Accelerated aging of ipê seeds under controlled conditions of storage¹

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ABSTRACT - This research was aimed at studying effects of storage and accelerated aging on germination and profile of storage proteins in *Handroanthus albus* seeds. These were stored into a cold chamber (± 8 °C; RH ± 40%) and after periods of 0, 3, 6, 9, and 12 months of storage, were subjected to accelerated aging for 0, 24, 48, 72, and 96 hours. Relationships between germination and proteins profile were assessed. Germination test was performed at 25 °C, under constant light. For protein extraction, 125 mg of seeds were macerated in 2 mL of extraction buffer (1M Tris-HCl; pH 8.8) and applied to SDS-PAGE polyacrylamide gel at 80 V .15 h⁻¹. Twelve month storage, combined with 72 hours accelerated aging have increased germination in approximately 65% when compared to non-aged seeds or to seeds with 24 h of accelerated aging. Besides beneficial effects, degradation and synthesis of different proteins were observed. It was concluded that germination of *Handroanthus albus* seeds, when not subjected to accelerated aging, is favored by storage in cold chamber during three to six months, or from nine to 12 months when subjected to accelerated aging process. Storage proteins may be associated to those increases, and hence further studies are needed.

Index terms: yellow "ipê", protein, Tabebuia alba.

Envelhecimento acelerado de sementes de ipê em condições controladas de armazenamento

RESUMO - Objetivou-se estudar o efeito do armazenamento e do envelhecimento acelerado na germinação e no perfil das proteínas de reserva de sementes de *Handroanthus albus*. As sementes foram armazenadas em câmara fria e ao 0, 3, 6, 9 e 12 meses submeteu-se ao envelhecimento acelerado por 0, 24, 48, 72 e 96 h, e avaliou-se a germinação e o perfil proteico. O teste de germinação foi realizado sob luz constante a 25 °C. Para a extração das proteínas, utilizaram-se 125 mg de sementes maceradas com 2 mL do tampão de extração (1M tris HCl pH 8,8) e aplicadas em gel SDS-PAGE (3% e 15%) a 80 V .15 h⁻¹. O armazenamento por 12 meses associado a 72 h de envelhecimento acelerado aumentaram a germinação em aproximadamente 65% em relação às sementes não envelhecidas e envelhecidas por 24 h. Associado a este efeito benéfico, observou-se a degradação e síntese de diferentes proteínas. Conclui-se que a germinação de sementes de *Handroanthus albus* não envelhecidas artificialmente é favorecida pelo armazenamento em câmara fria por três ou seis meses, e por nove ou 12 meses, associado ao envelhecimento acelerado. As proteínas de reserva podem estar associadas a este acréscimo e necessitam ser mais bem estudadas.

Termos para indexação: ipê-amarelo, proteínas, Tabebuia alba.

Introduction

The tree of clustered blossoming commonly called

[&]quot;Yellow Ipê" [Handroanthus albus (Cham.) Mattos; Syn: Tabebuia alba (Cham.) Sandwith], popularly also known by its other common names "Golden Ipê" and "Ipê From

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the Mountain" is a native Brazilian tree, which occurs in the states of Rio de Janeiro and Minas Gerais and spreads until the state of Rio Grande do Sul (Grose and Olmstead, 2007; Lorenzi, 2002). Due to its exuberant yellow blooming is widely used in landscaping, particularly for ornamentation of sidewalks of streets and avenues; and is also used for recomposing ciliary woods in areas free of inundations (Carvalho, 2003).

To preserve the species and meet the needs of reforestation programs and urban landscaping, seed conservation becomes a necessity. On seed storage, however, it was found a wide variation on physiological quality of seeds in the genus *Handroanthus* (Carvalho et al., 1976; Kano et al, 1978; Maeda and Matthes, 1984; Figliolia, 1988; Figliolia et al., 1988; Cunha et al., 1992; Kageyama et al., 1992; Mello and Eira, 1995), making difficult the establishment of cultivation techniques for recomposition of degraded areas and seed conservation in germplasm banks (Cabral et al., 2003; Pinto et al., 1986).

In a general way, research works related to seed deterioration during storage stumbles on limitations in the time necessary on performing these studies. One of the technical procedures used to allow for deterioration studies is by artificially aging seeds.

The accelerated aging test consists on subjecting seeds to elevated temperature and relative humidity (42 °C; 100% RH), simulating normal storage conditions, but with an increased speed of deterioration, thus allowing the monitoring of processes involved. Such technique has been used for studying changes on the biochemical, cytological, and physiological conditions of seeds during deterioration process (Jeng and Sung, 1994; Khan et al., 1996; Spinola et al., 2000) as compared to the natural aging process (Ganguli and SenMandi, 1990; Aiazzi et al., 1996; Camargo et al., 2000; Machado-Neto et al., 2001).

