

# Production of PHB Scaffolds Reinforced with HAp Through Electrospinning

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Electrospinning, an economical technique, is widely used for biomedical scaffold fabrication, crucial in tissue and organ regeneration, particularly with biomaterials. Polymers, either pure or reinforced with ceramics, aid in cell proliferation and tissue formation. Polyhydroxybutyrate (PHB) is a promising biopolymer for tissue engineering, offering biocompatibility comparable to petroleum-derived polymers. Combining PHB with hydroxyapatite (HAp) enhances mechanical strength and osteoconductivity. This study aims to produce electrospun PHB microfibrillar webs reinforced with HAp for scaffold fabrication. Morphological variations are analyzed through manipulation of electrospinning parameters. The study observed microfibrillar webs with diameters ranging from 2 to 9  $\mu\text{m}$ . Mechanical and microstructural evaluations demonstrate superior strength of PHB/HAp microfibrillar webs compared to pure PHB, 1.23 MPa and 0.58 MPa respectively, demonstrating the efficacy of HAp reinforcement. These findings highlight the potential of PHB/HAp microfibrillar webs in bone tissue engineering.

**Keywords:** *Electrospinning, Hydroxyapatite, Polyhydroxybutyrate, Scaffold, Tissue engineering.*

## 1. Introduction

Among the known methods for obtaining nanofibers, electrospinning has garnered significant attention in the academic world in recent decades. This is evidenced by the substantial increase in publications on the subject, numbering around 2000 articles per year<sup>1</sup>. Electrospinning, known for its simple configuration, has found diverse applications, including filtration<sup>2-4</sup>, employment in the textile industry<sup>5-7</sup>, catalysis<sup>8-10</sup>, tissue engineering<sup>11-13</sup>, wound healing<sup>14-16</sup>, drug delivery<sup>17-19</sup> and water decontamination<sup>20-22</sup>.

Within the various polymeric materials suitable for producing electrified fibers, polyhydroxybutyrate (PHB) has gained attention and extensive study due to its biodegradability, biocompatibility, non-toxicity, and mechanical properties closely resembling those of polymers like polyethylene (PE)<sup>23-25</sup>. However, similar to other organic polymers, PHB has limited bioactivity and is not suitable for applications involving high mechanical stresses. One solution to this challenge involves combining this polyhydroxyalkanoate with a bioceramic to enhance its strength and osteoconductivity compared to the pure polymer<sup>26-28</sup>.

Hydroxyapatite (HAp) has demonstrated its potential as a bioceramic that can improve the bioactivity and mechanical properties of PHB<sup>29</sup>. With distinct properties depending on its preparation method, hydroxyapatite is a calcium phosphate compound with the chemical formula  $\text{Ca}_{10}(\text{PO}_4)_6(\text{OH})_2$ . This member of the apatite family exhibits properties that render

it suitable for various applications, including serving as a synthetic bone substitute, creating extracellular supports/matrixes for tissue engineering involving cytokines, bone, and cartilage cells, and acting as a promoter for improving cell adhesion and dissemination on the surfaces of membranes designed for tissue engineering applications<sup>30-33</sup>.

Numerous studies have concentrated on producing PHB/HAp scaffolds for applications in bone tissue engineering, with the aim of achieving bone regeneration at sites affected by damage or injury<sup>29,34</sup>. Other research has investigated the electrospinning of PHB and HAp in combination with polymers like polylactic acid (PLLA)<sup>35</sup> and poly(lactic acid-co-glycolic acid) (PLGA)<sup>36,37</sup>.

The electrospinning machine used in this study offers the advantage of cost-effectiveness and ease of replication. This makes it pedagogically suitable and accessible not only for novice researchers in the field of electrospinning but also for research groups operating under budget constraints. Its total cost was US\$172.65, making it intriguing for two key reasons: it competes favorably with similar electrospinning machines developed in contemporary studies focused on the construction and application of such equipment and proves exceptionally cost-effective when compared to commercially available laboratory-grade electrospinning machines (with prices ranging from US\$15,595 to US\$60,000)<sup>38-41</sup>.

This study aims to produce microfibrillar webs of PHB and PHB/HAp through electrospinning, creating three-dimensional supports/scaffolds. The electrospinning process

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was carried out using a low-cost machine manufactured at the Materials Science Graduate Institute of the Federal University of São Francisco Valley (UNIVASF). Various electrospinning parameters, including solution flow rate, solution concentration, distance between the capillary tube and collector, rotating collector speed, and capillary tube diameter, were adjusted to determine the optimal configuration for fiber production. The effects of these parameters were investigated using scanning electron microscopy (SEM).

## 2. Materials and Methods

### 2.1. Electrospinning machine

The electrospinning machine, developed and constructed at the Materials Science Graduate Institute of the Federal University of São Francisco Valley, consists of three essential components: a high-voltage power supply, a controlled injection system (CIS), and a rotating drum collector. The rotating drum is mounted on a mobile base with rails and has a nylon body with its surface covered by an aluminum plate.

A potentiometer is used to control the rotational speed of the drum. The DC motor has a 9V input voltage that can be continuously adjusted from 0 to 9 V, corresponding to an angular velocity ranging from 0 to 580 rpm. The high-voltage source supply in the electrospinning system incorporates a BSC25-0111 106-18G flyback high-voltage transformer and a Huntkey LW-6350HG switched-mode power supply, which together elevate the input voltage from 12 V to 20 kV. The controlled injection system (CIS) consists of a stepper motor coupled to a gear reducer, which, through a spindle, converts the motor's angular motion into linear movement to operate the syringe trigger.

