

EFFECTIVENESS OF HOME BLEACHING AGENTS IN DISCOLORED TEETH AND INFLUENCE ON ENAMEL MICROHARDNESS

Carina Sinclér DELFINO¹, Michelle Alexandra CHINELATTP, Laíse Daniela CARRASCO-GUERISOLI¹, Andriago Reis BATISTA³, Izabel Cristina FRÖNER⁴, Regina Guenka PALMA-DIBB⁵

1- DDS, MSc, Graduate student, Department of Restorative Dentistry, Ribeirão Preto Dental School, University of São Paulo, Ribeirão Preto, SP, Brazil.

2- DDS, MSc, PhD, Lab Technician, Department of Restorative Dentistry, Ribeirão Preto Dental School, University of São Paulo, Ribeirão Preto, SP, Brazil.

3- Undergraduate student, Department of Restorative Dentistry, Ribeirão Preto Dental School, University of São Paulo, Ribeirão Preto, SP, Brazil.

4- DDS, MSc, PhD, Associate Professor of Endodontics, Department of Restorative Dentistry, Ribeirão Preto Dental School, University of São Paulo, Ribeirão Preto, SP, Brazil.

5- DDS, MSc, PhD, Associate Professor of Restorative Dentistry, Department of Restorative Dentistry, Ribeirão Preto Dental School, University of São Paulo, Ribeirão Preto, SP, Brazil.

Corresponding address: Profa. Dra. Regina Guenka Palma-Dibb - Faculdade de Odontologia de Ribeirão Preto - USP - Departamento de Odontologia Restauradora - Av. do Café, s/nº. CEP: 14040-904. Ribeirão Preto - SP, Brazil - Phone: 55-16-3602-4016 - Fax: 55-16-3633-0999 - e-mail: rgpalma@forp.usp.br

Received: August 26, 2008 - Modification: November 10, 2008 - Accepted: November 30, 2008

ABSTRACT

Objectives: This study evaluated the effectiveness of different home bleaching agents on color alteration and their influence on surface and subsurface microhardness of discolored bovine enamel. Material and Methods: Forty-five fragments of bovine incisors were randomly allocated into 3 groups (n=15) according to the bleaching agent: 10% carbamide peroxide gel (CP10), 16% carbamide peroxide gel (CP16) and 6.5%-hydrogen-peroxide-based strip (HP6.5). Before bleaching treatment, initial values of Knoop surface microhardness and color (CIEL*a*b*) were obtained and the fragments were artificially stained in hemolyzed rat blood. Then, bleaching treatments were performed over a 21-day period. Color changes (ΔE) were assessed at 7, 14 and 21 days, and final surface microhardness reading was done after 21 days. Thereafter, the fragments were bisected to obtain subsurface microhardness. Data were subjected to ANOVA and Tukey's tests ($\alpha=5\%$). Results: Color changes produced by CP16 were similar to those of CP10, and the color changes produced by these materials were significantly superior to those produced by HP6.5. Color changes at 21 days were superior to 7 days and similar to 14 days. The time did not influence color changes for CP16, which showed similarity between the 14- and 21-day results. No statistically significant differences were found among the home bleaching agents for surface and subsurface microhardness. Conclusions: Microhardness of bovine enamel was not affected by the bleaching agents. The 16% carbamide peroxide gel was the most effective for bleaching the stained substrate.

Key words: Tooth bleaching. Dental enamel. Microhardness. Color change, CIEL*a*b*.

INTRODUCTION

Tooth color is currently believed to be one of the most important esthetic concerns for patients¹. Vital tooth bleaching has widely been performed, and several materials and techniques have been presented for this purpose¹⁶. Vital tooth bleaching is considered a safe, effective, minimally invasive, non-destructive and well-accepted procedure for the treatment of discolored teeth^{27,29}.

A number of methods and approaches have been described for the bleaching of vital teeth¹⁷. However, basically, there are three techniques: in-office or power bleaching, mass market bleaching products, and dentist-

supervised home bleaching¹⁵. Since its introduction by Haywood and Heymann¹⁴ (1989), several products have been employed for home bleaching. These products are mainly available in gels or strips containing several concentrations of carbamide peroxide or hydrogen peroxide²⁹. Carbamide peroxide gel at a 10% concentration is one of the most commonly employed home bleaching agents due to its safety and effectiveness¹². Although different concentrations of carbamide and hydrogen peroxides have shown similar results^{10,18,29}, increased concentrations and application times may cause enamel surface alterations, such as loss of mineral content and microhardness^{2,22}.

