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PPM1B and P-IKK β expression levels correlated inversely with rat gastrocnemius atrophy after denervation

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Abstract

Activated inhibitor of nuclear factor- κ B kinase β (IKK β) is necessary and sufficient for denervated skeletal muscle atrophy. Although several studies have shown that Mg²⁺/Mn²⁺-dependent protein phosphatase 1B (PPM1B) inactivated IKK β , few studies have investigated the role of PPM1B in denervated skeletal muscle. In this study, we aim to explore the expression and significance of PPM1B and phosphorylated IKK β (P-IKK β) during atrophy of the denervated gastrocnemius. Thirty young adult female Wistar rats were subjected to right sciatic nerve transection and were sacrificed at 0 (control), 2, 7, 14, and 28 days after denervation surgery. The gastrocnemius was removed from both the denervated and the contralateral limb. The muscle wet weight ratio was calculated as the ratio of the wet weight of the denervated gastrocnemius to that of the contralateral gastrocnemius. RT-PCR and Western blot analysis showed that mRNA and protein levels of PPM1B were significantly lower than those of the control group at different times after the initiation of denervation, while P-IKK β showed the opposite trends. PPM1B protein expression persistently decreased while P-IKK β expression persistently increased for 28 days after denervation. PPM1B expression correlated negatively with P-IKK β expression by the Spearman test, whereas decreasing PPM1B expression correlated positively with the muscle wet weight ratio. The expression levels of PPM1B and P-IKK β were closely associated with atrophy in skeletal denervated muscle. These results suggest that PPM1B and P-IKK β could be markers in skeletal muscle atrophy.

Key words: Muscular atrophy; Denervation; PPM1B; P-IKK β ; IKK β ; RT-PCR; Western blot

Introduction

Muscle atrophy can be caused by many genetic and environmental factors, including denervation (1,2). Currently, no effective treatment exists that can reverse the progression of muscle atrophy once initiated. Many strategies that efficiently promote muscle growth cannot prevent the muscle atrophy process (3), suggesting that diverse mechanisms maintain the muscle mass in different contexts. In cases of acute muscle atrophy, active elimination of the myonuclei is considered to be important when load is lacking; in cases of chronic muscle atrophy, the impairment of muscle repair is important (3). Targeting the proper molecules during muscle atrophy is critical to the reversal or cure the progression of atrophy. An understanding of the molecular mechanisms underlying the initiation and progression of muscle atrophy may have therapeutic implications for clinical treatment or drug targets.

Many signaling pathways and molecules, including the IGF-1/PI3K/Akt pathway, muscle ring finger 1, and muscle

atrophy F-box, both of which are ubiquitin ligases (4), mediate muscle atrophy caused by denervation or disuse. Previous studies have suggested that the inhibitor of the nuclear factor- κ B kinase β (IKK β /NF- κ B) pathway may be important in the development of muscle atrophy in denervated muscles (5-7), and that the phosphorylated inhibitor of nuclear factor- κ B kinase β (P-IKK β) might be the critical arbiter in this process (5,8-10). IKK β phosphorylation and activation is an important step in the activation of NF- κ B by tumor necrosis factor- α (TNF- α) (11). Upon stimulation by TNF- α , interleukin-1 (IL-1), and other cytokines, IKK β undergoes rapid phosphorylation and activation, whereas the I κ B kinase (IKK) complex undergoes ubiquitination for proteasome degradation. This process exposes the nuclear localization signal area of NF- κ B and leads to its nuclear transport, which in turn regulates the expression of downstream genes and results in muscle atrophy (9).