After harvest, both during storage or after accelerated aging, seeds may present physiological changes such as formation of chemical compounds and modifications on proteins stored in the seed (Dell'Aquila and Spada, 1994; Bettey and FinchSavage, 1998; Dell'Aquila and Di Turi, 1999; Korotaeva et al., 2001; Al-Niemi and Stout, 2002; Gashaw and Michelsen, 2002; Sun et al., 2002) what can favor the aging process (Shibata et al., 2011). These accumulated reserves are responsible for the vital functions of seeds, besides directly affecting their potential of storage (Marcos-Filho, 2005).

Therefore, the objective of this research work was to quantify effects of storage periods and of accelerated aging time on germination and on reserve proteins profile in seeds of *Handroanthus albus* aiming at improving the use and conservation of seeds of this species.

Material and Methods

The experimental work was carried out at the Laboratory for Analyses of Genetics and Seeds, State University of Santa Catarina, Campus of Lages, State of Santa Catarina, Southern Brazil. For that, *H. albus* seedpods were collected from three different mother-plants at the beginning of dehiscence, with the aid of a pruning shears. The extraction and processing of seeds were manually performed, discarding the visually damaged seeds.

The so processed seeds were stored into transparent polyethylene bags (22 cm x 32 cm in dimension, and 20 μ m thick), which were then placed into a cold chamber (\pm 8 °C; RH \pm 40%) on top of cardboard boxes. At each three months storage period (0, 3, 6, 9, and 12 months) the percentage of germination and the total proteins profile of seeds subjected or not to accelerated aging were assessed.

For performing the test of accelerated aging, after homogenization of samples, 200 seeds of each sample were evenly distributed, on a single layer, on top of nylon screens (5 mm mesh), placed on the upper part of gerbox type germination boxes containing 40 mL of distilled water at the bottom. A BOD type seed germinator, adjusted to 42 °C and 100% RH, was used for aging seeds during times of 0 (control), 24, 48, 72, and 96 h.

Assessments of moisture content were performed by the oven method at 105 ± 3 °C, for 24 h, (Brasil, 2009) by using two replications of 1 g seeds each.

Physiological quality of seeds was assessed by the germination test, using a completely randomized experimental design, with the treatments arranged into a 5 x 5 factorial scheme [5 storage periods (0, 3, 6, 9, and 12 months) x 5 incubation periods (0, 24, 48, 72, and 96 h)], with four replications of 25 seeds each, for each set of factors (storage period x incubation time). Seeds were evenly distributed on top of sheets of germination paper (Germitest®), moistened with sterile distilled water, in the ratio of 3 mL of water per 1 g of dry paper. The set (seeds + paper) were made into rolls and then placed into a BOD type seed germinator, at 25 °C, under constant light. The first count of germination was performed at the 14th day after starting the test and the final counting was performed at the 28th day (Silva et al., 2011), by assessing normal and abnormal seedling and non-germinated seeds, according to Brasil (2009).

Data were transformed into arc sine $\sqrt{x/100}$ and treatment means were compared by the Tukey test, at 5% probability.

For electrophoretic analysis of proteins, 125 mg of seeds for each aging time was used. For that, seeds were macerated with 2 mL of extraction buffer containing 1M tris-HCL, pH 8.8, for 10 min. and afterwards centrifuged at 14000 rpm, for 20 min., at 4 °C, according to methodological standardization proposed by Shibata et al.(2011). The supernatant was poured into micro-tubes and pellet was discarded. For applying into the gel, into the supernatant were added 25 μ L of proteinic extract + 25 μ L of sample buffer (5 mM glycerol, 1.73 mM sodium dodecyl sulfate (SDS), 1.54 mM tris-HCl, pH 6.8, 0.036 mM bromophenol blue, and 0.15 mM β-mercapto-ethanol). To make up the volume to 25 mL, distilled water was added, and the solution was then placed into water-bath with boiling water, for 5 min. A 20 µg sample of protein was applied to polyacrylamide gel SDS-PAGE at 15% (separator gel) and at 3% (concentrator gel) according to Laemmli (1970), and in agreement with changes performed by Ma and Bliss (1978). The electrophoretic system containing the samples were placed into a buffer solution with pH of 8.3 (50 mM Tris, 0.384 M Glycine, 5mM ethylenediaminetetraacetate (EDTA), pH 8.8 and 0.25% SDS). The electrophoretic run was performed at 80 V, for 15 hours. Afterwards, the gels were stained with 0.1% Coomassie blue, according to Alfenas (1998), for 12 h and discolored with a solution of 10% acetic acid, 45% methanol, and 45% distilled water.

The analysis of protein fractions bands was qualitative, where the presence or absence of bands and the increases or decreases of their intensity were visually identified for each different seed quality assessment methods used.