Thus, considering the divergences on the effects of

bleaching agents on properties of hard tooth tissues, the purposes of this study were to evaluate the effectiveness of different home bleaching agents and their effects on surface and subsurface microhardness of bovine enamel. The hypotheses tested in this study were that the type of bleaching agent influences the color alteration in stained teeth, but does not influence the surface and subsurface microhardness of bovine enamel.

MATERIAL AND METHODS

Enamel Specimen Preparation

Forty-five freshly extracted sound bovine incisors were selected and stored under refrigeration in a saturated thymol solution until preparation for testing. Teeth had their roots removed 2 mm apically to the cemento-enamel junction in a sectioning machine with a water-cooled diamond saw (Minitom, Struers A/S, Copenhagen, D2610, Denmark). The buccal surfaces of the crowns were cut longitudinally and transversely, using double-faced diamond discs mounted at a low-speed handpiece in order to obtain fragments with dimensions of 11 mm x 11 mm. To obtain plane surfaces, required for microhardness tests, the external enamel surfaces of the fragments were flattened in a water-cooled polishing machine (Politriz; Struers A/S) using 600-, 1000- and 1200-grit silicon carbide abrasive papers sequentially, followed by a final polishing with a 0.3- μ m-gamma alumina suspension on felt wheel. The specimens were kept in an ultrasonic bath (Sonic Clean, D.M.C. Equipamentos Ltda., São Carlos, SP, Brazil) in distilled and deionized water for 10 min to remove polishing debris.

Staining Procedure

Artificial staining of the specimens was performed

following a modification⁵ of the method proposed by Freccia and Peters⁸. Blood samples collected from adult male Wistar rats (weighing 200 to 250 g) were heparinised to avoid coagulation, and were centrifuged at 10,000 rpm for 10 min. The blood serum was discarded and 40 mL of distilled and deionized water were added to 60 mL of the precipitated blood. This mixture was centrifuged at 10,000 rpm for 20 min, resulting in a rich-haemoglobin hemolyzate blood solution. Specimens were immersed in this solution for 4 days, performing a centrifugation cycle (10,000 rpm, 20 min) at every 24 h. After 4 days, the specimens were removed, washed in distilled water, dried with absorbent paper and kept at 37°C in 100% relative humidity for 15 days.

Bleaching Treatment

After staining, the specimens were randomly assigned to 3 groups (n=15) according to the bleaching treatment: 10% carbamide peroxide gel (CP10), 16% carbamide peroxide gel (CP16) and 6.5%-hydrogen-peroxide-based strip (HP6.5). The bleaching treatment was performed over 21 days, according to the manufacturers' instructions. Specifications of the materials used in this phase are given in Figure 1. For each specimen of the groups treated with the bleaching gels (CP10 and CP16), a tray was fabricated using a low-density polyethylene sheet and a vacuum-thermoforming machine. Every day, a thin layer of each gel was applied on the enamel surface and the tray was placed on each specimen, which was stored for 8 h at 37°C in a recipient containing artificial saliva (pH 7.0 – Figure 1). After 8 h, the gel was removed from enamel surface with running distilled water for 15 s.

Enamel surfaces of the group treated with the bleaching strip (HP6.5) were covered with the strip for 30 min twice a day. After each application, the strips were removed from enamel surfaces, which were rinsed with running distilled

Bleaching agent	Composition	Manufacturer
CP10 Opalescence Whitening Gel	10% Carbamide Peroxide, Carbopol, Glycerin, Flavoring	Ultradent Products Inc., South Jordan, Utah, USA
CP16 Clarigel Gold 16%	Sodium Benzoate, EDTA, Carbopol, 16% Carbamide Peroxide, Sodium Fluoride, Mint Flavor	Dentsply Indústria e Comércio Ltda, Petrópolis, RJ, Brazil
HP6.5 Crest Whitestrips Classic	Purified Water, Glycerin, 6.5% Hydrogen Peroxide, Carbopol, Sodium Hydroxide, Sodium Acid Pyrophosphate, Sodium Stannate	Procter & Gamble Company, Cincinnati, Ohio, USA
Artificial saliva	K ₂ HPO ₄ , 70% Sorbitol, NaF, KCl, NaCl, MgCl ₂ ·6H ₂ O, CaCl ₂ ·2H ₂ O, Nipagin, Sodium Benzoate, Hydroxyethylcellulose	Department of Biochemistry, School of Pharmaceutical Sciences of Ribeirão Preto, University of São Paulo, Ribeirão Preto, SP, Brazil

FIGURE 1- Materials employed

water for 15 s.

In all groups, when the specimens were not in contact with the bleaching agents, they were kept at 37°C immersed in artificial saliva, which was changed daily.

