

# Phlebotominae sand flies (Diptera: Psychodidae): potential vectors of American cutaneous leishmaniasis agents in the area associated with the Santo Antônio Hydroelectric System in Western Amazonian, Brazil

Allan Kardec Ribeiro Galardo<sup>[1]</sup>, Clícia Denis Galardo<sup>[1]</sup>, Guilherme Abbad Silveira<sup>[2]</sup>,  
Kaio Augusto Nabas Ribeiro<sup>[2]</sup>, Andréa Valadão Híjar<sup>[3]</sup>, Liliane Leite Oliveira<sup>[3]</sup>  
and Thiago Vasconcelos dos Santos<sup>[4]</sup>

[1]. Laboratório de Entomologia Médica, Instituto de Pesquisas Científicas e Tecnológicas do Estado do Amapá, Macapá, Amapá, Brasil. [2]. Santo Antônio Energia S/A, Porto Velho, Rondônia, Brasil. [3]. Saneamento Ambiental Projetos e Operações Ltda, Rio de Janeiro, Brasil. [4]. Instituto Evandro Chagas, Secretaria de Vigilância em Saúde, Ministério da Saúde, Ananindeua, Pará, Brasil.

## ABSTRACT

**Introduction:** An entomological study was conducted as part of a vector-monitoring program in the area associated with the Santo Antônio hydroelectric system in State of Rondônia, Western Amazonian Brazil. **Methods:** Fourteen sampling sites were surveyed to obtain data on the potential vectors of *Leishmania* spp. in the area. Sand flies were collected from 2011 to 2014 during the months of January/February (rainy season), May/June (dry season), and September/October (intermediary season) using light traps arranged in three vertical strata (0.5, 1, and 20m). **Results:** A total of 7,575 individuals belonging to 62 species/subspecies were collected. The five most frequently collected sand flies were *Psychodopygus davisii* (Root) (36.67%), *Trichophoromyia ubiquitalis* (Mangabeira) (8.51%), *Nyssomyia umbratilis* (Ward & Fraiha) (6.14%), *Bichromomyia flaviscutellata* (Mangabeira) (5.74%), and *Psychodopygus complexus* (Mangabeira) (5.25%). These species have been implicated in the transmission of American cutaneous leishmaniasis agents in the Brazilian Amazon region and described as potential vectors of this disease in the study area. **Conclusions:** Additional surveillance is needed, especially in areas where these five species of sand fly are found.

**Keywords:** Vectors. Cutaneous leishmaniasis. Amazonian region.

## INTRODUCTION

Phlebotominae sand flies (Diptera: Psychodidae: Phlebotominae) belong to a group of insects with public health relevance that are vectors of certain pathogens, including agents of leishmaniasis<sup>(1)(2)</sup>. Currently, there are 274 recognized species of sand fly in Brazil, and approximately 10% of these species have been implicated in the transmission of American cutaneous leishmaniasis (ACL)<sup>(3)(4)</sup>. Most of these species are endemic to the Brazilian Amazon region. The high biodiversity of the region may contribute to the variety of ecological systems that *Leishmania* Ross parasites and their respective mammalian reservoir hosts inhabit<sup>(5)</sup>.

In the State of Rondônia in Western Amazonian Brazil, ACL is considered to be a major public health problem, with an annual incidence of 115 cases per 100,000 inhabitants. From 2011

to 2013, 2,945 confirmed autochthonous cases of the disease were reported<sup>(6)</sup>. In Rondônia, at least four *Leishmania* species have been associated with human infection, including *Leishmania (Viannia) braziliensis* Vianna, *Leishmania (Viannia) lainsoni* Silveira, Shaw, Braga & Ishikawa, *Leishmania (Viannia) guyanensis* Floch, and *Leishmania (Leishmania) amazonensis* Lainson & Shaw<sup>(7)(8)</sup>.

Previous studies have compiled interesting data on sand fly populations in State of Rondônia<sup>(9)(10)(11)(12)(13)(14)(15)(16)</sup>. These studies and secondary data compiled in earlier checklists<sup>(17)(18)</sup> identified 111 Phlebotominae species in State of Rondônia; at least 18 of these species have been associated with *Leishmania* infections<sup>(14)</sup>.

As in other states of the Brazilian Amazon region, the territory of Rondônia contains infrastructure such as hydroelectric systems that impact the environment by suppressing the growth of vegetation. Because this human activity might influence the emergence of unknown epidemiological profiles of ACL, it is necessary to improve current knowledge of *Leishmania* species diversity and the temporal/spatial distribution of vector fauna in these areas using surveillance programs aimed at monitoring the transmission of disease. Thus, the aim of the present study was to provide data on the sand fly fauna of an area associated with a hydroelectric system in western Amazonian Brazil and identify potential vectors of ACL agents in this area.

**Correspondence author:** Dr. Thiago Vasconcelos dos Santos. Laboratório de Leishmanioses Prof. Dr. Ralph Lainson/Instituto Evandro Chagas/SVS/MS. Rodovia BR 316, Km 07, 67030-000 Ananindeua, Pará, Brasil.