Results and Discussion

At beginning of experiment, seeds of *H. albus* had moisture content of 7.9%, which remained constant during the following months.

Handroanthus albus seed germination was favored by storage periods of three to six months with values of 88% and 84%, respectively (Figure 1). Percent germination of freshly harvested seeds was 72% and had an increase during the six initial months of storage, but from this point on a gradual reduction on germination was observed, reaching 18% at 12 months. Changes on percent of germination have already been found by other authors for different species within the same genus. Kano et al., (1978)

have observed that the germination of seed of the Golden Trumpet Tree *Tabebuia chrysotricha* (Mart. Ex DC.) after 15 days storage under dry chamber conditions, has decreased percentage of germination from 78% to 63% in 30 days, and again increased to 85% in 63 days. For seeds of the Purple Trumpet Tree [*Tebebuia impetiginosa* (Mart. Ex DC) Stanley], Mello and Eira (1995) have observed a 15% increase on germination of seeds stored during 18 months at -29 °C.

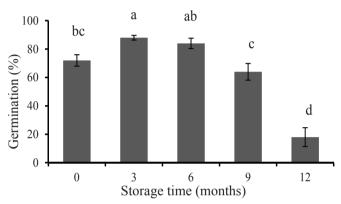


Figure 1. Percent germination of seeds of *Handroanthus albus* as a function of storage periods under cold chamber conditions (±8 °C; RH±40%) during 0 (control), 3, 6, 9, and 12 months. Letters on top of columns are referent to comparison of treatment means by Tukey test, at 5% probability. Columns topped by the same letter do not differ statistically between each other.

Among factors causing changes on percent germination during storage are: initial quality of seeds; maturation degree at harvest moment; presence or absence of microorganisms; environmental conditions, and packages used (Carvalho and Nakagawa, 2000; Marcos-Filho, 2005). Other factors causing changes on germination are the phenolic compounds, which are present in the seed tegument and can act as germination inhibitors (Bewley e Black, 1994), or the presence of proteins during storage that can be expressed under stress conditions, such as high temperatures (Vierling, 1991).

Based on changes of percent of germination during the storage process, biochemical changes on proteins profile and their close correlations with the increases or decreases of seed germination were analyzed. For the results on profile of proteins (Figure 2), it was found a 60 kDa band for storage periods of six and nine months that coincided with the decrease of 20% on germination percentage. For

the remaining bands, the general profile of proteins did not differ statistically among storage periods, with emphasis in the high intensity of proteins with 50.6 kDa, 40.6 kDa, and 27.8 kDa. Such abundance on proteins profile was a particular characteristic of the assessments performed within this study, since only in a small number of studies this correlation is done. Some research works have already mentioned the presence of 10 kDa proteins in seeds freshly harvested or after storage (Silva et al., 2011) as well during their development, as reported by Carvalho et al. (2008) in a study carried out with *Tabebuia serratifolia*.

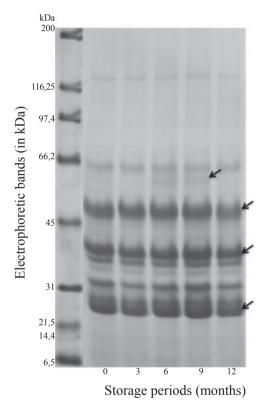


Figure 2. Electrophoretic profile of reserve proteins of seeds of *Handroanthus albus* as a function of storage periods during 0 (control), 3. 6, 9, and 12 months under cold chamber conditions (±8 °C; RH±40%).

To better understand changes during storage, the accelerated aging test was performed for each storage period, aiming at a more detailed assessment of changes of physiological and biochemical characteristics of seeds based on the profile of reserve proteins.

The accelerated aging process did not induce changes on germination of freshly harvested seeds or on those seeds stored for three months (Figure 1A and 1B). After storage periods of nine and 12 months, however, the accelerated aging favored seed germination (Figure 1E).

Increases on germination were also observed for seed of the noble wood tree *Cedrella fissilis* Vellozo after 72 h of accelerated aging (Borges et al., 1990). Likewise, Schmidt (2000) has stressed that the high moisture content during accelerated aging can activate the mechanism of cell repair, what was also observed for seeds of peanuts (*Arachis hypogacea* L.)

For all storage periods, data on artificial aging of 72 and 96 h showed a percent of germination around 80%, thus demonstrating that accelerated aging favored seed germination when compared to seeds stored for 12 months or non-aged seeds (18%). The remaining periods of accelerated aging for seeds subjected to distinct storage periods have presented irregular behavior, with increases and decreases depending on the period of storage assessed (Figure 3).

