

# Larval endoparasitoids (Hymenoptera) of frugivorous flies (Diptera, Tephritoidea) reared from fruits of the cerrado of the State of Mato Grosso do Sul, Brazil

Manoel A. Uchôa-Fernandes<sup>1</sup>Rosa M. da S. Molina<sup>1,2</sup>Isaias de Oliveira<sup>1,2</sup>Roberto A. Zucchi<sup>3</sup>Nelson A. Canal<sup>4</sup>Norma B. Díaz<sup>5</sup>

**ABSTRACT.** This paper presents a five years survey of endoparasitoids obtained from the larvae of frugivorous Tephritidae and Lonchaeidae flies. The insects were reared from cultivated and wild fruits collected in areas of the cerrado in the State of Mato Grosso do Sul, Brazil. The flies obtained from 14 host fruit species were eight *Anastrepha* species, *Ceratitis capitata* (Wiedemann, 1824) (Tephritidae); *Dasiops* sp. and *Neosilba* spp. (Lonchaeidae). Eleven parasitoid species were collected: Braconidae - *Asobara anastrephae* (Muesebeck, 1958), *Doryctobracon areolatus* (Szépligeti, 1911), *D. fluminensis* (Costa Lima, 1938), *Opius bellus* Gahan, 1930 and *Utetes anastrephae* (Viereck, 1913); Figitidae - *Aganaspis nordlanderi* Wharton, 1998, *Lopheucoila anastrephae* (Rhöwer, 1919), *Odontosoma anastrephae* (Borgmeier, 1935) and *Trybliographa infuscata* Gallardo, Díaz & Uchôa-Fernandes, 2000 and, Pteromalidae - *Spalangia gemina* Boucek, 1963 and *S. endius* Walker, 1839. In all cases only one parasitoid emerged per puparium. *D. areolatus* was the most abundant and frequent parasitoid of fruit fly species, as was *L. anastrephae* in *Neosilba* spp. larvae. This is the first record of *A. nordlanderi* in the midwestern Brazilian region.

**KEYWORDS.** Biocontrol; fruit fly; Lonchaeids; parasitoids.

## INTRODUCTION

The flies of the Tephritidae and Lonchaeidae families (Tephritoidea) are the most important insect pests of fruit and vegetables grown in the world, specially in the Neotropical region. The control of such flies by parasitoids has been considered an important component of integrated pest management (IPM) and also one of the safest methods for man and environment. The biological control program of fruit flies around the world have used exotic parasitoids (HERNÁNDEZ-ORTIZ *et al.* 1994; SALLES 1996) which, besides enduring environmental adaptation difficulties (usually being species from another country or continent), may cause contrary effects

upon the native parasitoids. More attention must be given to the use of native parasitoids in such program, mainly when they are abundant and have a performance similar or only somewhat inferior to that found in effective exotic species.

There is little research on native parasitoids of frugivorous tephritoids in Brazil (LEONEL Jr. *et al.* 1995, 1996; CANAL *et al.* 1995; SALLES 1996; AGUIAR-MENEZES & MENEZES 1997), in spite of their enormous potential as biological control agents of fruit flies (LÓPEZ *et al.* 1999) and frugivorous lonchaeids. The aim of this study was to survey native parasitoids of fruit flies [*Anastrepha* spp., *Ceratitis capitata* (Wied.)] and frugivorous lonchaeids (*Neosilba* spp. and *Dasiops* sp.) obtained in the laboratory in cultivated and wild fruit species from cerrado

1. Laboratório de Insetos Frugívoros, Departamento de Ciências Biológicas, Universidade Federal de Mato Grosso do Sul. Caixa Postal 241, 79804-970 Dourados-MS, Brazil. E-mail:uchoa@ceud.ufms.br

2. Trainee biologist.

3. Departamento de Entomología, Fitopatología e Zoología Agrícola, ESALQ-USP. Caixa Postal 9, 13418-900 Piracicaba-SP, Brazil. E-mail:razucchi@esalq.usp.br

4. Facultad de Agronomía, Universidad del Tolima. Apartado Aéreo 546, Ibagué - Tolima, Colômbia.

5. Departamento Científico de Entomología, Museo de La Plata. Pasaje del Bosque, 1900 La Plata, Argentina.

vegetation of the State of Mato Grosso do Sul (midwestern Brazil).