**Phone:** 55 91 3214-2003

**e-mail:** thiagovasconcelos@iec.pa.gov.br

**Received** 16 March 2015

**Accepted** 24 April 2015

## METHODS

### Study area

Porto Velho (8° 45' 43" S, 63° 54' 14" W) is the capital of and largest city in the State of Rondônia. The average altitude ranges from 70 to 600m above sea level. The climate has been classified as tropical super humid (medium annual temperature 25.5°C), and the predominant vegetation is Amazonian tropical rain forest. This area is directly impacted by the construction of the Santo Antônio Hydroelectric System in the Madeira River. Porto Velho, which has an estimated population of 495,000 inhabitants, experienced 314 reported autochthonous cases of ACL in 2011, 2012, and 2013, accounting for more than 10% of the cases of ACL in the State of Rondônia<sup>(6)</sup>. Fourteen collection sites were selected as priority areas for a survey based on the recommendations of previous studies conducted as part of the Basic Environmental Plan (BEP) of the Santo Antônio Hydroelectric System (Table 1). These sites are located in the *terra firme* forest, which is characterized by distantly spaced apart trees (open canopy) with an average of 40m of canopy. Larger trees with a canopy of up to 55m such as *Castanha-do-Pará* (Lecythidaceae), *Muiaracatiara* (Anacardiaceae), *Tauari* (Lecythidaceae) and *Angelim* (Fabaceae) were also observed. The composition of palms (Arecaceae) is variegated in these dense forest environments.

### Sampling

Before and after the dam was filled, eleven samples were collected between January 2011 and June 2014 during the rainy (January/February), dry (May/June), and intermediary

(September/October) Amazonian seasons using light traps from the Center of Diseases Control (CDC) that were run from 06:00 p.m. to 06:00 a.m. for two consecutive nights. To ensure that the sampling effort was sufficient, a cumulative curve was constructed to calculate the Chao 1, Chao 2, Jackknife 1, and Jackknife 2 diversity indices using the data from all 121 sampled points. The sampling points comprised three traps positioned in different vertical strata, i.e., 0.5, 1, and 20m into the tree canopy, providing 72h of sampling time per collection site. Individual data on the vertical stratification were not available, and each collection site provided a single trapping sample. All collections were approved by the Brazilian Institute of Environment and Renewable Natural Resources (Authorization IBAMA n. 219/2013).

### Sand fly processing

The collected individuals were identified according to their external morphological characteristics. Specimens that could not be identified in the field were stored in 70% alcohol, transported to the laboratory and mounted on glass slides with Berlese liquid (G.B.I. Laboratories, Manchester, England) using a modified version of the Young & Duncan<sup>(1)</sup> temporary mounting technique. The phylogenetic taxonomic criteria of Galati<sup>(19)</sup> were used to identify species, and generic abbreviations proposed by Marcondes<sup>(20)</sup> were used.

### Acquisition of secondary data

Data on the average rainfall during the 2012-2013 period were obtained from the Porto Velho automatic station (National Institute of Meteorology, www.inmet.com.br).

TABLE 1- Collection sites used to catch sand flies in the area associated with the Santo Antônio Hydroelectric System.

| Collection sites | Coordinates  |               | Localities                           |
|------------------|--------------|---------------|--------------------------------------|
|                  | South        | West          |                                      |
| P1               | 09°24'34.4"  | 064°44'39.2"  | Sítio Samaúma I/Jacy Paraná          |
| P2               | 09°25'09.2"  | 064°25'29.6"  | Alto Rio Jacy Paraná                 |
| P3               | 09°15'45.2"  | 064°24'19.2"  | Bairro Velha Jacy/Jacy Paraná        |
| P4               | 09°15'02.5"  | 064°24'11.7"  | Jacy Paraná/distrito                 |
| P5               | 09°05'41.3"  | 064°23'20.5"  | Assentamento Joana D'Arc             |
| P6               | 09°01'59.3"  | 064°08'54.7"  | Reassentamento Morrinhos             |
| P7               | 09°04'03.3"  | 064°10'49.3"  | Reassentamento Santa Rita            |
| P8               | 08° 51'55.8" | 064° 03'38.3" | Reassentamento Vila Nova do Teotônio |
| P9               | 08°46'13.8"  | 064°04'41.2"  | Vila Franciscana                     |
| P10              | 08°45'37.1"  | 064°01'42.4"  | Sítio São Domingos                   |
| P11              | 08°48'36.8"  | 063°56'19.7"  | Vila Santo Antônio                   |
| P12              | 08°45'45.0"  | 063°59'37.8"  | Entorno do Canteiro                  |
| P13              | 08°46'21.0"  | 063°55'52.0"  | Reassentamento Novo Engenho Velho    |
| P14              | 08°35'01.5"  | 063°43'59.3"  | Cujubim Grande                       |

## RESULTS

A total of 7,575 phlebotomine sand flies belonging to 64 taxa (62 species/subspecies) were collected over the course of 11,888h of sampling (Table 2). Fifteen genera were identified, including *Psychodopygus* Mangabeira (13 spp.), *Evandromyia* Mangabeira (10 spp.), *Psathyromyia* Barreto (8 spp.), *Trichophoromyia* Barreto (6 spp.), *Nyssomyia* Barreto (6 spp.), *Micropygomyia* Barreto (5 spp.), *Migonemyia* Galati (3 spp.), *Bichromomyia* Artemiev (2 spp.), *Sciopemyia* Barreto (2 spp.),

*Viannamyia* Mangabeira (1941) (2 spp.), *Pressatia* Mangabeira (2 spp.), *Pintomyia* Costa Lima (1 spp.), *Lutzomyia* França (1 spp.), *Trichopygomyia* Barreto (1 spp.), and *Brumptomyia* França and Parrot (1 spp.).

