

# Effect of six tropical tanniferous plant extracts on larval exsheathment of *Haemonchus contortus*

Efeito de seis extratos de plantas taniníferas tropicais sobre o desembainhamento larvar de *Haemonchus contortus*

Lorena Mayana Beserra de Oliveira<sup>1</sup>; Claudia Maria Leal Bevilaqua<sup>1\*</sup>; Iara Tersia Freitas Macedo<sup>1</sup>;  
Selene Maia de Moraes<sup>1</sup>; Maria Vivina Barros Monteiro<sup>1</sup>; Claudio Cabral Campello<sup>1</sup>;  
Wesley Lyevertton Correia Ribeiro<sup>1</sup>; Emanuelle Karine Frota Batista<sup>2</sup>

<sup>1</sup>Programa de Pós-Graduação em Ciências Veterinárias, Universidade Estadual do Ceará – UECE

<sup>2</sup>Departamento de Veterinária, Universidade Federal do Piauí – UFPI

Received December 7, 2010

Accepted January 24, 2011

## Abstract

Tanniferous plants represent a promising alternative for controlling gastrointestinal nematodes of small ruminants. This experiment evaluated the effects of extracts from the leaf and stem of *Anadenanthera colubrina*, *Leucaena leucocephala* and *Mimosa tenuiflora* on larval exsheathment of *Haemonchus contortus* *in vitro* and verified the role of tannins in this process. Third-stage larvae of *H. contortus* were incubated with extracts for 3 hours and were exposed to sodium hypochlorite solution. The extracts were tested at 300 µg.mL<sup>-1</sup> and accompanied by controls: phosphate buffer solution (PBS) and polyvinyl polypyrrolidone (PVPP). The larval exsheathment was evaluated for 60 minutes, and the results were subjected to the Kruskal-Wallis test ( $p < 0.05$ ). The six extracts blocked larval exsheathment. After PVPP addition, a tannin inhibitor, the exsheathment percentage was similar to the PBS ( $p > 0.05$ ), except for *L. leucocephala* and *M. tenuiflora* leaf extracts. However, pre-incubation with PVPP of these two extracts significantly changed larval exsheathment when compared to the non-treated extracts ( $p < 0.05$ ). These results suggest that *A. colubrina*, *L. leucocephala* and *M. tenuiflora* could be useful in gastrointestinal nematode control and that tannins are probably the main compounds involved in the observed effects. However, *in vivo* and toxicological studies should be conducted.

**Keywords:** *In vitro*, tannins, polyvinyl polypyrrolidone, helminth control, phytotherapies.

## Resumo

Plantas taniníferas representam uma promissora alternativa de controle dos nematóides gastrintestinais de pequenos ruminantes. Esse experimento avaliou *in vitro* os efeitos dos extratos das folhas e caules de *Anadenanthera colubrina*, *Leucaena leucocephala* e *Mimosa tenuiflora* sobre o desembainhamento larvar de *Haemonchus contortus* e verificou o papel dos taninos nesse processo. Larvas de terceiro estágio de *H. contortus* foram incubadas com 300 µg.mL<sup>-1</sup> de extrato por 3 horas e expostas a uma solução de hipoclorito de sódio. O ensaio foi acompanhado por controles: solução salina tamponada com fosfato (PBS) e polivinilpolipirrolidona (PVPP). O desembainhamento larvar foi avaliado durante 60 minutos e os resultados submetidos ao teste Kruskal-Wallis ( $p < 0,05$ ). Os seis extratos bloquearam o desembainhamento larvar. Após adição de PVPP, um inibidor de taninos, o percentual de desembainhamento foi similar ao PBS ( $p > 0,05$ ), exceto nos extratos das folhas de *L. leucocephala* e *M. tenuiflora*. Entretanto, a pré-incubação desses dois extratos com PVPP alterou significativamente o desembainhamento quando comparado com extratos não-tratados ( $p < 0,05$ ). Esses resultados sugerem que *A. colubrina*, *L. leucocephala* e *M. tenuiflora* podem ser úteis no controle de nematóides gastrintestinais e que os taninos são provavelmente os principais compostos envolvidos nos efeitos. Contudo, estudos toxicológicos e *in vivo* são necessários.

