

Division - Soil Processes and Properties | Commission - Soil Biology

Chemical, Biological, and Biochemical Parameters of the Soil P Cycle After Long-Term Pig Slurry Application in No-Tillage System

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Received: February 1, 2017

Approved: July 07, 2017

How to cite: Tiecher T, Tiecher TL, Mallmann FJK, Zafar M, Ceretta CA, Lourenzi CR, Brunetto G, Gatiboni LC, Santos DR. Chemical, biological, and biochemical parameters of the soil P cycle after long-term pig slurry application in no-tillage system. Rev Bras Cienc Solo. 2017;41:e0170037.
<https://doi.org/10.1590/18069657rbc20170037>

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ABSTRACT: In Southern Brazil, agricultural soils are often treated over long periods with annual pig slurry (PS) applications, and their influence on soil nutrient dynamics should be monitored. This study aimed to evaluate the P forms accumulated in the soil and some biological and biochemical parameters of the P cycle after 19 successive PS applications over eight years, in a no-tillage (NT) system. Pig slurry was applied at rates of 0, 20, 40, and 80 m³ ha⁻¹ per crop. Several P forms (available P, microbial P, phosphatase enzyme, total organic and inorganic P) in the 0.00-0.05 and 0.05-0.10 m soil layers were analyzed. Increases up to 5,523 % in available P, 308 % in microbial P, 104 % in phosphatase enzyme activity, 91 % in organic P, 251 % in inorganic P, and 19 % in Total-P were observed in the 0.00-0.05 m soil layer in response to the highest PS rate compared to the treatment with no PS. Results also indicated that P added via PS was preferentially accumulated in the inorganic fractions (83-98 %). The increase in organic P was limited by the increase in soil organic carbon. The increase in soil microbial biomass-P was more influenced by available P, while the increase in phosphatase enzyme activity was more related to the increase in organic P and C. Therefore, PS applied to the soil affected the chemical, biological, and biochemical components of the soil P cycle. Results indicate that even half of the PS rate normally used by farmers (20 m³ ha⁻¹) poses a high risk of pollution of water bodies by runoff from these soils.

Keywords: organic phosphorus, inorganic phosphorus, microbial biomass phosphorus, acid phosphatase.

INTRODUCTION

Brazil is the fourth largest producer and exporter of pork meat in the world. Pig production has increased from 368 million tons in 1960 to 3,643 million tons in 2015 (ABPA, 2016). Currently, the swine production industry sustains 610 thousand direct and indirect jobs, and there are 24 thousand pig producers in Brazil, of which 80 % are concentrated in the three southernmost states of Brazil (Paraná - PR, Santa Catarina - SC, and Rio Grande do Sul - RS) (ABPA, 2016). In Southern Brazil, 63 % of the national pig production is concentrated in an area that represents only 6 % of the national territory (513 million km²). These three states are also responsible for more than 80 % of pork exports from Brazil. This farming activity is widespread, profitable, and popular among small farmers in southern Brazil, where pig slurry (PS) application on agricultural fields is a common practice of nutrient management (Grohskopf et al., 2016; Hentz et al., 2016; Lourenzi et al., 2016; Couto et al., 2017). The use of manures as alternative to mineral P fertilizer is becoming an increasingly essential practice in order to decrease the demand of mineral P fertilizers in agriculture, in view of the expected global rock phosphate scarcity in the coming decades (Cordell et al., 2009). The field-applied P contained in PS in Brazil is estimated at around 123 thousand tons per year (Tiecher et al., 2014), representing about 3.2 % of the annual consumption of P contained in phosphate fertilizers in Brazil (3,860 thousand tons per year) (IFA, 2013).

On farms, PS is often stored in anaerobic dunghills and later applied as nutrient source on pastures and annual crops. Generally, PS is applied in limited areas because pig farms in Southern Brazil normally have small areas. Hence, repeated PS applications on the same fields result in the accumulation of nutrients and heavy metals in the topsoil, especially when the soil is not plowed (Ceretta et al., 2010a; Giroto et al., 2010; Guardini et al., 2012a; Xue et al., 2013). In this context, P accumulation in the soil surface is of particular concern, due to the high environmental risk of eutrophication of water bodies (Panuelas et al., 2009; Wang et al., 2013).

Higher soil P availability results in higher uptake by plants, which may increase the yield and dry matter production (Ceretta et al., 2005; Adeli et al., 2008). In this process, part of the inorganic P absorbed by plants is incorporated into organic compounds. After plant senescence and later decomposition in the soil, the soil organic P content increases (Ceretta et al., 2010a; Tiecher et al., 2012a). In addition, part of the P contained in PS is already in organic form, thus directly increasing the organic P fraction (McDowell and Stewart, 2005). However, to become available to plants, the organic phosphorus must be mineralized first because plants only absorb P as inorganic orthophosphate. This process is catalyzed by periplasmic and extracellular phosphatase enzymes, which are either secreted by plant roots and soil flora and microorganisms to meet their P requirement, or passively released after cell death and lysis (Quiquampoix and Mousain, 2005). Generally, plants and soil microorganisms respond to inorganic P starvation by an intensification of the phosphatase enzyme activity in the soil, while high levels of soil available P tend to decrease phosphatase enzyme activity (Gatiboni et al., 2008a).

Soil microorganisms affect the soil P dynamics not only by mineralization of soil organic P, but also by immobilizing soluble phosphate from the soil solution, thus preventing P sorption by soil inorganic colloids (Nziguheba and Bünemann, 2005). Later, after cell death and lysis, P release can be more synchronized with the plant demand (Martinazzo et al., 2007). Accordingly, P stored in the microbial biomass is considered potentially available for plants. In this setting, microbiological and biochemical parameters, such as P stored in the microbial biomass and acid phosphatase enzyme activity should be investigated to improve the understanding of soil P dynamics under PS application, as these parameters are sensitive to changes in land use and management (Gatiboni et al., 2008b; Tiecher et al., 2012a).

Accordingly, our hypothesis is that P applied via PS is accumulated in the soil mainly in inorganic forms, increasing P availability, which in turn may stimulate P accumulation in soil microbial biomass and suppress phosphatase enzyme activity in the soil. These effects will be the greater, the higher the PS rate applied. Therefore, this study aimed to evaluate the impact of successive PS applications on P forms in the soil and on some biological and biochemical parameters related to the soil P cycle.