Mg²⁺/Mn²⁺-dependent protein phosphatase 1B (PPM1B

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or PP2C) is a member of the Ser/Thr protein phosphatase (PP2C) family (12-15). There are 6 different splice variants of PPM1B, which show a broad specificity for different functions. PPM1B is highly expressed in heart and skeletal muscle and often negatively regulates cell reaction pathways (16-18). For instance, PPM1B can dephosphorylate cyclin-dependent kinase, thus regulating stress response, cell cycle, and actin cytoskeleton changes. PPM1B can suppress the kinase cascade effects of stress-induced p38 and c-Jun N-terminal kinases. PPM1B can also dephosphorylate and inactivate transforming growth factor- β (TGF- β)-activated kinase 1 and mitogen-activated protein kinase kinase (MKKK) induced by stress, IL-1 stimulation, and/or TGF- β treatment to down-regulate the activity of the NF- κ B signaling pathway (12,19-24). In HeLa cells, PPM1B was shown to dephosphorylate serine 177 and 181 in P-IKK β , thereby reducing the activity of IKK β (12). Moreover, PPM1B overexpression can block the activation of the IKK β /NF- κ B pathway induced by TNF- α treatment (12). However, the roles of PPM1B in denervated skeletal muscle cells have not been fully investigated.

In this study, using a rat model of denervation-induced skeletal muscle (gastrocnemius) atrophy, we examined variations in the expression levels of PPM1B and P-IKK β . We found that their expression levels correlated inversely with muscle atrophy after denervation. The results suggest that PPM1B and P-IKK β could be markers in skeletal muscle atrophy.

Material and Methods

Animals and surgery

Thirty young adult female Wistar rats (250-300 g) were provided by the Experimental Animal Center of Shanxi Medical University. The animals were randomly and evenly assigned to 5 groups of 6 rats each, including a control group (0 day) and 2-, 7-, 14-, and 28-day groups.

For surgery, the rats were anesthetized with an intraperitoneal injection of 10% chloral hydrate (2 mL/kg). After the animals were sedated, a posterior incision was made in the right lower limb for sciatic nerve transection (removal of 1.0 cm). The proximal end of the nerve was turned over and sutured to the surrounding tissues to prevent reinnervation of the gastrocnemius muscle. The control group was anesthetized for sham surgery, in which the nerve was exposed but not transected. The animals were sacrificed at 0 (control), 2, 7, 14, or 28 days after denervation surgery. The bilateral gastrocnemius muscles were harvested for analysis.

This study was approved by the Ethics Committee for the Scientific Use of Animals of Shanxi Medical University.

Reagents

In addition to the materials mentioned in the text, the following reagents and kits were used: real-time PCR (RT-

PCR) kit (MBI, China), DNA and protein markers (Sigma, USA), protein extraction reagent (Bio Co., Ltd., China), and β -actin (C4) antibody (Santa Cruz, USA).

RT-PCR

The RT-PCR primers were designed with the Primer 3.0 software package and were synthesized by Shanghai Biological Engineering Co., Ltd. (China). The following primers were used: rat PPM1B, upstream = 5'-gggtcgtt agcagtgctctcgt-3', downstream = 5'-gctcctcattgctcatcac atc-3'; GAPDH, upstream = 5'-tgaacgggaagctcactgg-3', downstream = 5'-tccaccacctgtgtcgtga-3'. Total mRNA was extracted with the Trizol reagent (Shanghai Sangon Technology Service, China) for reverse transcription synthesis of cDNA. The resulting cDNA was amplified by PCR with real-time monitoring of the fluorescence accumulation. The Ct value was set as the number of cycles needed for the required threshold of fluorescence. The $2^{-\Delta\Delta C_t}$ method was used for data analysis (25).