**Palavras-chave:** *In vitro*, taninos, polivinilpolipirrolidona, controle de helmintos, fitoterápicos.

\*Corresponding author: Claudia Maria Leal Bevilaqua  
Programa de Pós-Graduação em Ciências Veterinárias,  
Universidade Estadual do Ceará – UECE, Av. Dedé Brasil 1700,  
Campus do Itaperi, CEP 60740-093, Fortaleza - CE, Brazil;  
e-mail: claudiamlb@yahoo.com.br

## Introduction

Gastrointestinal nematode parasitism remains one of the main threats to the farming of small ruminants in tropical countries. *Haemonchus contortus* is one of the most significant species because of its high prevalence and pathogenicity (HOUNZANGBE-ADOTE et al., 2005). Control of these parasites has been based on the administration of synthetic anthelmintics. However, the development of populations resistant to all of the drug families currently available (TORRES-ACOSTA; HOSTE, 2008) and concerns regarding drug residue in food and in the environment have stimulated the search for alternative control strategies (ATHANASIADOU et al., 2008).

An alternative for control of gastrointestinal nematode parasites that is being researched is the use of tanniferous plants. Promising results to keep eggs from hatching, as well as the development, motility and exsheathment of larvae, have been obtained after incubation of tannin-rich plant extracts with nematodes (BARRAU et al., 2005; BRUNET et al., 2007; MOLAN; FARAJ, 2010). Reduction of nematode egg excretion and worm burden have been also recorded in goats and sheep fed with tanniferous plants (MARLEY et al., 2003; LANGE et al., 2006; HECKENDORN et al., 2007; MINHO et al., 2008, 2010). Most studies have focused on forage legumes belonging to the Fabacea family, such as *Hedysarum coronarium*, *Onobrychis viciifolia*, *Lotus corniculatus*, *Lotus pedunculatus* and *Lespedeza cuneata* (HOSTE et al., 2006). Nevertheless, in many small ruminant production systems, cultivated forage plants are not the main feed source because browser plants (bushes, trees or shrubs) contribute significantly to the nutrition of these animals during a prolonged dry period (MANOLARAKI et al., 2010), which occurs in tropical countries (SANON et al., 2008).

The aim of this study was to evaluate the effect of leaf and stem extracts of *Anadenanthera colubrina*, *Leucaena leucocephala* and *Mimosa tenuiflora* on the exsheathment of infective *H. contortus* larvae *in vitro*. The role of tannins in this process was analyzed after polyvinyl polypyrrolidone (PVPP), a tannin inhibitor, was added. These plants were chosen because of their prevalence in semi-arid regions of Brazil and other tropical countries; these species are also used in small ruminants' feed and are tannin-rich (KANANI et al., 2006; MONTEIRO et al., 2006; RIVERA-ARCE et al., 2007).

## Material and Methods

### 1. Plant and extract preparation

Aerial parts of *A. colubrina*, *L. leucocephala* and *M. tenuiflora* were collected in December 2009 in Fortaleza, State of Ceará, located in northeastern Brazil. Prior to the beginning of the trial, samples of the plants were identified and authenticated by botanists at the Herbarium Prisco Bezerra of the Universidade Federal do Ceará, Brazil. A voucher specimen of each species was deposited under the numbers 46022, 45875 and 46018, respectively.

*A. colubrina*, *L. leucocephala* and *M. tenuiflora* extracts were prepared as described by Barrau et al. (2005). Leaves (150 g) and stems (150 g) of each plant were separately air-dried at

room temperature, crushed to a powder and then mixed with 2 × 900 mL of acetone/water (70:30). The acetone was removed under low pressure using a rotary evaporator, and the aqueous solution was washed 4 times with 500 mL methylene chloride to remove chlorophyll and lipids. The remaining fractions, called leaf extract and stem extract of *A. colubrina*, *L. leucocephala* and *M. tenuiflora*, were freeze-dried and kept refrigerated at 4 °C in airtight containers until use.

### 2. Chemical analysis of extracts

The phytochemical test to detect the presence of hydrolyzable or condensed tannins was performed, following the method described by Matos (2009). The extracts were diluted in alcohol, three drops of ferric chloride (FeCl<sub>3</sub>) solution were added and the mixture was shaken. The formation of a dark blue precipitate indicated the presence of hydrolyzable tannins, and a green precipitate indicated the presence of condensed tannins. Samples were compared to a control, e.g., a mixture of water and ferric chloride solution.