MATERIALS AND METHODS

Study site, and experimental design and description

The study was carried out on a Typic Hapludalf soil (Soil Survey Staff, 2014), or *Argissolo Vermelho Distrófico arênico* (Santos, 2013), in Santa Maria, Rio Grande do Sul, Brazil (29° 42' 52" S, 53° 42' 10" W; 90 m a.s.l.). The soil is well-drained and has strong clay eluviation. The parent material is upper Triassic sandstone, called Santa Maria formation (Silva, 1997). The soil mineralogical composition includes interstratified illite-smectite, kaolinite-smectite, hydroxy-aluminum vermiculite, and kaolinite clay minerals (Bortoluzzi et al., 2008). The climate is subtropical (Cfa, Köppen classification system) with annual means of around 1,600 mm precipitation and temperature of 19 °C. Mean monthly rainfall and temperatures during the experimental period (2000-2008) are listed in figure 1.

According to the recommendation of the regional committee for soil chemistry and fertility (CQFS-RS/SC, 2016), there are two possible ways to determine correct PS rates for application as fertilizer: (i) based on previous manure chemical analysis, which is time-consuming and expensive, or (ii) evaluation of manure density to estimate N, P, and K contents. In both cases, the rate is defined as the amount of PS required to meet N, P or K demands. In order to meet the other nutrient requirements, complementary mineral fertilization is recommended. In practice, however, local farmers do not follow this recommendation and use the soil for easy disposal of PS. In this scenario, the PS rate commonly used is about 40 m³ ha⁻¹ per crop, which often exceeds the nutritional requirements of plants. Therefore, in order to investigate the impact of these management practices in the soil P dynamics, we tested three application rates: a 40 m³ ha⁻¹ standard rate, as usually applied by pig farmers, and half of and double the standard rate, plus a control treatment without PS application.

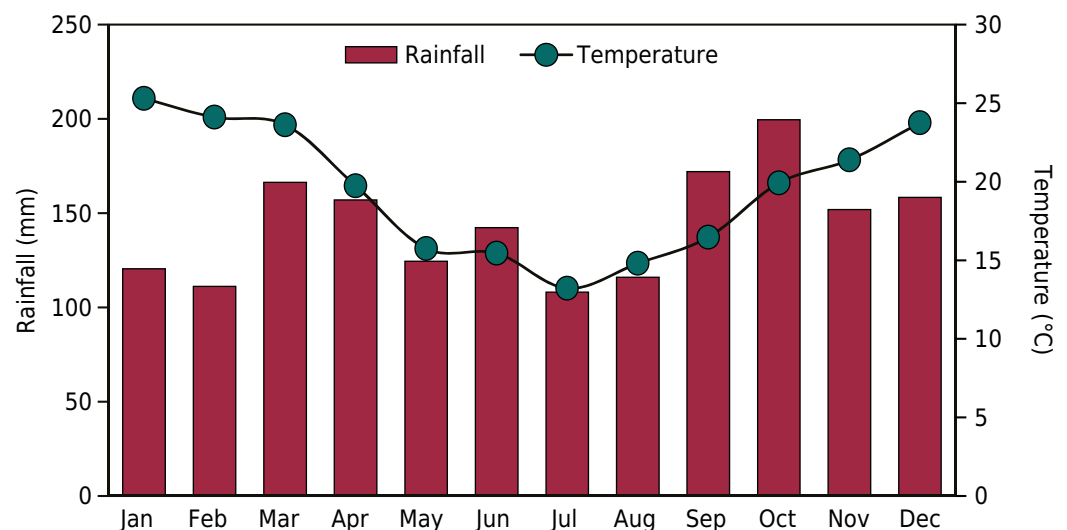


Figure 1. Monthly average temperature and rainfall in Santa Maria, Rio Grande do Sul, Southern Brazil.

The experimental area was cultivated in a no-tillage (NT) system since 1992 with annual crops and fertilization according to official recommendations (CQFS-RS/SC, 2016). In May 2000, the experiment began with the application of PS rates of 0, 20, 40, and 80 m³ ha⁻¹ on 12 m² plots (4 × 3 m). The experiment was arranged in a randomized block design with three replications and conducted until January 2008 under NT. Prior to the experiment, soil samples were taken between 0.00 to 0.10 m depth at different locations in the field and then mixed to blend a composite soil sample, and analyzed for physical and chemical properties. The background soil sample contained 170 g kg⁻¹ clay, 300 g kg⁻¹ silt, and 530 g kg⁻¹ sand (sandy-loam soil). The chemical properties were determined as follows: pH 4.7 (pH in water at a ratio of 1:1), 9.3 g kg⁻¹ of total organic carbon; 15 mg kg⁻¹ of available P, and 96 mg kg⁻¹ of available K (extracted by Mehlich-1); 0.8 cmol_c kg⁻¹ of Al³⁺, 2.7 cmol_c kg⁻¹ of Ca²⁺, and 1.1 cmol_c kg⁻¹ of Mg²⁺ (extracted by KCl 1.0 mol L⁻¹), and cation exchange capacity of 9.6 cmol_c kg⁻¹. The crop sequence, the dry matter and P content of the PS used in each crop, and the amount of P applied (kg ha⁻¹) per PS rate are shown in table 1.

Soil sampling and analysis

In January 2008, three bulk soil samples were collected from each plot at two layers: from 0.00-0.05 and 0.05-0.10 m. The soil samples were air-dried, ground, sieved through a 2-mm mesh, and stored at 4 °C before use. Approximately 200 g of soil was moistened to 80 % field capacity and incubated at 25 °C for 21 days to achieve the soil biological equilibrium, and then the P stored in the soil microbial biomass and acid phosphatase enzyme activity were analyzed.