Color Measurement

The color of the specimens was measured after the artificial staining (baseline) and 7, 14 and 21 days after the beginning of the bleaching treatment. Before each color measurement, specimens were rinsed with water and dried with absorbent paper. Color was measured over a white background employing a color spectrophotometer (Color guide 45/0, PCB 6807; BYK-Gardner, GmbH Geretsried, Germany), which records the color variables L*, a*, b* according to the CIEL*a*b* (Commission Internationale de l’Eclairage L*, a*, b*) color system¹⁸, where L* stands for luminosity dimension or whiteness, ranging from 0 (pure black) to 100 (reference white), a* for green-red contrast (-a*= green and +a*= red) and b* for blue-yellow contrast (-b*= blue and +b*= yellow). Color change (ΔE) was calculated from L*, a* and b* values employing the following formula: $\Delta E = [(\Delta L^*)^2 + (\Delta a^*)^2 + (\Delta b^*)^2]^{1/2}$. Positive ΔL* means the specimens became whiter, while negative ΔL* means specimens became darker.

Microhardness Testing

Knoop microhardness measurements of the enamel surface were performed before the staining procedure (initial values) and following the bleaching treatment (after 21 days) using a microhardness tester (HMV-2000; Shimadzu Corporation, Kyoto, Japan). The specimens were individually fixed in the device in such a way that the test surface was perpendicular to the micro-indenter tip. Three indentations (100 g load, 30 s) equally spaced over a circle and not closer than 1 mm to the adjacent indentations or the margin of the specimen were taken and the average was

calculated. The post-bleaching measurements were repeated for all the specimens at locations near to the previous series of indentations.

Microhardness of enamel subsurface was obtained after measuring surface microhardness post-bleaching. Specimens were individually included in acrylic resin blocks to facilitate that they were perpendicularly bisected by halves. One cross-sectioned face of each specimen was ground and polished following the protocol initially described. Measurements of subsurface microhardness were accomplished at distances of 50 μm, 100 μm, 150 μm and 200 μm from the external enamel surface. At each distance, three linearly 100-μm-spaced indentations were performed and the mean was calculated. Settings for load and penetration were equal to the one employed for surface microhardness.

Statistical Analysis

Data from color change and microhardness testing were analyzed for homogeneity and normality and were subjected to ANOVA and Tukey’s test at a 5% significance level.

RESULTS

Color Change

Table 1 displays the mean values (and standard deviations) of color change (ΔE*, ΔL*, Δa*, and Δb* values) at each time point. The ΔE* values produced by the bleaching agent CP16 were similar to those of CP10, and the ΔE* values produced by both CP16 and CP10 were significantly superior to those of HP6.5. The ΔE* values obtained at each time point revealed that the color changes at 21 days were superior to those observed at 7 days, but similar were similar to those occurred at 14 days. There was no significant difference between 7 and 14 days. The bleaching agents x time interaction showed that, for CP16, there was no

TABLE 1 - Mean values and standard deviations of ΔE, ΔL*, Δa*, and Δb* according to bleaching time and agent

		7 days	14 days	21 days
ΔE	CP10	10.22 (3.60)	10.72 (3.38)	11.63 (3.34)
	CP16	12.35 (3.16)	12.92 (3.32)	14.05 (3.61)
	HP6.5	7.14 (3.22)	7.51 (3.24)	9.37 (3.52)
ΔL*	CP10	8.36 (4.69)	8.36 (4.46)	9.04 (4.46)
	CP16	11.75 (3.54)	11.27(4.24)	12.39(4.35)
	HP6.5	5.81 (3.74)	5.44(3.99)	7.23(4.58)
Δa*	CP10	0.39 (0.82)	0.30 (0.72)	0.29 (1.05)
	CP16	-0.52 (0.87)	-0.68 (0.89)	-0.61 (0.88)
	HP6.5	0.21 (0.91)	0.06 (0.85)	0.02 (0.75)
Δb*	CP10	-4.54 (2.47)	-5.61 (2.50)	-6.19 (2.73)
	CP16	-3.76 (1.51)	-5.49 (1.68)	-5.80 (2.13)
	HP6.5	-3.57 (0.79)	-4.47 (1.25)	-4.92 (1.97)

TABLE 2- Mean Knoop microhardness values and standard deviations of enamel surface before and after bleaching treatment with different agents

Agent	Before bleaching	After bleaching
CP10	194.58 (6.34)	187.08 (4.42)
CP16	175.13 (5.27)	181.71 (5.75)
HP6.5	212.46 (3.23)	192.79 (3.10)

TABLE 3- Mean Knoop microhardness values and standard deviations at different distances from enamel surface, before and after bleaching treatment with different agents

Agent	50 μ m	100 μ m	150 μ m	200 μ m
CP10	176.13 (5.10)	184.38 (5.62)	183.69 (3.53)	179.13 (5.51)
CP16	186.09 (6.60)	180.59 (7.76)	170.59 (7.46)	166.80 (6.17)
HP6.5	180.45 (5.94)	186.00 (6.75)	182.15 (5.77)	174.86 (3.48)

statistically significant difference between 14 and 21 days. For both CP10 and HP6.5, the ΔE^* values were statistically significant only at 21 days.