The content of total phenol was determined using the Folin-Ciocalteu spectrophotometric method (BONOLI et al., 2004). Briefly, 100 µL of each extract solution (150 ppm) was shaken for 1 minute with 500 µL of Folin-Ciocalteu reagent and 6 mL of distilled water. Then, 2 mL of 15% sodium carbonate (Na<sub>2</sub>CO<sub>3</sub>) was added and the mixture was shaken again for 30 seconds. The solution was adjusted to 10 mL by adding distilled water. After 2 hours, the absorbance at 750 nm was evaluated using a spectrophotometer. The readings, with three replicates per sample, were performed with a negative control. The total phenol content was assessed by plotting the gallic acid calibration curve. For the total tannin content, 100 mg.mL<sup>-1</sup> PVPP was used to isolate these metabolites from extract. Then total tannins were measured as the difference of total phenol before and after treatment with PVPP.

### 3. Larval artificial exsheathment assay

To recover *H. contortus* third-stage larvae (L3), feces were collected directly from the rectum of an experimentally infected sheep and were processed according to Roberts and O'Sullivan (1950). The isolated of *H. contortus* is benzimidazole-resistant and has been maintained in experimentally infected animals for about five years. The larvae were stored at 4 °C. Twenty-four hours before the experiment, 2- to 3-months-old larvae were brought to 25 °C. The viability and proportion of exsheathed larvae were verified under microscope.

The test was performed according to Bahuaud et al. (2006). The extracts were diluted in phosphate buffer solution (PBS), and the concentration evaluated was 300 µg.mL<sup>-1</sup>. Then, approximately 1,000 ensheathed L3 were incubated for 3 hours at 20 °C with each extract. The larvae were washed and centrifuged three times in PBS (pH 7.2). Thereafter, the L3 were subjected to the process of artificial exsheathment by incubation in a solution of sodium hypochlorite (2% w/v) diluted 1:300 in PBS and divided in 7 aliquots. The kinetics of larval exsheathment were monitored under a microscope, and exsheathed larvae were counted at 0, 10, 20, 30, 40, 50 and 60 minutes after contact with the hypochlorite

solution. The identification and count of ensheathed and exsheathed larvae were realized after the addition of drops of lugol. PBS was used as a negative control. Six replicates were performed for each extract and for the control.

#### 4. Effects of PVPP addition to plant extracts on larval exsheathment

To evaluate the role of tannins in the assay, PVPP, a specific inhibitor of tannins, was added to leaf and stem extracts of *A. colubrina*, *L. leucocephala* and *M. tenuiflora*. PVPP is capable of forming bonds with tannins and can precipitate them from solutions. Comparisons of the results obtained with extracts with or without PVPP confirmed the involvement of these metabolites. Each extract was pre-incubated overnight with 50 mg.mL<sup>-1</sup> PVPP (ALONSO-DÍAZ et al., 2008a; 2008b) and then tested as previously described. A negative control with PBS was included in the test.

#### 5. Statistical analysis

The percentage of larval exsheathment in each treatment was determined by the Equation 1:

$$\left( \frac{\text{Number of exsheathed L3}}{\text{Number of ensheathed L3} + \text{Number of exsheathed L3}} \right) \times 100 \quad (1)$$

Data were initially subjected to the Shapiro-Wilk and Bartlett tests to confirm the normality of distribution and the homogeneity of variance between treatments, respectively. Even after data transformation, there was no homoscedasticity; then, the Kruskal-Wallis test ( $p < 0.05$ ) was applied using SAS (2002). Data were expressed as mean  $\pm$  standard error (S.E.).

## Results

### 1. Chemical analysis of extracts

The yields of *A. colubrina*, *L. leucocephala* and *M. tenuiflora* leaf extracts, after lyophilization, were 9, 8 and 11%, respectively, whereas the stem extract yields were 11, 13 and 17%, respectively. In all extracts, the presence of condensed tannins was detected. Table 1 shows the total phenols and tannins of the extracts.

