

# Swine manure on the control of damping-off in beetroot seedlings

Ivã Tavares Butrinowski<sup>1,2</sup>, Rosangela Dallemole-Giaretta<sup>2</sup>, Idalmir dos Santos<sup>2</sup>,  
Betania Brum De Bortoli<sup>2</sup>, Paula Steilmann<sup>2</sup>, Rafael Baldin<sup>2</sup>

<sup>1</sup> This study is part of the Master's dissertation of the first author. <sup>2</sup>Universidade Tecnológica Federal do Paraná (UTFPR), Câmpus Pato Branco (Câmpus PB), Programa de Pós-Graduação em Agronomia (PPGAG). Via do Conhecimento, s/n, KM 01, Fraron, CEP 85503-390, Pato Branco, PR, Brasil.

Corresponding author: Paula Steilmann (paulasteilmann@gmail.com)

Data de chegada: 10/09/2018. Aceito para publicação em: 15/05/2019

10.1590/0100-5405/213796

## ABSTRACT

Butrinowski, I.T.; Dallemole-Giaretta, R.; Santos, I.; De Bortoli, B.B.; Steilmann, P.; Baldin, R.. Swine manure on the control of damping-off in beetroot seedlings. *Summa Phytopathologica*, v.45, n.4, p.393-398, 2019.

The aim of this study was to evaluate the effect of swine manure (SM) doses applied to soils showing pH of 4.8 and 7.2 on the control of damping-off in beetroot seedlings caused by *Rhizoctonia solani*. To set the trial, plastic bags were filled with 4 kg soil (pH levels of 4.8 and 7.2) and 15 g *R. solani* inoculum kg soil<sup>-1</sup>. This mixture was moistened, homogenized and kept in a greenhouse for seven days. Following this period, SM doses (0, 5, 10, 15 and 20%) were incorporated into the bags, which were again stored in a greenhouse. After seven days, part of the soil from each plastic bag was separately placed into 16 cells of a 128-cell polystyrene tray, and two beetroot seeds were

sown per cell. Seedling emergence and damping-off were evaluated for 21 consecutive days. The other part of the soil was used for microbial activity quantification based on the CO<sub>2</sub> release method at 7, 14 and 21 days. The trial was conducted in a completely randomized design, with four replicates per treatment, and repeated twice. All tested SM doses reduced the number of damped-off beetroot seedlings in both trials, and the greatest disease control was provided by treatments that had SM doses of 15% and 20% applied to soil showing pH level of 7.2. In addition, regardless of the pH level, all tested SM doses increased soil microbial activity.

**Keywords:** soil borne pathogen, organic matter, *Beta vulgaris*.

## RESUMO

Butrinowski, I.T.; Dallemole-Giaretta, R.; Santos, I.; De Bortoli, B.B.; Steilmann, P.; Baldin, R.. Chorume de suíno no controle de tombamento de plântulas em beterraba. *Summa Phytopathologica*, v.45, n.4, p.393-398, 2019.

O objetivo deste estudo foi avaliar o efeito de doses de chorume de suíno (CS), aplicadas em solos com pH 4,8 e 7,2, no controle do tombamento de plântulas de beterraba causado por *Rhizoctonia solani*. Para montagem do ensaio foram acondicionados em sacos plásticos, 4 g de solo (pH 4,8 e 7,2) e 15 g de inóculo de *R. solani* kg de solo<sup>-1</sup>. Em seguida, essa mistura foi umedecida, homogeneizada e mantida em casa de vegetação por sete dias. Após esse período, em cada saco foram incorporadas, as doses do CS (0, 5, 10, 15 e 20%), sendo os sacos novamente armazenados em casa de vegetação. Após sete dias, parte do solo de cada saco plástico foi acondicionada separadamente em 16 células de bandeja de isopor de 128

células, e semeadas duas sementes de beterraba por célula. A emergência e o tombamento das plântulas foram avaliados por 21 dias consecutivos. A outra parte do solo foi utilizada para quantificação da atividade microbiana, por meio do método de desprendimento de CO<sub>2</sub>, aos 7, 14 e 21 dias. O ensaio foi conduzido em delineamento inteiramente casualizado, com quatro repetições por tratamento, e repetido duas vezes. Todas as concentrações de CS testadas reduziram o número de plântulas de beterraba tombadas, em ambos os ensaios, tendo o maior controle da doença sido obtido nos tratamentos com as doses 15 e 20% de CS, em solo com pH 7,2. Além disso, independentemente do pH, todas as doses de CS testadas aumentaram a atividade microbiana do solo.

