

Profile of thyroid hormones in breast cancer patients

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Abstract

Estrogen involvement in breast cancer has been established; however, the association between breast cancer and thyroid diseases is controversial. Estrogen-like effects of thyroid hormone on breast cancer cell growth in culture have been reported. The objective of the present study was to determine the profile of thyroid hormones in breast cancer patients. Serum aliquots from 26 patients with breast cancer ranging in age from 30 to 85 years and age-matched normal controls (N = 22) were analyzed for free triiodothyronine (T₃F), free thyroxine (T₄F), thyroid-stimulating hormone (TSH), antiperoxidase antibody (TPO), and estradiol (E₂). Estrogen receptor β (ER β) was determined in tumor tissues by immunohistochemistry. Thyroid disease incidence was higher in patients than in controls (58 vs 18%, P < 0.05). Subclinical hyperthyroidism was the most frequent disorder in patients (31%); hypothyroidism (8%) and positive anti-TPO antibodies (19%) were also found. Subclinical hypothyroidism was the only dysfunction (18%) found in controls. Hyperthyroidism was associated with postmenopausal patients, as shown by significantly higher mean T₃ and T₄ values and lower TSH levels in this group of breast cancer patients than in controls. The majority of positive ER β tumors were clustered in the postmenopausal patients and all cases presenting subclinical hyperthyroidism in this subgroup concomitantly exhibited ER β -positive tumors. Subclinical hyperthyroidism was present in only one of 6 premenopausal patients. We show here that postmenopausal breast cancer patients have a significantly increased thyroid hormone/E₂ ratio (P < 0.05), suggesting a possible tumor growth-promoting effect caused by this imbalance.

Key words

- Thyroid hormone
- Estrogen receptor
- Breast cancer
- Hyperthyroidism

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Growing and developing breasts require the coordinated action of several hormones such as prolactin, estrogen (E₂), progesterone, adrenal steroids, insulin, and growth and thyroid hormones (1). E₂ is considered to

be a potent mitogen for the normal mammary gland, whereas thyroid hormones appear to stimulate lobular development, contributing to the differentiation of normal breast tissue (2). The biological activity of thyroid hor-

mones and E_2 is only manifested in cells expressing thyroid hormone (TR) and estrogen receptors (ER), respectively, that belong to the nuclear receptor superfamily. These receptors share a common mechanism of action whereby hormone-receptor complexes bound to cis acting DNA elements enhance or repress transcription of target genes (3).

As the half-site of the consensus sequence of nucleotide bases constituting the thyroid hormone response element is identical to the ER response element half-site, TR has been shown to bind to ER response elements in addition to their cognate response elements (4). Since various isoforms of both ER and TR exist, combinations of different ER and TR isoforms could lead to different transcriptional end points (5). There are four major TR subtypes, TR α 1, α 2, β 1, and β 2, that are encoded by different genes (6). Two ER isoforms, α and β , arising from two differential genes exist in vertebrates (5).

The involvement of E_2 in breast cancer growth has been established. About one third of breast cancers maintain E_2 dependence for growth and the concentration of ER in malignant breast tissues is an indicator of their hormonal dependence (3). However, the relationship between breast cancer and thyroid hormone is controversial. Even though many studies have shown that thyroid diseases are common in women with breast cancer, other reports have not confirmed this association (reviewed in 7,8). Almost every form of thyroid disease including hyperthyroidism has been identified in association with breast cancer (9-11). Moreover, hyperthyroidism accounts for 2% of all patients presenting adult gynecomastia (12) and it has also been suggested that free triiodothyronine (T_3) plays an important role in the physiology of fibrocystic breast disease (13). Consistent with the proposal that thyroid hormones act on the breast, TR have been described in breast cancer (14).

There are reports on interference between estradiol and thyroid hormones. Previous

studies suggested a cross talk between ER and TR in neuroendocrine tissues leading to inhibition of estrogenic effects by thyroid hormone (15). On the other hand, physiological concentrations of T_3 , the more active form of thyroid hormone, significantly enhance estradiol growth stimulation of a number of human breast carcinoma cell lines (16). In T47D breast cancer cells, E_2 and T_3 similarly regulate cell cycle progression and proliferation raising the p53 level and causing hyperphosphorylation of pRb (17).

