

# Antioxidants in yacon products and effect of long term storage

## *Antioxidantes em produtos de yacon e efeito de armazenamento a longo prazo*

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### Abstract

Yacon (*Smallanthus sonchifolius* (Poepp. and Endl.) H. Robinson) is a storage root originally grown in the Andean highlands. The fresh roots are perishable and quickly turn brown during handling and processing. Aiming to prolong shelf-life and to preserve the antioxidant compounds in yacon roots, 3 mm thick yacon slices were dried in a drying cabinet at 40, 50, and 60 °C to a moisture content of 10-14%, and yacon strips were sun dried to a moisture content of 15-20%. The total phenolic content was measured by the Folin-Ciocalteu method, and the quenching capacity was evaluated by measuring the amount of DPPH (1,1-diphenyl-2-picrylhydrazyl) inhibited in samples after drying and after 7 months of storage. The results showed that it is possible to preserve the antioxidant capacity in yacon after cabinet or sun drying. Both yacon chips and strips presented total phenolic content values similar to those of fresh yacon roots. Both products also showed a high inhibition capacity of DPPH (1,1-diphenyl-2-picrylhydrazyl). A significant decrease in the phenolic content was observed in the yacon chips after storage, which indicates that the sun dried strips are more suitable for storage.

**Keywords:** yacon; phenolic content; drying; storage; antioxidant capacity.

### Resumo

Yacon (*Smallanthus sonchifolius* (Poepp. e Endl.) H. Robinson) é uma cultura de raiz de reserva, originária dos planaltos andinos. As raízes frescas são perecíveis e escurecem facilmente durante o manuseio e o processamento. Com o objetivo de aumentar o tempo de prateleira e a preservação de antioxidantes nas raízes frescas, rodela fina (chips) de raízes de 3 mm de tamanho foram secas numa estufa a 40, 50 e 60 °C e pedaços de raízes também foram secas ao sol até apresentarem umidade de 10-14% e 15-20%, respectivamente. O conteúdo de fenóis totais foi determinado pelo método de Folin-Ciocalteu, e a estabilidade oxidativa foi avaliada através da determinação da quantidade de DPPH (1,1-diphenyl-2-picrylhydrazyl) inibida nas amostras após a secagem e durante os 7 meses de armazenamento. Os resultados mostram que a secagem das raízes na estufa e ao sol permitiu manter a capacidade antioxidante das raízes de yacon. Ambos os produtos de secagem apresentaram teor de compostos fenólicos similares ao das raízes frescas e elevada inibição de 1,1-diphenyl-2-picrylhydrazyl. Foi observada uma redução significativa do teor de fenóis em chips após o armazenamento o que indica que os pedaços secos ao sol são mais adequados para o armazenamento.

**Palavras-chave:** yacon; compostos fenólicos; secagem; armazenamento; capacidade antioxidante.

## 1 Introduction

Yacon (*Smallanthus sonchifolius* (Poepp. and Endl.) H. Robinson) is a storage root originally grown in the Andean highlands covering regions from Ecuador, Peru, Bolivia, and northwestern Argentina (GRAU; REA, 1997). Fresh yacon roots are highly perishable and tend to quickly brown during post-harvest handling and processing. One option to prolong shelf-life of yacon roots is to process them into juice, syrup, or dehydrated products such as chips and strips (SEMINARIO; VALDERRAMA; MANRIQUE, 2003). During dehydration, enzymatic activity can be inhibited by heating and addition of inhibitors. However, dehydration can cause changes in the nutritional and organoleptic properties. The drying processes have to be well defined and optimized in order to retain as much as possible the nutritional properties.

The main cause for browning in vegetables is the presence of polyphenol oxidases, which have been extensively studied

and also found in yacon roots (MAYER, 1986; QUEIROZ et al., 2008; YORUK; MARSHALL, 2003; YOSHIDA et al., 2002). Neves and Da Silva (2007) studied the properties of polyphenol oxidases from yacon roots as well as its inactivation and response to inhibitors. The authors concluded that they are relatively stable at 60-70 °C and are progressively inactivated as the temperature reaches 80 and 90 °C.