**Palavras-chave:** Patógeno habitante de solo, matéria orgânica, *Beta vulgaris*.

*Rhizoctonia solani* Kunh is a soil borne necrotrophic pathogen that causes damping-off and root and collar rot in adult plants of numerous crops of economic importance (1). In addition, diseases caused by this fungus are difficult to control due to its high saprophytic ability and sclerotium production in the soil (1).

Incorporation of organic compounds into the soil is an alternative method for the management of soilborne pathogens (3). Potential organic compounds include products derived from composted plants (6), composted or non-composted forest substrates (7) and swine manure (5, 13).

Swine manure (SM) is used in agriculture as a source of nutrients

and organic matter. Its application for plant disease control is rarely studied; however, SM is known to have potential action in the management of soil borne pathogens due to the release of volatile compounds such as fatty acids, nitrous acid and ammonia (5, 20).

Considering the socioeconomic and environmental importance of the final destination of SM, its contribution to the control of soil borne pathogens and the few studies on its application to soils of different pH levels to control damping-off caused by *R. solani*, this study evaluated the effect of different SM doses applied to soils showing pH levels of 4.8 and 7.2 on the control of rhizoctoniosis in beetroots under greenhouse conditions.

## MATERIAL AND METHODS

### Inoculation of the fungus

To obtain *R. solani* inoculum, ten disks of seven-day-old culture of the pathogen (5 mm diameter), previously grown in potato-sucrose-agar medium (PSA) at  $\pm 24$  °C, were placed in a glass Erlenmeyer flask containing 150 g rice and 200 ml distilled water, which were previously autoclaved for 20 minutes at 120 °C. The flasks were kept in a growth chamber at  $\pm 24$  °C for 15 days. Then, the substrate was removed from the Erlenmeyer flasks and allowed to dry on plastic trays (35 x 20 cm), at room temperature ( $\sim 24$  °C), for eight days. After drying, the rice substrate containing the fungus was ground in a blender (Britania®, slow speed) for 2 minutes and stored in flasks kept in the dark, at 10 °C, until use.

### Obtaining the soil

The soil used in the experiments was collected from an area under no-tillage, at a depth of 0 to 20 cm, and classified as “Latossolo Vermelho Distroférico”, according to Embrapa classification (9). Chemical analysis of the soil showed the following values: pH in water of 4.8, organic matter of 50.93 g.dm<sup>-3</sup>, nutrients: Mg (2.0 cmol<sup>(+)</sup>.dm<sup>-3</sup>, K (0.35 cmol<sup>(+)</sup>.dm<sup>-3</sup>, P (3.28 mg).dm<sup>-3</sup>, Cu (3.15 mg).dm<sup>-3</sup> and Zn (4.46 mg).dm<sup>-3</sup>, and soil base saturation of 59%, according to the methodology described by Pavan et al. (17).

### Swine manure

Swine manure (SM) was obtained from “Universidade Tecnológica Federal do Paraná”, located in Dois Vizinhos, Paraná State, Brazil. The chemical components of SM were analysed according to the method described by Embrapa (8). The following nutrient values were found in the sample: nitrogen (N) 5.30%, phosphorus (P) 0.75%, potassium (K) 0.51%, calcium (Ca) 1.72%, magnesium 0.46% and dry matter (Ms) 7%.

### In vivo test

The assay was conducted in a greenhouse, in a completely randomised 2 x 5 bi-factorial design, using two soil pH levels (4.8 and 7.2) x five SM doses (0, 5, 10, 15 and 20%, calculated as a function of total soil weight per treatment), and four replicates per treatment.

To adjust the soil pH to 7.2, for each treatment, 5 kg sieved soil and 50 g dolomitic limestone kg<sup>-1</sup> soil were added to a 50-L plastic bag. Before the assay was set up, the pH of the soil was tested again to confirm that it had the desired pH (4.8 and 7.2).

To set the trial, 4 kg dry unsterilized soil (pH 4.8 and 7.2), 15 g *R. solani* inoculum kg<sup>-1</sup> soil and 200 mL water were placed, separately, in 50-L black plastic bags. Then, they were manually homogenized and the mixture was stored in a greenhouse at  $\pm 25$  °C.

After seven days, the respective SM doses were added to separate plastic bags, manually homogenized and stored again in a greenhouse for seven days. Then, half of the soil of the respective treatments was placed in polystyrene trays containing 128 cells, of which 16 cells represented one experimental unit. In each cell, two seeds (monogerm) of ‘Katrina’ beetroot were sown and irrigated when needed.

