

Larvicidal activity of oil-resin fractions from the Brazilian medicinal plant *Copaifera reticulata* Ducke (Leguminosae-Caesalpinoideae) against *Aedes aegypti* (Diptera, Culicidae)

Atividade larvívica das frações do óleo-resina da planta medicinal brasileira *Copaifera reticulata* Ducke (Leguminosae-Caesalpinoideae) sobre o *Aedes aegypti* (Diptera, Culicidae)

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ABSTRACT

Oil-resin fractions from *Copaifera reticulata* *Ducke* (Leguminosae-Caesalpinoideae) were evaluated for larvicidal activity on third larval instars of *Aedes aegypti*, in searching for alternative control methods for this mosquito. The bioactive fractions were chemically monitored by thin-layer chromatography, ¹H and ¹³C nuclear magnetic resonance and mass spectrometry. Bioassays were performed using five repetitions, at a temperature of 28 ± 1 °C, relative humidity of 80 ± 5% and light and dark cycles of 12h. Mortality was indicated by darkening of the cephalic capsule after 24h of exposure of the larvae to the solutions. The most active fractions were CRM₁₋₄ (sesquiterpenes) and CRM₅₋₇ (labdane diterpenes), which showed LC₅₀ values of 0.2 and 0.8ppm, respectively.

Key-words: *Copaifera reticulata*. *Aedes aegypti*. Terpenoids. Larvicidal activity.

RESUMO

A atividade larvívica das frações do óleo-resina de *Copaifera reticulata* *Ducke* (Leguminosae-Caesalpinoideae) foi avaliada em larvas de 3º estágio de *Aedes aegypti*, na busca de alternativas para o controle desse mosquito. As frações bioativas foram monitoradas quimicamente através de cromatografia de camada delgada, analisada por ressonância magnética nuclear de hidrogênio (¹H e ¹³C) e espectrometria de massas. Os bioensaios foram realizados à temperatura de 28±1°C, 80±5% de umidade relativa e fotofase de 12h, com cinco repetições. A mortalidade foi determinada através do escurecimento da cápsula cefálica, após 24h de exposição das larvas às soluções. As frações mais ativas foram CRM₁₋₄ (sesquiterpenos) e CRM₅₋₇ (diterpeno labdano), que mostraram os valores de CL₅₀ de 0,2 e 0,8ppm, respectivamente.

Palavras-chaves: *Copaifera reticulata*. *Aedes aegypti*. Terpenoides. Atividade larvívica.

Aedes aegypti (Lin), a mosquito that is disseminated around the world, has medical importance because it is a vector for dengue in Asia and for dengue and yellow fever in Africa and the Americas¹⁶. According to the World Health Organization²⁷, around 2.5 billion people are exposed to dengue transmission risk, in the urban environment of the cosmopolitan region. This problem could increase through adaptation of the mosquito to

polluted water¹⁹, which could make control a big challenge in the near future.

There is still no vaccine available for dengue prevention. Dengue control has been limited to combating the vector using synthetic and biological insecticides. However, due to continuous use, the vector has become resistant to some chemical products^{2 3 9 11 12}. This factor, allied to the growth of environmental

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concerns about the ecological damage produced by synthetic insecticides, has motivated scientific research towards finding active products of botanical origin, and a number of species have been investigated with this purpose^{5 6 7 13 14 15 17 20 21 23}.

Our own screening of plant extracts for their larvicidal activity has revealed that *Magonia pubescens* St Hil (Sapindaceae) and *Copaifera reticulata* (Leguminosae) were the most active extracts^{20 22}. *Copaifera reticulata* is a plant found in the tropical region of Latin America, with widespread distribution in the Brazilian Amazon forest, particularly in the States of Pará and Amazonas⁸. It is also known as *copaibeira* and *pau-d'oleo*, and is one of the principal sources of copaiba oil-resin, which is widely exported from Brazil. Local populations in this region have been using this oil-resin as a primary therapeutic source for several ethno-pharmacological indications^{1 4 8 26}.

A study on *Copaifera reticulata* would therefore be expected to have great potential for finding bioactive compounds, since its extract has been shown to present good larvicidal activity. The present paper describes the activity and chemical composition of fractions derived from *Copaifera reticulata* against third-instar *Aedes aegypti* larvae.