## MATERIAL AND METHODS

The fruits were collected from January 1993 to March 1997, from "Cerrado" vegetation of Anastácio ( $20^{\circ} 31' 36''$  S /  $55^{\circ} 50' 12''$  W, 170 m), Aquidauana ( $20^{\circ} 39' S$  /  $55^{\circ} 19' 50''$  W, 173 m), Rochedo ( $19^{\circ} 57' 30''$  S /  $54^{\circ} 53' 10''$  W, 398 m) and Terenos ( $20^{\circ} 26' 12''$  S /  $54^{\circ} 04' 54''$  W, 308 m) municipalities and brought to the laboratory. The parasitoids were recovered from frugivorous larvae of the families Tephritidae and Lonchaeidae. Each sample was placed in a plastic container, from which the 3<sup>rd</sup>-instar larvae exited and fell into plastic trays with water. Larvae were recovered every 12h by pouring the material through a sieve with a mesh of 1 mm diameter, so tephritids and lonchaeids maggots were kept separate in different containers, in order to pupate and allow the emergence of flies or their parasitoids (UCHÔA-FERNANDES & ZUCCHI 1999). Only *Spondias lutea* L., 1758 (Anacardiaceae) and *Terminalia catappa* L., 1758 (Combretaceae) data, from fruit collected in 1993, were obtained by the usual methodology that consisted of placing fruits over trays containing a fine layer of sterilized sand. In this case, the puparia were recovered through sand sifting.

The viability of 3<sup>rd</sup>-instar larvae (L3) and the percentage of emergence of frugivorous flies were calculated according to the following formula, adapted from NASCIMENTO *et al.* (1984):

$$\% \text{ V.L3} = \frac{\text{Number of emerged flies} \times 100}{\text{Total No. of L3} - \text{No. of emerged parasitoids}}$$

% V.L3 (Percentage of the viability of third instar larvae).

The combined larval parasitism rate (total parasitism) by hymenopterans was calculated using the formula:

$$\% \text{ C.P.} = \frac{\text{N.R.P.} \times 100}{\text{N.L3}}$$

% C.P. = The percentage of total parasitism rate;  
N.R.P. = Number of recovered parasitoids;  
N. L3 = The number of 3<sup>rd</sup>-instar larvae of frugivorous flies.

The voucher specimens of tephritids, frugivorous lonchaeids and their parasitoids were deposited in the Coleção Zoológica, Universidade Federal de Mato Grosso do Sul, Campo Grande (ZUFMS) and Departamento de Entomologia, Fitopatologia e Zoologia Agrícola, Escola Superior de Agricultura "Luiz de Queiroz", Universidade de São Paulo, Piracicaba (ESALQ). The herborized samples of the host plants were deposited in the Herbário Central, Universidade Federal de Mato Grosso do Sul, Campo Grande and in the Coleção de Botânica, Departamento de Biologia, Universidade de São Paulo, São Paulo.

## RESULTS AND DISCUSSION

From January 1993 to March 1997, 283 samples were collected, totaling 648.66 kg of biomass and 20,166 fruits of 14 plant species; 193 corresponded to *Citrus spp.* (Rutaceae) and 90 to other fruits. Some 11,298 mature larvae of Tephritidae were obtained, emerging as: 4,814 adults of *Anastrepha spp.*, 2,637 *Ceratitis capitata*, 225 braconids and 23 pteromalids. Also, 11,246 larvae of Lonchaeidae were recovered, giving one adult of *Dasiops sp.*, 7,677 *Neosilba spp.* and 279 Eucoilinae (Figitidae) parasitoids (Table I). In all cases only one parasitoid emerged per puparium.

### Parasitoid species composition (Table I)

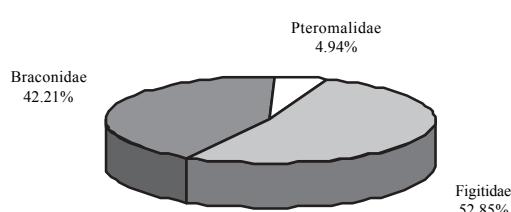
#### Braconidae

*Asobara anastrephae* (Muesebeck) (5 specimens), *Doryctobracon areolatus* (Szépligeti) (166 specimens), *D. fluminensis* (Costa Lima) (12 specimens), *Opius bellus* Gahan (17 specimens), *Utetes anastrephae* (Viereck) (23 specimens) and two unidentified braconids were obtained. All recovered braconids were associated with Tephritidae larvae (*Anastrepha spp.* and *C. capitata*). Generally, the braconid species found in this survey correspond to those already registered in other Brazilian regions (CANAL *et al.* 1995; LEONEL Jr *et al.* 1995, 1996; SALLES 1996; AGUIAR-MENEZES & MENEZES 1997). However, *D. fluminensis* which after its description by COSTA LIMA (1938) has not been collected since, was found in 1993 in Fazenda Ranchinho, municipality of Rochedo, Mato Grosso do Sul. In this survey, *D. fluminensis* was obtained from larvae feeding on cassava fruits, *Manihot esculenta* Crantz, 1766 (Euphorbiaceae), probably parasiting *A. montei* Costa Lima, 1934.