Emergence and damping-off were daily evaluated for 21 days, and their percentages were calculated based on the number of seeds that were sown and the number of seedlings exhibiting damping-off.

The other half of the soil was placed in separate 1-kg pots and sown with the beetroot seeds to quantify microbial activity based on CO<sub>2</sub> release, according to the methodology described by Grisi (11). Thus, 100 g soil was collected from each pot at 7, 14 and 21 days after the

incorporation of SM. Each soil sample was sieved and placed in separate 2-L plastic pots. A Petri dish containing 10 mL potassium hydroxide (KOH), at 0.5 normal (N), was deposited on the soil of each pot. The pots were hermetically sealed to prevent both air ingress and egress and stored in the dark, for 15 days, at 24 °C. Two pots containing only Petri dishes with KOH left under the same conditions were used as control. The trial was conducted in a completely randomised design, using four replicates per treatment.

Subsequently, the KOH in the dishes was titrated with 0.1 Mol hydrochloric acid (HCl), using phenolphthalein and methyl orange as indicators, two drops/sample, respectively. The obtained results were employed in the following formula: (Treatment - Control) \* Molecular weight CO<sub>2</sub> \* mol HCL, yielding the quantity of mg CO<sub>2</sub>/100g soil. Both experiments were repeated twice.

### Statistical analysis

The obtained data were verified according to analysis of variance and transformed if necessary. Analysis of variance was performed at 5% probability of error. If an interaction between factors was significant, regression analysis of SM doses was carried out for each pH level (4.8 and 7.2); in the absence of interaction, F test was used to verify the difference between the means of levels, while polynomial regression analysis was adopted for SM doses.

## RESULTS

There was a significant interaction between pH levels x SM doses for beetroot seedling emergence in both trials (Tables 1 and 2). In the first trial, emergence percentage was highest for SM doses of 6.16% and 5.65% in soil showing pH of 4.8 and 7.2, respectively: 97.41% and 96.85% beetroot seedling emergence. On the other hand, the highest SM dose reduced beetroot seedling emergence by more than 45% at both pH levels (Figure 1A). In the second trial, SM doses of 10.18% and 6.49% in soil showing pH of 4.8 and 7.2, respectively, resulted in maximum beetroot seedling emergence: higher than 99.40% (Figure 2A).

For the variable damping-off, there was no significant interaction between pH levels x SM doses. However, a difference between crops was observed for the isolated factors (Tables 1 and 2). In trial I, the lowest percentage of beetroot seedling damping-off (17.37%) was obtained for the SM dose of 13.56% (Figure 1B). In trial II, the fit of the equation was linear, and beetroot seedling damping-off decreased from SM dose of 10% (Figure 2B).

Considering the different pH levels, soil showing pH of 7.2 had 21.84% and 16.56% less beetroot seedling damping-off in trials I and II, respectively, compared to soil showing pH of 4.8 (Table 3).

For microbial activity, the factor SM dose was only significant at 7 and 14 days (trials I and II) and at 21 days (trial II) (Tables 1 and 2). In trial I, microbial activity (8.42 mg CO<sub>2</sub>/100 g soil) was maximal for SM dose of 13.64% at 7 days and (8.85 mg CO<sub>2</sub>/100 g soil) for SM dose of 12.24% at 14 days (Figure 1C and 1D).

However, in the second trial, there was a significant interaction between factors (pH x SM dose) at 7 days, when all tested SM doses, regardless of the soil pH, increased microbial activity. SM doses of 17.14% and 13.20% led to CO<sub>2</sub> releases of 8.43 and 8.52 mg CO<sub>2</sub>/100 g soil at pH levels of 4.8 and 7.2, respectively (Figure 2C). At 14 days, the highest microbial activity (8.89 mg CO<sub>2</sub>/100 g soil) was obtained for SM dose of 13.1% applied to the soil (Figure 2D). At 21 days, SM dose of 12.50% had the best result for microbial activity (8.73 mg CO<sub>2</sub>/100 g soil) (Figure 2E).

**Table 1.** Degrees of freedom (DF) and mean squares of analysis of variance for the variables: emergence (%) and damping-off (%) of beetroot seedlings, and microbial respiration at 7, 14 and 21 days after the incorporation - DAI (mg CO<sub>2</sub>/100 g of soil), in a bi-factorial trial, using two soil pH levels (4.8 and 7.2) and five swine manure (SM) doses (0, 5, 10, 15 and 20%) for the first trial conducted in completely randomised design.