Microhardness Test

The means and standard deviations of surface microhardness and subsurface microhardness are shown in Table 2 and Table 3, respectively. The analysis of data did not reveal significant differences among the studied factors neither in the surface nor in the subsurface microhardness.

DISCUSSION

When the objective of an *in vitro* study is to evaluate the effectiveness of bleaching agents, it is very difficult to observe differences in the effect of bleaching techniques on samples without staining⁷. Thus, the present study used the method proposed by Freccia and Peters⁸ (1982) for artificial staining of extracted teeth. In addition to being reliable, safe and easily reproducible, this method simulates one of the main causes of intrinsic tooth discoloration that is the oxidation of hemoglobin inside dentinal tubules³.

A number of methods are available for evaluating the efficacy of bleaching products⁴. Shade guides, photography, colorimeters or computer digitization can be employed to assess tooth color changes^{20,25,28}. Tooth color determination employing the commonly cited shade-based guides have limitations as they are subjected to examiner and environmental factors that can potentially influence classification of color^{9,19} and hamper the visualization of color variation existent among the thirds of a tooth crown²⁶. The CIEL*a*b* three-dimensional color space system is the most frequently quoted index employed in dental bleaching research and can be generated from colorimeters,

spectrophotometers or digital image analysis^{7,11,28}.

In the present study, the 16% carbamide peroxide bleaching agent was more effective than the 10% carbamide peroxide gel and the strip containing 6.5% hydrogen peroxide. The reason why 16% carbamide peroxide gave the highest change in L^* values is probably its extended contact time (8 h/day) with enamel compared to the considerably shorter contact time of the 6.5% hydrogen peroxide gel (30 min twice a day).

It is known that the outcome of a bleaching procedure

depends mainly on the concentration of the bleaching agent, the ability of the agent to reach the chromophore molecules, and the duration and frequency that the agent is in contact with chromophore molecules⁶. Moreover, the difference among the bleaching systems could be explained by the different kinetics of hydrogen peroxide and carbamide peroxide. Hydrogen peroxide acts as a strong oxidizing agent through the formation of free radicals²³, reactive oxygen molecules, and hydrogen peroxide anions¹³. These reactive molecules attack the long-chained, dark-colored chromophore molecules and split them into smaller, less colored and more diffusible molecules. Carbamide peroxide also yields urea that theoretically can be further decomposed to carbon dioxide and ammonia. The high pH of ammonia facilitates the bleaching procedure²⁴.

The influence of hydrogen peroxide and carbamide peroxide on enamel and dentin properties, such as surface morphology and chemistry, surface and subsurface ultrastructure and microhardness, has been extensively investigated in the literature. According to a recent review¹⁶, the most frequently employed technique for evaluating the effects of peroxide and bleaching products on enamel and dentin has been surface microhardness. Surface microhardness testing is a simple method for determining the mechanical properties of enamel and dentin surfaces and is related to a loss or gain of mineral of the dental structure¹⁶. On account of the acidic properties of the bleaching agents, changes in the mineral content of dental hard tissues may occur, leading a decrease in the microhardness values after the bleaching treatment²².

However, in the present study, neither the enamel surface microhardness nor the enamel subsurface microhardness was affected by the different bleaching agents employed. As the specimens were maintained in artificial saliva during and after the bleaching treatment,

it may be assumed that this solution promoted a continuous mineral intake due to the presence of calcium and phosphate ions in its composition, justifying the absence of microhardness changes. This also demonstrates that peroxide bleaching products have no significant effects on microhardness, and contrasting results, in general, reflect some methodological limitations that do not represent the clinical situation or employ highly acidic agents¹⁶. Furthermore, decrease in microhardness can be recovered in a post-bleaching period, following remineralization from saliva storage^{21,22}.

CONCLUSION

It may be concluded that the 16% carbamide peroxide gel was the most effective in bleaching stained bovine enamel without causing changes in the microhardness of this substrate.

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