*Doryctobracon areolatus* was the most abundant braconid, achieving 31.38% of the parasitoid adults and 74.67% of the braconids (Table I). That species shows a worldwide distribution and has been considered an important native parasitoid species attacking the genus *Anastrepha* in neotropical countries. Surveys carried out in Brazil and in other countries have shown that *D. areolatus* is the most dominant, frequent and abundant species among the parasitoids of fruit flies (JIRÓN & MEXZON 1989; HERNÁNDEZ-ORTIZ *et al.* 1994; OVRUSKI & FIDALGO 1994; CANAL *et al.* 1995; LEONEL Jr. *et al.* 1995, 1996; SALLES 1996; AGUIAR-MENEZES & MENEZES 1997; LÓPEZ *et al.* 1999).

#### Figitidae

Four species in four genera of Eucoilinae were found in the present study: *Aganaspis nordlander* Wharton (16 specimens), *Lopheucoila anastrephae* (Rhower) (230 specimens), *Odontosema anastrephae* (Borgmeier) (6 specimens), *Trybliographa infuscata* Gallardo, Díaz & Uchôa-Fernandes (24 specimens) and three unidentified specimens. This is the first record of *A. nordlander* in the midwestern Brazilian region. *T. infuscata* was described from specimens

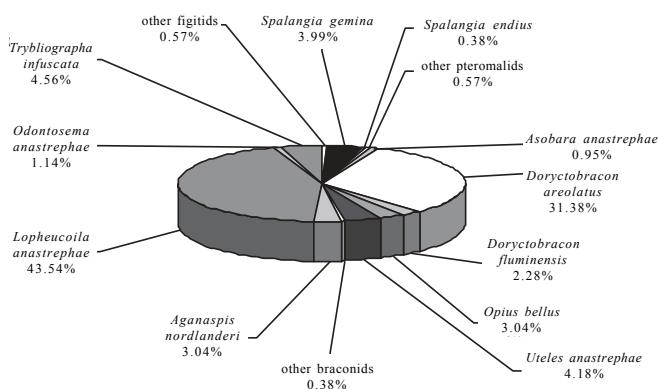


**Fig. 1.** Families of hymenopteran parasitoids ( $n= 527$ ) reared from frugivorous larvae in 14 host fruit species from the cerrado, Mato Grosso do Sul, Brazil (January 1993 to March 1997).

parasitising larvae of *Neosilba* sp. infesting oranges in Anastácio, Terenos, and from *Caryocar brasiliense* Camb. 1828, fruits ("pequi") (Caryocaraceae), collected in Aquidauana (GALLARDO *et al.* 2000). The adults of *Trybliographa* are larval endoparasitoids that emerged from *Neosilba* spp. puparia.

All eucoelines were associated with the Lonchaeidae larvae of the genus *Neosilba*. These solitary endoparasitoids lay their eggs in the larval stages of Cyclorrhaphous Diptera and emerge as adults from the puparium of their hosts. They are important natural enemies of phytophagous dipterans (WHARTON *et al.* 1998). Two species of Eucoilinae have been used as biological control agents of some economically important fruit flies: *Aganaspis daci* (Weld, 1951) (oriental species) and *A. pelleranoi* (Brèthes, 1924) (neotropical species) and, this latter species has been mass reared and released in orchards in Argentina and Mexico for the control of tephritid pests (WHARTON *et al.* 1998).

The eucoelines made up almost 53% of the total of the recovered parasitoids (Fig. 1). *L. anastrephae* was the species with the highest abundance and the most frequent in the samples, totaling 82.44% of the eucoelines collected (Fig. 2). This parasitoid species has high potential as a biological control agent for lonchaeid pests of the genus *Neosilba*, specially in *Citrus* orchards in Mato Grosso do Sul, where the parasitism of this species is constant, in spite of the frequent insecticide sprays.