Causes of variation	DF	Mean squares		GL	Mean squares		
		Emergence <sup>1</sup>	Damping-off		Microbial respiration (mg CO <sub>2</sub> 100g <sup>-1</sup> soil <sup>-2</sup> )		
					7 DAI	14 DAI	21 DAI
pH	1	0.0006 <sup>ns</sup>	466.8989*	1	0.9118 <sup>ns</sup>	0.8875 <sup>ns</sup>	0.0975 <sup>ns</sup>
Swine manure dose	4	0.5060*	1266.3326*	4	7.1475*	2.4151*	0.5883 <sup>ns</sup>
pH x SM dose	4	0.0479*	33.3858 <sup>ns</sup>	4	0.7316 <sup>ns</sup>	0.1124 <sup>ns</sup>	0.0990 <sup>ns</sup>
Residual	30	0.0169	28.3752	20	0.3324	0.3739	0.2082
Overall mean		81.25	27.85		7.63	8.28	8.36
C.V. (%)		11.00	19.12		7.56	7.38	5.45

<sup>1</sup> Arcosene transformation. \* Significant at 5% probability of error. <sup>ns</sup> Not significant at 5% probability of error.

**Table 2.** Degrees of freedom (DF) and mean squares of analysis of variance for the variables: emergence (%), damping-off (%) of beetroot seedlings and microbial respiration at 7, 14 and 21 days after the incorporation - DAI (mg CO<sub>2</sub>/100 g of soil), in a bi-factorial trial, using two soil pH levels (4.8 and 7.2) and five swine manure (SM) doses (0, 5, 10, 15 and 20%) for the second trial conducted in completely randomised design.

Causes of variation	DF	Mean squares		DF	Mean squares		
		Emergence <sup>1</sup>	Damping-off		Microbial respiration (mg CO <sub>2</sub> 100g <sup>-1</sup> soil <sup>-2</sup> )		
					7 DAI	14 DAI	21 DAI
pH	1	0.0000 <sup>ns</sup>	161.8855*	1	0.4060 <sup>ns</sup>	1.3062 <sup>ns</sup>	0.0740 <sup>ns</sup>
Swine manure dose	4	0.00017*	501.9845*	4	6.7578*	4.3314*	30.524*
pH x SM dose	4	0.00005*	58.1467 <sup>ns</sup>	4	1.2317*	0.3674 <sup>ns</sup>	0.3529 <sup>ns</sup>
Residual	30	0.00001	31.5516	20	0.3133	0.4040	0.2272
Overall mean		93.59	22.32		7.56	8.12	8.14
C.V. (%)		3.12	25.17		7.40	7.83	5.86

<sup>1</sup> Arcosene transformation. \* Significant at 5% probability of error. <sup>ns</sup> Not significant at 5% probability of error.

**Table 3.** Comparison of means of the soil pH levels for the variable (%) damping-off in beetroot seedlings (Trial I and II), in a bi-factorial experiment, using two soil pH levels (4.8 and 7.2) and five swine manure (SM) doses (0, 5, 10, 15 and 20%), conducted in a completely randomised design.

Soil pH	% Damping-off	
	Trial I	Trial II
pH 4.8	31.27 a*	24.33 a
pH 7.2	24.44 b	20.30 b

\* Means not followed by the same letter, in the column, differ from each other at 5% probability of error, according to F test.

