

Triterpenes and flavonoids from the roots of *Mauritia flexuosa*

Hector H. F. Koolen,¹ Elzalina R. Soares,¹ Felipe M. A. da Silva,¹ Antonia Q. L. de Souza,² Edson Rodrigues Filho,³ Afonso D. L. de Souza^{*1}

¹Departamento de Química, Universidade Federal do Amazonas, Brazil,

²Escola Superior de Ciências da Saúde, Universidade do Estado do Amazonas, Brazil,

³Departamento de Química, Universidade Federal de São Carlos, Brazil.

Abstract: *Mauritia flexuosa* L. f., Arecaceae, is an endemic species of South America. This species was studied with the intent to isolate the constituents of its roots. After the fractionation of the *n*-hexane and methanolic extracts from the roots of *M. flexuosa*, six triterpenes were obtained: friedelin, taraxerone, lupenyl acetate, lupenone, betulin and betulinic acid, along with three flavonoids: rutin, quercitrin and quercetin. All the compounds were identified by analysis of NMR and MS data and comparison with the literature. All those compounds are been reported for the first time in *Mauritia*, and the chemosystematic significance of the flavonoids isolated in this genus is discussed.

Introduction

The Arecaceae family is one of the biggest vegetal families of the world and by its morphological aspects is the most characteristic of the tropical flora (Galotta & Boaventura, 2005). This family comprises 1500 species distributed in 200 genera (Uhl & Dransfield, 1987). The chemical composition of the Arecaceae plants includes: diterpenes, triterpenes and their methyl esters, steroids (including brassinosteroids), proanthocyanidines, flavonoids (derived from kaempferol, quercetin, tricetin and luteolin), saponins and rarely alkaloids (pyrimidinics) (Piozzi et al., 1981; Harborne et al., 1994; Holdsworth et al., 1997; El Dib et al., 2004). The species *Mauritia flexuosa* L. f., popularly named buriti or miriti, is a large palm tree distributed throughout the Amazon region (Passos & Mendonça, 2006). While this palm is found in several vegetal formations, it is achieved more commonly growing up in flooded areas forming typical homogeneous groups (Ruiz & Alencar, 2004). This species has high economic potential, due mainly to its fruits that have valuable oil for the cosmetic industries (França et al., 1999). The oil extracted from the buriti fruits is popularly used against burns and as a potent vermifuge (Passos & Mendonça, 2006), and those activities are attributed principally to the carotenoids and the ascorbic acid, principal compounds in the oil (França et al., 1999). The buriti fruit showed also

previously moderate antimicrobial activity against several bacteria (Silveira et al., 2005). No previous phytochemical studies were carried out with any species of the *Mauritia* genus.

Material and Methods

General

Melting points were measured on Buchi 545 B apparatus, ¹H-NMR spectra was recorded on a Bruker DRX-400 MHz spectrometer, MS spectra was obtained from a Thermo LCQ *Fleet* apparatus. TLC was performed using silica gel 60 PF₂₅₄ (Merck). The fractionation was performed in chromatographic columns using normal phase silica 230-400 mesh (Merck) and Sephadex LH-20 (Merck) and spots were visualized under UV light and after sprayed vanillin-H₂SO₄ reagent followed by heating at 100 °C.

Plant material

The botanical material of *Mauritia flexuosa* L. f., Arecaceae, was collected in the green area of the Universidade Federal do Amazonas, Manaus (Brazil), and authenticated by the Herbarium of the Universidade Federal do Amazonas where a voucher specimen (no.



Short Communication

Received 25 May 2011

Accepted 3 Aug 2011

Available online 16 Nov 2011

Keywords:

Arecaceae

flavonoids

Mauritia flexuosa

triterpenes

ISSN 0102-695X

<http://dx.doi.org/10.1590/S0102-695X2011005000201>

695X2011005000201

7282) is deposited.