The phenolic content in yacon roots has been also investigated. Yan et al. (1999) found that yacon roots contain two major antioxidant compounds, identified as chlorogenic acid and tryptophan (48.5 and 14.6 µg.g<sup>-1</sup>, respectively; values on a fresh weight basis). Five caffeic acid derivatives isolated from yacon roots have been identified: chlorogenic acid, 3,5-dicaffeoylquinic acid; 2,4- or 3,5-dicaffeoylaltronic acid; 2,5-dicaffeoylaltronic acid; and 2,3,5- or 2,4,5-tricaffeoylaltronic acid (SIMONOVSKA et al., 2003; TAKENAKA et al., 2003). It

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is equally important and necessary to determine the amount of phenols that remain after processing. The aim of the present study was to evaluate the effects of drying and storage on the retention of antioxidant capacity in yacon chips and strips.

## 2 Materials and methods

Fresh yacon roots were purchased from a local market. The samples came from Locotal, in the province of Chapare (Cochabamba, Bolivia). Roots were stored at room temperature until processing. Anhydrous sodium carbonate and gallic acid were obtained from Riedel-de-Haën (Seelze, Germany). Folin-Ciocalteu reagent and 1,1-diphenyl-2-picrylhydrazyl (DPPH) were obtained from Sigma-Aldrich (St. Louis, MO). The Total Antioxidant Status kit (TAS) was obtained from Randox Laboratories (Crumlin, UK). Ethanol was obtained from Industrias La Bélgica (Santa Cruz, Bolivia).

Yacon roots were washed in running tap water, peeled manually while being immersed in distilled water, and cut into 3 mm thick slices using an industrial cutter (Metalúrgica SKYMPSEN, Brazil). The chips were immersed in a lemon juice and distilled water solution, pH 3, for 10 minutes. Finally, the chips were dried at 40, 50, and 60 °C in a drying cabinet (Binder - FD 53, Tuttlingen, Germany) to a moisture content of 10-14%. Antioxidant capacity was measured in the fresh dried chips. The chips were then stored at room temperature in polyethylene bags for 7 months for further antioxidant capacity analysis. The same procedure described above was followed for the dried strips, with the exception that the roots were cut into 3 cm thick longitudinal pieces, placed on trays, and dried in a solar dryer for 5 days or until final moisture content about 15-20%.

Approximately 300 g of dried chips or strips were ground in a knife mill (Retsh, Haan, Germany). A total of 500 mg of ground sample was transferred into 125 mL flasks, and 20 mL of extraction solvent (1:1 v/v ethanol:water) was added. The extraction was carried out for 2 hours at 30 °C in a shaker incubator (Innova 4080, Edison, USA). Finally, the extracts were filtered and diluted with the extraction solvent to 25 mL.

Total phenolic content was determined using the Folin-Ciocalteu method (SINGLETON; ROSSI, 1965), with the following modification: 3.16 mL of water and 200 µL of Folin-Ciocalteu reagent were added to 40 µL of extract. After one minute, 600 µL of a 1.9 mol.L<sup>-1</sup> Na<sub>2</sub>CO<sub>3</sub> (saturated solution) were added and mixed. Absorbance was measured at 765 nm after 2 hours using a spectrophotometer (Unicam, Leeds, UK). Results were expressed as mg of gallic acid equivalents GAE.g<sup>-1</sup> of sample.

The DPPH assay is based on the determination of scavenging free 1,1-diphenyl-2-picrylhydrazyl (DPPH) radicals (MOLYNEUX, 2004; PRIOR; WU; SCHAICH, 2005). A DPPH solution was prepared with 2.5 mg of DPPH in 50 mL of ethanol. A blank was prepared mixing 1 mL of extraction solvent with 1 mL of DPPH solution. For sample analysis, 1 mL of the sample extract was mixed with 1 mL of DPPH. The reactions were allowed to proceed for 30 minutes at 20 °C in the dark. The absorbance of the blank (A<sub>0</sub>) and the sample extract (A<sub>s</sub>) were measured at 523 nm.

The results were expressed in terms of percentage of DPPH reduction. Q, refers to inhibition or quenching and is defined by the following Equation 1:

$$Q = \frac{A_0 - A_s}{A_0} \times 100 \quad (1)$$

Total antioxidant status (TAS) was measured with a kit from Randox Laboratories (Crumlin, UK) following the instructions of manufacturer. The method is based on the 2,2'-azino-bis-(3-ethylbenzthiazoline-6-sulphonic acid) assay (ABTS assay) (PRIOR et al., 2005). The results were expressed in mmol trolox equivalents.L<sup>-1</sup>.