The sampling sufficiency curve showed stabilization, reaching an asymptote for the four analyzed diversity indices (Figure 1). The presence of six species (*Trichophoromyia eurypyga* (Martins, Falcão & Silva), *Pressatia choti* (Floch & Abonnenc), *Sciopemyia fluviatilis* (Floch & Abonnenc), *Evandromyia pinottii* (Damasceno & Arouck), *Nyssomyia yuilli pajoti* (Abonnenc, Léger & Fauran), and *Trichophoromyia ruii*

TABLE 2 - Sand fly species from the eleven collection sites associated with the Santo Antônio Hydroelectric System in State of Rondônia, Western Amazonian Brazil, from 2011 to 2014.

| Species                                          | Total | Percentage | Species                                               | Total        | Percentage    |
|--------------------------------------------------|-------|------------|-------------------------------------------------------|--------------|---------------|
| 1 <i>Psychodopygus davisi</i>                    | 2,778 | 36.67      | 35 <i>Evandromyia evandroi</i>                        | 17           | 0.22          |
| 2 <i>Trichophoromyia ubiquitalis</i>             | 645   | 8.51       | 36 <i>Evandromyia walkeri</i>                         | 17           | 0.22          |
| 3 <i>Nyssomyia umbratilis</i>                    | 465   | 6.14       | 37 <i>Psathyromyia inflata</i>                        | 16           | 0.21          |
| 4 <i>Bichromomyia faviscutellata</i>             | 435   | 5.74       | 38 <i>Micropygomyia longipenis</i>                    | 16           | 0.21          |
| 5 <i>Psychodopygus complexus</i>                 | 398   | 5.25       | 39 <i>Psychodopygus clausirei</i>                     | 15           | 0.20          |
| 6 <i>Sciopemyia sordellii</i>                    | 266   | 3.51       | 40 <i>Evandromyia monstrosa</i>                       | 13           | 0.17          |
| 7 <i>Psychodopygus corossoniensis</i>            | 191   | 2.52       | 41 <i>Trichophoromyia brachipyga</i>                  | 12           | 0.16          |
| 8 <i>Trichophoromyia castanheirai</i>            | 167   | 2.20       | 42 <i>Evandromyia bacula</i>                          | 11           | 0.15          |
| 9 <i>Evandromyia saulensis</i>                   | 152   | 2.01       | 43 <i>Psathyromyia dreisbachi</i>                     | 11           | 0.15          |
| 10 <i>Nyssomyia antunesi</i>                     | 149   | 1.97       | 44 <i>Evandromyia williamsi</i>                       | 11           | 0.15          |
| 11 <i>Micropygomyia trinidadensis</i>            | 141   | 1.86       | 45 <i>Psathyromyia lutziana</i>                       | 10           | 0.13          |
| 12 <i>Trichophoromyia eurypyga</i>               | 138   | 1.82       | 46 <i>Lutzomyia migonei</i>                           | 10           | 0.13          |
| 13 <i>Psychodopygus chagasi</i>                  | 138   | 1.82       | 47 <i>Bichromomyia olmeca nociva</i>                  | 10           | 0.13          |
| 14 <i>Viannamyia furcata</i>                     | 150   | 1.98       | 48 <i>Viannamyia tuberculata</i>                      | 9            | 0.12          |
| 15 <i>Evandromyia infraspinoza</i>               | 120   | 1.58       | 49 <i>Evandromyia begoniae</i>                        | 11           | 0.14          |
| 16 <i>Nyssomyia yuilli yuilli</i>                | 102   | 1.35       | 50 <i>Evandromyia brachypalla</i>                     | 8            | 0.11          |
| 17 <i>Psychodopygus hirsutus hirsutus</i>        | 90    | 1.19       | 51 <i>Lutzomyia carvalhoi</i>                         | 8            | 0.11          |
| 18 <i>Pressatia choti</i>                        | 89    | 1.17       | 52 <i>Psathyromyia brasiliensis</i>                   | 5            | 0.07          |
| 19 <i>Psychodopygus amazonensis</i>              | 76    | 1.00       | 53 <i>Psathyromyia</i> sp. ( <i>shannoni</i> complex) | 5            | 0.07          |
| 20 <i>Lutzomyia aragaoi</i>                      | 73    | 0.96       | 54 <i>Psathyromyia dendrophila</i>                    | 4            | 0.05          |
| 21 <i>Trichopygomyia trichopyga</i>              | 72    | 0.95       | 55 <i>Evandromyia pinottii</i>                        | 4            | 0.05          |
| 22 <i>Psychodopygus carrerai carrerai</i>        | 65    | 0.86       | 56 <i>Pintomyia damascenoi</i>                        | 3            | 0.04          |
| 23 <i>Psychodopygus leonidasdeanei</i>           | 55    | 0.73       | 57 <i>Pintomyia gruta</i>                             | 2            | 0.03          |
| 24 <i>Psychodopygus paraensis</i>                | 51    | 0.67       | 58 <i>Micropygomyia micropyga</i>                     | 2            | 0.03          |
| 25 <i>Psychodopygus geniculatus</i>              | 50    | 0.66       | 59 <i>Micropygomyia oswaldoi</i>                      | 2            | 0.03          |
| 26 <i>Sciopemyia fluviatilis</i>                 | 49    | 0.65       | 60 <i>Psychodopygus paraensis</i>                     | 2            | 0.03          |
| 27 <i>Nyssomyia shawi</i>                        | 43    | 0.57       | 61 <i>Psathyromyia scaffii</i>                        | 2            | 0.03          |
| 28 <i>Nyssomyia anduzei</i>                      | 32    | 0.42       | 62 <i>Nyssomyia yuilli pajoti</i>                     | 2            | 0.03          |
| 29 <i>Micropygomyia rorotaensis</i>              | 32    | 0.42       | 63 <i>Psychodopygus</i> sp.                           | 2            | 0.03          |
| 30 <i>Psychodopygus ayrozai</i>                  | 32    | 0.42       | 64 <i>Brumptomyia travassosi</i>                      | 1            | 0.01          |
| 31 <i>Trichophoromyia readyi</i>                 | 23    | 0.30       | 65 <i>Trichopygomyia dasypodogeton</i>                | 1            | 0.01          |
| 32 <i>Psychodopygus squamiventris sensu lato</i> | 27    | 0.35       | 66 <i>Psathyromyia triacantha</i>                     | 1            | 0.01          |
| 33 <i>Psychodopygus lainsoni</i>                 | 19    | 0.25       | 67 <i>Trichophoromyia ruii</i>                        | 1            | 0.01          |
| 34 <i>Lutzomyia</i> sp.                          | 18    | 0.24       |                                                       |              |               |
|                                                  |       |            | <b>Total</b>                                          | <b>7,575</b> | <b>100.00</b> |