MATERIALS AND METHODS

Plant material. Crude oil from *Copaifera reticulata* was obtained as an exudate from direct perforation of its trunk, in Jacundá, Pará, Brazil (October 2001). The trunk was perforated at 70cm from the ground. The oil-resin exudate was collected, filtered through nylon cloth, and stored in amber flasks until use. The borehole in the plant was sealed with clay.

Extraction and separation. An oil sample (186.5g) was subjected to liquid-liquid partition with *n*-hexane and methanol (1:1), thus resulting in two extracts: hexane soluble (162.3g) and methanol soluble (22.8g). These extracts were subsequently chromatographed on silica gel CC (70-230 mesh), using *n*-hexane, dichloromethane and methanol, by means of the gradient polarity elution technique. Fractions from the hexane extract were named CRH₁ to CRH₈, and those from the methanol extract were named CRM₁ to CRM₈. The active fractions were re-chromatographed on a silica flash CC (230-400 mesh), using an *n*-hexane, ethyl acetate and methanol gradient. The fractions were analyzed by thin layer chromatography (TLC) using sulfuric acid solution of vanillin as the revealing reagent. Fractions with similar TLC patterns (retention factor) were combined for subsequent bioassays.

Analyses of bioactive subfractions. The bioactive subfractions were monitored by TLC and analyzed by ¹H nuclear magnetic resonance (¹H NMR) in a Bruker ARX-200 spectrometer, using deuterated chloroform (CDCl₃) as the solvent and tetramethylsilane (TMS) as the internal standard. The non-polar fractions (from the hexane extract) were analyzed by gas chromatography coupled to mass spectrometry (GC-MS), using electron impact.

Bioassays. For the larvicidal activity assays, third-instar larvae of *Aedes aegypti* were used. These were obtained from a cyclic colony that has been maintained for more than 10 years

at the Institute of Tropical Pathology and Public Health, Federal University of Goiás, at a temperature of 28 ± 1°C, relative humidity of 80 ± 5% and light and dark cycles of 12h¹⁸. Each fraction containing test compounds was weighed, dissolved previously in dimethylsulphoxide (DMSO), and the volume was made up with distilled water to obtain a stock solution (500ppm). The non-lethal concentration of DMSO that was used to prepare the solutions was determined previously, in experiments with *Aedes aegypti* larvae, to be 16.2% (v/v). From the stock solution, a series of dilutions was prepared in order to obtain the desired concentrations. The bioassays were carried out as five replicates in a climate-controlled environment similar to that of the colony. Twenty third-instar larvae of *Aedes aegypti* were placed in a 25ml test solution. Control assays were conducted using the same number of larvae in a DMSO-distilled water solution. The larva mortality after 24h was recorded. Total absence of larval movement as well as dark body color and cephalic capsule were used as indicative of death. The lethal concentrations and their respective confidence intervals were calculated by data interpolation, by means of Probit analyses using the Statistical Analyses System (SAEG).

RESULTS AND DISCUSSION

Liquid-liquid partition of *Copaifera reticulata* oil-resin (186.5g) resulted in 162.3g of hexane extract and 22.8g of methanol extract, which both showed larvicidal activity against *Aedes aegypti*. The bioassay-guided fractionation of these extracts produced eight fractions for each one. Two hexane (CRH₁, CRH₅) and two methanol (CRM₁, CRM₅) fractions showed the highest toxicity against *Aedes aegypti* larvae. The lethal concentrations (LC), with their respective confidence intervals, are shown in Table 1. There was no mortality in the control group.

The results showed that the two fractions CRH₁ (LC₅₀ 2.3ppm) and CRM₁ (LC₅₀ 0.8ppm) are more active than the oil-resin (LC₅₀ 8.9ppm) (Table 1). The two fractions CRH₅ (LC₅₀ 13.9ppm) and CRM₅ (LC₅₀ 10.5ppm) were slightly less active. Examination of these fractions by TLC and ¹H NMR showed their terpenoid nature and GC/MS analysis (NIST library) verified that they

Table 1 - Chemical constituents of the fractions obtained from *Copaifera reticulata* and their activity against third-instar larvae of *Aedes aegypti*, after 24h of exposure.