The polymer solution was loaded into 5 mL syringes using needles as capillary tubes. The entire CIS is controlled via Arduino. The developed electrospinning machine had a cost of US\$172.65 (approximately R\$851,17), as indicated in Table 1. The schematic design of the electrospinning

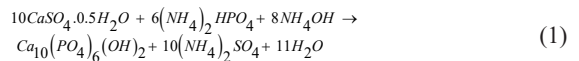
**Table 1.** Costs of the built-up electrospinning machine.

| Materials   | Cost (R\$)    |
|---|---------------|
| Wooden Board  | 60,00         |
| Drawer Slide Rails  | 25,00         |
| ATX Power Supply + Cooler + Heatsink  | 19,90         |
| Flyback Transformer BSC25   | 40,00         |
| Breadboard  | 15,00         |
| LCD Display   | 10,90         |
| Arduino Uno R3  | 48,90         |
| Stepper Motor 28ybt-48 + Drive  | 20,90         |
| Nylon Billet (55mm x 500mm)   | 52,32         |
| Vise (Acme Thread)  | 45,45         |
| U Metal Clamp 1 1/4   | 4,00          |
| Steel Round Bar 6mm   | 5,00          |
| Stainless Steel Tube 2" x 250mm   | 13,35         |
| 9V DC Motor   | 7,00          |
| Others (wires, jumpers, resistors, capacitors, syringes, needles, and screws) | 33,45         |
| Labor   | 450,00        |
| <b>Total</b>  | <b>851,17</b> |

machine can be seen in Figure 1a, while the constructed prototype is depicted in Figure 1b.

### 2.2. Hydroxyapatite synthesis

Hydroxyapatite was produced following the methodology described by Álvares<sup>42</sup>. The wet precipitation method was employed, utilizing gypsum (calcium sulfate hemihydrate,  $\text{CaSO}_4 \cdot 0.5\text{H}_2\text{O}$ ) as the calcium source, along with ammonium hydroxide ( $\text{NH}_4\text{OH}$ ) and dibasic ammonium phosphate ( $(\text{NH}_4)_2\text{HPO}_4$ ) as reagents. The molar ratio of Ca/P was calculated to be 1.67 using Equation 1.



A solution containing 0.2 mol/L of calcium sulfate hemihydrate ( $\text{CaSO}_4 \cdot 0.5\text{H}_2\text{O}$ ) was continuously stirred while dibasic ammonium phosphate ( $(\text{NH}_4)_2\text{HPO}_4$ ) at a concentration of 0.12 mol/L was added at a rate of 20 mL/min. The pH of the reaction was controlled and maintained at 9.5 by adding ammonium hydroxide ( $\text{NH}_4\text{OH}$ ) at a concentration of 3 mol/L, as needed. After the addition of all reagents, stirring was discontinued, and the solution was allowed to stand for 48 hours. The entire process was carried out at room temperature. After this elapsed time, the solution's pH was adjusted to 7.0 by washing it with deionized water, followed by vacuum filtration of the hydroxyapatite. Subsequently, all the hydroxyapatite was oven-dried for 12 hours at 100 °C. After drying, the material was ground to a fine powder using a mortar and pestle and subjected to calcination for 2 hours at 900 °C.