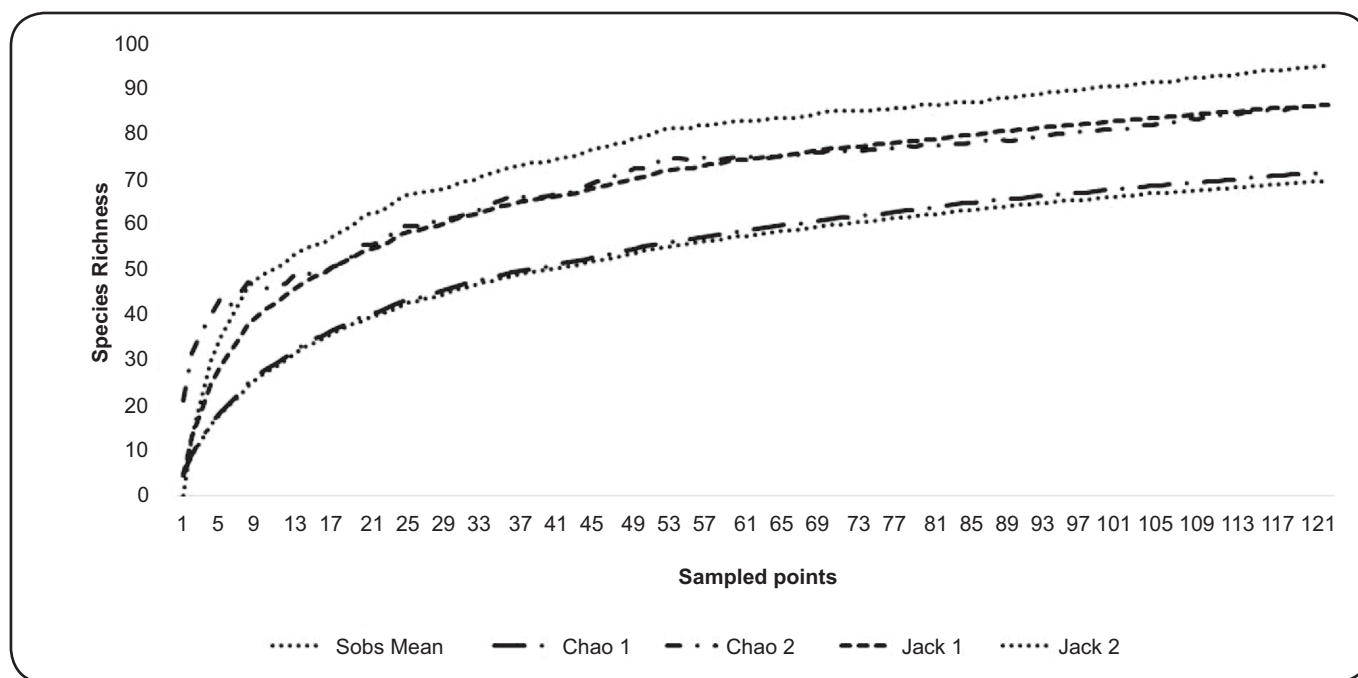


FIGURE 1 - Accumulation curve for sand fly species richness in 121 sites sampled from 2011 to 2014.

(Arias & Young) was documented for the first time in State of Rondônia.

As shown in **Figure 2**, an increasing number of sand flies was collected between the rainy and dry seasons (i.e., January/June), and the number of sand flies collected tended to decrease during the intermediary season (July/October). In 2012-2013, the first and second years after the dam was filled, the number of individuals collected increased during all of the collection seasons; thereafter, the number plateaued until June 2014.

The five most frequently collected species of sand flies have been implicated in the epidemiology of ACL in the Brazilian Amazonian region. These species included *Psychodopygus davisi* (Root) (36.7%), *Trichophoromyia ubiquitousalis* (Mangabeira) (8.51%), *Nyssomyia umbratilis* (Ward & Fraiha) (6.14%), *Bichromomyia flaviscutellata* (Mangabeira) (5.74%), and *Psychodopygus complexus* (Mangabeira) (5.25%). The frequency with which each species was encountered in 2012 and 2013 was compared with the rainfall; the population size of these insects was found to increase at the beginning of the dry season (May/June) when pluviometric precipitation began to decrease (**Figure 3**).

## DISCUSSION

A total of 118 species were identified in State of Rondônia, including the present six new records. Two previous studies conducted in this state had identified 22 new records<sup>(14)</sup> <sup>(16)</sup>. Therefore, it appears that new records of phlebotomine sand flies are continuously being discovered in Rondônia. The Chao/Jackknife indices calculated in the present study indicated that the sampling effort was sufficient to completely estimate the species richness in the studied region.

Several recent studies have described a high level of biodiversity of sand flies (>50 species registered in some of environments) in areas associated with hydroelectric systems in Brazil<sup>(21)</sup> <sup>(22)</sup> <sup>(23)</sup>. The present study is the first to provide a longitudinal evaluation of sand fly populations before and after the filling of a dam. The results of this study indicate that the filling of the dam may be associated with an immediate increase in the density of the sand fly population.