**Fig. 2.** Percent of the species of hymenopterans ( $n= 527$ ) parasiting larvae of frugivorous flies from 14 host fruit species from the cerrado, Mato Grosso do Sul, Brazil (January 1993 to March 1997).

All the eucoelines registered in this study were associated with lonchaeids (Table I). Based on a survey in the region of Pelotas (Southern Brazil) related to native parasitoids of *A. fraterculus*, *Odontosema* sp. was listed among the parasitoids complex of the South American fruit fly (SALLES 1996). Although considering that the author applied the traditional methodology of fruit fly collecting, which kept all the insects together in the same container, probably *Odontosema* was parasiting Lonchaeidae larvae. As WHARTON *et al.* (1998) pointed out, if the larvae or pupae of frugivorous insects were kept together in the same rearing recipient until the emergence of adults, it will be possible to make a wrong association among the hosts and parasitoids.

In this survey, *O. anastrephae* emerged from larvae of *Neosilba* spp. infesting fruits of "pequi" (*C. brasiliense*). Although the eucoelines have been characterized as larval endoparasitoids of lonchaeids, other authors have found species of *Odontosema* and *Aganaspis* parasiting *C. capitata* and *Anastrepha* spp. larvae, as well as lonchaeids (WHARTON *et al.* 1981, 1998). WHARTON *et al.* (1998) pointed out that the records of the genus *Lopheucoila* and *Trybliographa* attacking tephritids in the New World need confirmation. In fact, our results show that *L. anastrephae* and *T. infuscata* were recovered only from *Neosilba* larvae (Lonchaeidae). Nevertheless, in contrast to the results obtained by WHARTON *et al.* (1998), who found a preference of *Odontosema* for tephritids, with only 2% of this species being recovered from frugivorous lonchaeids, we found that *O. anastrephae* was associated only with *Neosilba* spp. larvae. According to LÓPEZ *et al.* (1999), in Mexico, *A. pelleranoi* and *O. anastrephae* were obtained from pupae of *Anastrepha* spp. in guavas. Although these authors have associated those parasitoids with *Anastrepha* species, they did not identify previously the larvae that exited from fruits. So, it is possible that that parasitoid, in fact, was parasitising lonchaeids, which also infest guavas.

#### Pteromalidae

Two species were reared, *Spalangia gemina* Boucek (21 specimens) and *S. endius* Walker (2 specimens). The pteromalids seem to be associated with pupae of Tephritidae. The species of the genus *Spalangia* attack exclusively puparia of dipterans (V. A. Costa, personal communication). In this study, probably the parasitism occurred in the puparia collected in the field, because fruits (*S. lutea* and *T. catappa*) from which these parasitoids emerged, were kept when traditional methodology was still being used (in 1993), i.e., all larvae were kept together in the same container.

The average of parasitism in the 14 fruit species was 3.03%. When the parasitism was calculated individually for the 14 fruit species, the indices ranged from 0.07% to 14.37%. The highest level was found in tangerine araçá (*Psidium* sp.), sugar-apple and orange, in decreasing order. In Guatemala, the combined percentage of parasitism of *Anastrepha* spp. and *C. capitata* varied from 0.04 to 7.85%, in 14 sampled species of fruits (ESKAFI 1990).

The fruits of *C. reticulata* (tangerine) and of *Solanum*

**Table I.** Infestation indices of frugivorous larvae (Diptera: Tephritidae and Lonchaeidae) and their association with parasitoids (Hymenoptera) in 14 host plants sampled from the Cerrado of Mato Grosso do Sul, Brazil (January 1993 to March 1997)