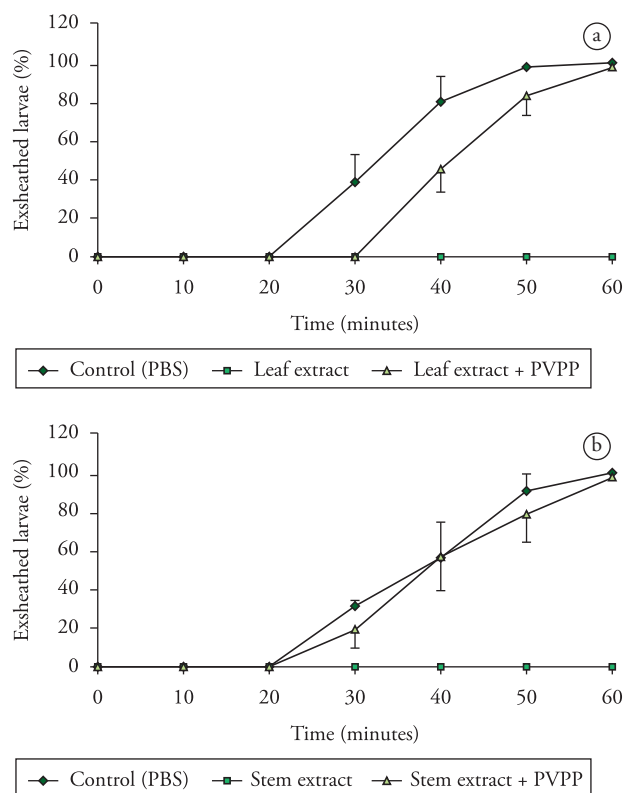
### 2. Larval artificial exsheathment assay

Figures 1, 2 and 3 demonstrate the results of the larval artificial exsheathment assay. In the negative control, >99% of *H. contortus* L3 were exsheathed 60 minutes after contact with the sodium hypochlorite solution. Moreover, 3 hours of contact with the leaf and stem extracts of *A. colubrina*, *L. leucocephala* and *M. tenuiflora* completely inhibited the process, as almost all of the larvae were ensheathed and the percentage obtained was different from the PBS ( $p < 0.05$ ).

**Table 1.** Total phenol and tannin contents of leaf and stem extracts of *Anadenanthera colubrina*, *Leucaena leucocephala* and *Mimosa tenuiflora*.

Plant	Total phenols	Total tannins
<i>Anadenanthera colubrina</i>		
Leaf extract	48.23	40.68
Stem extract	99.58	78.44
<i>Leucaena leucocephala</i>		
Leaf extract	58.81	45.21
Stem extract	37.66	24.07
<i>Mimosa tenuiflora</i>		
Leaf extract	78.73	30.13
Stem extract	147.91	138.85

Results expressed as mg gallic acid per g of extract.

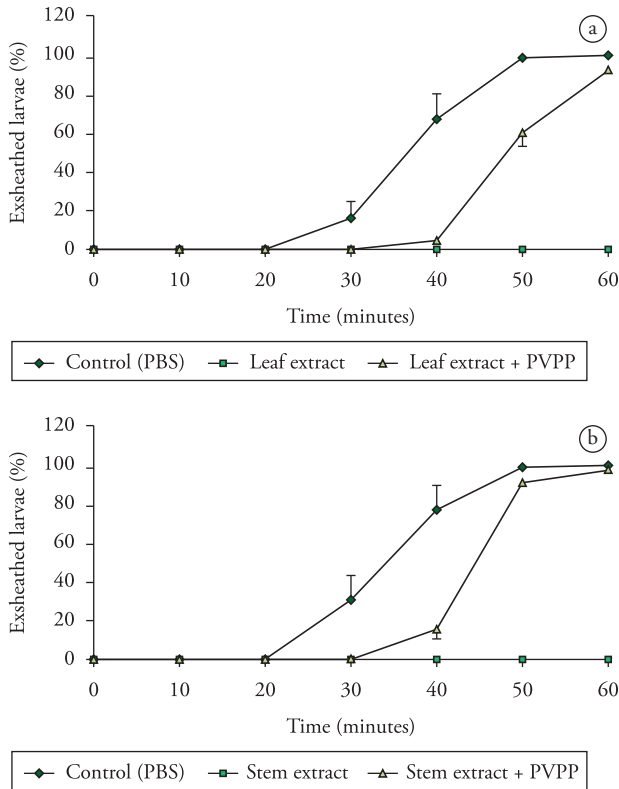


**Figure 1.** Effect of leaf extract (a) and stem extract (b) of *Anadenanthera colubrina*, with or without addition of PVPP, on the artificial *in vitro* exsheathment of *Haemonchus contortus* third-stage larvae.