Fractions	Chemical class	LC ₅₀ (95% CI) ppm	LC ₉₀ (95% CI) ppm
Oil-resin		8.9 (6.9 – 10.8)	59.4 (41.9 – 102.2)
CRH ₁	monoterpenes	2.3 (0.9 – 3.4)	8.8 (7.3 – 11.5)
CRH ₄	sesquiterpenes	22.6 (22.5 – 26.4)	43.9 (40.2 – 49.1)
CRH ₅	sesquiterpenes	13.9 (11.2 – 15.3)	40.9 (34.5 – 52.4)
CRM ₁	mono-, di-, sesquiterpenes	0.8 (0.01 – 2.5)	8.1 (7.9 – 11.9)
CRM ₃	diterpenes	17.3 (15.3 – 19.1)	61.1 (50.4 – 80.3)
CRM ₄	diterpenes	15.5 (11.8 – 18.6)	93.1 (67.2 – 163.0)
CRM ₅	diterpenes	10.5 (9.1 – 11.7)	21.4 (18.6 – 26.5)

95% CI - confidence interval at 95% probability

LC₅₀ - lethal concentration required to kill 50% of the population exposed

LC₉₀ - lethal concentration required to kill 90% of the population exposed

ppm - parts per million

consisted of monoterpenes and sesquiterpenes. CRH₁, which is rich in monoterpenoids that are mainly from the pinane and para-menthane groups, was four times more active than the oil-resin. CRM₁, which was 11 times more active than the oil-resin, consisted mainly of sesquiterpenoids from the caryophyllene, bisabolane, cadinane and copaene groups. It was refractionated to give, among other subfractions, CRM₁₋₄ with an LC₅₀ of 0.2ppm and an NMR spectrum that confirmed the presence of sesquiterpenoids (Table 2). CRM₅ was rather less active than the oil-resin and, on further fractionation, gave the subfraction CRM₅₋₇ with an LC₅₀ of 0.8ppm, such that the TLC and NMR (¹H and ¹³C) data showed the presence of clerodane and labdane diterpenoids (Table 2).

Table 2 - Chemical constituents of the subfractions CRM₁ and CRM₅ from *Copaifera reticulata* and their activity against third-instar larvae of *Aedes aegypti*, after 24h of exposure.

Fractions	Chemical class	LC ₅₀ (95% CI) ppm	LC ₉₀ (95% CI) ppm
CRM ₁₋₂	monoterpenes	27.5 (12.1 – 41.9)	> 100
CRM ₁₋₃	monoterpenes	3.9 (2.1 – 5.5)	37.8 (26.6 – 69.4)
CRM ₁₋₄	sesquiterpenes	0.2 (0.1 – 1.2)	12.2 (6.1 – 22.2)
CRM ₁₋₅	sesquiterpenes	7.5 (4.9 – 15.7)	65.1 (59.4 – 85.2)
CRM ₁₋₆	sesquiterpenes	>100	>100
CRM ₁₋₇	sesquiterpenes	> 100	>100
CRM ₁₋₈	labdane diterpenes	> 100	> 100
CRM ₁₋₉	labdane diterpenes	1.3 (0.2 – 2.5)	>100
CRM ₅₋₇	labdane diterpenes	0.8 (0.1 – 1.9)	8.2 (6.5 – 11.3)
CRM ₅₋₈	labdane diterpenes	7.9 (7.1 – 8.4)	12.5 (11.3 – 15.1)
CRM ₅₋₉	furane labdane diterpenes	32.3 (30.4 – 34.2)	56.2 (51.8 – 62.1)
CRM ₅₋₁₀	furane labdane diterpenes	10.6 (9.7 – 11.5)	21.6 (19.4 – 25.2)
CRM ₅₋₁₁	clerodane diterpenes	6.2 (5.7 – 6.8)	11.1 (9.9 – 13.0)
CRM ₅₋₁₄	clerodane diterpenes	7.3 (5.1 – 9.4)	68.5 (48.4 – 117.2)

95% CI - confidence interval at 95% probability

LC₅₀ - Lethal concentration required to kill 50% of the population exposed

LC₉₀ - Lethal concentration required to kill 90% of the population exposed

ppm - parts per million

Summarizing these results, CRH₁ consisted mainly of monoterpenoids, CRM₁₋₄ consisted mainly of sesquiterpenoids and CRM₅₋₇ consisted of diterpenoids. These were, respectively, 4.44 and 13 times more active than the oil-resin.