Table 1. Cropping history, chemical composition of PS, and amount of P applied by PS at each application during experiment, and the total amount of dry matter (DM) applied by PS, and total amount of shoot yield in eight experimental years in Santa Maria, Rio Grande do Sul, Southern Brazil

Crop	Year	Dry matter	P ⁽³⁾	P contained in pig slurry		
				20 m ³ ha ⁻¹	40 m ³ ha ⁻¹	80 m ³ ha ⁻¹
		%	g kg ⁻¹	kg ha ⁻¹		
<i>Avena strigosa</i>	2000	1.9	24.0	9.0	18.0	36.0
<i>Zea mays</i>	2000/2001	9.9	21.0	42.0	84.0	168.0
<i>Raphanus sativus</i>	2001	0.8	41.0	3.4	6.8	13.6
<i>Avena strigosa</i>	2001	1.1	15.0	3.3	6.6	13.2
<i>Zea mays</i>	2001/2002	3.7	16.0	12.0	24.0	48.0
<i>Raphanus sativus</i>	2002	1.2	13.0	3.1	6.2	12.4
<i>Avena strigosa</i>	2002	0.5	21.2	2.0	4.0	8.0
<i>Pennisetum americanum</i>	2002	6.7	11.9	15.9	31.8	63.6
<i>Phaseolus vulgaris</i>	2002/2003	4.8	29.5	28.4	56.8	113.6
<i>Avena strigosa</i> + <i>Vicia sativa</i>	2003	2.4	34.1	16.2	32.4	64.8
<i>Zea mays</i>	2003/2004	2.1	33.3	13.8	27.6	55.2
<i>Avena strigosa</i> + <i>Vicia sativa</i>	2004	1.0	42.2	8.4	16.8	33.6
<i>Zea mays</i>	2004/2005	7.2	55.8	80.3	160.6	321.2
<i>Avena strigosa</i>	2005	6.8	29.5	40.1	80.2	160.4
<i>Pennisetum americanum</i>	2005	8.9	44.3	60.2	120.4	240.8
<i>Phaseolus vulgaris</i>	2005/2006	12.1	47.9	119.3	238.6	477.2
<i>Avena strigosa</i>	2006	9.9	53.7	106.7	213.4	426.8
<i>Zea mays</i>	2006/2007	1.9	51.8	19.8	39.6	79.2
<i>Avena strigosa</i>	2007	3.5	57.7	40.6	81.2	162.4
Total-P applied (kg ha ⁻¹)	2000-2007	-	-	624.5	1249.0	2498.0
Total DM applied by PS (Mg ha ⁻¹) ⁽¹⁾	2000-2007	-	-	17	35	69
Total shoot DM yield (Mg ha ⁻¹) ⁽¹⁾⁽²⁾	2000-2007	-	-	92	109	147

⁽¹⁾ Data from Lourenzi et al. (2014b). ⁽²⁾ Total shoot dry matter (DM) yield in the control treatment (0 m³ ha⁻¹ PS) was 58 Mg ha⁻¹. ⁽³⁾ Content of P in the PS dry matter.

In chemical analysis, soil pH was determined in distilled water with a glass electrode (pH in water at a ratio of 1:1 v/v), exchangeable Ca^{2+} , Mg^{2+} , and Al^{3+} extracted by KCl 1.0 mol L^{-1} (soil:extractant ratio of 1:20) whereas available K^{+} was extracted by Mehlich-1 (soil:extractant ratio of 1:10) as described by Tedesco et al. (1995). Exchangeable Al^{3+} in the KCl extract was determined by titrating with NaOH $0.0125 \text{ mol L}^{-1}$, Ca^{2+} and Mg^{2+} were determined by atomic absorption spectrophotometry and K^{+} by flame photometry. The potential acidity (H+Al) was calculated as a function of pH SMP (Kaminski et al., 2001). The Cu and Zn contents were determined by extraction with HCl 0.1 mol L^{-1} (CQFS-RS/SC, 2016). These soil chemical properties are presented in table 2.

Available P (Av-P) was estimated by extraction with Mehlich-1 (soil:extractant ratio of 1:10) and determined by the molybdenum-blue method (Murphy and Riley, 1962). The soil microbial biomass P (SMB-P) was estimated by the chloroform fumigation extraction method, after inorganic P extraction with anion exchange resin membrane (Hedley and Stewart, 1982). The P content stored in the soil microbial biomass (SMB) was calculated as proposed by Morel et al. (1996).

The acid phosphatase enzyme activity (P-ase) was estimated by measuring the release of *p*-nitrophenol from *p*-nitrophenyl phosphate, after exposure to soil in a modified universal buffer (MUB) at pH 6.5, as described by Tabatabai and Bremner (1969).

Organic P (Org-P) estimated by the ignition method was calculated by the difference between the amount of P extracted with $0.5 \text{ mol L}^{-1} \text{ H}_2\text{SO}_4$ from ignited ($550 \text{ }^\circ\text{C}$, 2h) and non-ignited soil samples (Olsen and Sommers, 1982). Total-phosphorus (Total-P) was estimated by digestion with H_2SO_4 and H_2O_2 in the presence of saturated MgCl_2 (Brookes and Powlson, 1981; Hedley et al., 1982). The content of inorganic P (Inorg-P) was obtained by the difference between the content of Total-P and Org-P estimated by the ignition method. The accumulation of P added via PS in inorganic form to the soil was calculated according to equation 1:

$$\text{Accumulation of inorganic P (\%)} = \frac{(\text{Inorganic } P_{PS} - \text{Inorganic } P_0)}{(\text{Total } P_{PS} - \text{Total } P_0)} \times 100 \quad \text{Eq. 1}$$

Table 2. Chemical properties of soil treated with 19 successive PS applications over 8 years, in Santa Maria, Rio Grande do Sul, Southern Brazil

Soil property	Soil layer							
	0.00-0.05 m				0.05-0.10 m			
	PS rate ($\text{m}^3 \text{ ha}^{-1}$)							
	0	20	40	80	0	20	40	80
pH(H_2O) ⁽¹⁾	5.2	5.4	5.4	5.5	5.4	5.3	5.3	5.5
Ca^{2+} ($\text{cmol}_c \text{ dm}^{-3}$) ⁽²⁾	3.4	3.9	4.7	5.2	3.0	3.0	3.6	4.1
Mg^{2+} ($\text{cmol}_c \text{ dm}^{-3}$) ⁽²⁾	2.2	2.7	3.3	4.1	1.8	2.1	2.7	3.9
Al^{3+} ($\text{cmol}_c \text{ dm}^{-3}$) ⁽²⁾	0.3	0.2	0.2	0.2	0.7	0.6	0.3	0.2
K^{+} ($\text{cmol}_c \text{ dm}^{-3}$) ⁽³⁾	0.2	0.2	0.2	0.3	0.0	0.1	0.1	0.2
H+Al ($\text{cmol}_c \text{ dm}^{-3}$)	4.4	4.5	4.8	4.5	5.3	5.2	4.8	4.7
Effective CEC ($\text{cmol}_c \text{ dm}^{-3}$)	6.1	7.0	8.3	9.8	5.4	5.8	6.7	8.3
Potential CEC ($\text{cmol}_c \text{ dm}^{-3}$)	10.2	11.3	13.0	14.1	10.1	10.3	11.1	12.7
Al saturation (%)	4.7	2.7	2.0	2.0	12.6	11.6	4.3	2.2
Base saturation (%)	57.1	60.0	62.9	68.2	47.4	49.6	57.8	63.4
Available Cu (mg kg^{-1}) ⁽⁴⁾	1.7	11.0	20.5	42.8	1.8	4.1	6.8	11.4
Available Zn (mg kg^{-1}) ⁽⁴⁾	5.3	16.9	35.1	79.7	2.0	6.0	9.5	20.6