The storage period of 12 months, when associated to the accelerated aging of 72 h induced a 65% increase on percent of germination, in relation to non-aged seeds or seeds aged for a 24 h period. These results show the dependence on artificial aging of seeds during a 72 h period in order to achieve such increase on germination percentage when these seeds are stored for long periods (Figure 3E).

Because of high influence of artificial aging on germination rate of seeds stored for 12 months, the profile of proteins was assessed within the different aging periods.

An increase on intensity of bands of 30.2 kDa and 27.8 kDa was observed, coinciding with an increase on percent of germination (34%, between 48 and 72 h of aging time) and a decrease on intensity for the bands of 39 kDa and 32 kDa, as a function of the increased aging time (from 24 h until 96 h) (Figure 4). A band of 19 kDa was also detected. In a general way, this low molecular weight protein may correspond to a particular group of proteins denominated "thermal shock proteins" that are directly related to thermic stresses (Waters et al., 1996), such as the high temperature (42 °C) to which seeds were subjected during the aging process.

In studies with seeds of Crimson clover (*Trifolium incarnatum* L.) and Perennial Ryegrass (*Lolium perenne* L.), Ching and Schoolcraft (1968) reported reduction on protein content only after seed viability loss and that for both species the protein loss was dependent on the severity of storage conditions. Pereira (1980), however, did not find a defined trend on the behavior of proteins during storage

of seeds of the hardwood Rubber Tree [*Hevea brasiliensis* (Willd.) Muell.-Arg.], and observed an intermittent variation among treatments.

The variation found on protein profile within this study may be a response of seeds to extreme temperatures. Under such conditions, the thermal shock protein is produced. This protein is synthesized in temperatures higher than 30 °C and contributes for the maintenance of the integrity of cellular membranes (Marcos-Filho, 2005). Such protein has

considerable heterogeneity in isoelectric points, molecular weight, stability, and expression level (Lee et al., 1994). In order to correlate such proteins with results herein obtained, further detailed studies are necessary. For example: analysis of protein profiles by 2-D electrophoresis; immunolocalization of specific antibodies at reaction level; and gene expression, aiming at verifying specific involvements of this group of proteins with characteristics of germination of *H. albus* seeds.

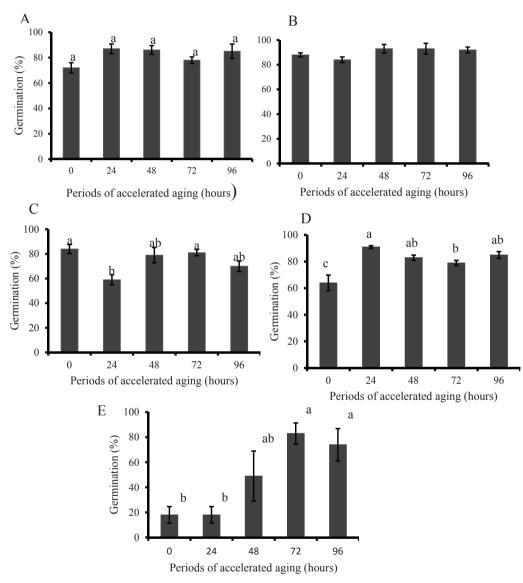
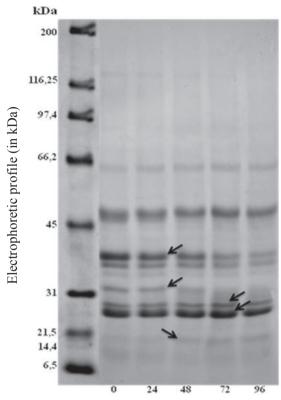


Figure 3. Percentage of germination of freshly harvested seeds of *Handroanthus albus* (A) and after storage periods of 3 (B), 6 (C), 9 (D), and 12 (E) months under cold chamber conditions (±8 °C; RH±40%), in function of accelerated aging periods of 0 (control), 24, 48, 72, and 96 hours. Letters on top of columns are referent to comparison of treatment means by Tukey test, at 5% probability. Columns topped by the same letter do not differ statistically between each other.



Periods of accelerated aging (hours)

Figure 4. Electrophoretic profile of reserve proteins of seeds of *Handroanthus albus* stored during 12 months under cold chamber conditions (±8 °C; RH ± 40%) and then subjected to different periods of accelerated aging of 0 (control), 24, 48, 72, and 96 hours. The electrophoretic run was performed in polyacrylamide gel SDS-PAGE (at 80 V; for 15 h).

Conclusions

The germination of not artificially aged seeds of *Handroanthus albus* is favored by storage in cold chamber during three or six months.

Storage of seeds of *Handroanthus albus* during nine or 12 months when associated to accelerated aging of 24 or 48 hours, respectively, increases germination rate. The proteins of 39 kDa, 32 kDa, 30.2 kDa, 27.82 kDa, and 19 kDa may be associated to these increases and need to be better studied for this species.

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