Western blot

Rat denervated gastrocnemius muscles were dissected, frozen in liquid nitrogen, pulverized, and homogenized in 20 mM Tris-HCl, 4% (w/v) SDS, 1 mM phenylmethylsulfonyl fluoride, and 1 μ m each of leupeptin and pepstatin A. Protein concentrations were determined using an SDS-compatible protein assay. Protein samples were mixed with loading buffer, subjected to SDS-PAGE (10%), and transferred electrophoretically to Immobilon-NC membranes. Gels with identical samples were stained with Coomassie brilliant blue and used as an additional control of equilibration of protein loading. Samples were transferred to a nitrocellulose membrane, which was blocked in Blotto buffer containing 10% skim milk in Tris-buffered saline with Tween 20 (TBST) and incubated overnight at 4°C with a rabbit anti-mouse IKK β or P-IKK α / β antibody (1:1000; Cell Signaling, USA) or goat anti-mouse PPM1B polyclonal antibody (1:1000; Santa Cruz). After washing the membrane 4 times with TBST for 15 min each, the membrane was incubated with horseradish peroxidase-labeled goat anti-rabbit IgG secondary antibody (1:1000; Beijing Zhongshan Golden Bridge Biotechnology Co., Ltd., China) at room temperature for 2 h, followed by another set of four 15-min washes with TBST. The membrane was processed for enhanced chemiluminescence using appropriate reagents (Santa Cruz) and was imaged by gray-scale scanning using a GS-800 Densitometer (Bio-Rad, USA).

Wet weight of the gastrocnemius muscle

The bilateral gastrocnemius muscles were harvested entirely from the starting point of the femoral condyle to the end point of the calcaneal tubercle and were measured immediately by an accurate electronic balance (1/10,000, Mettler Toledo AL104, Switzerland). The ratio between the wet weights of the denervated

and contralateral muscles was calculated.

Statistical analysis

SPSS 13.0 was used for statistical analyses, and the results are reported as the means \pm SD. The ImageJ software was used for Western blot image analysis. Analysis of variance (ANOVA) was used for comparison of many groups. The Dunnett method was used to compare two groups, and the Spearman method was used for correlation analysis. Differences with a P value <0.05 were considered to be statistically significant.

Results

Denervated rat gastrocnemius muscle shows temporal changes of PPM1B mRNA expression

Quantitative analysis revealed a significant and consistent decrease in the PPM1B mRNA turnover measured by RT-PCR during the denervation phase of skeletal muscle, which started as early as 2 days after denervation surgery and lasted at least 28 days. The PPM1B mRNA expression decreased to 41% of that observed in the control group at 2 days, and to 7% of control at 28 days. The mRNA expression levels were significantly different from those of the control group at all times tested (Table 1).

The PPM1B, P-IKK β , and IKK β protein levels show time-dependent changes in the rat denervated gastrocnemius

The Western blot results for the PPM1B protein revealed trends that were similar to but smaller than the mRNA results. The PPM1B protein level began to decrease at 2 days after denervation ($P > 0.05$). Significant changes compared to the control group were evident from 7 days through 28 days after denervation. The PPM1B protein level in the denervated groups decreased from 56% of that in the control group at 7 days to 24% of that in the control group at 28 days (Figure 1, Table 1).

In contrast, the P-IKK β protein

levels began to increase soon after denervation, reaching levels 137% of those in the control group at 2 days. The increase lasted for at least 28 days, when the P-IKK β level of the denervated group was 174% of that in the control group. The P-IKK β protein levels were significantly different from control at all times studied (Figure 1, Table 1). The IKK β protein expression consistently decreased with time after denervation ($P < 0.05$ at all times compared to control); however, the magnitude of the changes was smaller than that observed for PPM1B or P-IKK β (Figure 1, Table 1).

The wet weight ratio of the gastrocnemius muscle varies with time after denervation

The muscle wet weight ratio was calculated as the ratio of the wet weight of the injured side to that of the contralateral muscles. After denervation, the muscle wet weight

Table 1. Effects of muscle denervation on PPM1B mRNA turnover and protein levels of PPM1B, P-IKK β , and IKK β and the muscle wet weight ratio.