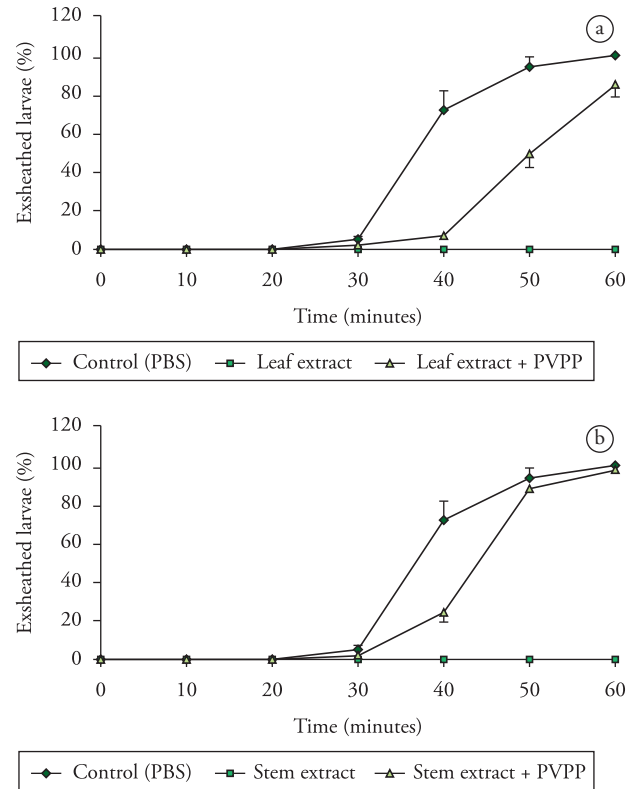
### 3. Effects of PVPP addition to plant extracts on the larval exsheathment

Pre-incubation of *A. colubrina* leaf and stem extracts with PVPP completely reversed the inhibitory effect on larval exsheathment; 60 minutes after the addition of sodium hypochlorite solution, the percentage obtained was similar to negative control ( $p > 0.05$ ) (Figures 1a and b).

After the addition of PVPP to *L. leucocephala* and *M. tenuiflora* leaf extracts, the percentage of *H. contortus* larval exsheathment



**Figure 2.** Effect of leaf extract (a) and stem extract (b) of *Leucaena leucocephala*, with or without addition of PVPP, on the artificial *in vitro* exsheathment of *Haemonchus contortus* third-stage larvae.



**Figure 3.** Effect of leaf extract (a) and stem extract (b) of *Mimosa tenuiflora*, with or without addition of PVPP, on the artificial *in vitro* exsheathment of *Haemonchus contortus* third-stage larvae.

obtained 60 minutes after the addition of sodium hypochlorite solution was 92.27 and 84.50%, respectively. This change was significant when compared with extracts not treated with PVPP ( $p < 0.05$ ). However, the results remained significantly different from the negative control values ( $p < 0.05$ ) (Figures 2a and 3a). Pre-incubation of *L. leucocephala* and *M. tenuiflora* stem extracts with PVPP completely reversed the inhibitory effect, as there were no significant differences when compared to the results obtained with PBS ( $p > 0.05$ ) (Figures 2b and 3b).

## Discussion

The main objective of this study was to demonstrate *in vitro* that *A. colubrina*, *L. leucocephala* and *M. tenuiflora* extracts inhibited *H. contortus* larval exsheathment. Leguminosae is the third largest family of angiosperms, comprising 650 genera and about 18,000 species, including three subfamilies: Caesalpinoideae, Mimosoideae and Papilionoideae (BOUCHENAK-KHELLADI et al., 2010). Most studies have been conducted with species of the Papilionoideae subfamily, whereas the Mimosoideae subfamily has not been well studied. This study was the first to evaluate the antiparasitic effects of *A. colubrina* and *M. tenuiflora*. With respect to *L. leucocephala*, the results obtained confirm those demonstrated by Alonso-Díaz et al. (2008b). These authors demonstrated 90% efficacy on *H. contortus* larval exsheathment using 1,200  $\mu\text{g}\cdot\text{mL}^{-1}$  of *L. leucocephala* leaf extract, while this

study obtained 100% efficiency on larval exsheathment using 300  $\mu\text{g}\cdot\text{mL}^{-1}$ . This difference could be attributed to the plant chemical composition. Genetic factors, climate, soil, harvest time and solar radiation can influence the composition and concentration of secondary metabolites (GOBBO-NETO; LOPES, 2007).