Host Fruits n (kg)	Sites (samples)	Index Larvae/ fruit (n)	Index Larva/kg of fruit	3 <sup>rd</sup> instar recovered larvae (n)	Taxa of Frugivorous flies (n)	Emerged flies		Taxa of Parasitoids	Parasitoids and Parasitism	
						n	Viability (%)		(n)	% total
<b>Anacardiaceae</b>										
Hog plum, <i>Spondias lutea</i> L. 640 (7,43)	Aqid. (3)	2.86	246.84	1,834 Teph.	<b>Tephritidae</b> <i>Anastrepha</i> spp. <i>A. obliqua</i> (581)	1,145	63.54	<b>Braconidae</b> <i>Asobara anastrephae</i> (1) <i>Doryctobracon areolatus</i> (3) <i>Uteles anastrephae</i> (7) <b>Pteromalidae</b> <i>Spalangia gemina</i> (19) <i>S. endius</i> (2)	11	1.74
Red mombin, <i>S. purpurea</i> L. 484 (6.88)	Anast. Aqid. (8)	1.18	83.43	570 Teph.	<b>Tephritidae</b> <i>Anastrepha</i> spp. (226) <i>A. obliqua</i> (99) <i>A. sororcula</i> (2) <i>A. turpiniæ</i> (1) <i>Ceratitis capitata</i> (3)	229	40.17	<b>Braconidae</b> <i>A. anastrephae</i> (1) <i>U. anastrephae</i> (2)	3	0.52
				4 Lonch.	<b>Lonchaeidae</b> <i>Neosilba</i> sp. (1) <i>Dasiops</i> sp. (1)	2	50.00			
<b>Annonaceae</b>										
Sugar-apple, <i>Annona squamosa</i> L. 39 (5.70)	Anast. (4)	1.10	7.54	43 Lonch.	<b>Lonchaeidae</b> <i>Neosilba</i> sp.	41	95.35	<b>Figitidae</b> <i>Lopheucoila anastrephae</i> (2)	2	4.65
<b>Duguetiaceae</b>										
<i>Duguetia furfuracea</i> St. Hil. 204 (11.46)	Aqid. Terenos (8)	1.15	20.42	234 Lonch.	<b>Lonchaeidae</b> <i>Neosilba</i> sp.	200	85.47	<b>Figitidae</b> Non-identified	3	1.28
<b>Caryocaraceae</b>										
<i>Caryocar brasiliense</i> Camb. 562 (77.25)	Aqid. Rochedo (15)	7.06	51.35	1 Teph.	<b>Tephritidae</b> <i>A. sororcula</i> (1)	1	100.00			
				3,966 Lonch.	<b>Lonchaeidae</b> <i>Neosilba</i> sp.	2,913	73.45	<b>Figitidae</b> <i>L. anastrephae</i> (10) <i>Odontosema anastrephae</i> (6) <i>Trybliographa infuscata</i> (6) <b>Pteromalidae</b> <i>S. gemina</i> (2)	22	0.55
									2	0.07
<b>Combretaceae</b>										
Tropical almond, <i>Terminalia catappa</i> L. 443 (15.32)	Aqid. (10)	6.23	180.16	2,595 Teph.	<b>Tephritidae</b> <i>Anastrepha</i> spp. (3) <i>A. zenilde</i> (1) <i>C. capitata</i> (2,131)	2,134	82.23			
				165 Lonch.	<b>Lonchaeidae</b> <i>Neosilba</i> sp.	140	84.85			
<b>Euphorbiaceae</b>										
Cassava fruits, <i>Manihot esculenta</i> Crantz 1,760 (4.12)	Faz. Ranchinho, Rochedo (4)	0.36	154.85	638 Teph.	<b>Tephritidae</b> <i>Anastrepha</i> spp. <i>A. montei</i> (76) <i>A. pickeli</i> (1)	140	21.94	<b>Braconidae</b> <i>Doryctobracon fluminensis</i> (12)	12	1.88
<b>Mimosaceae</b>										
Inga, <i>Inga laurina</i> (Sw.) 1,545 (11.43)	Aqid. (6)	0.35	47.59	77 Teph.	<b>Tephritidae</b> <i>C. capitata</i>	50	64.93	<b>Braconidae</b> Non-identified (1)	1	0.18
				467 Lonch.	<b>Lonchaeidae</b> <i>Neosilba</i> spp.	199	42.61			
<b>Myrtaceae</b>										
Araça, <i>Psidium</i> sp. 1,313 (10.38)	Aqid. (5)	0.60	76.01	789 Teph.	<b>Tephritidae</b> <i>Anastrepha</i> spp. <i>A. sororcula</i> (167) <i>A. striata</i> (79) <i>A. fraterculus</i> (3)	517	65.53	<b>Braconidae</b> <i>D. areolatus</i> (63) <i>Opium bellus</i> (4)	67	8.49