## DISCUSSION

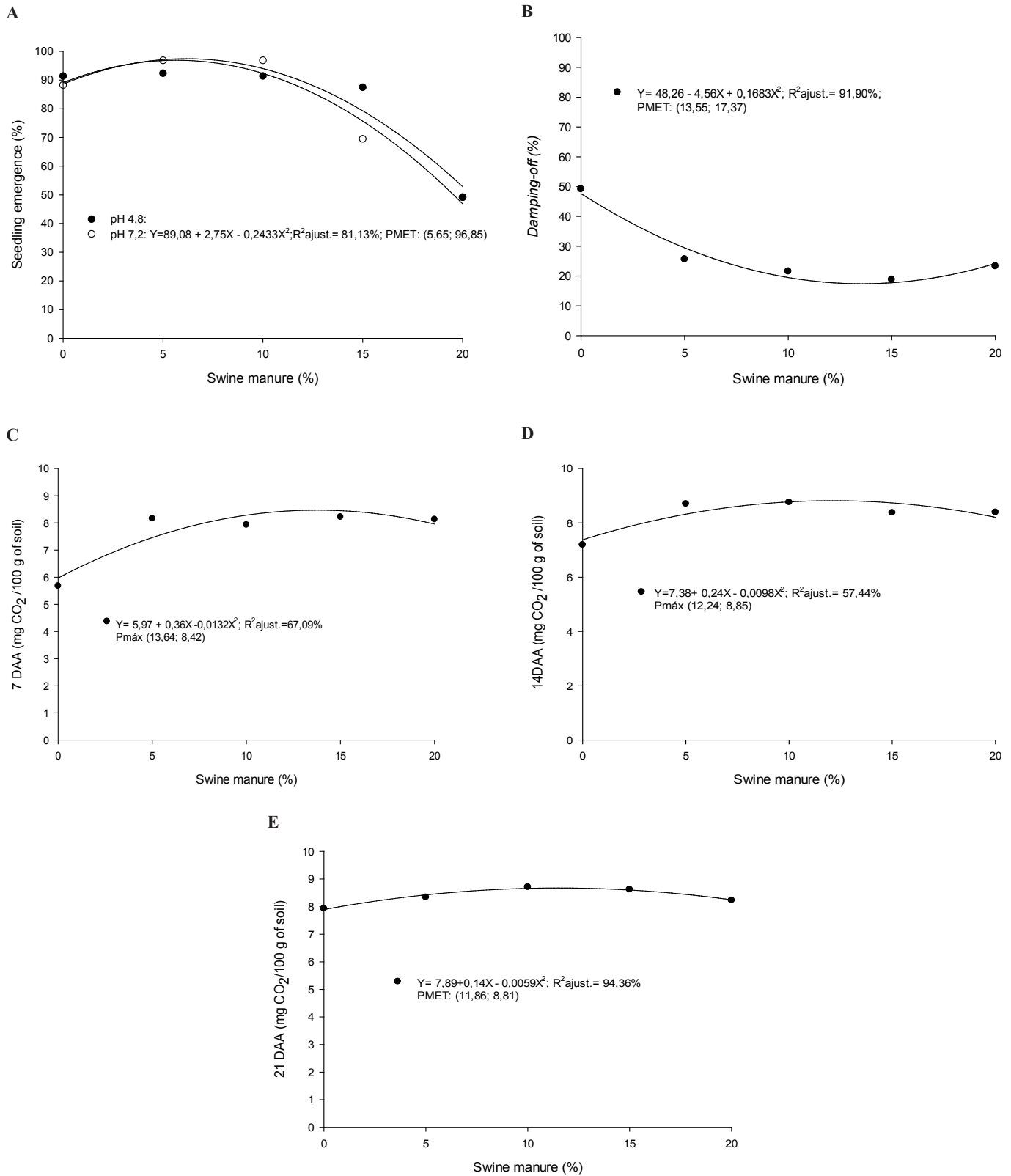
Addition of SM to the soil controlled damping-off caused by *R. solani* in beetroots and increased microbial activity in the soil; besides, SM doses up to 15% did not affect beetroot seedling emergence.

Swine manure (SM) is an organic compound rich in nitrogen, which leads to release of ammonia when added to soil showing alkaline pH (5, 13). On the other hand, in soils showing acidic pH, it leads to the release of volatile fatty acids, especially acetic acid, propionic acid and