The second objective of this study was to determine if tannins have an important role in the antiparasitic activity of *A. colubrina*, *L. leucocephala* and *M. tenuiflora* on *H. contortus* larvae. Condensed tannins are likely the main substances involved in the antiparasitic activity of these plants because the addition of ferric chloride solution during phytochemical analysis produced a green precipitate, indicating the predominance of these metabolites. Two hypotheses, based on the direct and indirect effects of tannins, have been proposed to explain their antiparasitic effect. However, the exact mechanisms of action of these metabolites remain obscure and could differ depending on the parasite, its stage of development and possibly, the biochemical characteristics of the forage species (HOSTE et al., 2006).

The fact that tanniferous plant extracts demonstrated activity against nematodes *in vitro* supports the hypothesis of a direct effect for tannins. The assay conducted in this study has been routinely used in other experiments. Bahuaud et al. (2006), Brunet et al. (2007), Alonso-Díaz et al. (2008a, 2008b) found retardation or inhibition of *H. contortus* and *Trichostrongylus colubriformis* larval exsheathment after incubation with tanniferous plant extracts. The assumption is that condensed tannins complex with the

sheath proteins of nematodes, which have high proline content and prevent exsheathment (BRUNET et al., 2007).

A relationship between the total phenol and tannin contents and effectiveness of *A. colubrina*, *L. leucocephala* and *M. tenuiflora* extracts on *H. contortus* larval exsheathment could not be established because the results were similar for different concentrations of these metabolites. This observation supports the hypothesis that the chemical structure of condensed tannins is one of the factors modulating their antiparasitic efficacy (HOSTE et al., 2006). *In vitro* experiments showed that prodelphinidin monomers and flavan-3-ol gallates were more active on the egg hatching, larvae motility and exsheathment of *H. contortus*, *Teladorsagia circumcincta*, *T. colubriformis* than procyanidins monomers and flavan-3-ol (MOLAN et al., 2003; BRUNET; HOSTE, 2006).

The supposition that the tannins in *A. colubrina*, *L. leucocephala* and *M. tenuiflora* extracts are primarily responsible for *H. contortus* larval exsheathment inhibition was confirmed by pre-incubation of extracts with PVPP, a synthetic polymer that has the ability to bind tannins and to inactivate their biological properties. Although the addition of this polymer to some plant extracts did not result in obtaining values similar to PBS, the substantial restoration observed indicates an involvement of tannins in this process. However, the action of other secondary metabolites should not be discarded. Barrau et al. (2005) showed that flavonoid glycosides could also interfere with the biology of *H. contortus* and emphasized the similarity between the chemical structures of tannins and flavonoids glycosides, suggesting a similar or close mechanism of action for both types of compounds.

The results of this study indicate that *A. colubrina*, *L. leucocephala* and *M. tenuiflora* extracts can be used to control gastrointestinal nematodes of small ruminants. However, *in vivo* studies should be conducted to evaluate the toxicity and anthelmintic activity of these plants.

## Acknowledgements

The authors would like to thank Banco do Nordeste (BNB) for financial support and *Coordenação de Aperfeiçoamento de Pessoal de Nível Superior* (CAPES) for a scholarship. Dr. Bevilaqua has a grant from the *Conselho Nacional de Desenvolvimento Científico e Tecnológico* (CNPq).