We have demonstrated that in breast cancer cell lines, T_3 at supraphysiologic concentrations and in the absence of E_2 mimics the effects of E_2 , possibly through the ER (18). To continue our study of the association between thyroid hormones and breast cancer, the aim of the present investigation was to analyze the thyroid hormone profile of breast cancer patients.

Patients included in the present study ($N = 26$) were newly diagnosed and submitted to surgery at Hospital das Clínicas, School of Medicine, UNESP, Botucatu, SP, Brazil. All cases were classified as stage II. Ages ranged from 30 to 85 years, and 20 patients were menopausal (amenorrhea for at least one year). The study was approved by the Hospital Ethics Committee and all patients signed an informed consent form.

Patients were excluded for the following reasons: radio- or chemotherapy before surgery, hormonal replacement, any kind of previously diagnosed thyroid disease, chronic kidney failure, or recent elevation of serum creatinine to values greater than normally expected for that particular age. Other exclusion factors were: abnormal hepatic function shown by AST, ALT, bilirubin, and/or alkaline phosphatase concentrations higher than twice the normal upper limit; use of blocking agents, aspirin, heparin, phenytoin, steroids, or dopamine, taken one month before starting or during the study; use of iodine contrasts for a 6-month period before and during the study.

Serum aliquots were analyzed for T₃, free thyroxine (T₄F), thyroid-stimulating hormone (TSH), E₂, and antiperoxidase antibody (anti-TPO) using commercially available kits (DPC, Los Angeles, CA, USA). The normal ranges were 1.4-4.4 pmol/ml for T₃F, 0.8-2.0 ng/dl for T₄F, and 0.3-5.0 μU/ml for TSH. Serum determinations were performed prior to surgery, radiotherapy, and chemotherapy. After surgery, new serum determinations were performed in order to confirm the previous data. For the control group, 22 women aged 30 to 85 years were selected (4 premenopausal and 18 postmenopausal women) whose recent mammograms indicated absence of breast cancer. These mammograms were performed in the same week when anamnesis and blood samples were collected. There was no difference in E₂ levels between controls and patients. Premenopausal patients presented E₂ levels of 17.63 ± 33.66 and controls 24.80 ± 3.65 pg/ml. The mean values found in postmenopausal control women and in breast cancer patients were 21.80 ± 12.00 and 13.03 ± 46.99 pg/ml, respectively.

The presence of ER in tumors was determined by immunohistochemical staining using a polyclonal rabbit anti-human ERβ antibody (Upstate Biotechnology, Inc., Lake Placid, NY, USA). Biotinylated secondary antibodies (anti-mouse IgG or anti-rabbit IgG) were obtained from Vector Laboratories (Burlingame, CA, USA). The reactions were developed with the avidin-biotin-peroxidase complex. Tumors known to be positive for the studied marker were considered to be positive controls.

Mean serum thyroid hormone values were 3.56 ± 3.14 pmol/ml for T₃F, 1.40 ± 1.64 ng/dl for T₄F and 1.36 ± 0.63 μU/ml for TSH in breast cancer patients and 2.87 ± 3.12 pmol/ml, 1.10 ± 0.83 ng/dl and 2.41 ± 0.35 μU/ml, respectively, in controls. Anti-TPO antibodies were not found in controls. T₃F levels were significantly higher in patients than in controls (P < 0.001, Mann-Whitney test).

In Table 1, patients and controls were classified by menopausal status, and T₃F, T₄F and TSH levels were compared between the two groups. There was a clear association between hyperthyroidism and menopausal status, with postmenopausal patients presenting a statistically significant subclinical hyperthyroidism compared to controls. TSH levels in postmenopausal patients were lower than in both controls and premenopausal patients; 8 of 20 patients presented subclinical levels, defined by TSH < 0.4 μU/ml. Nonparametric analysis of variance was used for the two-factor model (P < 0.05; Mann-Whitney test). Postmenopausal patients had a significantly higher T₃F/E₂ ratio than controls.