The proportion of sand fly populations that had been disturbed by environmental changes tended to decline over time, as evidenced by comparison with subsequent collections, suggesting an apparent reestablishment of the ecological system. The factors associated with these fluctuations have not been well defined. Environmental changes influence the biodiversity, population size, and proportion of dominant species of sand fly<sup>(24)</sup>. Long-term surveys are needed to evaluate whether sand fly populations are able to be replenished after a long period. A preliminary analysis found no correlation between the number of cases of ACL and variations in the number of flies (data not shown).

*Psychodopygus davisi* is a medically important species of sand fly. The anthropophilic behavior and widespread distribution of these insects in the Amazon basin<sup>(25)</sup> and previous records of natural *Leishmania* infections with this species suggest that this species is a potential vector of ACL agents in the studied region. This species has been shown to carry *L. (V.) braziliensis* and *L. (V.) naiffi* Lainson and Shaw in Serra dos Carajás<sup>(26)</sup> (Southern State of Pará, Brazil) and *L. (V.) braziliensis* in State of Rondônia<sup>(14)</sup> <sup>(27)</sup>. Recently, two females of this species from the area associated with the Belo Monte Hydroelectric System in State of Pará were found to carry peripylarian *Leishmania*-like flagellates (unpublished



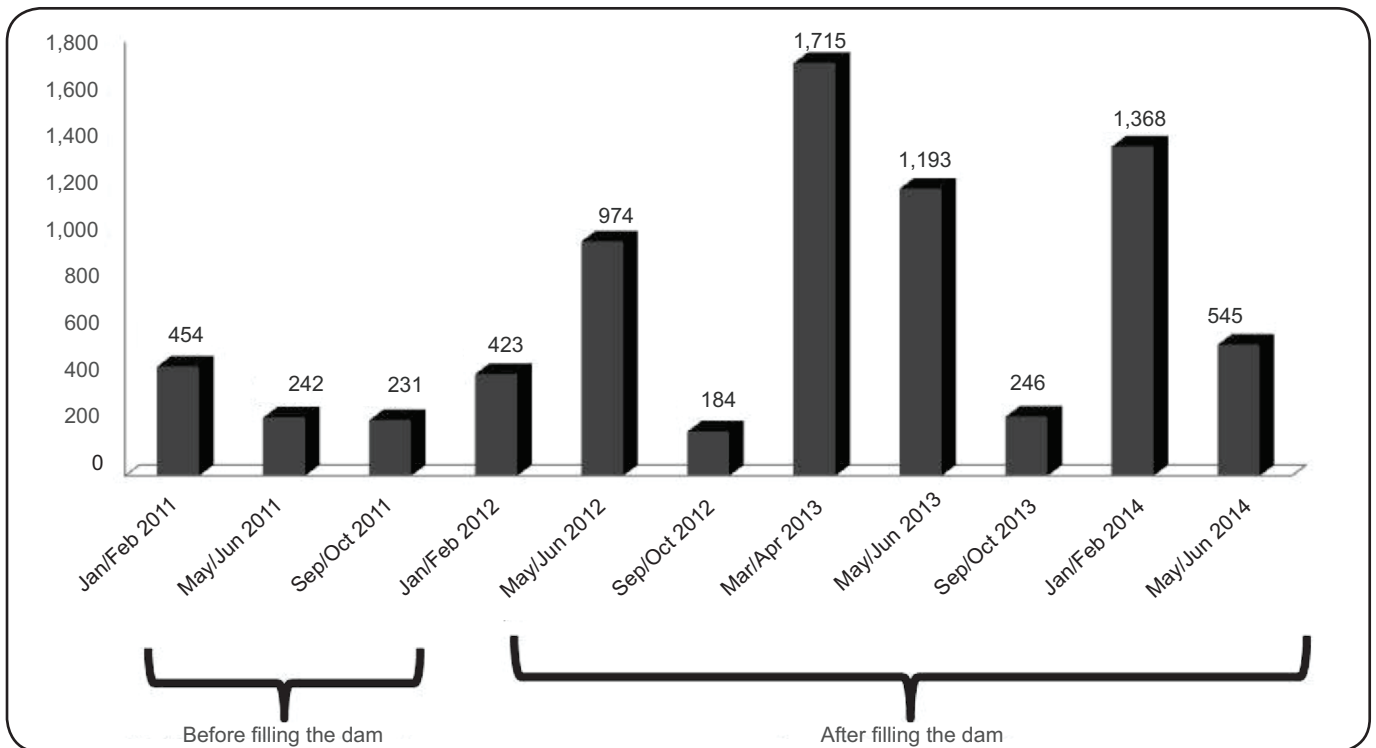


FIGURE 2 - The seasonal variation in number of sand flies collected during the rainy, intermediary, and dry seasons from 2011 to 2014 in the area associated with the Santo Antônio Hydroelectric System.

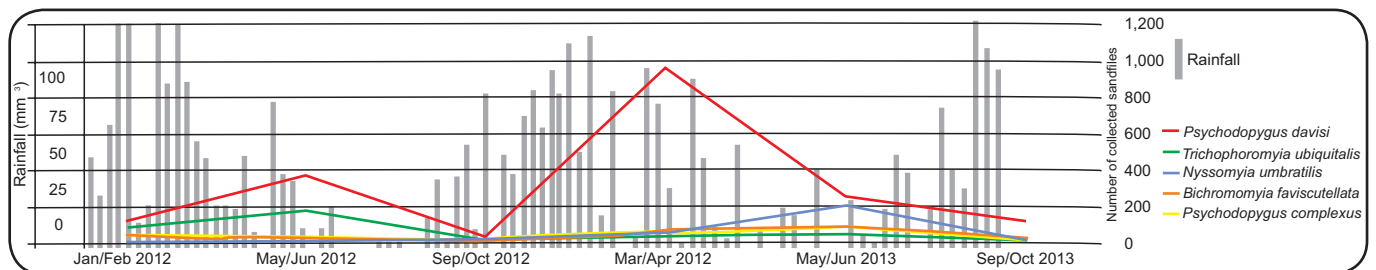


FIGURE 3 - Frequency with which *Psychodopygus davisii*, *Trichophoromyia ubiquitalis*, *Nyssomyia umbratilis*, *Bichromomyia flaviscutellata*, and *Psychodopygus complexus* were encountered and rainfall during the entomological collections of 2012 and 2013.