⁽¹⁾ Soil:water ratio of 1:1. ⁽²⁾ Extracted by 1.0 mol L^{-1} KCl (soil: extractant ratio 1:20). ⁽³⁾ Extracted by Mehlich-1 (soil:extractant ratio 1:10). ⁽⁴⁾ Extracted by HCl 0.1 mol L^{-1} (soil:extractant ratio 1:10).

where P_0 is the P concentration in the soil in the control treatment ($0 \text{ m}^3 \text{ ha}^{-1}$ of PS) and P_{PS} is the P concentration in the soil treated with PS rates (20, 40, and $80 \text{ m}^3 \text{ ha}^{-1}$).

The total organic carbon (TOC) content was determined by wet oxidation with $\text{K}_2\text{Cr}_2\text{O}_7$ and H_2SO_4 (Walkley and Black, 1934), with modifications proposed by Tedesco et al. (1995), and the result was corrected to the TOC content obtained by dry combustion (elementary analyzer) using a correction factor (Rheinheimer et al., 2008a).

Statistical analyses

The data were submitted to analysis of variance (ANOVA). When treatment effects were significant at $p < 0.05$ by the F test, regression equations were adjusted for PS rates and the differences between soil layers were compared by the Tukey test at $p < 0.05$. We also performed Principal Component Analysis (PCA) using the variables: Av-P, SMB-P, P-ase, Org-P, Inorg-P, Total-P, and TOC. Later cluster analysis was performed using the method of k-means. All statistical analyses were performed using software R (R Core Team, 2013).

RESULTS

Successive PS applications altered the P content and forms and some biological and biochemical parameters of the P cycle in the soil (Table 3). The results indicated significant interactions for all forms of soil P with the factors PS rates (R) and soil depth (D) (Table 3). However, for the ratios SMB-P/Org-P and SMB-P/Total-P, no effect of interaction between the factors R and D was observed.

Pig slurry applications increased the levels of Av-P, SMB-P, P-ase, Inorg-P, Total-P, and TOC (Figure 2), and the ratios of Av-P/Total-P, SMB-P/Org-P, and P-ase/Org-P in both soil layers (Table 4). On the other hand, Org-P increased with increasing PS rates applied only on the soil surface layer (0.00-0.05 m) (Figure 2d), and SMB-P/Total-P ratio in the evaluated layers was not increased by PS applications (Table 4).

Highest increases were observed in the 0.00-0.05 m soil layer. Compared to the control treatment with application of $80 \text{ m}^3 \text{ ha}^{-1}$ of PS, the increases were: 5,523 % for Av-P; 308 % for SMB-P; 104 % for P-ase; 91 % for Org-P; 251 % for Inorg-P; 194 % for Total-P; and 56 % for TOC.

Table 3. Analysis of variance (ANOVA) for different P pools in response to different PS rates application (R), soil layers (D) and their interaction (R × D)

Variable	PS rate (R)	Soil depth (D)	R × D
Av-P	***	***	*
SMB-P	***	**	*
P-ase	***	**	***
Org-P	**	*	**
Inorg-P	***	*	*
Total-P	***	*	*
Av-P/Total-P	***	*	*
SMB-P/Org-P ratio	**	ns	ns
SMB-P/Total-P ratio	ns	ns	ns
P-ase/Org-P ratio	ns	*	*
Org-P/Total-P ratio	***	*	**
Inorg-P/Total-P ratio	***	***	**
TOC	**	**	**

Av-P: available P; SMB-P: microbial P; P-ase: acid phosphatase enzyme activity; Org-P: organic P; Inorg-P: inorganic P; Total-P: total-phosphorus; Av-P/Total-P: available P/total-phosphorus ratio; TOC: total organic carbon; * Significant at $p < 0.05$; ** : significant at $p < 0.01$; ***: significant at $p < 0.001$.