**Table I.** Cont.

Host Fruits n (kg)	Sites (samples)	Index Larvae/ fruit (n)	Index Larva/kg of fruit	3 <sup>rd</sup> instar recovered larvae (n)	Taxa of Frugivorous flies (n)	Emerged flies		Taxa of Parasitoids	Parasitoids and Parasitism	
						n	Viability (%)		(n)	% total
Guava, <i>Psidium guajava</i> L. 1,728 (89.78)	Anast. Aqid. Terenos (13)	2.53	48.62	3,993 Tephr.	<b>Tephritidae</b> <i>Anastrepha</i> spp. (2,494) <i>A. sororcula</i> (994) <i>A. turpiniæ</i> (161) <i>A. striata</i> (50) <i>A. obliqua</i> (26) <i>A. zenildae</i> (19) <i>A. fraterculus</i> (9)  <i>C. capitata</i> (382)	2,876	72.02	<b>Braconidae</b> <i>D. areolatus</i> (93) <i>U. anastrephae</i> (12) <i>O. bellus</i> (6) <i>A. anastrephae</i> (3) Non-identified (1)	115	2.75
				372 Lonch.	<b>Lonchaeidae</b> <i>Neosilba</i> spp.	292	78.49	<b>Figitidae</b> <i>L. anastrephae</i> (5)	5	
Guavira, <i>Campomanesia sessiflora</i> (Berg.) 8,179 (19.66)	Anast. Aqid. Rochedo Terenos (9)	0.10	42.52	712 Tephr.	<b>Tephritidae</b> <i>Anastrepha</i> spp. (286) <i>A. sororcula</i> (141) <i>A. zenildae</i> (4) <i>A. obliqua</i> (1) <i>C. capitata</i> (5)	291	40.87	<b>Braconidae</b> <i>D. areolatus</i> (7) <i>O. bellus</i> (7) <i>U. anastrephae</i> (2)	16	1.91
				112 Lonch.	<b>Lonchaeidae</b> <i>Neosilba</i> sp.	72	64.28			
<b>Rutaceae</b>										
Orange, <i>Citrus sinensis</i> L. 2,346 (357.26)	Anast. Aqid. Terenos (179)	2.43	15.95	89 Tephr.	<b>Tephritidae</b> <i>A. turpiniæ</i> (2) <i>C. capitata</i> (66)	68	76.40			
				5,611 Lonch.	<b>Lonchaeidae</b> <i>Neosilba</i> sp.	3,360	60.97	<b>Figitidae</b> <i>L. anastrephae</i> (164) <i>T. infuscata</i> (18) <i>A. nordlanderi</i> (12)	194	3.40
Tangerine, <i>Citrus reticulata</i> (L.) 134 (21.60)	Anast. Aqid. Terenos (14)	2.65	16.43	355 Lonch.	<b>Lonchaeidae</b> <i>Neosilba</i> sp.	213	60.00	<b>Figitidae</b> <i>L. anastrephae</i> (47) <i>A. nordlanderi</i> (4)	51	14.37
<b>Solanaceae</b>										
<i>Solanum viarum</i> Dun. 789 (10.39)	Aqid. (5)	0.37	27.81	289 Lonch.	<b>Lonchaeidae</b> <i>Neosilba</i> sp.	246	85.12	<b>Figitidae</b> <i>L. anastrephae</i> (2)	2	0.69
<b>TOTALS / MEANS</b>	4 sites (283)	2.07	72.82	11,298 Tephr. 11,246 Lonch	<i>Anastrepha</i> 4,814 <i>C. capitata</i> 2,637  <i>Neosilba</i> 7,677 <i>Dasiops</i> 1	7,451 Tephr.  7,678 Lonch.	44.83% T  55.76% L	<b>Braconidae</b>  <b>Pteromalidae</b>  <b>Figitidae</b>	527	3.03
14 plant species 20,166 fruits 648,66 kg of biomass										

Anast. (Anastácio); Aquid. (Aquadauana); Tephr. and T (Tephritidae) and Lonch. and L (Lonchaeidae).

*viarum* Dun. (Solanaceae) were infested exclusively by *Neosilba* spp. In oranges, 98% of adult tephritoids that emerged also belonged to this genus and all recovered parasitoids in these hosts were Figitidae (Eucoilinae). This suggests a specificity of this parasitoid subfamily to the lonchaeids. The economic importance of *Neosilba* species (as *Silba*) in *Citrus* and in other fruit trees has been pointed out in other Brazilian

regions (MALAVASI & MORGANTE 1980; RAGA *et al.* 1996, 1997).