observations). These findings and data from the present study further suggest that *Ps. davisii* is involved in the transmission of *Leishmania* (*Viannia*) parasites (mainly *L. (V.) naiffi*) in the Amazon region<sup>(14)</sup>. *Trichophoromyia ubiquitalis*, *Nyssomyia umbratilis*, *Bichromomyia flaviscutellata*, and *Ps. complexus* are also likely involved in the transmission of agents of ACL in Brazilian Amazonia.

*Trichophoromyia ubiquitalis* is the only sand fly vector associated with the transmission of *L. (V.) lainsoni*<sup>(28)</sup>, a parasite recently identified in State of Rondônia<sup>(8)</sup>. However, *Trichophoromyia* species are also thought to be carriers of *L. (V.) lainsoni*, as deoxyribonucleic acid (DNA) of this parasite was found in *Trichophoromyia auraensis* specimens in the Peruvian Amazon<sup>(29)</sup>. Furthermore, two pooled samples of *Th. ubiquitalis* were recently observed to carry the DNA of an ambiguously identified *Leishmania* (*Viannia*) parasite [most likely *L. (V.) braziliensis*] in the rural area of Cerrado in the State of Mato Grosso<sup>(30)</sup>.

In the present study, *Th. ubiquitalis*, a known vector of *L. (V.) lainsoni*, was the second most frequently identified species of sand fly in the study area. These observations are in conflict with the hypothesis that other species more anthropophilic than *Th. ubiquitalis* transmit *L. (V.) lainsoni*-like parasites in State of Rondônia<sup>(8)</sup> which was suggested by the fact that *Th. ubiquitalis* was infrequently encountered in entomological studies conducted in the municipality of Monte Negro<sup>(14)</sup>. Although it has previously been suggested that *Th. ubiquitalis* does not exhibit high levels of anthropophilic behavior under field conditions<sup>(28)</sup>, previous studies have described the intradomiciliar transmission of *L. (V.) lainsoni* via indoor-collected *Th. ubiquitalis* in an island of the metropolitan area of Belém, State of Pará (unpublished observations). These findings suggest that *Th. ubiquitalis* is a putative vector of *L. (V.) lainsoni* in State of Rondônia.

*Nyssomyia umbratilis* has been identified as the main vector of *L. (V.) guyanensis* in northern Brazil, particularly in the Guianan ecoregion complex<sup>(31)</sup>. The findings of the present study suggest that *Ny. umbratilis* is a frequently encountered species in the Porto Velho municipality, which is consistent with findings of Azevedo et al<sup>(13)</sup>. These authors found that *Ny. umbratilis* was the second most (19.9%) prevalent species in the Samuel Ecological Station, accounting for 48.5% of the specimens collected in the canopy at 20m. Although the distribution of *L. (V.) guyanensis* near the southern Amazon River remains unknown, recent findings related to the circulation of a *L. (V.) guyanensis*-like parasite in State of Rondônia<sup>(8)</sup> suggest that there is a transmission cycle for this parasite/vector in the studied region. However, additional observations are needed to confirm this hypothesis.

*Bichromomyia flaviscutellata*<sup>(2) (7)</sup>, the major vector of *L. (L.) amazonensis*, was frequently encountered in the present survey. This was surprising because light traps were used for collection, and these species typically cannot be collected in large numbers without the use of rodent-baited traps<sup>(32)</sup>. This unexpected result could be explained by the fact that the traps were placed 50cm above the ground at each CDC collection station where low-flying specimens may have been attracted to the light and intercepted.

*Bichromomyia olmeca nociva* (Young & Arias), a suspected vector<sup>(33)</sup> closely related to *Bichromomyia* species which are associated with the epidemiology of *L. (L.) amazonensis*, was detected in this study. This species is similar to *Bi. reducta* (Felicianelli, Ramirez Pérez & Ramirez), a species that was not observed in the present survey but has previously been described as a natural vector of *L. (L.) amazonensis* in studies conducted in Cachoeira Samuel, State of Rondônia<sup>(12)</sup>. Because *Bi. flaviscutellata* was detected most frequently in the present study, this species is suspected to be the main species involved in the transmission of ACL attributed to *L. (V.) amazonensis* in Rondônia<sup>(7)</sup>.

*Psychodopygus complexus* is currently recognized to be associated with *L. (V.) braziliensis*-associated ACL in areas where the closely related species *Psychodopygus wellcomei* is absent<sup>(34)</sup>. In the present study, *Ps. complexus* was unambiguously identified in the collection because of the absence of *Ps. wellcomei* males. However, the transmission of *L. (V.) braziliensis* by *Ps. complexus* cannot be assumed because no reports of natural infection with this species or evidence of ecological association between the presence of this species and ACL have been reported in the study region. *L. (V.) braziliensis* is widely distributed throughout Brazil, and several species of sand fly that were not encountered in this study, including *Ps. wellcomei* Fraiha, Shaw & Lainson, *Migonemyia migonei* (França), *Nyssomyia whitmani* (Antunes & Coutinho), and *Ny. intermedia* (Lutz & Neiva)<sup>(2) (7)</sup>, have been implicated in the transmission of these agents.