Table 2 shows that 58% of the patients with breast cancer (15 patients) presented some kind of thyroid hormone-related pathology, whereas in the control group hypothyroidism (18%) was the only thyroid disorder observed. Subclinical hyperthyroid-

Table 1. Serum thyroid hormone levels in breast cancer patients and control individuals classified by menopausal status.

	Postmenopausal women		Premenopausal women	
	Breast cancer (N = 20)	Control (N = 18)	Breast cancer (N = 6)	Control (N = 4)
T ₃ F	3.91 ± 1.64*	2.84 ± 0.82	3.33 ± 0.97	2.97 ± 0.69
T ₄ F	1.43 ± 0.63*	1.10 ± 0.20	1.31 ± 0.33	1.30 ± 0.30
TSH	1.00 ± 2.11*	2.86 ± 3.12	1.82 ± 2.44*	1.14 ± 0.26
T ₃ F/E ₂	0.33 ± 2.64*	0.13 ± 0.06	0.18 ± 0.36	0.14 ± 0.03

Data are reported as median ± total semi-range. T₃F = free triiodothyrosine (pmol/ml); T₄F = free thyroxine (ng/dl); TSH = thyroid-stimulating hormone (μU/ml); T₃F/E₂ = ratio of free triiodothyrosine and estrogen (E₂).

*P < 0.05 compared to control group (Mann-Whitney test).

Table 2. Distribution of thyroid hormone dysfunction in patients with breast cancer and controls.

	Breast cancer	Control
Thyroid dysfunction	15 (58%)*	4 (18%)
Absence of thyroid dysfunction	11 (42%)	18 (82%)

*P < 0.05 compared to control group (Fisher's exact test).

ism was the most frequent disorder, being present in 8 of the 26 (31%) breast cancer patients. Seven of these were postmenopausal patients, whereas only 1 (15%) of the six premenopausal patients had hyperthyroidism. Hypothyroidism was detected only in the postmenopausal group (2 cases) whereas anti-TPO antibodies were identified in two and three post- and premenopausal patients, respectively. Thyroid dysfunction was found to be significantly higher in patients with breast cancer. Data analysis was performed by Fisher's exact test ($P < 0.05$).

ER β immunostaining of tumor tissues from all patients indicated ER+ in 17/26 (65%) cases, 14 of which were classified as postmenopausal; this group included all patients with subclinical hyperthyroidism. On the other hand, patients displaying TPO antibodies had tumors classified as ER negative, independent of menopausal status and in the absence of thyroid hormone dysfunction.

Considering the control and experimental patients as a whole, 12.5% presented subclinical hypothyroidism, a higher occurrence than described in population studies (4.7%) (19). However, most of our patients were postmenopausal and hypothyroidism is a common hormonal alteration in patients over 50 years of age (20). This higher incidence may also derive from the fact that all the patients in this study came from an area known to be endemic for low iodine ingestion.

In conclusion, subclinical hyperthyroid-

ism was the only statistically significant thyroid alteration found in our breast cancer population. This, together with the fact that the majority of patients with subclinical hyperthyroidism were postmenopausal, showed that the normal ratio between thyroid hormone and E₂ serum concentration was enhanced in these patients due to increased serum thyroid hormone and reduced E₂ concentration. We may speculate that subclinical hyperthyroidism in postmenopausal patients contributes to breast tumor growth. This might result from an E₂-like effect through the interactions between T₃/T₄ and the ER since the tumors in this patient subgroup expressed ER β +. These interactions may imply binding of T₃ to ER, as reported for breast cancer cell lines (18). Alternatively, thyroid hormones might stimulate the transcription of ER β -dependent genes through a combination of ER β -TR α 1, as suggested by Vasudevan et al. (5). Dinda et al. (17) postulated that the estrogen-like effects of T₃ were mediated by TR interaction with ER response element. Therefore, another possibility is that T₃ bound to the T₃ receptor, independently of ER, stimulates ER response element function, promoting transcription of target genes that lead to cellular proliferation.

Our results suggest the existence of a biological link between breast cancer in postmenopausal women and subclinical hyperthyroidism. However, further studies are required to confirm this association.

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