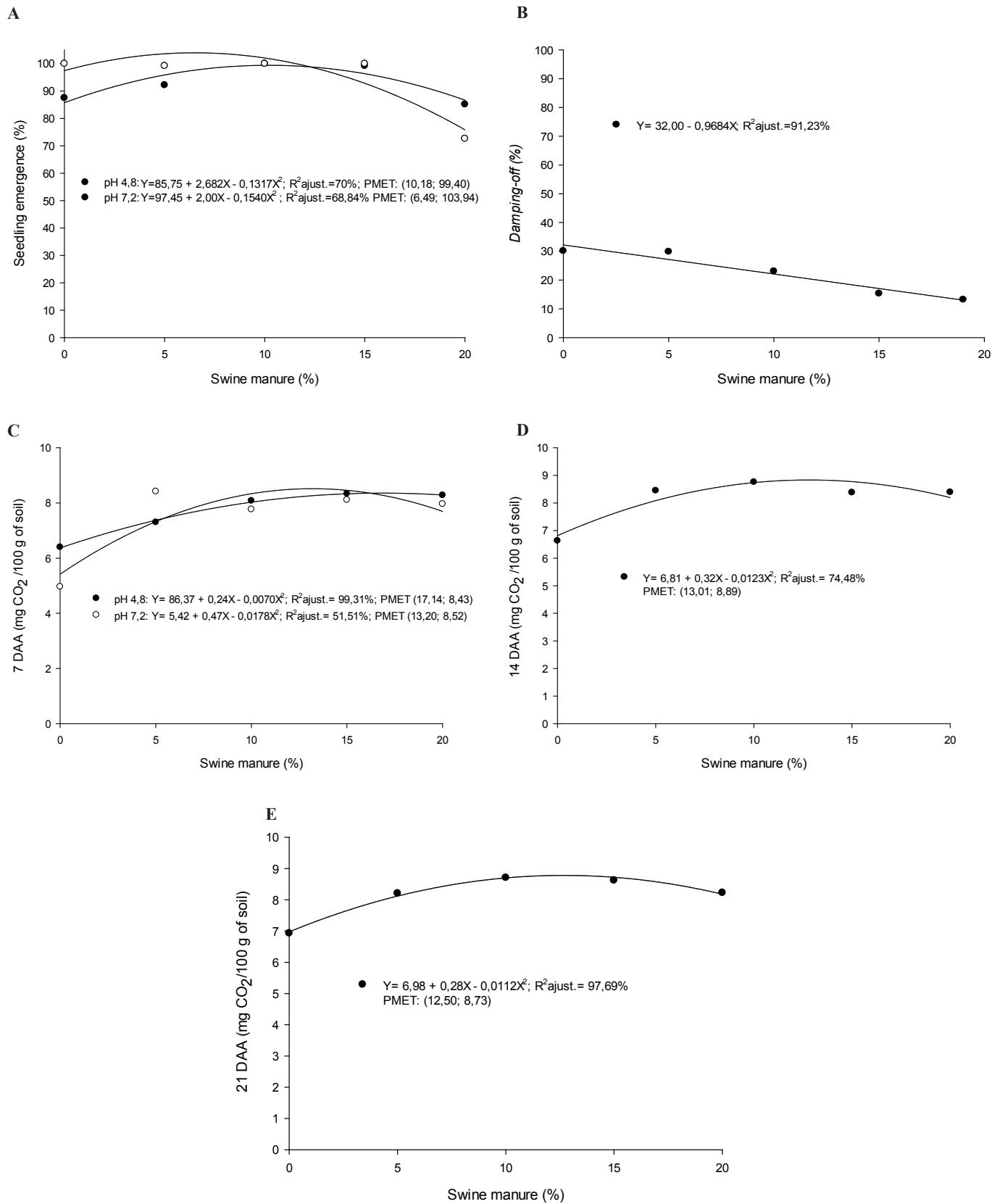
isobutyric acid (19). These compounds were responsible for reducing the damping-off caused by *R. solani* in the beetroots of the present study when the highest SM doses were added to the soil, particularly soil showing pH of 7 (Figures 1 and 2; Table 3). Antifungal activity of SM was also reported against the pathogens *Verticillium dahlia* Kleb (4, 5), *Sclerotium rolfsii* Sacc (16) and *Pythium aphanidermatum* (Edson) Fitzp (15).

Swine manure (SM) also stimulated microbial activity in the soil, especially when applied at higher doses (Figure 1C, 1D and Figure 2C, 2D and 2E). The increased microbial activity promoted antagonism among the microorganisms present in the soil, which is another factor responsible for reducing the damping-off caused by *R. solani* in the beetroot seedlings of the present study (Figures 1B and 2B). Antagonistic effect on this pathogen is reported for numerous microorganisms such as actinomycetes (3), *Pseudomonas fluresces* (21), *Bacillus subtilis* (12) and *Trichoderma* sp. (14).

Therefore, based on these results, SM can be employed by farmers in the management of areas infested with this pathogen. In small farms, in areas cultivated with vegetables, the farmer can apply SM with watering cans or hoses coupled to reservoirs. However, in larger areas, pig SM can be applied to the soil with a liquid manure distributor coupled to a tractor or tanker truck. Thus, repeated SM application to the soil enhances the control of soil borne pathogens by multiple



**Figure 1.** Emergence (%) of seedlings (A), damping-off (%) of seedlings (B), microbial respiration (mg CO<sub>2</sub>/100 g of soil) at 7 (C), 14 (D) and 21 days after swine manure incorporation - DAI (E), using five swine manure doses (0, 5, 10, 15 and 20%) and two soil pH levels (pH 4.8 and 7.2) for the first trial conducted in completely randomised design.



**Figure 2.** Emergence (%) of seedlings (A), damping-off (%) of seedlings (B), microbial respiration (mg CO<sub>2</sub>/100 g of soil) at 7 (C), 14 (D) and 21 days after swine manure incorporation - DAI (E), using five swine manure doses (0, 5, 10, 15 and 20%) and two soil pH levels (pH 4.8 and 7.2) for the second trial conducted in a completely randomised design.

mechanisms of action.