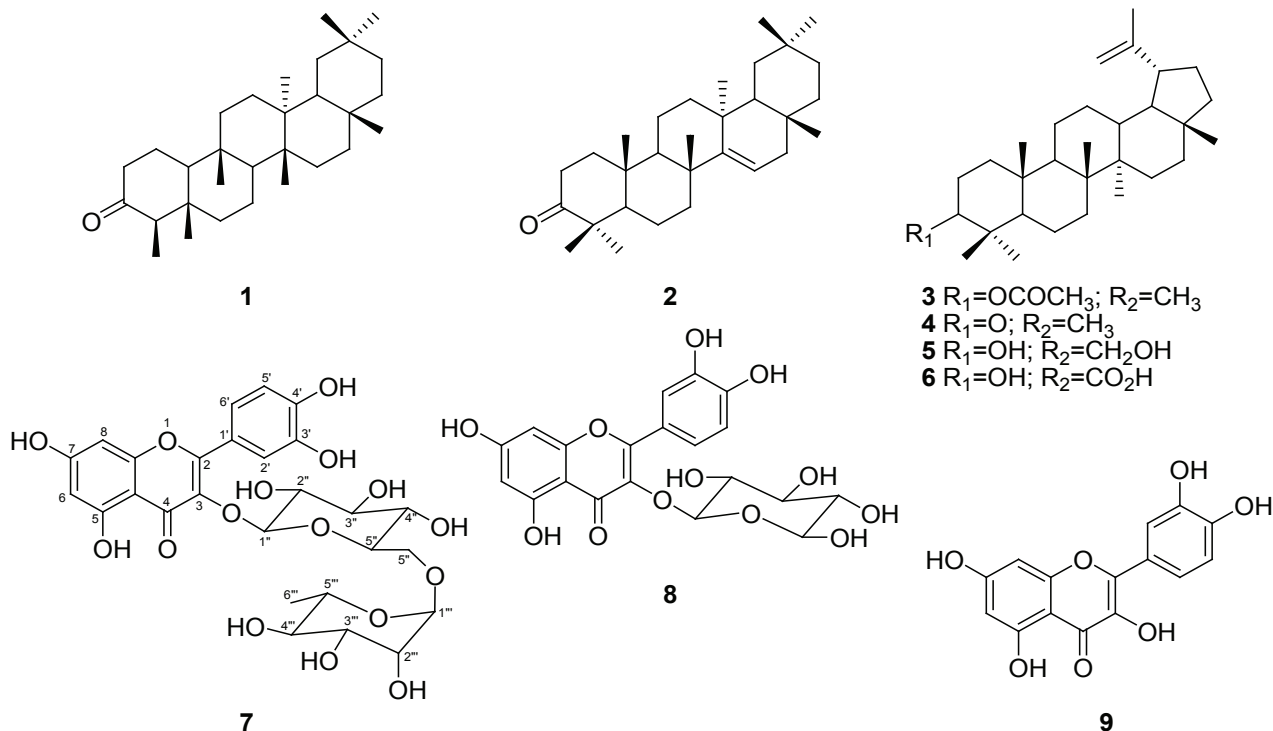
Extraction and isolation

Roots were shade dried for a week and powdered. The powder material (900 g) was extracted at room temperature with *n*-hexane and methanol for about 72 h for each solvent. Every extract was filtered and concentrated *in vacuo* using rotary flash evaporator. The hexane extract (5 g) was fractionated on an open normal silica column (h x Ø = 40 x 2 cm) eluted with hexane: CH₂Cl₂ (9:1, 8:2, 1:1, 3:7) and CH₂Cl₂:AcOEt (9:1, 8:2, 7:3, 1:1, 3:7, 1:9) affording friedelin **1** (21.0 mg), taraxerone **2** (17.4 mg), lupenyl acetate **3** (15.8 mg), lupenone **4** (13.2 mg), betulin **5** (23.0 mg) and betulinic acid **6** (26.6 mg). The methanolic extract (10 g) was separated on a Sephadex LH-20 column (h x Ø = 50 x 3.5 cm) using MeOH in isocratic mode yielding rutin **7** (10.6 mg), quercitrin **8** (4.6 mg) and quercetin **9** (21.9 mg).

Results and Discussion

The triterpenes **1-6** were identified by comparison of their NMR data with the literature (Fulgentius et al., 1990; Klass & Tinto, 1992; Silva et al., 2000; Souza et al., 2001; Saraiva et al., 2006). Biological activities were previously reported for the isolated triterpenes: anti-AIDS for betulinic acid,

herbicide for lupenone and fibrinolytic for betulin (Connolly & Hill, 2010). The flavonoids **7-9** were identified based on NMR data (Hansen et al., 1999; Galotta et al., 2008) and typical fragmentation by CID experiments with ESI-MS (Fabre et al., 2001). Some biological activities reported for the isolated flavonoids are: antidepressant for rutin (Nolder & Schotz, 2002), anti-inflammatory for rutin and quercetin (Afanaseva et al., 2001) and antioxidant for all of them (Galotta et al., 2008). The flavonol quercetin is ubiquitous in monocotyledons, so it cannot be used as marker of the *Arecaceae* family (Silva, 2007). The chemosystematic of the *Arecaceae* family is very complex and by earlier studies it was suggested that the negatively charged flavones and tricetin are possible markers for this family (Williams & Harborne, 1973). Although those authors investigated only leaves of the *Arecaceae* plants, the present research on *Mauritia flexuosa* roots revealed the flavonoids along to triterpenes as compound classes to be considered in future chemosystematic studies of *Arecaceae*. Flavonol *O*-glycosides are rarely reported in *Arecaceae* trees: rutin was previously isolated only from *Chamaeropsis* and *Phoenix* species (Harborne et al., 1974; Asami et al., 1991) while quercitrin was only isolated in the *Phoenix* genus (Hasan et al., 2010). This is the first phytochemical study of *Mauritia flexuosa*, a new source of all the compounds isolated. These are novelty in the genus and their taxonomic significance needs further investigation.