	Days after denervation				
	0	2	7	14	28
PPM1B mRNA	1.00	0.41 \pm 0.08*	0.32 \pm 0.03*	0.14 \pm 0.04*	0.07 \pm 0.02*
PPM1B	1.57 \pm 0.07	1.55 \pm 0.05	0.88 \pm 0.04*	0.59 \pm 0.06*	0.38 \pm 0.04*
P-IKK β	0.73 \pm 0.04	1.00 \pm 0.05*	1.14 \pm 0.07*	1.26 \pm 0.04*	1.27 \pm 0.06*
IKK β	1.58 \pm 0.08	1.43 \pm 0.05*	1.33 \pm 0.05*	1.31 \pm 0.07*	1.18 \pm 0.06*
Wet weight ratio	1.01 \pm 0.02	0.99 \pm 0.03	0.85 \pm 0.04*	0.52 \pm 0.08*	0.33 \pm 0.05*

PPM1B = Mg²⁺/Mn²⁺-dependent protein phosphatase 1B; IKK β = inhibitor of nuclear factor- κ B kinase β ; P-IKK β = phosphorylated IKK β . *P < 0.05 compared to the control group at 0 days (Dunnett test).

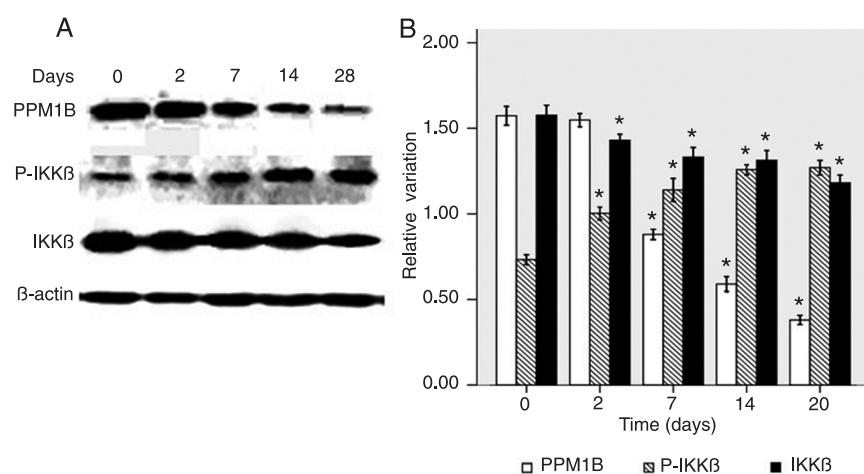


Figure 1. A, Western blot showing the effect of denervation of rat gastrocnemius muscle on protein levels of PPM1B, P-IKK β , and IKK β . B, Gray-scale analysis of staining of PPM1B, P-IKK β , and IKK β proteins in Western blot. Data are reported as means \pm SD. PPM1B = Mg²⁺/Mn²⁺-dependent protein phosphatase 1B; IKK β = inhibitor of nuclear factor- κ B kinase β ; P-IKK β = phosphorylated IKK β . *P < 0.05 compared to the control group at 0 days (Dunnett test).

ratio decreased with time. The differences compared to control were significant starting at 7 days after denervation. At 7, 14, and 28 days, the muscle wet weight ratios of the denervated groups were 85, 51, and 33% of those in the control group, respectively (Table 1).

Changes in the PPM1B and P-IKK β expression levels correlated with the muscle wet weight ratio

The muscle wet weight ratio is a reliable indicator of muscle atrophy, with lower ratio values indicating more severe atrophy. We performed a correlation analysis by the Spearman method between changes in the muscle wet weight ratio and changes in the mRNA turnover rates of PPM1B and P-IKK β . Changes in the PPM1B mRNA and protein expression levels correlated positively with changes in the muscle wet weight ratio ($r = 0.950$, $P < 0.01$ for mRNA; $r = 0.897$, $P < 0.01$ for protein). Changes in the level of the P-IKK β protein correlated negatively ($r = -0.884$, $P < 0.01$) with changes in the muscle wet weight ratio after muscle denervation. Changes in the protein expression levels of PPM1B and P-IKK β correlated negatively ($r = -0.878$, $P < 0.01$; Figure 2).