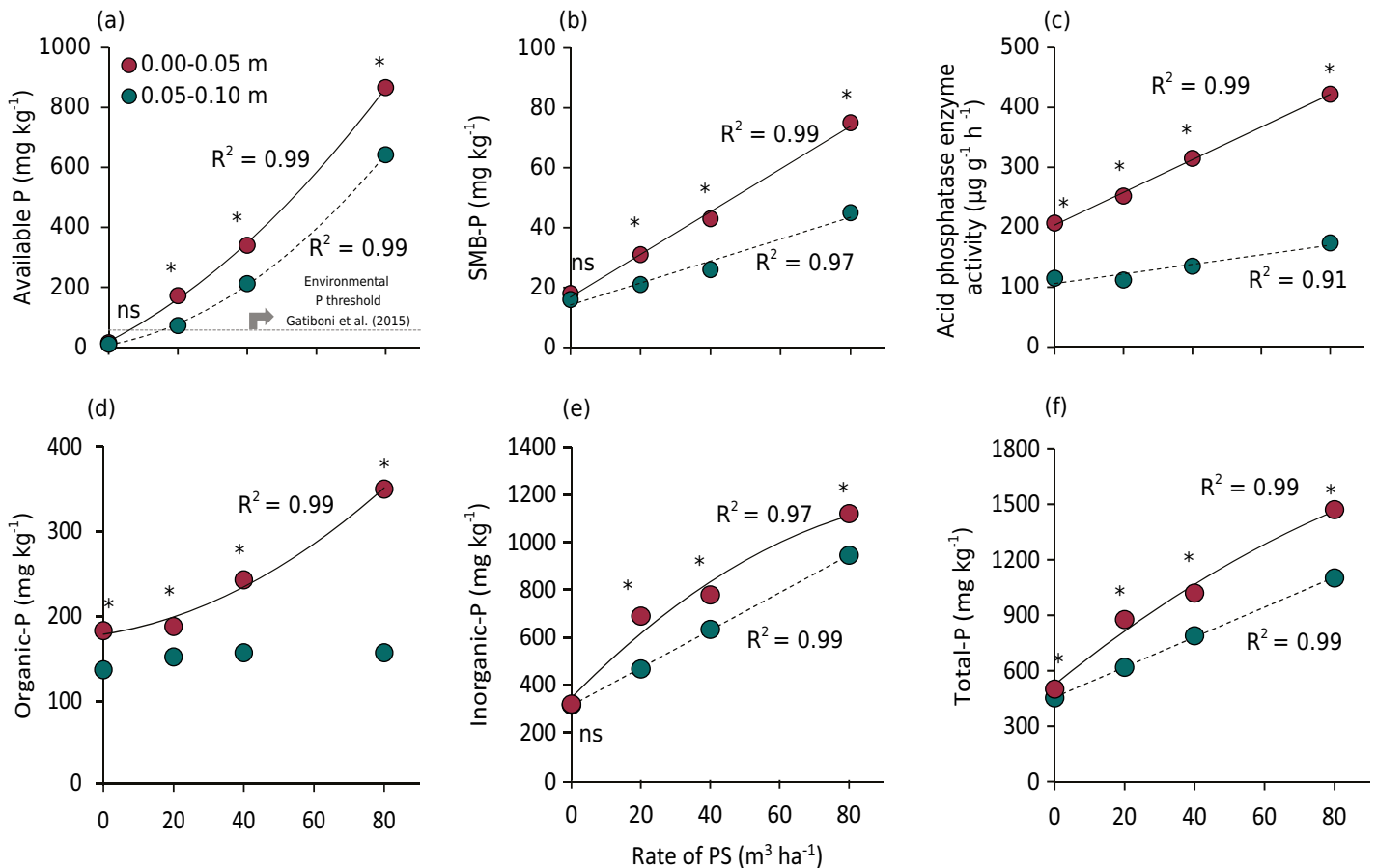


Figure 2. Levels of available P (a), SMB-P (b), acid phosphatase enzyme activity (c), organic P (d), inorganic P(e), and Total-P (f) in soil treated with 19 PS applications over 8 years, in Santa Maria, Rio Grande do Sul, Southern Brazil. Asterisks indicate significant difference ($p < 0.05$) between soil layers.

Phosphorus added as PS was mostly accumulated in the soil in inorganic form (83-98 % - Table 4). In this experiment, the increase in Total-P was followed by an increase in Inorg-P proportion and a decrease in Org-P proportion ($R^2 = 0.58$ - Figure 3a). However, the variation in the P accumulation form in the soil was better explained when taking into account the variation of TOC content ($R^2 = 0.87$ - Figure 3b), highlighting that the increase in Org-P was limited by the increase in organic carbon in the soil.

There were significant correlations ($p < 0.001$) among all variables in this study (Table 5), and the highest correlations ($R^2 > 0.8$) were found among P-ase, TOC, and Org-P, and among Av-P, Inorg-P, and Total-P. For SMB-P, the highest correlation was obtained with Av-P ($R^2 = 0.87$), and P-ase showed the highest correlation with TOC ($R^2 = 0.94$). The SMB-P content was less correlated with P-ase ($R^2 = 0.62$), Org-P ($R^2 = 0.56$), and TOC ($R^2 = 0.56$), and P-ase had a weaker correlation with Av-P ($R^2 = 0.57$) and Inorg-P ($R^2 = 0.45$).

Together, the first two principal components (Figure 4) explained 95.4 % of the total variation in both soil layers (0.00-0.05 and 0.05-0.10 m). The first principal component explained 84.4 % and the second principal component 10.9 % (Table 6). The contribution of variables in the first principal component was on average 14.3 ± 0.7 % for each variable. The variable contribution in the second principal component was 22.3 % due to TOC, 20.0 % due to Inorg-P, 18.3 % due to P-ase, and 16.4 % due to Org-P (Table 6).

Cluster analysis clearly formed three distinct groups (Figure 4). Group 1 contained the 0.00-0.05 m soil layer treated with $80 \text{ m}^3 \text{ ha}^{-1}$ PS. Group 2 comprised the surface soil layer treated with 20 and $40 \text{ m}^3 \text{ ha}^{-1}$ PS and the 0.05-0.10 m layer treated with $80 \text{ m}^3 \text{ ha}^{-1}$ PS. Group 3 comprised both soil layers of the control treatment and the 0.05-0.10 m soil layer treated with PS rates of 20 and $40 \text{ m}^3 \text{ ha}^{-1}$.

Table 4. Ratios of Av-P/Total-P, SMB-P/Total-P, SMB-P/Org-P, P-ase/Org-P, Org-P/Total-P, Inorg-P/Total-P, total organic carbon, and accumulation of P added via PS in inorganic form in soil treated with 19 PS applications over 8 years, in Santa Maria, Rio Grande do Sul, Southern Brazil

Soil layer	PS rate				Regression	R ²
	0	20	40	80		
m	m ³ ha ⁻¹					
	Av-P/Total-P ratio (%)					
0.00-0.05	2 a	9 a	17 a	24 a	$\hat{y} = 3.18 + 0.277x$	0.97
0.05-0.10	2 a	6 b	9 b	19 b	$\hat{y} = 1.68 + 0.215x$	0.99
	SMB-P/Total-P ratio (%)					
0.00-0.05	4 a	4 a	4 a	5 a	-	ns
0.05-0.10	3 a	3 a	3 a	4 a	-	ns
	SMB-P/Org-P ratio (%)					
0.00-0.05	10 a	17 a	18 a	22 a	$\hat{y} = 11.86 + 0.135x$	0.88
0.05-0.10	12 a	14 a	17 a	28 a	$\hat{y} = 10.06 + 0.215x$	0.95
	P-ase/Org-P ratio					
0.00-0.05	1.1 a	1.3 a	1.3 a	1.2 a	$\hat{y} = 1.151 + 0.0086x - 0.0001x^2$	0.80
0.05-0.10	0.8 b	0.7 b	0.9 b	1.1 a	$\hat{y} = 0.820 - 0.0037x - 0.00009x^2$	0.94
	Org-P/Total-P ratio (%)					
0.00-0.05	36 a	21 a	24 a	24 a	$\hat{y} = 34.83 - 0.572x + 0.0055x^2$	0.78
0.05-0.10	30 b	25 a	20 b	14 b	$\hat{y} = 29.12 - 0.194x$	0.97
	Inorg-P/Total-P ratio (%)					
0.00-0.05	64 b	79 a	76 a	76 b	$\hat{y} = 65.17 + 0.572x - 0.0055x^2$	0.78
0.05-0.10	70 a	76 a	80 a	86 a	$\hat{y} = 70.86 + 0.195x$	0.97
	Total organic carbon (g kg ⁻¹)					
0.00-0.05	15 a	17 a	18 a	23 a	$\hat{y} = 14.36 + 0.102x$	0.98
0.05-0.10	10 b	11 b	11 b	12 b	$\hat{y} = 10.12 + 0.027x$	0.96
	Accumulation of P added via PS in inorganic form (%)					
0.00-0.05	-	98	88	83	$\hat{y} = 101.34 - 0.247x$	0.89
0.05-0.10	-	92	95	97	$\hat{y} = 90.35 + 0.087x$	0.94