Rutin (**7**): $^1\text{H NMR}$ (400 MHz, CD_3OD) δ_{H} 1,00 (*d*, $J = 6,2$ Hz, H-6''); 3,04-3,11 (*m*, H-4'', H-4'''); 3,22-3,38 (*m*, H-2'', H-3'', H-5'', H-2''', H-5'''); 3,29 (*m*, H-6''); 3,71 (*m*, H-6''); 4,39 (*s*, H-1'''); 5,09 (*d*, $J = 7.1$ Hz, H-1'') 6,19 (*d*, $J = 2.1$ Hz, H-6); 6,38 (*d*, $J = 2.1$ Hz, H-8); 6,86 (*d*, $J = 8.5$ Hz, H-5'); 7,64 (*dd*, $J = 2.1$ and 8.5 Hz, H-6'); 7,67 (*d*, $J = 2.1$ Hz, H-2'). ESI-MS (product ions, 15 eV) m/z : 609 [M-H]⁻, 301 [M-H-Glu-Rha]⁻, 179, 151. mp: 197-199 °C.

Quercitrin (**8**): $^1\text{H NMR}$ (400 MHz, CD_3OD) δ_{H} 1,18 (*d*, $J = 1,2$ Hz, H-6''); 3,31 (*m*, H-4''); 3,55 (*m*, H-2''); 3,66 (*m*, H-3''); 4,08 (*d*, $J = 1,2$ Hz, H-1''); 4,23 (*m*, H-5''); 6,20 (*d*, $J = 1,8$ Hz, H-6); 6,42 (*d*, $J = 1,8$ Hz, H-8); 6,81 (*d*, $J = 8,2$ Hz, H-5'); 7,55 (*dd*, $J = 2,1$ e 8,5 Hz, H-6'); 7,68 (*d*, $J = 2,1$ Hz, H-2'). ESI-MS (product ions, 15 eV) m/z : 447 [M-H]⁻, 301 [M-H-Rha]⁻, 179, 151, 121. mp: 193-195 °C.

Quercetin (**9**): $^1\text{H NMR}$ (400 MHz, CD_3OD) δ_{H} 6,20 (*d*, $J = 1,8$ Hz, H-6); 6,42 (*d*, $J = 1,8$ Hz, H-8); 6,81 (*d*, $J = 8,2$ Hz, H-5'); 7,55 (*dd*, $J = 2,1$ e 8,5 Hz, H-6'); 7,68 (*d*, $J = 2,1$ Hz, H-2'). ESI-MS (product ions, 15 eV) m/z : 301 [M-H]⁻, 179, 151, 121, 107. mp: 315-318 °C.