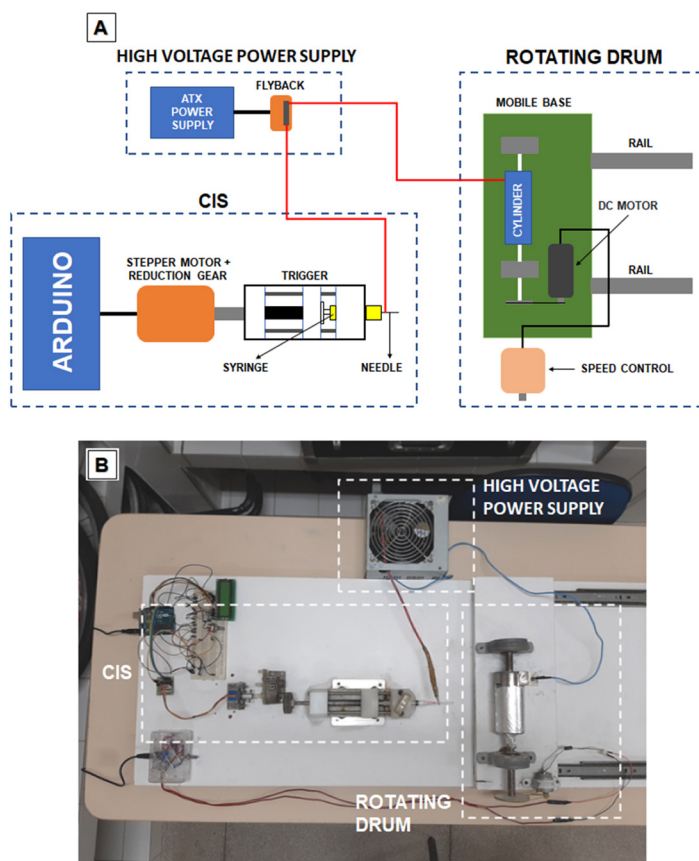
### 2.3. Polymeric solution

Polymer solutions were prepared using the following materials: PHB powder obtained from PHB Industrial (SP) with a purity exceeding 95% and an average molecular weight of approximately 600,000 g/mol, and chloroform ( $\text{CHCl}_3$ ) of PA ACS grade with a purity of 99.8% and a molecular weight of 119.38 g/mol, supplied by Metaquímica. Pure PHB solutions were prepared by magnetic stirring at 3000 rpm while heating to 60 °C for 3 hours until the polymer completely dissolved. PHB/HAp solutions were formulated by adding hydroxyapatite powder, which had been ground and immersed in a chloroform solution in an ultrasonic bath for 30 minutes, to the PHB solution. The mixture was then subjected to magnetic stirring at 3000 rpm and heated to 60 °C for 3 hours until complete dissolution of the polymer.

### 2.4. Fiber's production

#### 2.4.1. Study of the electrospinning parameters for the production of electrospun PHB and PHB/HAp microfibrinous webs

Initial experiments were carried out to determine the optimal spinning distances as a function of sample concentrations, select the most suitable polymer solution concentrations, and define the spacing between the capillary tube and collector for PHB fiber production. Additionally, the influence of varying this parameter on the morphology of the electrospun



**Figure 1.** The constructed electrospinning machine. a) Schematic diagram of the electrospinning machine; b) The built prototype of the electrospinning machine.

microfibrinous webs was analyzed. Distances of 200, 250, and 300 mm from the capillary tube (with an internal diameter of 0.80 mm) to the rotating collector were tested, with the collector speed set at 580 rpm. For each of these distances, fibers were produced using solutions with concentrations of 10.0%, 12.5%, 15.0%, 17.5%, and 20.0% (m/m). The flow rate, set in the Controlled Injection System (CIS), and used for all microfibrinous webs produced in this step, was 1.64 mL/h. The room temperature and humidity were maintained at 27 °C and 65%, respectively. The produced samples were designated according to the following standard: AXXXYYY. In this nomenclature, “A” represents the abbreviation of the word “Sample” in Portuguese, while “XXX” and “YYY” correspond to digits signifying the solution concentration and the distance between the capillary tube and the rotating collector, respectively.

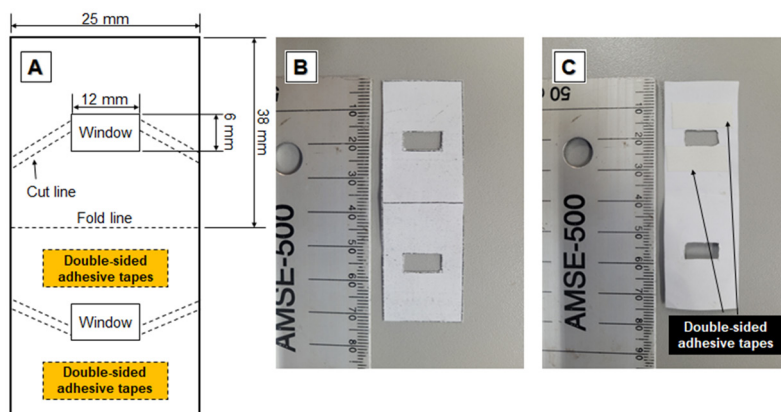
Initial tests revealed that for distances up to 200 mm, there was ineffective solvent evaporation during spinning, resulting in the production of plastic films instead of microfibrinous webs. Conversely, distances approaching 300 mm occasionally caused the electrospun jet to deviate significantly from the direction of the rotating collector. The optimal concentration/distance combination was determined to be 15.0% (m/m) at a distance of 250 mm. Consequently, these values were held constant for the analysis of the effects of varying parameters on the produced microfibrinous webs, including

drum rotating speed (290 and 580 rpm), solution flow rate (2.45 and 3.27 mL/h), and internal diameter of the capillary tube (0.80 and 1.20 mm). The voltage applied during fiber production remained constant at 20 kV.

During the production process, it was determined that among all the parameter combinations assessed, the following configuration demonstrated enhanced processability and yielded superior qualitative results for the production of pure PHB fibers: a 250 mm distance between the capillary tube and the rotating collector, a collector speed set at 580 rpm, a solution flow rate of 2.45 mL/h, an internal diameter of the capillary tube measuring 0.80 mm, and an applied voltage of 20 kV. Consequently, this configuration was adopted for the production of PHB/HAp microfibrinous webs. The production of PHB/HAp microfibrinous webs maintained a PHB concentration of 15.0% (m/m) and an HAp concentration of 2% (m/m). Environmental conditions during production were within a temperature range of 26.0°C to 28.5°C and a humidity range of 30% to 60%.

#### 2.4.2. Structural characterization

Fourier Transform Infrared Spectroscopy (FTIR) was employed to characterize both the PHB and the hydroxyapatite produced. For the analysis, the samples were blended with KBr, and pellets were prepared using a press before being subjected to the FTIR technique.



**Figure 2.** Template used in tensile tests: a) Schematic design of the template for sample accommodation; b) Front view of the template; c) Back view of the template where the microfibrous webs are accommodated.