The mean of 3<sup>rd</sup>-instar larvae viability in the 14 host fruit species was 44.83% for Tephritidae and 55.76% for Lonchaeidae. Considering the number of 3<sup>rd</sup>-instar larvae per kg of fruit the highest infestation levels were: *Spondias lutea*, *Terminalia catappa*, *Manihot esculenta*, *S. purpurea*, *Psidium* sp. and *Caryocar brasiliense*, respectively. These results show a trend

of higher infestation levels in lighter and smaller fruits (Table I), according to the same observations of MALAVASI & MORGANTE (1980).

The tephritids colonized 10 host species, and the lonchaeids 11 among the 14 fruit species sampled. Anacardiaceae, Myrtaceae and Euphorbiaceae (cassava fruit) were infested mainly by tephritids, in which the infesting larvae were parasitized by braconids.

*Terminalia catappa* fruits were mainly attacked by *C. capitata*. This host plant is native to the Malay Peninsula (LIQUIDO *et al.* 1991) and *C. capitata* has become successfully adapted to Tropical almond, in other Brazilian regions (RONCHI-TELES & SILVA 1996; SILVA *et al.* 1998).

Acknowledgements. To José Zorandir Nogueira (*in memoriam*) and his wife, Dina Cândida Fajardo, for their friendship and incentive in the collecting of fruits at their farm, Fazenda Ranchinho, Rochedo, Mato Grosso do Sul; Dr. Valmir A. Costa, Instituto Biológico, Campinas, São Paulo, for the identification of the Pteromalidae species; IBAMA-Mato Grosso do Sul (Jacob Ronaldo Kuffner) and MAARA-Mato Grosso do Sul (Celso Luiz Antoniallii) for the donation of two vehicles to the Universidade Federal de Mato Grosso do Sul for assisting the "Projeto Moscas Frugívoras" field work; Coordenadoria de Pesquisa, Pró-Reitoria de Pesquisa e Pós-Graduação, Universidade Federal de Mato Grosso do Sul for the constant support; Conselho Nacional de Desenvolvimento Científico e Tecnológico-CNPq for the grant (1992 to 1994, Process No. 301067-92-9) and Coordenação de Aperfeiçoamento de Pessoal de Nível Superior-CAPES for the scholarship to the first author (1995 to 1997); Ubirazilda Maria Resende, for helping in the identification of the plant species and Professor Elsbeth A. Flunker (WI, USA) for the final revision of the English version.