All analyses were carried out in duplicate if not stated otherwise. Arithmetic mean values and standard deviations are shown for all measurements. Analysis of variance (ANOVA) was used for statistical analysis using Minitab (version 14, Minitab Inc. USA). Significant differences between means were evaluated using *t*-test. The level of significance was set at *p* < 0.05.

## 3 Results and discussion

The content of phenolic compounds in yacon chips and strips after drying was in the range of 6.8 to 10.1 mg GAE.g<sup>-1</sup> with significant lower values for chips dried at 40 and 60 °C. Similar behaviour was found when the percentage of DPPH quenched was determined. Chips dried at 40 and 60 °C inhibited 75% of DPPH, whereas chips dried at 50 °C and strips inhibited 88 and 80% of DPPH, respectively (Table 1). The slight difference in antioxidant activity between strips and chips dried at 50 °C is due to differences in the drying conditions. The strips were submitted to long-term exposure to oxidative conditions during drying. Phenolic compounds could have been oxidised by phenol oxidases.

Compared to other known natural sources of antioxidants that have been studied, such as tomato (*Lycopersicon esculentum* Mill) (CHANG et al., 2006), the content of total phenolic compounds is higher for yacon products, while the capacity of quenching DPPH is higher for tomato. Vasco, Ruales and Kamal-Eldin (2008) classified seventeen different crops from Ecuador based on analyses of total phenolic compounds. According to Vasco's classification, yacon strips and chips belong to the group of "crops with high phenolic content". The total phenolic values obtained for yacon strips and chips are higher than those determined in plums

**Table 1.** Total phenolic content (mg GAE.g<sup>-1</sup>, wet basis) and percentage of quenching of DPPH (Q%) in yacon chips and strips after drying.

| Sample | Total phenolic content<br>(mg GAE.g <sup>-1</sup> ) | Q (%)<br>(quenching of DPPH) |
|--------|---|------------------------------|
| Strips | 9.7 ± 0.2 <sup>a</sup>                              | 79.7 ± 0.1 <sup>b</sup>      |
| Chips  |   |                              |
| 40 °C  | 7.1 ± 0.1 <sup>b</sup>                              | 75.4 ± 0.3 <sup>c</sup>      |
| 50 °C  | 10.1 ± 0.1 <sup>a</sup>                             | 88.1 ± 0.6 <sup>a</sup>      |
| 60 °C  | 6.8 ± 0.1 <sup>b</sup>                              | 74.6 ± 0.1 <sup>c</sup>      |

Values with different letter in the same column are significantly different (*p* < 0.05). Mean values ± standard deviation.

(*Prunus salicina* Lindl.), strawberry (*Fragaria ananassa* Duch.) and tomato (*Lycopersicon esculentum* Mill.)

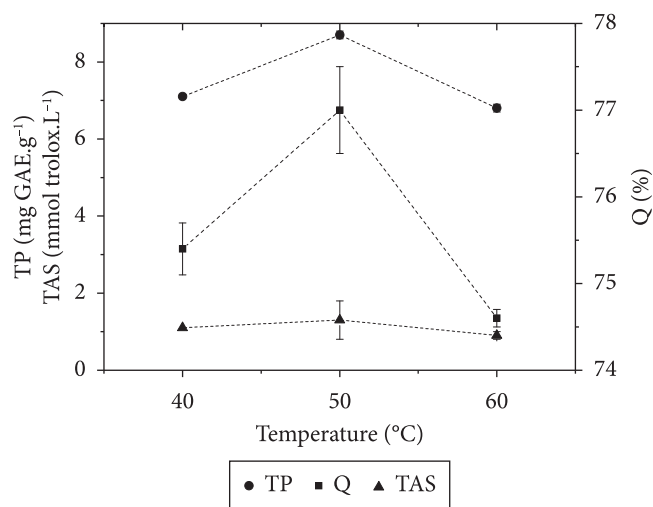
Furthermore, our results are similar to those reported by Lachman et al. (2007) with regard to the phenolic content. Lachman et al. (2007) determined the total phenolic content in fresh yacon roots using genotypes from Bolivia, Ecuador, New Zealand, and Germany reporting a total phenolic content from 5 to 15 mg GAE.g<sup>-1</sup> on dry matter basis. Therefore, drying yacon into chips or strips does not significantly affect the phenolic content compared with that in fresh yacon roots.