The measurements were carried out over a wavelength range between 4000 and 500  $\text{cm}^{-1}$ , by averaging 64 scans of each spectrum with a resolution of 1  $\text{cm}^{-1}$ . The acquired data were compared with literature values for PHB and HAp. The spectrometer utilized in this analysis was the IRPrestige-21 by Shimadzu.

The Tescan Vega3 XMU scanning electron microscope, operated at an accelerating voltage of 10 kV, was utilized for imaging the microfibrous webs and observing their morphology. Prior to microscopy, all samples underwent a gold coating process using the Quorum Q150R ES Metallizer.

The micrograph was analyzed using the DiameterJ 1.018 plugin within the ImageJ 1.51 software. Scanning electron microscope images were segmented and processed for the purpose of measuring the average fiber diameter, as well as assessing the sample's porosity and fiber alignment.

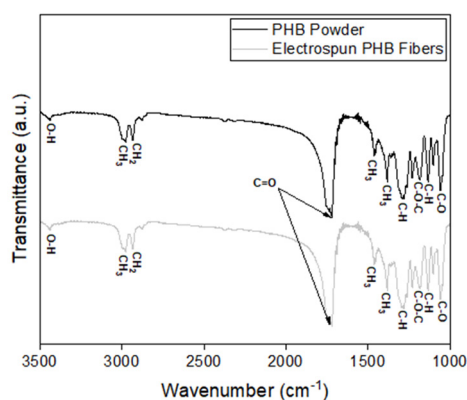
### 2.4.3. Tensile testing

The fiber tensile tests were carried out using the EMIC DL 10000 universal testing machine. Data acquisition was performed using TESC 3.04 software, and a 500 N load cell was utilized in the universal machine. All tests were carried out at a displacement rate of 1 mm/min, following the methodology described by Boakye et al.<sup>43</sup>. Each sample underwent five tests. To secure the microfibrous webs, double-sided adhesive tapes were employed. Templates were used to centrally position the microfibrous webs within the template openings, which were created by printing and precise cutting. After securing the templates in the grips of the testing machine, the lateral regions were trimmed along the cutting lines, and the tests commenced. For reference, Figure 2 illustrates the template model used along with its dimensions.

## 3. Results and Discussion

### 3.1. Characterization of raw materials

Fourier Transform Infrared Spectroscopy (FTIR) was employed to characterize PHB both before and after electrospinning to assess potential structural changes.



**Figure 3.** FTIR spectra were obtained for PHB samples in two different states: PHB powder before electrospinning and electrospun PHB fibers.

Figure 3 presents the FTIR spectrum for PHB. Characteristic PHB bands are evident in both the spectrum of PHB powder and the PHB fibers obtained through the electrospinning process, as documented in previous studies by Barbosa<sup>44</sup>, El-Hadi et al.<sup>45</sup>, and Furukawa et al.<sup>46</sup>. The identified spectral bands were as follows: 3435  $\text{cm}^{-1}$  for O-H stretching; 2972  $\text{cm}^{-1}$  for C-H stretching; 1726  $\text{cm}^{-1}$  for C=O stretching; 1460 and 1382  $\text{cm}^{-1}$  for asymmetric and symmetric  $\text{CH}_2$  deformations, respectively; 1286  $\text{cm}^{-1}$  for C-H deformation; 1282 and 1226  $\text{cm}^{-1}$  for the stretching vibrations of C-O-C; 1176 and 1106  $\text{cm}^{-1}$  for C-O-C stretching; 1131  $\text{cm}^{-1}$  for  $\text{CH}_2$  rocking; and 1054  $\text{cm}^{-1}$  for C-O stretching.

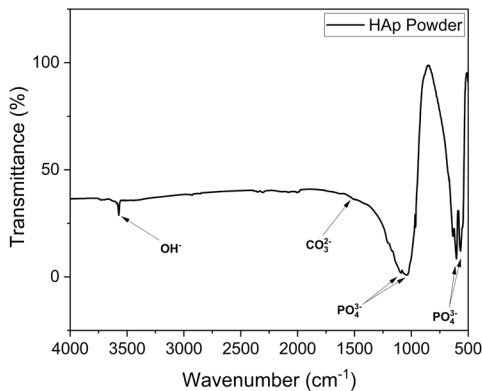
The hydroxyapatite produced through wet precipitation underwent Fourier Transform Infrared Spectroscopy (FTIR) analysis to characterize and identify its functional groups. The observed spectral bands were as follows: 3570  $\text{cm}^{-1}$  for (-OH) groups; 1650  $\text{cm}^{-1}$  for ( $\text{CO}_3^{2-}$ ) groups; 1036  $\text{cm}^{-1}$  and 1095  $\text{cm}^{-1}$  ( $\nu_3$ ), as well as 568  $\text{cm}^{-1}$  and 600  $\text{cm}^{-1}$  ( $\nu_4$ ), which are indicative of the ( $\text{PO}_4^{3-}$ ) ions. The FTIR spectrum obtained for HAp is presented in Figure 4, where the characteristic bands for hydroxyapatite are observed as shown in Ślósarczyk et al.<sup>47</sup> and Rehman and Bonfield<sup>48</sup>.