Terpenoids are often reported as candidate insecticides that could be an effective alternative for insect control, with a lower impact on human health, household animals and the environment²⁵. The terpenoids reported from *Copaifera reticulata* oil-resin were much more active than the terpenoids reported in the literature. For example, the sesquiterpenes *E*-nerolidol, farnesol and nerolidol (the first of these isolated from *Myroxylon balsamum* and the other two obtained from commercial sources) showed LC₅₀ values of 6.0, 13.0 and 17.0ppm, respectively²³. The monoterpene isolated from oil produced by *Tagetes minuta* was active at concentrations above 40ppm¹⁰, and the diterpene isolated from *Melantheria albinervia* showed an LC₁₀₀ of 62.5ppm²⁴. Furthermore, a triterpene isolated from *Azadirachta indica* showed an LC₅₀ of 21ppm¹⁷. All of these concentrations were much higher than those observed for fractions of *C. reticulata* oil-resin.

The results described in the present paper suggest that some oil-resin fractions (CRM₁₋₄ and CRM₅₋₇) of *Copaifera reticulata* are comparatively more active against *Aedes aegypti* larvae. These results encourage the search for new natural larvicides. However, further investigations on larvicidal mechanisms of action and the effects of subfractions on non-target organisms and the environment in general, as well as formulations for improving larvicidal potency and stability, are needed for them to be used in practice as naturally occurring mosquito larval control agents.

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REFERENCES

- Basile AC, Sertie JAA, Freitas PCD, Zanini AC. Anti-inflammatory activity of oleoresin from Brazilian *Copaifera*. Journal of Ethnopharmacology 22:101-109, 1988.
- Braga IA, Lima JBP, Soares SS, Valle D. *Aedes aegypti* resistance to temephos during 2001 in several municipalities in the states of Rio de Janeiro, Sergipe and Alagoas, Brazil. Memórias do Instituto Oswaldo Cruz 99:99-203, 2004.
- Carvalho LAF, Silva IG. Atividade larvicida do temephos a 1% sobre o *Aedes aegypti* (Lin.,1762) em diferentes criadouros artificiais. Revista de Patologia Tropical 28:211-232, 1999.
- Cascon V, Gilbert B. Characterization of the chemical composition of oleoresins of *Copaifera guianensis* Desf., *Copaifera duckei* Dwyer and *Copaifera multijuga* Hayne. Phytochemistry 55:773-778, 2000.
- Cheng SS, Chang HT, Ghang ST, Tsai KH, Chen WJ. Bioactivity of selected plant essential oils against the yellow fever mosquito *Aedes aegypti* larvae. Bioresource Technology 89:99-102, 2003.
- Choochoe W, Kanjanapothi D, Panthong A, Taesotikul T, Jitpakdi A, Chaitong U, Pitasawat B. Larvicidal, adulticidal and repellents effects of *Kaempferia galanga*. Southeast Asian Journal Tropical Medicine Public Health 30:470-476, 1999.
- Ciccio G, Coussio J, Mongelli E. Insecticidal activity against *Aedes aegypti* larvae of some medicinal South American plants. Journal of Ethnopharmacology 72:185-189, 2000.
- Corrêa MP. Dicionário das Plantas Úteis do Brasil e das Exóticas Cultivadas. Instituto Brasileiro de Desenvolvimento Florestal, Ministério da Agricultura, Brasília, Volume V, 1984.
- Failloux AB, Ung A, Raymond M, Pasteur N. Insecticide susceptibility in mosquitoes (Diptera: Culicidae) from French Polynesia. Journal of Medical Entomology 31:639-644, 1994.
- Green MM, Singer JM, Sutherland DJ, Hibben CR. Larvicidal activity of *Tagetes minuta* (Marigold) toward *Aedes aegypti*. Journal of the American Mosquito Control Association 7:282-286, 1991.
- Lima JBP, Cunha MP, Silva RC, Galarido AK, Soares SS, Braga IA, Ramos RP, Valle D. Resistance of *Aedes aegypti* to organophosphates in several municipalities in the State of Rio de Janeiro and Espírito Santo, Brazil. American Journal of Tropical Medicine and Hygiene 68:329-333, 2003.
- Macoris ML, Andrighetti MTM, Takaku L, Glasser CM, Garbeloto VC, Bracco JE. Resistance of *Aedes aegypti* from the state of São Paulo, Brazil, to organophosphates insecticides. Memórias do Instituto Oswaldo Cruz 98:703-708, 2003.
- Monzon RB, Alviro JP, Luczon LL, Morales AS, Mutuc FE. Larvicidal potential of five Philippine plants against *Aedes aegypti* and *Culex quinquefasciatus*. Southeast Asian Journal of Tropical Medicine and Public Health 25:755-759, 1994.
- Park IK, Lee SG, Shin SC, Park JD, Ahn YJ. Larvicidal activity of isobutylamides identified in *Piper nigrum* fruits against three mosquito species. Journal of Agricultural and Food Chemistry 50:1866-1870, 2002.