## References

ALONSO-DÍAZ, M. A. et al. Effects of four tropical tanniferous plant extracts on the inhibition of larval migration and the exsheathment process of *Trichostrongylus colubriformis* infective stage. **Veterinary Parasitology**, v. 153, n. 1-2, p. 187-192, 2008a. <http://dx.doi.org/10.1016/j.vetpar.2008.01.011>

ALONSO-DÍAZ, M. A. et al. *In vitro* larval migration and kinetics of exsheathment of *Haemonchus contortus* larvae exposed to four tropical tanniferous plant extracts. **Veterinary Parasitology**, v. 153, n. 3-4, p. 313-319, 2008b. <http://dx.doi.org/10.1016/j.vetpar.2008.01.042>

ATHANASIADOU, S.; HOUDIJK, J.; KYRIAZAKIS, I. Exploiting synergisms and interactions in the nutritional approaches to parasite control in sheep production systems. **Small Ruminant**

**Research**, v. 76, n. 1-2, p. 2-11, 2008. <http://dx.doi.org/10.1016/j.smallrumres.2007.12.016>

BAHUAUD, D. et al. Effects of four tanniferous plant extracts on the *in vitro* exsheathment of third-stage larvae of parasitic nematodes.

**Parasitology**, v. 132, n. 4, p. 545-554, 2006. PMID:16388690. <http://dx.doi.org/10.1017/S0031182005009509>

BARRAU, E. et al. Effect of bioactive compounds from Sainfoin (*Onobrychis viciifolia* Scop.) on the *in vitro* larval migration of *Haemonchus contortus*: role of tannins and flavonol glycosides. **Parasitology**, v. 131, n. 4, p. 531-538, 2005. PMID:16174418. <http://dx.doi.org/10.1017/S0031182005008024>

BONOLI, M. et al. Antioxidant Phenols in Barley (*Hordeum vulgare* L.) Fluor: Comparative Spectrophotometric Study among Extraction Methods of Free and Bound Phenolic Compounds. **Journal of Agricultural and Food Chemistry**, v. 52, n. 16, p. 5195-5200, 2004. PMID:15291496. <http://dx.doi.org/10.1021/jf040075c>

BOUCHENAK-KHELLADI, Y. et al. The evolutionary history and biogeography of Mimosoideae (Leguminosae): An emphasis on African acacias. **Molecular Phylogenetics and Evolution**, v. 57, n. 2, p. 495-508, 2010. PMID:20696261. <http://dx.doi.org/10.1016/j.ympev.2010.07.019>

BRUNET, S. et al. The kinetics of exsheathment of infective nematode larvae is disturbed in presence of tannin-rich plant (sainfoin) both *in vitro* and *in vivo*. **Parasitology**, v. 134, n. 9, p. 1253-1262, 2007. PMID:17346358. <http://dx.doi.org/10.1017/S0031182007002533>

BRUNET, S.; HOSTE, H. Monomers of Condensed Tannins Affect the Larval Exsheathment of Parasitic Nematodes of Ruminants. **Journal of Agricultural and Food Chemistry**, v. 54, n. 20, p. 7481-7487, 2006. PMID:17002411. <http://dx.doi.org/10.1021/jf0610007>

GOBBO-NETO, L.; LOPES, N. P. Plantas medicinais: fatores de influência no conteúdo de metabólitos secundários. **Química Nova**, v. 30, n. 2, p. 374-381, 2007. <http://dx.doi.org/10.1590/S0100-40422007000200026>

HECKENDORN, F. et al. Individual administration of three tanniferous forage plants to lambs artificially infected with *Haemonchus contortus* and *Cooperia curticei*. **Veterinary Parasitology**, v. 146, n. 1-2, p. 123-134, 2007. PMID:17336459. <http://dx.doi.org/10.1016/j.vetpar.2007.01.009>

HOSTE, H. et al. The effects of tannin-rich plants on parasitic nematodes in ruminants. **Trends in Parasitology**, v. 22, n. 6, p. 256-261, 2006. <http://dx.doi.org/10.1016/j.pt.2006.04.004>

HOUNZANGBE-ADOTE, M. S. et al. *In vitro* effects of four tropical plants on three life-cycle stages of the parasitic nematode, *Haemonchus contortus*. **Research in Veterinary Science**, v. 78, n. 2, p. 155-160, 2005. PMID:15563923. <http://dx.doi.org/10.1016/j.rvsc.2004.05.009>

KANANI, J.; LUKEFAHR, S. D.; STANKO, R. L. Evaluation of tropical forage legumes (*Medicago sativa*, *Dolichos lablab*, *Leucaena leucocephala* and *Desmanthus bicornutus*) for growing goats. **Small Ruminant Research**, v. 65, n. 1-2, p. 1-7, 2006. <http://dx.doi.org/10.1016/j.smallrumres.2005.04.028>