### 3.2. PHB fiber production and analysis

#### 3.2.1. Microstructural characterization of the microfibrinous webs

At the outset, microfibrinous webs were fabricated by systematically adjusting both the polymer concentration and the distance between the capillary tube and the collector to determine the optimal configuration for subsequent tests. Consequently, microfibrinous webs were produced with variations in distance, spanning from 200 to 300 mm, and concentrations ranging between 10% and 20% (m/m). Micrographs of these samples, captured using scanning electron microscopy, are presented in Figures 5 and 6, illustrating different concentrations of the polymer solution and varying distances between the capillary tube and the rotating collector, respectively. The fibers exhibit a low surface roughness, and their diameters display uniformity in size.

The average diameters of the fibers produced in this work are presented in Tables 2 and 3. In Table 2, it is observed that as the solution concentration increased for a fixed distance,



**Figure 4.** FTIR spectrum of HAp powder (produced through the wet precipitation method) after calcination at 900°C.

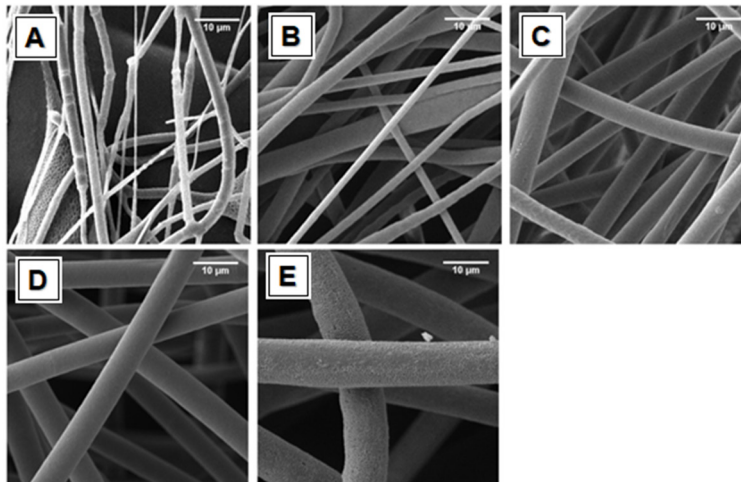
there was also an increase in the average diameter of the fibers produced. This effect of solution concentration at a constant distance between the capillary tube and rotating collector is consistent with the findings of Sukigara et al.<sup>49</sup>, Bhardwaj and Kundu<sup>50</sup>, Matabola and Moutloali<sup>51</sup>, and Neo et al.<sup>52</sup>. Table 3 indicates that increasing the distance from the collector to the capillary tube resulted in an increase in fiber diameters, which is consistent with the results reported for fibers produced by Cramariuc et al.<sup>53</sup>, Chen et al.<sup>54</sup>, and Yördem et al.<sup>55</sup>. Nevertheless, previous studies, such as those by Kameoka and Craighead<sup>56</sup>, Chowdhury and Stylios<sup>57</sup>, and Bosworth and Downes<sup>58</sup>, have reported decreases in average fiber diameters as the deposition distance on the rotary collector increases. However, when considering the errors associated with diameter measurements across these

**Table 2.** Average diameters of electrospun fibers at a fixed distance of 300 mm between the capillary tube and the collector for various concentrations.

| Sample  | Solution Concentration (m/m) | Distance (mm) | Average diameters ( $\mu\text{m}$ ) |
|---------|------------------------------|---------------|-------------------------------------|
| A100300 | 10.0%                        | 300           | $2.10 \pm 0.98$                     |
| A125300 | 12.5%                        | 300           | $2.91 \pm 0.26$                     |
| A150300 | 15.0%                        | 300           | $4.77 \pm 1.89$                     |
| A175300 | 17.5%                        | 300           | $6.29 \pm 1.00$                     |
| A200300 | 20.0%                        | 300           | $9.61 \pm 1.50$                     |

**Table 3.** Average diameters of electrospun fibers at a fixed concentration of 17.5% (m/m) for various distances.

| Sample  | Solution Concentration (m/m) | Distance (mm) | Average diameters ( $\mu\text{m}$ ) |
|---------|------------------------------|---------------|-------------------------------------|
| A175200 | 17.5%                        | 200           | $5.68 \pm 0.81$                     |
| A175250 | 17.5%                        | 250           | $5.74 \pm 1.51$                     |
| A175300 | 17.5%                        | 300           | $6.29 \pm 1.00$                     |



**Figure 5.** Micrographs of the PHB fibers for a fixed distance of 300 mm at the concentrations: a) 10.0%(m/m); b) 12.5%(m/m); c) 15.0%(m/m); d) 17.5%(m/m); and e) 20.0%(m/m).

studies, the range of variation remains minimal. Consequently, drawing a definitive conclusion about the relationship between wire diameters and deposition distances becomes challenging. Therefore, in this study, it can be concluded that the distance between the collector and the capillary tube had no significant effect on fiber diameters. While an analysis of the relationship between electric field voltage and deposition distances could provide further insights, such a study was not carried out due to the construction characteristics of the electrospinning machine.