Means followed by the same letter in the same column comparing soil layer in each treatment are not significantly different at $p < 0.05$ by the Tukey test; ns: not significant. Av-P: available P; SMB-P: microbial P; P-ase: acid phosphatase enzyme activity; Org-P: organic P; Inorg-P: inorganic P; Total-P: total-phosphorus; Av-P/Total-P: available P/ total-phosphorus ratio.

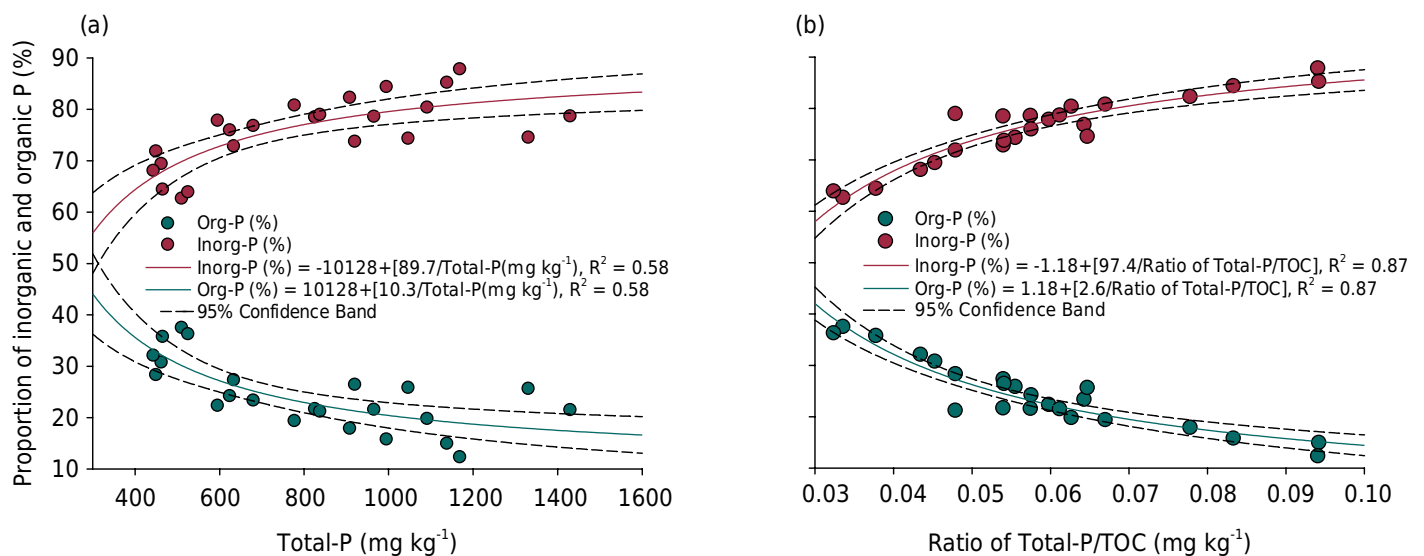


Figure 3. Effect of increase in the total-P content (a) and increase in the ratio total-P/TOC (b) in the soil and the proportion of inorganic and organic P.

DISCUSSION

Phosphorus application through PS to the soil surface at amounts exceeding the P crop uptake increased the P levels in all pools evaluated. Highest increases were observed in Av-P and Inorg-P, because between 60 % (Cassol et al., 2001) and 99 % (Tiecher et al., 2014) of P contained in PS is in inorganic forms. This increase was up to 12.5-fold in the Av-P/Total-P ratio in the 0.00-0.05 m soil layer where PS was applied at the maximum rate (Table 4). Moreover, it induced high linear correlation coefficients between Av-P × Total-P, Av-P × Inorg-P, and Inorg-P × Total-P (Table 5). Similar changes also occurred in the 0.05-0.10 m soil layer, but were less distinctive. This is because under repeated P soil application through PS, the ability of plants to absorb P from deeper soil layers and deposit it

Table 5. Linear correlation (r^2) coefficients between Av-P, SMB-P, P-ase, Org-P, Inorg-P, and Total-P in response to 19 successive PS applications over 8 years, in Santa Maria, Rio Grande do Sul, Southern Brazil

	Av-P	SMB-P	P-ase	Org-P	Inorg-P	Total-P
SMB-P	0.87*					
P-ase	0.57*	0.62*				
Org-P	0.55*	0.56*	0.81*			
Inor-P	0.88*	0.75*	0.45*	0.45*		
Total-P	0.90*	0.80*	0.57*	0.60*	0.98*	
TOC	0.52*	0.56*	0.94*	0.86*	0.44*	0.57*

Av-P: available P; SMB-P: microbial P; P-ase: acid phosphatase enzyme activity; Org-P: organic P; Inorg-P: inorganic P; Total-P: total-phosphorus; Av-P/Total-P: available P/total-phosphorus ratio; TOC: total organic carbon. * Significant at $p < 0.001$.