With regards to yacon chips, the results showed that total phenolic content and the percentage of DPPH inhibited decreased in chips dried at 40 and 60 °C (Figure 1). The temperatures used for drying in the present study, are in the range at which polyphenol oxidases in yacon roots are active (NEVES; DA SILVA, 2007). Neves and da Silva (2007) found that the optimum temperature of polyphenol oxidases activity in yacon is 30 °C, and their enzymatic activity progressively decreases as the temperature reaches 80 °C. Therefore, when drying at 40 °C, there is a reduction in phenolic content and in the percentage of DPPH inhibited (Figure 1). Similarly, chips dried at 60 °C had lower phenolic content compared to those dried at 50 °C. An increase in temperature shortens drying time and inactivates polyphenol oxidases to some extent, but it may also cause a loss of thermo sensitive antioxidant compounds during processing (BROOKS; EL-HANA; GHALY, 2008; CHANG et al., 2006; ZANONI et al., 1998). Despite the differences in total phenolic content (TP) and quenching of DPPH in yacon chips, no significant differences were found in the total antioxidant status (TAS) (Figure 1).

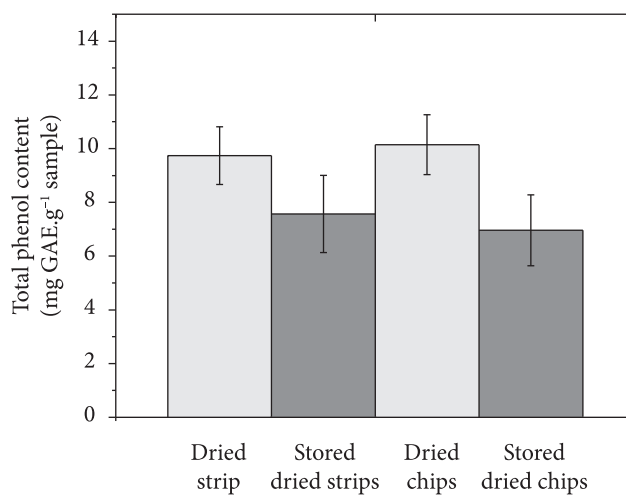
To summarise, the antioxidant capacity was less affected when yacon was processed into chips and dried at 50 °C. Negi and Roy (2001) also found lower levels of antioxidant compounds when sun drying was used compared to cabinet drying. The time that the drying process takes can indirectly affect the retention of phenolic compounds. Depending on the geometry of the samples, the drying time can be reduced. This is a reason why to process yacon into chips; increasing surface area per weight results in a faster decrease in moisture content over time (BROOKS; EL-HANA; GHALY, 2008).

Storage time is another factor that can affect retention of antioxidant capacity in dried strips and chips. Strips and chips were packaged in polyethylene bags and stored for 7 months at 20 °C in a dark and ventilated room.

A difference in total phenolic content was found between freshly dried strips and chips compared with samples stored for 7 months (Figure 2). The percentage of phenolic compounds lost in chips and strips was 31 and 20%, respectively. Unlike what happens to strips, the decrease of phenolic compounds in chips was significant. A likely explanation for this is the large surface area per mass unit that chips have in comparison to strips. There could be an interaction between the surface of chips and the remaining oxygen inside the polyethylene bag. As a result, a more extensive oxidation of phenolic compounds occurs in comparison to that in strips during storage.



**Figure 1.** Antioxidant capacity determined as Total Phenol content (TP ●), percentage of DPPH inhibited (Q ■) and Total Antioxidant Status (TAS ▲) in yacon chips at different drying temperatures. Mean values on fresh weight basis and standard error of the mean from two replicates.



**Figure 2.** Total phenol content (mg GAE.g<sup>-1</sup> of sample) determined in dried yacon strips and chips before and after 7 months of storage. Error bars are standard error of the mean.

The time used in processing and/or storage affect the antioxidant level and antioxidant capacity in the product (NEGI; ROY, 2000, 2001; WU et al., 2004). Therefore, it is important and necessary to determine the content of antioxidants in the final product.

#### 4 Conclusions

Through drying, it is possible to retain antioxidant capacity in yacon roots. The retention of antioxidant activity depends on the drying method employed and the geometry of the samples. Processing yacon into chips and drying them in a drying cabinet at 50 °C, results in a better retention of antioxidant capacity.

However, strips have a more suitable shape than that of chips for storage purposes because the larger surface area of chips increases the exposure of phenolic compounds to oxidation by remaining oxygen inside the polyethylene bags. Nevertheless, this can be solved with the use of vacuum packaging.

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