On the other hand, although the highest SM doses applied to the soil were more efficient in reducing beetroot seedling damping-off, there was a reduction in seedling emergence in both trials (Figure 1A and 2A). Similar results were obtained by Manteli (15) for cucumber when doses higher than 10% were added to soils showing pH levels of 4.8 and 6.3, in the management of *P. aphanidermatum*, under greenhouse conditions. The reduced emergency observed in this study may have been due to the excess K (potassium) and N (nitrogen) present in the used SM (5.30% N and 0.51% K), which consequently decreased water absorption, germination of seeds, and development of seedlings. Salt excess may impair plant growth (2, 10, 18). In addition, the reduction in beetroot seedling emergence was also due to the surface sealing of the soil. In addition, the reduction in beetroot seedling emergence was also due to the surface sealing of the soil.

Based on the obtained results, application of SM at doses up to 10%, regardless of the soil pH, is efficient for rhizoctoniosis management, without affecting beetroot seedling emergence. Swine manure (SM) was not only a viable option for rhizoctoniosis control but also an inexpensive and easily accessible product, especially for regions where pig farming is predominant.

#### ACKNOWLEDGMENTS

The authors would like to thank “Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES)” for granting a scholarship to the first author.

#### REFERENCES

1. Amorin, L.; Rezende, J.A.M.; Bergamin Filho, A.; Camargo, L.E.A. **Manual de fitopatologia: doenças das plantas cultivadas**. 5.ed. Ouro Fino: Agronômica Ceres, 2016. v.2, 772p.
2. Andréo-Souza, Y.; Pereira, A.L.; Da Silva, F.F.S.; Riebeiro-Reis, R.C.; Evangelista, M.R.V.; De Castro, R.D.; Dantas, B.F. Efeito da salinidade na germinação de sementes e no crescimento inicial de mudas de pinhão-manso. **Revista Brasileira de Sementes**, Londrina, v.32, n.2 p.83-92, 2010. Available at: <<http://dx.doi.org/10.1590/S0101-31222010000200010>>. Accessed on: 22 ago. 2018.
3. Ascencion, L.C.; Liang, W.J.; Yen, T.B. Control of *Rhizoctonia solani* damping-off disease after soil amendment with dry tissues of *Brassica* results from increase in Actinomycetes population. **Biological Control**, San Diego, v.82, p.21-30, 2015. Available at: <<https://doi.org/10.1016/j.biocontrol.2014.11.010>>. Accessed on: 22 ago. 2018.
4. Conn, K.L.; Lazarovits, G. Soil factors influencing the efficacy of liquid swine manure added to soil to kill *Verticillium dahliae*. **Canadian Journal of Plant Pathology**, Canada, v.22, n.4 p.400-406, 2000. Available at: <<https://doi.org/10.1080/07060660009500459>>. Accessed on: 22 ago. 2018.
5. Conn, K.L.; Tenuta, M.; Lazarovits, G. Liquid swine manure can kill *Verticillium dahliae* microsclerotia in soil by volatile fatty acid, nitrous acid, and ammonia toxicity. **Phytopathology**, Ithaca, v.95, n.1, p.28-35, 2005. Available at: <<https://doi.org/10.1094/PHYTO-95-0028>>. Accessed on: 22 ago. 2018.
6. De Corato, U.; Viola, E.; Arcieri, G.; Valerio, V.; Zimbardi, F. Use of composted agro-energy co-products and agricultural residues against soil-borne pathogens in horticultural soil-less systems. **Scientia Horticulturae**, Amsterdam, v.210, n.10, p.166-179, 2016. Available at: <<https://doi.org/10.1016/j.scienta.2016.07.027>>. Accessed on: 22 ago. 2018.
7. Díez, R.M.A.; López Pérez, J.Á.; Terrón, P.U.; Pérez, A.B. **Biodesinfecção de solos y manejo agronômico**. Espana: Ministerio de Medio Ambiente e Medio Rural e Marino, 2010. 52p.
8. Embrapa. **Manual de análises químicas de solo, plantas e fertilizantes / editor técnico, Fábio César da Silva**. 2. ed. rev. ampl., Brasília, DF: Embrapa Informação Tecnológica, 2009. 627p.