## REFERENCES

- AGUAR-MENEZES, E. & E. B. MENEZES. 1997. Natural occurrence of parasitoids of *Anastrepha* spp. Schiner, 1868 (Diptera: Tephritidae) in different host plants, in Itaguai (RJ), Brazil. **Biological Control** **8**:1-6.
- CANAL, N. A. D.; R. A. ZUCCHI; N. M. SILVA & S. SILVEIRA NETO. 1995. Análise faunística dos parasitóides (Hymenoptera, Braconidae) de *Anastrepha* spp. (Diptera, Tephritidae) em Manaus e Iranduba, Estado do Amazonas. **Acta Amazonica** **25**(3/4): 235-246.
- COSTA LIMA, A. da. 1938. Vespas parasitas de moscas de frutas (Hymenoptera: Braconidae). **O Campo** **9**: 69-72.
- ESKAFI, F. M. 1990. Parasitism of fruit flies *Ceratitis capitata* and *Anastrepha* spp. (Diptera: Tephritidae) in Guatemala. **Entomophaga** **35**(3): 355-362.
- GALLARDO, F. E.; N. B. DÍAZ & M. A. UCHÔA-FERNANDES. 2000. A new species of *Trybliographa* (Hymenoptera: Figitidae: Eucoilinae) from Brazil associated with fruit infesting Lonchaeidae (Diptera). **Revista de la Sociedad Entomológica Argentina** **59**(1-4):21-24.
- HERNÁNDEZ-ORTIZ, V.; R. PÉREZ-ALONSO & R. A. WHARTON. 1994. Native parasitoids associated with the genus *Anastrepha* (Dip.: Tephritidae) in Los Tuxtlas, VeraCruz, Mexico. **Entomophaga** **39**(2): 171-178.
- JIRÓN, F. L. & R. G. MEXZON. 1989. Parasitoid hymenopterans of Costa Rica: geographical distribution of the species associated with fruit flies (Diptera: Tephritidae). **Entomophaga** **34**(1): 53-60.
- LEONEL JR., F. L.; R. A. ZUCCHI & R. A. WHARTON. 1995. Distribution and Tephritis hosts (Diptera) of braconid parasitoids (Hymenoptera) in Brazil. **International Journal of Pest Management** **41**(4): 208-213.
- LEONEL JR., F. L.; R. A. ZUCCHI & N. A. D. CANAL. 1996. Parasitismo de moscas-das-frutas (Diptera: Tephritidae) por Braconidae (Hymenoptera) em duas localidades do Estado de São Paulo. **Anais da Sociedade Entomológica do Brasil** **25**(2): 199-206.
- LIQUIDO, N. J.; L. A. SHINODA & R. T. CUNNINGHAM. 1991. Host plants of the mediterranean fruit fly (Diptera: Tephritidae): an annotated world review. **Entomological Society of América, Miscelaneous Publications** **77**, 52 p.
- LÓPEZ, M.; M. ALUJA & J. SIVINSKI. 1999. Hymenopterous larval-pupal and pupal parasitoids of *Anastrepha* flies (Diptera: Tephritidae) in Mexico. **Biological Control** **15**: 119-129.
- MALAVASI, A. & J. S. MORGANTE. 1980. Biologia de "moscas-das-frutas" (Diptera, Tephritidae). II: Índices de infestação em diferentes hospedeiros e localidades. **Revista Brasileira de Biologia** **40**(1): 17-24.
- NASCIMENTO, A. S.; A. L. M. MESQUITA & R. A. ZUCCHI. 1984. Parasitism of pupae of *Anastrepha* spp. (Dip., Tephritidae) by *Doryctobracon areolatus* (Szépligeti, 1911) (Hym., Braconidae) in *Citrus* and tropical fruits. **Anais da Academia de Ciências do Estado de São Paulo** **2**: 239-246.
- OVRUSKI, S. M. & P. FIDALGO. 1994. Use of parasitoids (Hym.) in the control of fruit flies (Dip.: Tephritidae) in Argentina: bibliographic review (1937-1991). **IOBC / WPRS Bulletin** **17**(6): 84-92.
- RAGA, A.; M. F. SOUZA FILHO; V. ARTHUR & A. L. M. MARTINS. 1996. Avaliação da infestação de moscas-das-frutas em variedades de café (*Coffea* spp.). **Arquivos do Instituto Biológico** **63**: 59-63.
- RAGA, A.; M. F. SOUZA FILHO; V. ARTHUR; M. E. SATO; L. A. MACHADO & A. BATISTA FILHO. 1997. Observações sobre a incidência de moscas-das-frutas (Diptera: Tephritidae) em frutos de laranja (*Citrus sinensis*). **Arquivos do Instituto Biológico** **64**: 125-129.
- RONCHI-TELES, B. & N. M. da SILVA. 1996. Primeiro registro de ocorrência da mosca-do-mediterrâneo, *Ceratitis capitata* (Wied.) (Diptera: Tephritidae) na Amazônia brasileira. **Anais da Sociedade Entomológica do Brasil** **25**(3): 569-570.
- SALLES, L. A. B. 1996. Parasitismo de *Anastrepha fraterculus* (Wied.) (Diptera: Tephritidae) por Hymenoptera, na região de Pelotas, RS. **Pesquisa Agropecuária Brasileira** **31**(11): 769-774.
- SILVA, J. G.; K. URAMOTO & A. MALAVASI. 1998. First report of *Ceratitis capitata* (Diptera: Tephritidae) in the Eastern Amazon, Pará, Brazil. **Florida Entomologist** **81**: 574-577.
- UCHÔA-FERNANDES, M. A. & R. A. ZUCCHI. 1999. Metodología de colecta de Tephritidae y Lonchaeidae (Diptera, Tephritoidea) y sus parasitoídes (Hymenoptera). **Anais da Sociedade Entomológica do Brasil** **28**(4): 601-610.
- WHARTON, R. A.; F. E. GILSTRAP; R. H. RHODE; M. FISCHEL & W. G. HART. 1981. Hymenopterous egg-pupal and larval-pupal parasitoids of *Ceratitis capitata* and *Anastrepha* spp. [Dip.: Tephritidae] in Costa Rica. **Entomophaga** **26**(3): 285-290.
- WHARTON, R. A.; S. M. OVRUSKI & F. E. GILSTRAP. 1998. Neotropical Eucoilidae (Cynipoidea) associated with fruit-infesting Tephritidae, with new records from Argentina, Bolivia and Costa Rica. **Journal of Hymenoptera Research** **7**(1): 102-115.

Received in 08.V.2002; accepted in 15.I.2003