The epidemiological background of the highly anthropophilic species *Ps. davisi* should be considered in the context of the transmission of various *L. (Viannia)* species in areas of the Amazon region such as Rondônia<sup>(14)</sup>. The evidence encountered

in the present study cannot exclude the possibility that this particular species of sand fly was involved in human infections recently attributed to *L. (V.) braziliensis*<sup>(8)</sup> in Rondônia.

The area associated with the Santo Antônio hydroelectric system currently has the highest recorded diversity of Phlebotominae in State of Rondônia. The seasonal fluctuations in sand fly populations from the studied region have been shown to be associated with the end of the rainy period, which is consistent with findings from other entomological studies. However, changes in the populations of these insects are also influenced by the filling of the dam. The increased number of sand flies was not found to be associated with an increase in the incidence of ACL.

*Psychodopygus davisi*, *Trichophoromyia ubiquitalis*, *Nyssomyia umbratilis*, *Bichromomyia flaviscutellata*, and *Ps. complexus* were the five most frequently collected species in this study, and they are all considered to be potential vectors of ACL agents in the Amazon region. The large number of *Ps. davisi* specimens collected in the present study suggests a need for the entomological surveillance of this sand fly species, as these insects are potential vectors of zoonotic cutaneous leishmaniasis in Rondônia.

## ACKNOWLEDGMENTS

The authors wish to thank to Redivaldo Francisco Almeida de Souza, Aderbal Amanajás Santana and Jorge Pereira Duarte for their technical support in the field and laboratory work. The authors also appreciate the cooperation of the *Instituto de Pesquisas Científicas e Tecnológicas do Estado do Amapá*.

## CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

## FINANCIAL SUPPORT

*Santo Antônio Energia S/A*.

## REFERENCES

1. Young DG, Duncan MA. Guide to the identification and geographic distribution of *Lutzomyia* sand flies in Mexico, the West Indies, Central and South America (Diptera: Psychodidae), Memoirs of the American Entomological Institute. Gainesville: Associated Publishers American Entomological Institute; 1994.
2. Ready P. Biology of phlebotomine sand flies as vectors of disease agents. *Annu Rev Entomol* 2013; 58:227-250.
3. Andrade AJ, Gurgel-Gonçalves R. New record and update on the geographical distribution of *Pintomyia monticola* (Costa Lima, 1932) (Diptera: Psychodidae) in South America. *Check List* 2015; 11:1566.
4. World Health Organization (WHO). Library Cataloguing-in-Publication Data. Control of the leishmaniasis: Report of a meeting of the WHO Expert Committee on the Control of Leishmaniasis. Technical Report Series n. 949. Geneva: WHO; 2010.