Discussion

We used a rat model of atrophy of the denervated gastrocnemius muscle to investigate underlying molecular changes in the IKK β /NF- κ B pathway. The mRNA- and/or protein-level changes of PPM1B, IKK β , and P-IKK β were analyzed by RT-PCR and/or Western blot at different times after denervation.

The protein expression levels of IKK β showed a sustained decrease from 2 to 28 days after denervation. The P-IKK β protein levels showed a sustained increase over the same period of time and correlated negatively with changes in the muscle wet weight ratio. The contrasting changes of IKK and P-IKK β after muscle denervation confirmed that the IKK β phosphorylation step in the IKK β /NF- κ B pathway is important in determining the progression of muscle atrophy. Moreover, the ratio between IKK β and P-IKK β could be considered a criterion for evaluating the progress of muscle atrophy.

Similarly, the PPM1B mRNA and protein expression levels in the gastrocnemius muscle displayed a sustained decrease that began 2 days after denervation surgery and lasted the entire 28-day observation period. The changes in PPM1B expression were positively and linearly correlated with the changes in the muscle wet weight ratio. Taken together, these results suggest that the altered expression levels of P-IKK β and PPM1B are closely associated with the development of skeletal muscle atrophy after denervation.

Prajapati et al. (26) have reported that PPM1B negatively

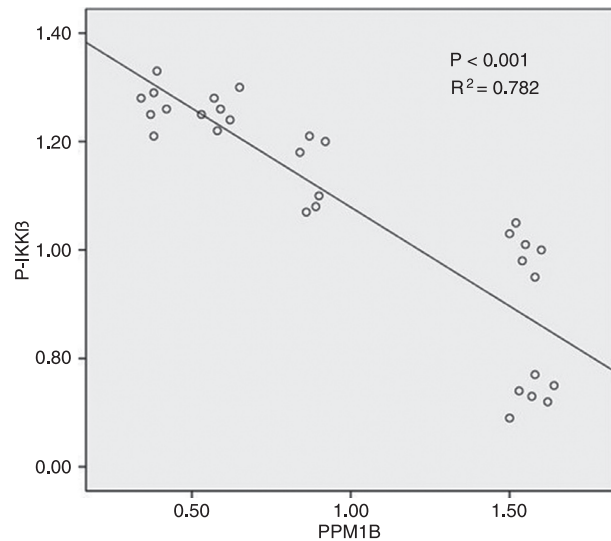


Figure 2. Correlation analysis of protein expression levels of Mg²⁺/Mn²⁺-dependent protein phosphatase 1B (PPM1B) and phosphorylated inhibitor of nuclear factor- κ B kinase (P-IKK β).

regulates IKK β kinase activity. Sun et al. (12) reported that TNF- α induces IKK β phosphorylation and activation. PPM1B bound to P-IKK β , subsequently dephosphorylating P-IKK β at Ser177 and Ser181. This process reduced the activity of IKK β and terminated IKK β -mediated NF- κ B activation. Our current results demonstrate that PPM1B expression correlated negatively with P-IKK β protein expression. As skeletal muscle atrophy increased, PPM1B expression gradually decreased, while P-IKK β expression gradually increased. However, we cannot conclude that the correlations indicate a cause/effect relationship, which mediates the pathogenesis of denervated atrophy. Further identification of PPM1B function will help elucidate the molecular mechanism of muscle atrophy after denervation.

The present study demonstrates some primary changes in PPM1B and the IKK β /NF- κ B pathway. After denervation, expression of PPM1B in skeletal muscle was down-regulated, expression of P-IKK β was up-regulated, and their expression levels correlated negatively with muscle atrophy. Thus, PPM1B might be involved in denervation-induced muscle atrophy. These results provide potential targets for molecular and genetic therapies or drug targets for pharmacological approaches to treat muscle atrophy.

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