References

- Afanaseva IB, Ostrakhovitch EA, Mikhalechik EV, Ibragimova, GA, Korkina L 2001. Enhancement of antioxidant and anti-inflammatory activities of bioflavonoid rutin by complexation with transition metals. *Biochem Pharmacol* 61: 677-684.
- Asami A, Hirai S, Shoji J 1991. Studies on the constituents of the Palmae plants VI. Steroids, saponins and flavonoids from the leaves of *Phoenix canariensis* hort. ex Chabaud, *P. humilis* Royle var. hanceana Becc., *P. dactylifera* L. and *Licuala spinosa* Wur MB. *Chem Pharm Bull* 39: 2053-2056.
- Connolly JD, Hill RA 2010. Triterpenoids. *Nat Prod Rep* 27: 79-132.
- El Dib R, Kaloga M, Mahamoud I, Soliman HSM, Moharram FA, Kolodziej H 2004. Sablacaurin A and B, two 19-nor-3,4-seco-lanostane-type triterpenoids from *Sabal causiarum* and *Sabal blackburniana*, respectively. *Phytochemistry* 65: 1153-1157.
- Fabre N, Rustan I, Hoffan E, Lequerc J 2001. Determination of flavone, flavonol and flavanone aglycones by negative ion liquid chromatography electrospray ion trap mass spectrometry. *J Am Soc Mass Spectrom* 12: 707-715.
- França LF, Reber G, Meireles MA, Machado NT, Brunner G 1999. Supercritical extraction of carotenoids and lipids from buriti (*Mauritia flexuosa*), a fruit from the Amazon region. *J. Supercrit. Fluids* 14: 247-256.
- Fulgentius N, Lugemwa T, Huang FH, Bentley D, Alfordx RA 1990. A Heliothis zea Antifeedant from the abundant birchbark triterpene betulin. *J Agric Food Chem* 38: 494-496.
- Galotta ALQ, Boaventura MA, Lima ARS 2008. Antioxidant and cytotoxic activities of "açai" (*Euterpe precatoria* Mart.). *Quim Nova* 31: 1427-1430.
- Galotta ALQ, Boaventura MA 2005. Constituintes químicos da raiz e do talo da folha do açai (*Euterpe precatoria* Mart., Arecaceae). *Quim Nova* 28: 610-613.
- Hansen SH, Jensen AG, Cornett C, Bjørnsdottir I, Taylor S, Wright B, Wilson D 1999. High-performance liquid chromatography on-line coupled to high field NMR and mass spectrometry for structure elucidation of constituents of *Hypericum perforatum* L. *Anal Chem* 71: 5235-5241.
- Hasan MM, Ahmed SW, Azhar I, Bano H 2010. Phytoconstituents isolated from *Phoenix sylvestris* Roxb. *J Basic Apply Sci* 6: 17-22.
- Harborne JB, Williams CA, Greenham J 1974. Distribution of charged flavones and caffeylshikimic acid in Palmae. *Phytochemistry* 13: 1557-1559.
- Harborne JB, Saito N, Detoni CH 1994. Anthocyanins of *Cephaelis*, *Cynomorium*, *Euterpe*, *Lavatera* and *Pinanga*. *Biochem Syst Ecol* 22: 835-836.
- Holdsworth DK, Jones RA, Self R 1997. Volatile alkaloids from *Areca catechu*. *Phytochemistry* 48: 581-582.
- Klass J, Tinto W 1992. Friedelane triterpenoids from Perztassa compta: Complete ^1H and ^{13}C assignments by 2D NMR spectroscopy. *J Nat Prod* 55: 1626- 1630.
- Noldner M, Schotz K 2002. Rutin is essential for the antidepressant activity of *Hypericum perforatum* extracts in the forced swimming test. *Planta Med* 68: 577-580.
- Passos MAB, Mendonça MS 2006. Epiderme dos segmentos foliares de *Mauritia flexuosa* L. f. (Arecaceae) em três fases de desenvolvimento. *Acta Amaz* 36: 431-436.
- Piozzi F, Passananti S, Nansini G 1981. Pterocarpol and triterpenes from *Daemonorops draco*. *Phytochemistry* 20: 514-516.
- Ruiz RR, Alencar JC 2004. Comportamento fenológico da palmeira pataua (*Oenocarpus bataua*) na reserva florestal Adolpho Ducke, Manaus, Amazonas, Brasil. *Acta Amaz* 34: 553-558.
- Saraiva RC, Pinto AC, Nunomura SM, Pohlit A 2006. Triterpenos e alcalóide do tipo cantinona dos galhos de *Simaba polyphylla* (Cavalcante) W.W. Thomas (Simaroubaceae). *Quim Nova* 29: 264-268. 2006.
- Silva AG 2007. A importância de flavonóides na taxonomia de Monocotiledôneas. *Natureza on line* 5: 44-47.
- Silva JRA, Rezende CM, Pinto AC, Pinheiro MLB, Cordeiro MC, Tamborini E, Young CM, Bolzani VS 2000. Ésteres triterpênicos de *Himatanthus sucuuba* (Spruce) Woodson. *Quim Nova* 21: 702-704.
- Silveira CS, Pessanha CM, Lourenço MCS, Neves Junior I, Menezes FS, Kaplan MAC 2005. Atividade

antimicrobiana dos frutos de *Syagrus oleracea* e *Mauritia vinifera*. *Acta Amaz* 15: 143-148.

Souza ADL, Da Rocha AFI, Pinheiro MLB, Andrade CHS, Galotta ALQ, Dos Santos MP 2001. Constituintes químicos de *Gustavia augusta* L. (Lecythidaceae). *Quim Nova* 24: 439-442.

Williams CA, Harborne JB 1973. Negatively charged flavones and tricin as chemosystematic markers in the Palmae. *Phytochemistry* 12: 2417-2430.

Uhl NW, Dransfield J 1987. *Genera Palmarum: A classification of Palms based on the work of Harold E. Moore Jr.*, Allen Press: Lawrence, Kansas.

***Correspondence**

Afonso D. L. de Souza
Departamento de Química, Universidade Federal do Amazonas
Av. Gal. Rodrigo Otávio, 3000, CEP 69077-000, Japiim, Manaus, Amazonas, Brazil
Souzadq@ufam.edu.br
Tel. +55 92 9132 5200