LANGHE, K. C. et al. Effect of sericea lespedeza (*Lepedeza cuneata*) fed as hay, on natural and experimental *Haemonchus contortus* infections in lambs. **Veterinary Parasitology**, v. 141, n. 3-4, p. 273-278, 2006. PMID:16857320. <http://dx.doi.org/10.1016/j.vetpar.2006.06.001>

MANOLARAKI, F. et al. Anthelmintic activity of some Mediterranean browse plants against parasitic nematodes. **Parasitology**, v. 137, n. 4, p. 685-696, 2010. PMID:19961649. <http://dx.doi.org/10.1017/S0031182009991399>

- MARLEY, C. L. et al. The effect of birdsfoot trefoil (*Lotus corniculatus*) and chicory (*Cichorium intybus*) on parasite intensities and performance of lambs naturally infected with helminth parasites. **Veterinary Parasitology**, v. 112, n. 1-2, p. 147-155, 2003. [http://dx.doi.org/10.1016/S0304-4017\(02\)00412-0](http://dx.doi.org/10.1016/S0304-4017(02)00412-0)
- MATOS, F. J. A. **Introdução à fitoquímica Experimental**. 3. ed. Fortaleza: UFC, 2009. 45 p.
- MINHO, A. P. et al. Effect of *Acacia molissima* tannin extract on the control of gastrointestinal parasites in sheep. **Animal Feed Science and Technology**, v. 147, n. 1-3, p. 172-181, 2008. <http://dx.doi.org/10.1016/j.anifeedsci.2007.09.016>
- MINHO, A. P. et al. Efficacy of condensed tannin presents in acacia extract on the control of *Trichostrongylus colubriformis* in sheep. **Ciência Rural**, v. 40, n. 6, p. 1360-1365, 2010. <http://dx.doi.org/10.1590/S0103-84782010005000088>
- MOLAN, A. L. et al. Effect of flavan-3-ols on *in vitro* egg hatching, larval development and viability of infective larvae of *Trichostrongylus colubriformis*. **International Journal for Parasitology**, v. 33, n. 14, p. 1691-1698, 2003. [http://dx.doi.org/10.1016/S0020-7519\(03\)00207-8](http://dx.doi.org/10.1016/S0020-7519(03)00207-8)
- MOLAN, A. L.; FARAJ, A. M. The effects of condensed tannins extracted from different plant species on egg hatching and larval development of *Teladorsagia circumcincta* (Nematoda: Trichostrongylidae). **Folia Parasitologica**, v. 57, n. 1, p. 62-68, 2010.
- MONTEIRO, J. M. et al. Use and traditional management of *Anadenanthera colubrina* (Vell.) Brenan in the semi-arid region of northeastern Brazil. **Journal of Ethnobiology and Ethnomedicine**, v. 2, n. 6, p. 1-7, 2006. <http://dx.doi.org/10.1186/1746-4269-2-6>
- RIVERA-ARCE, E. et al. Pharmacognostical studies of the plant drug *Mimosae tenuiflorae cortex*. **Journal of Ethnopharmacology**, v. 113, n. 3, p. 400-408, 2007. PMID:17709219. <http://dx.doi.org/10.1016/j.jep.2007.06.023>
- ROBERTS, F. H. S.; O'SULLIVAN, P. J. Methods for egg counts and larval cultures for strongyles infecting the gastrointestinal tract of cattle. **Australian Journal of Agricultural Research**, v. 1, n. 1, p. 99-102, 1950. <http://dx.doi.org/10.1071/AR9500099>
- SANON, H. O.; KABORÉ-ZOUNGRANA, C.; LEDIN, I. Nutritive value and voluntary feed intake by goats of three browse fodder species in the Sahelian zone of West Africa. **Animal Feed Science and Technology**, v. 144, n. 1-2, p. 97-110, 2008. <http://dx.doi.org/10.1016/j.anifeedsci.2007.10.004>
- SAS INSTITUTE. **SAS/STAT**. User's guide. Cary, NC, 2002.
- TORRES-ACOSTA, J. F. J.; HOSTE, H. Alternative or improved methods to limit gastro-intestinal parasitism in grazing sheep and goats. **Small Ruminant Research**, v. 77, n. 2-3, p. 159-173, 2008. <http://dx.doi.org/10.1016/j.smallrumres.2008.03.009>