### 3.2.2. Porosity of PHB microfibrous webs

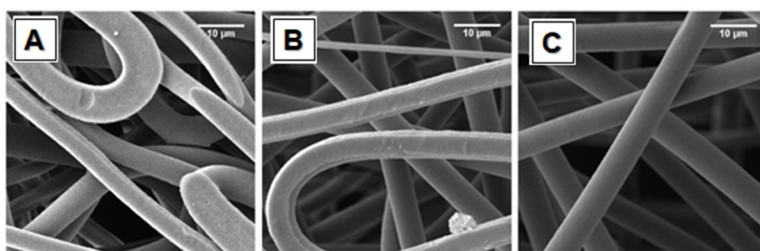
The percentage of porosity exhibited by the fiber assembly in each sample under different concentrations and at various distances between the capillary tube and rotary collector can be observed in Tables 4 and 5, respectively. Porosity indicates the density of fibers produced in the microfibrous webs. Thus, lower porosity means higher fiber density. Examining Table 4, we can observe that as the solution concentration increases, the porosity of the microfibrous webs decreases. The observed results suggest an increase in fiber density in the produced samples. Specifically, concerning the fixed distance between the capillary tube and the rotating collector, the decrease in porosity is directly correlated with a higher quantity of PHB in the solution, indicating a more substantial amount of polymer being electrospun. It's worth

**Table 4.** Porosity of microfibrous webs for a fixed distance of 300 mm between the capillary tube and collector and varying concentrations.

| Sample  | Solution Concentration (m/m) | Distance (mm) | Porosity (%) |
|---------|------------------------------|---------------|--------------|
| A100300 | 10.0%                        | 300           | 48.02        |
| A125300 | 12.5%                        | 300           | 33.64        |
| A150300 | 15.0%                        | 300           | 29.22        |
| A175300 | 17.5%                        | 300           | 32.08        |
| A200300 | 20.0%                        | 300           | 20.13        |

**Table 5.** Porosity of microfibrous webs for samples with a fixed concentration of 17.5% (m/m) at different distances.

| Sample  | Solution Concentration (m/m) | Distance (mm) | Porosity (%) | Average diameters of electrospun fibers ( $\mu\text{m}$ ) |
|---------|------------------------------|---------------|--------------|---|
| A175200 | 17.5%                        | 200           | 31.21        | $5.68 \pm 0.81$   |
| A175250 | 17.5%                        | 250           | 37.34        | $5.74 \pm 1.51$   |
| A175300 | 17.5%                        | 300           | 32.08        | $6.29 \pm 1.00$   |



**Figure 6.** Micrographs of the PHB fibers for a fixed concentration of 17.5% (m/m) at distances: a) 200 mm, b) 250 mm, and c) 300 mm.

noting that the 17.5% (m/m) sample did not conform to this observed pattern. Regarding the relationship between the porosity of PHB microfibrous webs and the distance between the capillary tube and the rotating collector, no significant influence of this parameter was noted. These findings are presented in Table 5.

### 3.2.3. Tensile characterization of PHB microfibrous webs

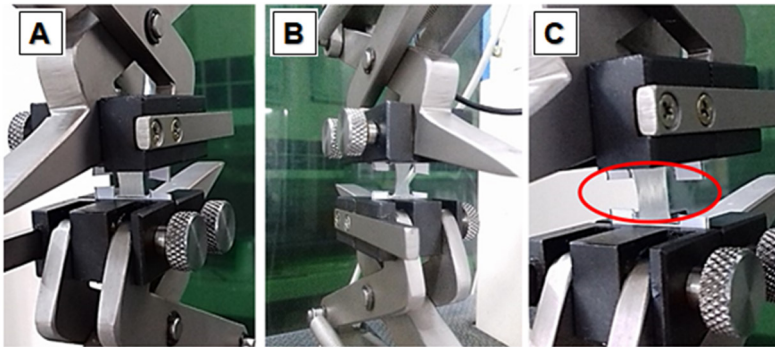
All the PHB microfibrous webs were tested until the complete rupture of all strands. A common characteristic among the tested samples was low elongation, indicating low ductility. Figure 7 illustrates the tensile test performed, and Table 6 presents the tensile strength and Young's modulus values of the tested fibers. It is evident that neither the tensile strength nor Young's modulus exhibited a noticeable trend or significant influence with variations in solution concentration and the distance between the rotary collector and capillary tube, consistent with prior findings by Asvar et al.<sup>59</sup>. This is suspected to be a consequence of the pronounced non-alignment exhibited by the electrospun fibers. The low rotation speed of the drum collector used in the experiments (580 rpm) induces fiber misalignment. Due to their different angles and subsequent rotation while attempting to align with the loading direction during tensile tests, the fibers experience mechanical failure, breaking at the junctions and cohesion points between them. Thus, no significant conclusions regarding the effect of varying solution concentration and the distance between the rotary collector and capillary tube on mechanical properties could be observed<sup>60-62</sup>.

However, by fixing the concentration at 20.0% (m/m) and varying only the distances, it was possible to evaluate the effect of changing the distance between the capillary tube and the collector on the stress-strain curves in Figure 8a. Similarly, it was possible to assess the impact of the concentration of the polymer solution by setting the distance at 250 mm and varying the concentration, as shown in Figure 8b.