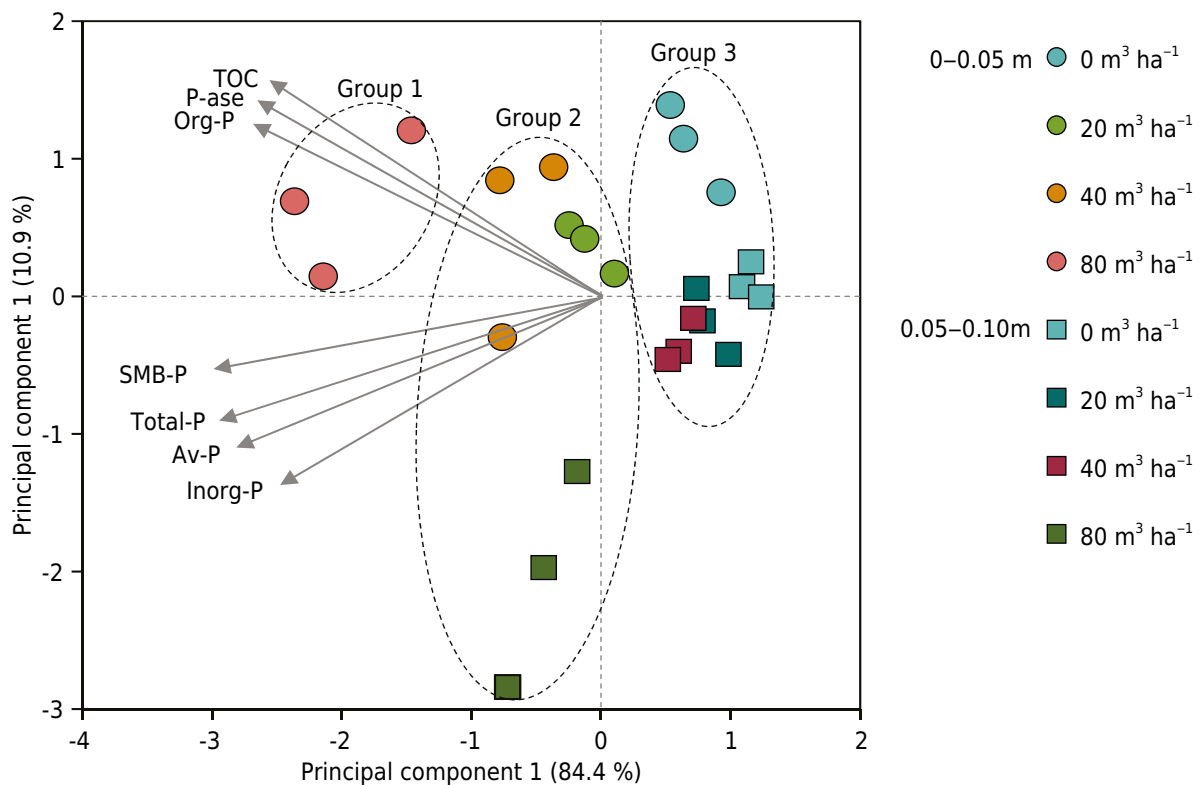


Figure 4. Projection of the variables related to the soil phosphorus cycle in the first factorial plan and principal component analysis of the 0.00-0.05 and 0.05-0.10 m layers of soil treated with 19 PS applications in 8 years, in Santa Maria, Rio Grande do Sul, Southern Brazil. Numbers in parentheses indicate percent of variation explained by each axis. Av-P: available phosphorus; SMB-P: microbial phosphorus; P-ase: acid phosphatase enzyme activity; Org-P: organic phosphorus; Inorg-P: inorganic phosphorus; Total-P: total-phosphorus; Av-P/Total-P: available phosphorus/total-phosphorus ratio; TOC: total organic carbon.

Table 6. Percentage explained by each principal component and percentage explained by each variable in each principal component. Bold numbers $>|0.7|$

Variable	Principal component 1	Principal component 2
Av-P	15.0	10.0
SMB-P	14.7	4.4
P-ase	13.9	18.3
Org-P	13.6	16.4
Inorg-P	13.8	20.0
Total-P	15.4	8.6
TOC	13.6	22.3
Explained variance (%)	84.4	10.9

Av-P: available P; SMB-P: microbial P; P-ase: acid phosphatase enzyme activity; Org-P: organic P; Inorg-P: inorganic P; Total-P: Total-P; Av-P/Total-P: available P/Total-P ratio; TOC: total organic carbon.

on the soil surface is exceeded by the transfer of P to the soil profile as a result of infiltration and biological activities. The increase in P in the 0.05-0.10 m layer varies according to the PS rates (Table 4). These results are similar to findings of Gatiboni et al. (2008b) for a Hapludalf under natural pasture, and of Guardini et al. (2012a,b) for a cultivated Hapludalf, treated with successive PS applications in Southern Brazil.

There are two other factors that increase the soil Av-P content after PS application. Firstly, P accumulation on the soil surface is a result of its absorption by plants in the deeper soil layers and its subsequent deposition on the soil surface through decomposition of plant residues (Rheinheimer and Anghinoni, 2001; Tiecher et al., 2012b). Secondly, the increased plant biomass production in treatments with PS application (Table 1) was coupled with the direct input of organic material by PS, which resulted in the increase of the TOC content of the soil (Table 4). Thus, the higher amount of low-molecular-weight organic acids released due to the increased TOC content may cause a competition effect with phosphate for the soil adsorption sites (Jones and Oburger, 2011). Moreover, the implementation of the NT system in subtropical regions has decreased soil erosion and changed the soil element dynamics, particularly those related to the carbon cycle (Mellek et al., 2010; Santos et al., 2011; Conceição et al., 2013).

In Southern Brazil, according to recommendations of CQFS-RS/SC (2016) for soils with a clay content of less than 20 %, the optimum amount of Mehlich-1 available P is 30 mg kg^{-1} for grain crops (e.g. soybean, maize, wheat). Below this range, there is a high probability of crop response to P addition. Only the treatment without PS application was below this limit (15.4 and 10.1 mg kg^{-1} in the 0.00-0.05 and 0.05-0.10 m soil layers - Figure 2a).

On the other hand, Gatiboni et al. (2015) proposed an environmental P threshold based on a simple model considering soil available P and clay content (P threshold in $\text{mg kg}^{-1} = 40 + \% \text{ of clay}$). Applying the threshold to this study, the maximum available P in the soil without causing environmental concerns is 57 mg kg^{-1} in the 0.00-0.10 m soil layer. The Av-P levels in the 0.00-0.10 m (considering the mean of the layers 0.00-0.05 and 0.05-0.10 m) in the treatments with PS applications of 20, 40, and $80 \text{ m}^3 \text{ ha}^{-1}$ are 2.1, 4.8, and 13.2 times higher, respectively, than this environmental critical level. In other words, even half of the rate normally applied by farmers ($20 \text{ m}^3 \text{ ha}^{-1}$) poses a high risk of environmental contamination with P. In subtropical regions such as Southern Brazil, the high precipitation often exceeds the capacity of water infiltration into the soil, resulting in runoff. However, according to Ceretta et al. (2010b) and Lourenzi et al. (2014a), if well managed, the runoff water from the soil surface under NT does not result in large soil losses by erosion, which would remove P as particulate P. Unfortunately, this is not the case of most fields in the state of Rio Grande do Sul, Southern Brazil, where cropland is unambiguously the main source delivering sediments into the rivers (Tiecher et al., 2015, 2016, 2017).