9. Embrapa. **Sistema brasileiro de classificação de solos**. 3.ed. Rio de Janeiro: Centro Nacional de Pesquisa de Solos, 2013. 353p.
10. Greenway, H.; Munns, R. Mechanisms of salt tolerance in nonhalophytes. **Annual Review of Plant Physiology**, Palo Alto, v.31, n.1, p.149-190, 1980. Available at: <<https://doi.org/10.1146/annurev.pp.31.060180.001053>>. Accessed on: 22 ago. 2018.
11. Grisi, B.M. Método químico de medição da respiração edáfica: alguns aspectos técnicos. **Ciência e Cultura**, São Paulo, v.30, n.1, p.82-88, 1978.
12. Khedher, S.B.; Kilani-Feki, O.; Dammak, M.; Jabnoun-Khiaredidine, H.; Daami-Remadi, M.; Tounsi, S. Efficacy of *Bacillus subtilis* V26 as a biological control agent against *Rhizoctonia solani* on potato. **Comptes Rendus Biologies**, Paris, v.338, n.12, p.784-792, 2015. Available at: <<https://doi.org/10.1016/j.crvi.2015.09.005>>. Accessed on: 22 ago. 2018.
13. Lazarovits, G. Management of soil-borne plant pathogens with organic soil amendment: a disease control strategy salvaged from the past. **Canadian Journal of Plant Pathology**, Canada, v.23, n.1, p.1-7, 2001. Available at: <<https://doi.org/10.1080/07060660109506901>>. Accessed on: 22 ago. 2018.
14. Malolepsza, U.; Nawrocka, J.; Szczech, M. *Trichoderma virens* 106 inoculation stimulates defence enzyme activities and enhances phenolic levels in tomato plants leading to lowered *Rhizoctonia solani* infection. **Journal Biocontrol Science and Technology**, United Kingdom, v.27, n.2, p.180-199, 2017. Available at: <<https://doi.org/10.1080/09583157.2016.1264570>>. Accessed on: 22 ago. 2018.
15. Manteli, C. **Efeito do chorume de suínos e do ph do solo sobre o tombamento de pepino causado por *Pythium* sp.** 2010. 70p. Dissertação (Mestrado em Agronomia)- Universidade Tecnológica Federal do Paraná, Pato Branco. Available at: <[http://repositorio.utfpr.edu.br/jspui/bitstream/1/228/1/PB\\_PPGA\\_M\\_Manteli,%20Claudia\\_2010.pdf](http://repositorio.utfpr.edu.br/jspui/bitstream/1/228/1/PB_PPGA_M_Manteli,%20Claudia_2010.pdf)>. Accessed on: 22 ago. 2018.
16. Morales, R.G.F.; Santos, I.; Danner, M.A. Efeito do chorume líquido de suínos na podridão do colo e tombamento de plântulas de feijoeiro causadas por *Sclerotium rolfsii*. **Fitopatologia Brasileira**, Brasília, DF, v.32, n.5, p.429-433, 2007. Available at: <<http://dx.doi.org/10.1590/S0100-41582007000500010>>. Accessed on: 22 ago. 2018.
17. Pavan, M.A.; Bloch, M.F.; Zempulski, H.C.; Miyazawa, M.; Zocoler, D.C. **Manual de análise química de solo e controle de qualidade**. Londrina: IAPAR, 1992. 40p. (Circular, 76).
18. Rebouças, M.A.; Façanha, J.G.V.; Ferreira, L.G.R.; Prisco, J.T. Crescimento e conteúdo de N, P, K e Na em três cultivares de algodão sob condições de estresse salino. **Revista Brasileira de Fisiologia Vegetal**, Campinas, v.1, n.1, p.79-85, 1989.
19. Tenuta, M.; Conn, K.L.; Lazarovits, G. Volatile fatty acids in liquid swine manure can kill microsclerotia of *Verticillium dahliae*. **Phytopathology**, Ithaca, v.92, n.5, p.548-552, 2002. Available at: <<https://doi.org/10.1094/PHYTO.2002.92.5.548>>. Accessed on: 22 ago. 2018.
20. Tenuta, M.; Lazarovits, G. Ammonia and nitrous acid from nitrogenous amendments kill the microsclerotia of *Verticillium dahliae*. **Phytopathology**, Ithaca, v.92, n.3, p.255-264, 2002. Available at: <<https://doi.org/10.1094/PHYTO.2002.92.3.255>>. Accessed on: 22 ago. 2018.
21. Yu, Y.; Jianga, C.; Wanga, C.; Chena, L.; Li, H.; Xu, Q. An improved strategy for stable biocontrol agents selecting to control rice sheath blight caused by *Rhizoctonia solani*. **Microbiological Research**, Jena, v.203, p.1-9, 2017. Available at: <<https://doi.org/10.1016/j.micres.2017.05.006>>. Accessed on: 22 ago. 2018.