15. Perich MJ, Wells C, Bertsch W, Tredway KE. Isolation of the insecticidal components of *Tagetes minuta* (Compositae) against mosquito larvae and adults. *Journal of the American Mosquito Control Association* 11:307-310, 1995.
16. Pinheiro FP, Corber SJ. Global situation of dengue and dengue haemorrhagic fever and its emergence in the Americas. *World Health Statistics Quarterly* 50:161-169, 1997.
17. Siddiqui BS, Afshan F, Ghiasuddin SF, Navqi SNH, Tariq RM. Two insecticidal tetranortriterpenoids from *Azadirachta indica*. *Phytochemistry* 53:371-376, 2000.
18. Silva HHG, Silva IG, Lira KS. Metodologia de criação, manutenção de adultos e estocagem de ovos de *Aedes aegypti* (Linnaeus, 1762) em laboratório. *Revista de Patologia Tropical* 27:51-63, 1998.
19. Silva HHG, Silva IG, Lira KS. Adaptação do *Aedes aegypti* em criadouros artificiais com água poluída. *Entomologia y Vectores* 6:383-391, 1999.
20. Silva HHG, Silva IG, Santos RMG, Rodrigues F^o E, Elias CN. Atividade larvicida de taninos isolados de *Magonia pubescens* St.Hil. (Sapindaceae) sobre o *Aedes aegypti* (Diptera, Culicidae). *Revista da Sociedade Brasileira de Medicina Tropical* 37:396-399, 2004.
21. Silva IG, Santos AH, Ferri PH, Alves RBN, Melo RL, Peixoto L, Silva HHG, Elias CN, Isac E, Lira KS, Camargo ME. Atividade larvicida do extrato bruto etanólico de *Magonia pubescens* St.Hil. (tingui-do-cerrado) sobre o *Aedes aegypti* (Lin.) em laboratório. *Revista de Patologia Tropical* 25:51-59, 1996.
22. Silva IG, Zanon VOM, Silva HHG. Larvicidal activity of *Copaifera reticulata* Ducke oil-resin against *Culex quinquefasciatus* Say (Diptera, Culicidae). *Neotropical Entomology* 32:729-732, 2003.
23. Simas NK, Lima EC, Conceição SR, Kuster RM, Oliveira Filho AM. Produtos naturais para o controle da transmissão da dengue - Atividade larvicida de *Myroxylon balsamum* (óleo vermelho) e de terpenóides e fenilpropanóides. *Química Nova* 27:46-49, 2004.
24. Slimestad R, Marston A, Mavi S, Hostettmann K. Larvicidal constituents of *Melantheria albinervia*. *Planta Medica* 61:562-563, 1995.
25. Viegas Jr C. Terpenes with insecticidal activity: an alternative to chemical control of insects. *Química Nova* 26:390-400, 2003.
26. Veiga Jr V, Pinto AC. O Gênero *Copaifera* L. *Química Nova* 25:273-286, 2002.
27. World Health Organization. Dengue/dengue haemorrhagic fever prevention and control. Regional Office for South-East Ásia 1-33, 2003.