5. Rangel EF, Lainson R. Proven and putative vectors of American cutaneous leishmaniasis in Brazil: aspects of their biology and vectorial competence. *Mem Inst Oswaldo Cruz* 2009; 104: 937-954.
6. Sistema de informação de agravos de notificação (SINAN). Secretaria de Vigilância e Saúde, Ministério da Saúde. Tab Data Systems (Internet). Brasília: Ministério da Saúde; 2014. (Cited 2014, October 22). Available at: <http://dtr2004.saude.gov.br/sinanweb/index.php>
7. Ministério da Saúde, Secretaria de Vigilância em Saúde. Manual de Vigilância da Leishmaniose Tegumentar Americana. Série A. Normas e Manuais Técnicos. 2<sup>nd</sup>. ed. Brasília: Editora do Ministério da Saúde; 2007.
8. Shaw JJ, Faria DL, Basano SA, Corbett CE, Rodrigues CJ, Ishikawa EA, et al. The aetiological agents of American cutaneous leishmaniasis in the municipality of Monte Negro, Rondônia State, western Amazonia, Brazil. *Ann Trop Med Parasitol* 2007; 101: 681-688.
9. Martins AV, Falcão AL, Silva JE. Notas sobre os flebotomos do território de Rondônia, com a descrição de seis espécies novas (Diptera, Psychodidae). *Rev Bras Biol* 1965; 35:1-20.
10. Martins AV, Williams P, Falcão AL. American Sand Flies (Diptera: Psychodidae, Phlebotominae), Academia Brasileira de Ciências, Rio de Janeiro, Brasil. 1978; 194 p.
11. Biancardi CB, Arias JR, Freitas RA, Castellon EG. The known geographical distribution of sand flies in the state of Rondônia, Brazil (Diptera: Psychodidae). *Acta Amaz* 1982; 12:167-179.
12. Freitas RA, Barret TV, Naiff RD. *Lutzomyia reducta* Feliciangeli et al., 1988, a host of *Leishmania amazonensis*, sympatric with two other members of the *Flaviscutellata* complex in southern Amazonas and Rondônia, Brazil (Diptera: Psychodidae). *Mem Inst Oswaldo Cruz* 1989; 84:363-369.
13. Azevedo ACR, Luz SLB, Vieira ML, Rangel EF. Studies on the sandfly fauna of Samuel Ecological Station Porto Velho municipality, Rondônia state, Brazil. *Mem Inst Oswaldo Cruz* 1993; 88:509-512.
14. Gil LHS, Basano AS, Souza AA, Silva MGS, Barata I, Ishikawa EA, et al. Recent observations on the sand fly (Diptera: Psychodidae) fauna of the State of Rondônia, Western Amazônia, Brazil: the importance of *Psychodopygus davisi* as a vector of zoonotic cutaneous leishmaniasis. *Mem Inst Oswaldo Cruz* 2003; 98:751-755.
15. Gil LHS, Araújo MS, Villalobos JM, Camargo LMA, Ozaki LS, Fontes CJF, et al. Species structure of sand fly (Diptera: Psychodidae) fauna in the Brazilian western Amazon. *Mem Inst Oswaldo Cruz* 2009; 104:955-959.
16. Teles CBG, Basano AS, Zagonel-Oliveira M, Campos JJ, Oliveira AFJ, Freitas RA, et al. Epidemiological aspects of American cutaneous leishmaniasis and phlebotomine sandfly population, in the municipality of Monte Negro, State of Rondônia, Brazil. *Rev Soc Bras Med Trop* 2013; 46: 60-66.
17. Bermudez H, Dedet JP, Falcão AL, Feliciangeli D, Rangel EF, Ferro C, et al. A programme for computer aided identification of phlebotomine sandflies of the Americas (CIPA) - Presentation of check-list of American species. *Mem Inst Oswaldo Cruz* 1993; 88:221-230.
18. Aguiar GM, Medeiros WM. Distribuição regional e habitats das espécies de flebotomíneos do Brasil. In: Rangel EF, Lainson R, editors. *Flebotomíneos do Brasil*. Chapter 3. Rio de Janeiro: Editora Fiocruz; 2003. p. 207-256.
19. Galati EAB. Morfologia e Taxonomia: Morfologia, terminologia de adultos e identificação dos táxons da América. In: Rangel EF, Lainson R, editors. *Flebotomíneos do Brasil*, Rio de Janeiro: Editora Fiocruz; 2003. p. 176.
20. Marcondes CB. A proposal of generic and subgeneric abbreviations for phlebotomine sandflies (Diptera: Psychodidae: Phlebotominae) of the world. *Entomol News* 2007; 118:351-356.
21. Vilela ML, Azevedo CG, Carvalho BM, Rangel EF. Phlebotomine fauna (Diptera: Psychodidae) and putative vectors of leishmaniasis in impacted area by hydroelectric plant, State of Tocantins, Brazil. *PLoS One* 2011; 6:e27721.
22. Barata RA, Ursine RL, Nunes FP, Morais DH, Araújo HS. Synanthropy of mosquitoes and sand flies near the Aimorés hydroelectric power plant, Brazil. *J Vector Ecol* 2012; 37:397-401.
23. Gomes AC, Galati EAB, Paula MB, Mucci LF. Phlebotomines in the area adjacent to the Porto Primavera dam, between São Paulo and Mato Grosso do Sul states, Brazil. *Rev Pat Trop* 2012; 41:215-221.
24. Teodoro U, Kühl JB, Santos DR, Santos C. Impact of environmental changes on sand fly ecology in southern Brazil. *Cad Saude Publica* 1999; 15:901-906.
25. Castellón EG, Arias JR, Freitas RA, Naiff RD. Os flebotomíneos da região Amazônica, estrada Manaus Humaitá, estado do Amazonas, Brasil (Diptera: Psychodidae: Phlebotominae). *Acta Amaz* 1994; 24:91-102.
26. Souza AAA, Silveira FT, Lainson R, Barata IR, Silva MGS, Lima JAN, et al. The Phlebotominae fauna of Serra dos Carajás, Pará, Brazil, and its possible implication in the transmission of American tegumentary leishmaniasis. *Rev Pan-Amaz Saude* 2010; 1:45-51.
27. Grimaldi Jr G, Momen H, Naiff RD, McMahon-Pratt D, Barret TV. Characterization and classification of leishmanial parasites from humans, wild mammals, and sand flies in the Amazon region of Brazil. *Am J Trop Med Hyg* 1991; 44:645-661.
28. Lainson R, Shaw JJ, Souza AAA, Silveira FT, Falqueto A. Further observations on *Lutzomyia ubiquitalis* (Psychodidae: Phlebotominae), the sandfly vector of *Leishmania (Viannia) lainsoni*. *Mem Inst Oswaldo Cruz* 1992; 87:437-439.
29. Valdivia HO, Santos MB, Fernandez R, Baldeviano GC, Zorrilla VO, Vera H, et al. Natural *Leishmania* infection of *Lutzomyia (Trichophoromyia) auraensis* in Madre de Dios, Peru, detected by a fluorescence resonance energy transfer-based real-time polymerase chain reaction. *Am J Trop Med Hyg* 2012; 87:511-517.
30. Thies SF, Ribeiro ALM, Michalsky EM, Miyazaki RD, Fortes-Dias CL, Fontes CJF, et al. Phlebotomine sandfly fauna and natural *Leishmania* infection rates in a rural area of Cerrado (tropical savannah) in Nova Mutum, State of Mato Grosso in Brazil. *Rev Soc Bras Med Trop* 2013; 46:293-298.
31. Rotureau B. Ecology of the *Leishmania* species in the Guianan ecoregion complex. *Am J Trop Med Hyg* 2006; 74:81-96.
32. Lainson R, Shaw JJ. Leishmaniasis in Brazil I. Observations on enzootic rodent leishmaniasis – Incrimination of *Lutzomyia flaviscutellata* (Mangabeira) as the vector in the lower Amazonian basin. *Trans R Soc Trop Med Hyg* 1968; 62:385-395.
33. Arias JR, Freitas RA, Naiff RD, Barret TV. Observations on the parasite *Leishmania mexicana amazonensis* and its natural infection of the sandfly *Lutzomyia olmeca nociva*. *Bull PAHO* 1987; 21:48-54.
34. Souza AAA, Ishikawa EAY, Braga R, Silveira FT, Lainson R, Shaw JJ. *Psychodopygus complexus*, a new vector of *Leishmania braziliensis* to humans in Pará State, Brazil. *Trans R Soc Trop Med Hyg* 1996; 90:112-113.