = 2.43 \cdot 10^{-9} \,\mathrm{m}^2.\mathrm{s}^{-1}$$
.

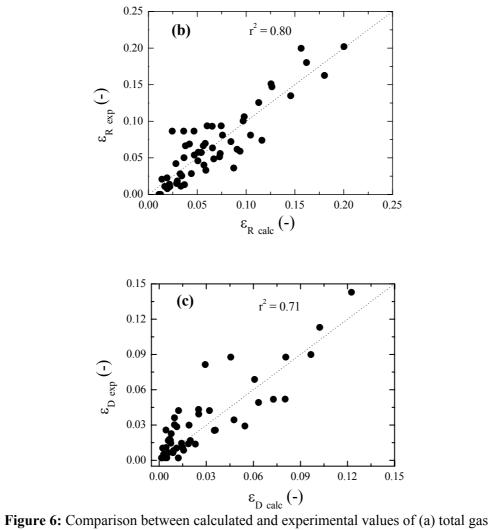
As shown by Equation (19), U_{LR} is affected by T (d_2/D_R+1), while the effect of B (d_1/D_R) is almost negligible. It seems that the circulation rate is much more affected by the increase in liquid height than the decrease in pressure drop, caused by the increase in B. Therefore k_L a should be affected more by T than by B, as shown by equation 20. Gravilescu and Tudose (1998c) point out that higher circulation rates lead to more small-size bubble entrainment. As a result k_L a increases more rapidly with T than the holdup due to the larger interfacial area.

There are some difficulties in comparing correlations obtained in this work with those presented in the literature, mainly due to the differences in geometrical characteristics between the bioreactors studied. In the present work the range of U_{GR} was quite narrow as compared with that found in the literature. For instance, Gravilescu and Tudose (1998a, b and c) vary U_{GR} from 0.02 to 0.10 m.s⁻¹, while in the present work the range varies between 0.016 and 0.040 m.s⁻¹, since higher U_{GR} could not be attained in the small-scale equipment utilized. However, use of correlations (13), (14), (15), (19) and (20) allows prediction of variable behavior in the concentric-tube airlift bioreactor for the gas-liquid system studied in the present work. This enables adoption of operational conditions that favor the development of processes involving industrially important aerobic microorganisms. Figure 8 shows a comparison of the experimental k_I a data with those predicted by equation (20). As shown by the correlation coefficient, there is an excellent agreement making it possible to define the proper geometry and operational conditions for a specific bioprocess.

The important conclusions drawn from this work are that the volumetric mass transfer coefficient is affected by even small changes in the geometrical relations, particularly by the top clearance, d₂. Also, the classical correlation with dimensionless numbers. namely the modified Sherwood number and the Froude number, together with the geometrical relations, enables prediction of the oxygen mass transfer in the reactor. The results showed that, even when working with the minimum liquid volume (d₂ = 0), the oxygen mass transfer is large enough to support a high growth rate of microorganisms with a high oxygen demand, which allows operation of the reactor as a fed-batch. Also, the experimental procedure was shown to be convenient for evaluation of mixing and mass transfer studies in bench-scale airlift reactors.



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holdup (ϵ_T) , (b) riser gas holdup (ϵ_R) and (c) downcomer gas holdup (ϵ_D)

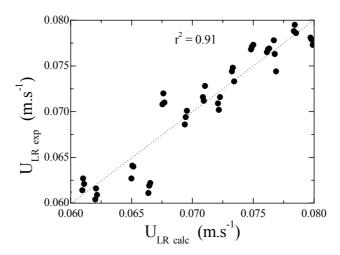


Figure 7: Comparison between calculated and experimental values of the liquid superficial velocity in the raiser (U_{LR})

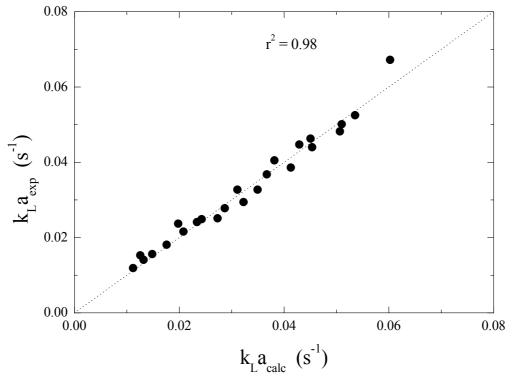


Figure 8: Comparison between calculated and experimental values of volumetric oxygen transfer coefficient (k_La)

ACKNOWLEDGEMENTS

The authors acknowledge the financial support received from CNPq (Conselho Nacional de Desenvolvimento Científico e Tecnológico, Brasilia DF, Brazil) and FAPESP (Fundação de Amparo à Pesquisa do Estado de São Paulo, Brazil).

NOMENCLATURE

A_{D}	Downcomer cross-sectional area (m ²)
A_R	Riser cross-sectional area (m ²)
В	Bottom spatial ratio (-)
C	Dissolved concentration of oxygen in the air-water system (mol.m ⁻³)
C*	Dissolved concentration of saturated oxygen in the air-water system (mol.m ⁻³)
d_1	Bottom clearance (m)
d_2	Top clearance (m)
$\mathrm{D_{L}}$	Diffusivity of oxygen in water (m ² .s ⁻¹)
D_D	Downcomer diameter (m)
D_R	Riser equivalent diameter $D_R = (4/\Pi (A_T - A_D))^{1/2} (m)$
Fr	Froude number (-)
g	Gravitational acceleration (m.s ⁻²)

$k_L a$	Volumetric oxygen transfer coefficient (s ⁻¹)
n	Number of moles of sodium sulfite
11	consumed (mole)
N_V	Volumetric oxygen transfer rate per unit
110	volume (mol.m ⁻³ .s ⁻¹)
R_{H}	Hydraulic radius (m)
Sh	Sherwood number (-)
T	* /
	Top spatial ratio (-)
$t_{\rm C}$	Circulation time (s)
$t_{ m D}$	Time in downcomer region (s)
t_R	Time in riser region (s)
$t_{\rm m}$	Mixing time (s)
U_{GR}	Air superficial velocity in the riser (m.s ⁻¹)
U_{LR}	Liquid superficial velocity in the
	downcomer (m.s ⁻¹)
V	Liquid volume (m ³)
$\overline{ extbf{V}}_{ ext{L}}$	Average liquid linear velocity (m.s ⁻¹)
_	
$ m V_{LR}$	Liquid linear velocity in the riser (m.s ⁻¹)
$V_{ m LD}$	Liquid linear velocity in the downcomer
LD	$(m.s^{-1})$
	(111.5)

Greek Letters

 $\epsilon_{\rm i}$ general gas holdup (-) gas holdup in the downcomer (-) ϵ_{D}

 ϵ_R gas holdup in the riser (-) ϵ_T total gas holdup (-)

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