By analyzing Figure 8, one can observe the fragility of PHB when subjected to mechanical stress, as also emphasized by

**Table 6.** Tensile properties of PHB microfibrous webs.

| Sample  | Solution Concentration (m/m) | Distance (mm) | Tensile Strength (MPa) | Young's Modulus (MPa) | Elongation at break (%) |
|---------|------------------------------|---------------|------------------------|-----------------------|-------------------------|
| A150200 | 15.0%                        | 200           | 0.90 ± 0.14            | 6.90 ± 0.83           | 32.56 ± 15.47           |
| A150250 | 15.0%                        | 250           | 0.83 ± 0.08            | 5.23 ± 0.14           | 63.56 ± 33.48           |
| A150300 | 15.0%                        | 300           | 1.04 ± 0.16            | 9.25 ± 0.37           | 64.30 ± 26.13           |
| A175200 | 17.5%                        | 200           | 0.92 ± 0.13            | 7.95 ± 0.87           | 95.55 ± 46.03           |
| A175250 | 17.5%                        | 250           | 0.74 ± 0.05            | 11.87 ± 0.77          | 89.77 ± 42.88           |
| A175300 | 17.5%                        | 300           | 0.89 ± 0.13            | 4.90 ± 0.62           | 95.53 ± 23.38           |
| A200200 | 20.0%                        | 200           | 0.97 ± 0.13            | 10.50 ± 0.99          | 84.21 ± 16.83           |
| A200250 | 20.0%                        | 250           | 0.82 ± 0.09            | 3.91 ± 0.69           | 89.70 ± 31.87           |
| A200300 | 20.0%                        | 300           | 0.74 ± 0.03            | 14.47 ± 0.22          | 31.39 ± 11.38           |

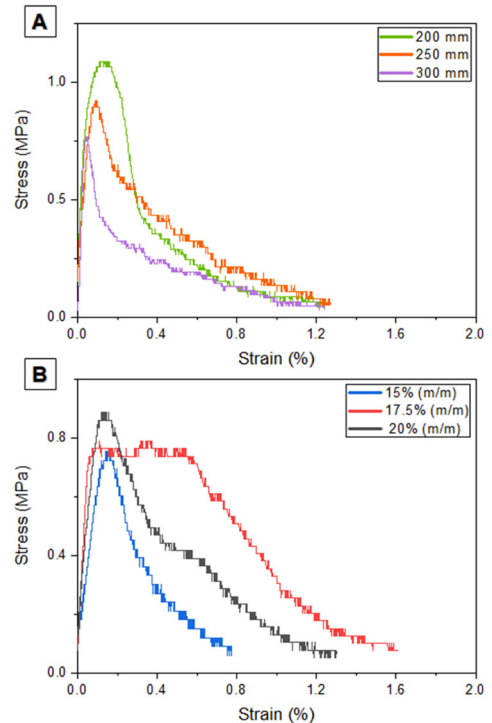
**Figure 7.** Tensile test performed on the electrospun PHB microfibrous webs. a) Microfibrous webs positioned before the start of the test; b) Behavior of the microfibrous webs during the test; and c) Rupture.

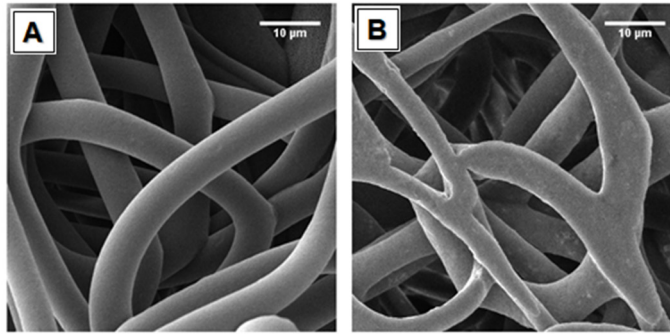
Arrieta et al.<sup>63</sup> and Ding et al.<sup>64</sup>. In Figure 8a, it is evident that, for a fixed solution concentration, an increase in the distance between the capillary tube and the rotating collector leads to a higher maximum stress supported by the samples. This behavior can be attributed to the decrease in fiber diameter with an increasing distance between the capillary tube and the collector. As the wire diameters become thinner, there is a more significant alignment of the molecular chains within the fibers, resulting in increased crystallinity<sup>65</sup>. Consequently, this molecular alignment provides enhanced resistance to tensile forces. The observed increase in tensile strength can also be attributed to the improved alignment of lamellae and fibrillar structures. These fibrillar structures exhibit a high degree of molecular orientation, as suggested by Kameoka and Craighead<sup>56</sup> and Baji et al.<sup>66</sup>. Figure 8b illustrates the impact of polymer solution concentration. As the concentration of the solution rises, there is a reduction in the ultimate tensile strength. Solutions with higher concentrations tend to produce wires with larger diameters, a phenomenon also noted in the study by Bhardwaj and Kundu<sup>50</sup>. Larger average fiber diameters correspond to lower tensile strength, as elucidated by Baji et al.<sup>66</sup>.

### 3.3. PHB/HAP fiber production and analysis

#### 3.3.1. Characterization of PHB/HAP microfibrous webs

The tensile strength results are presented in Table 7. It can be observed that the inclusion of hydroxyapatite

**Figure 8.** a) - Stress-strain curves for a 20% (m/m) sample at different distances from the collector; b) Stress-strain curves for a sample at a fixed distance of 250 mm at different concentrations.