The increase in SMB-P observed with the increase in PS rates (Table 4) is a beneficial effect, for delaying phosphate adsorption by the soil, and after cell death and lysis, it releases P to the soil solution for plant uptake (Martinazzo et al., 2007; Gatiboni et al., 2008a). The results indicated a high linear correlation coefficient between SMB-P \times Av-P and SMB-P \times Total-P, because the soil microbial biomass is a dynamic compartment that immobilizes P when P availability increases in the soil solution (Rheinheimer et al., 2008b).

The higher amount of organic residues (Table 1) and TOC (Table 4) resulting from PS applications favors the growth and development of soil microbial biomass. In addition, the intensified development of plant roots grown in soil with PS application may also result in an increased exudation of organic compounds (Souza et al., 2010), which serve as energy and carbon source for the soil microbial community (Rheinheimer et al., 2000). According to Dakora and Phillips (2002), fungi and plants are mainly responsible for the production of acid phosphatase in the soil. In addition, the organic material protects the enzymes and keeps them in their active forms through the formation of complexes with humic compounds (Quiquampoix and Mousain, 2005). Thus, the increase in P-ase in treatments with higher PS rates may be attributed to the increased biological activity (plants and microorganisms) and the further protection of the enzyme by soil organic matter in both soil layers. Furthermore, the high correlation between P-ase, TOC, and Org-P, and the low correlation between P-ase and SMB-P shows that biochemical mineralization of the soil organic P forms is not a specific function of the soil microorganisms.

The P-ase generally increases with decreasing soil Av-P content, acting as a compensatory mechanism of the low soil P availability (Gatiboni et al., 2008a). However, in this study, P-ase increased even where Av-P was high (Table 4). In this context, several controversial findings are reported in the literature, stating the complexity of the relationship between acid phosphatase enzyme activity and soil available P. To some extent, this increase is natural, especially in view of the complex interaction of biological, chemical, and biochemical mobilization processes of P in soils (Jones and Oburger, 2011). In our experiment, the higher biological activity due to PS addition increased the acid phosphatase enzyme activity in the soil. In addition, the increase in P-ase, even at higher soil available P, may be due to the presence of extracellular phosphomonoesterases stabilized by the soil (Nannipieri et al., 2011).

There is little information about the factors that favor an increase in soil organic P, but it usually occurs in systems where there is an increase in TOC and Total-P, or when they are not considerably altered (Nziguheba and Bünemann, 2005). This trend is clearly shown (Figure 3) by the influence of TOC and Total-P on the pattern of P form distribution in the soil, and that the increase in Org-P was limited by the increase in soil organic carbon. Although Org-P in PS is low (McDowell and Stewart, 2005), the transformation of inorganic into organic P is evident in soils in well-managed NT systems, where soil TOC is increased. Under such conditions, inorganic P is incorporated from the soil solution into organic forms by plants and soil microorganisms (Shen et al., 2011). Thus, the increase in the soil Av-P content (Figure 2a) and in the deposition of organic residues (Table 1), associated with an intensified microbial activity in the soil surface, resulted in an approximately two-fold increase in the content of soil Org-P in the 0.00-0.05 m soil layer in the treatment with the highest PS rate, compared to the control. Despite the large increase in Total-P in both soil layers, the increase in Org-P content in the 0.05-0.10 m soil layer was less evident. This effect is attributed to the small increase in the TOC content in this soil layer, a fact that also explains the close relationship between these two variables (Table 5).

The preferential accumulation of P supplied by PS in inorganic forms (Table 4) was in agreement with previous studies in Southern Brazil (Gatiboni et al., 2008b; Ceretta et al., 2010a; Guardini et al., 2012a). This can be explained by the fact that P contained in PS is mostly in inorganic forms [up to 99 % according to Tiecher et al. (2014)] and the remaining

P in highly labile organic forms, which are easily mineralized after application to the soil. Therefore, P applied in PS has the same pattern of adsorption by soil inorganic colloids as that of P applied in mineral fertilizers, with preferential accumulation in inorganic forms.

The combined analysis of the variables by Principal Component Analysis shows that the soil P dynamics are changed mainly in the soil surface layer and with increasing amounts of P applied by PS (Figure 4). The variables related to the P cycle in the treatment with highest PS rate were substantially altered, even in the 0.05-0.10 m layer. Furthermore, the loadings of the first principal component (explaining 84.4 % of the variation in the experiment) were very similar among all variables, indicating that all variables were equally impacted by PS application and also that the evaluated variables are strongly related with each other, as also shown by the correlation among all variables (Table 5). The figure 4 also shows two clusters of variables; the first is related to TOC, Org-P, and P-ase, and shows that P-ase depends on TOC and Org-P in soil under successive PS application. The second one is related to Av-P, Inorg-P, Total-P, and SMB-P, and shows that SMB-P depends on the availability of inorganic P forms.

CONCLUSIONS

Phosphorus applied in pig slurry is preferentially accumulated in the soil in inorganic form (83-98 %). The increase in organic P was limited by the increase in soil organic carbon. The increase in pig slurry rates also led to an increase in the levels of available P, soil microbial biomass-P content, and even in the acid phosphatase enzyme activity. These results show that pig slurry application to the soil affects the chemical, biological, and biochemical components of the soil P cycle. The increase in soil microbial biomass-P was more influenced by available P, while the increase in phosphatase enzyme activity was more related to the increase in organic P and C. Moreover, results indicate that even half of the pig slurry rates normally applied by farmers ($20 \text{ m}^3 \text{ ha}^{-1}$) poses a high risk of pollution of water bodies, due to runoff from the soil.

ACKNOWLEDGMENTS

The authors express their gratitude to the National Council for Scientific and Technological Development (CNPq) for financial support, and to the Coordination for the Improvement of Higher Education Personnel (CAPES), for supporting the first author with a doctoral scholarship.

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