**Figure 9.** Micrographs of PHB fibers: a) Pure PHB fibers (15.0% m/m) and b) PHB/HAp fibers (PHB 15.0% (m/m) / 2% (m/m) HAp).

particles in the PHB fibers significantly increased both the tensile strength and Young's modulus. This enhancement is attributed to favorable interactions between the polymeric matrix and the bioceramic, which is uniformly distributed within the fibers. This characteristic acts as a filler, facilitating the efficient transfer of tensile load from the polymer to the HAp, in agreement with the findings of Ramier et al.<sup>67</sup>.

The PHB/HAp microfibrinous webs exhibited a tensile strength of 1.23 MPa, representing a 112.06% increase compared to microfibrinous webs produced under the same parameters but without the addition of bioceramic. This demonstrates the excellent potential of HAp as a reinforcement for PHB in the construction of scaffolds for bone tissue engineering. Applications such as bone graft manufacturing become more viable with this enhancement. The properties of the fibers developed in this research closely resemble those reported by Chen et al.<sup>34</sup>, who utilized similar fibers for bone graft applications. The utilization of PHB/HAp fibers for bone regeneration is also widely advocated in the studies of Guan et al.<sup>68</sup>, Wang et al.<sup>69</sup>, Ye et al.<sup>70</sup>, and Doyle et al.<sup>71</sup>.

### 3.3.2. Microstructural characterization of PHB/HAp microfibrinous webs

The microstructure morphologies of the PHB/HAp fibers compared to the pure PHB fibers can be seen in Figure 9. Electrospinning was successful, resulting in the formation of long, continuous fibers for both microfibrinous webs. The pure PHB meshes in Figure 9a appear smoother and more uniform. In contrast, the PHB/HAp meshes in Figure 9b exhibit a rougher surface. This rougher surface is attributed to the aggregation of hydroxyapatite particles in distinct regions of the fiber surface, as noted by Guan et al.<sup>68</sup>.

The comparison of the average diameters of PHB and PHB/HAp fibers is presented in Table 8. It is evident that the average fiber diameter tends to decrease with the addition of hydroxyapatite to the polymer solution. This phenomenon can be attributed to the increased conductivity of the PHB/HAp solution compared to the PHB solution, resulting from the presence of calcium and phosphate ions from the bioceramic. Elevated conductivity in the solution corresponds to a higher capacity for electrical charge, leading to greater

**Table 7.** Mechanical properties comparison of PHB and PHB/HAp microfibrinous webs.

| Sample                         | Tensile Strength (MPa) | Young's Modulus (MPa) |
|--------------------------------|------------------------|-----------------------|
| PHB 15.0% (m/m)                | 0.58 ± 0.07            | 4.13 ± 0.90           |
| PHB 15.0% (m/m) / 2% HAp (m/m) | 1.23 ± 0.22            | 13.70 ± 2.94          |

**Table 8.** Average diameters presented by PHB and PHB/HAp fibers.

| Sample                         | Average diameters of electrospun fibers (µm) |
|--------------------------------|--|
| PHB 15.0% (m/m)                | 7.39 ± 0.70                                  |
| PHB 15.0% (m/m) / 2% HAp (m/m) | 6.00 ± 0.04                                  |

**Table 9.** Porosity presented by PHB and PHB/HAp microfibrinous webs.

| Sample                         | Porosity (%) |
|--------------------------------|--------------|
| PHB 15.0% (m/m)                | 58.72        |
| PHB 15.0% (m/m) / 2% HAp (m/m) | 54.47        |

stretching forces and, consequently, smaller fiber diameters, as reported by Ramier et al.<sup>67</sup>.

### 3.3.3. The porosity of PHB/HAp microfibrinous webs

The porosity of both PHB and PHB/HAp microfibrinous webs is presented in Table 9. Interestingly, the addition of hydroxyapatite to the solution resulted in a slight decrease in the fabric's fiber density, contrary to our expectations. Typically, the reduction in average fiber diameters due to the addition of HAp leads to lower fabric porosity, as described by Chen et al.<sup>34</sup>. Nonetheless, the microfibrinous webs developed in this study exhibited a consistently high level of porosity in the produced microfibrinous webs, averaging approximately 60% for all fiber samples. This high porosity is advantageous for facilitating adhesion, infiltration, and cell growth, particularly in the context of bone and cartilage tissue regeneration. It is worth noting that greater porosity in electrospun microfibrinous webs has been linked to increased cell infiltration, as demonstrated by Thorvaldsson et al.<sup>72</sup>.



## 4. Conclusions

The constructed electrospinning system demonstrated its efficiency as all PHB and PHB/HAp solutions used produced satisfactory electrospun fibers, facilitating the fabrication of scaffolds. These developed microfibrinous webs allowed for the investigation of how electrospinning parameters influence their morphology.

The variation in electrospinning parameters significantly influenced the properties of the electrospun fibers. It was observed that the porosity of the microfibrinous webs decreased with increasing concentration, indicating a higher fiber density in the produced samples.

Moreover, an increase in the concentration of the polymer solutions led to a decrease in the mechanical strength of the microfibrinous webs, attributed to the production of fibers with larger diameters when using solutions with high concentrations. The average diameter of the pure PHB fibers ranged from 2.10 to 9.61  $\mu\text{m}$ .

The addition of hydroxyapatite increased the tensile strength by 112.06% compared to pure PHB microfibrinous webs. The average diameter of the PHB/HAp fibers was smaller than that of the PHB fibers. The porosity of the microfibrinous webs decreased when HAp was incorporated into the fiber production.

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