



Potential Application in Biocatalysis of Mycelium-Bound Lipases from Amazonian Fungi

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Neste trabalho foram avaliados 212 fungos isolados de plantas da Região Amazônica quanto ao potencial de aplicação em biocatálise como fornecedores de lipases ligadas ao micélio. Estes isolados foram submetidos a ensaios de atividade hidrolítica e sintética. Quando submetidos ao ensaio com o substrato tributirina, 87% dos isolados apresentaram atividade hidrolítica. Destes, 30% mostraram bom crescimento em meio líquido indutor de lipase, e foram submetidos à avaliação sintética em reações de esterificação e transesterificação em solvente orgânico. Os nove fungos que apresentaram os melhores desempenhos sintéticos foram avaliados quanto a sua enantiosseletividade na reação de resolução do (*R,S*)-2-octanol. O isolado UEA_115 foi o biocatalisador mais versátil, apresentando bom desempenho nas reações de esterificação (conversão > 90%) e boa capacidade para a resolução do (*R,S*)-2-octanol (ee_s 29%; ee_p 99%; *c* 22%; *E* > 200). Com este estudo foi possível demonstrar o grande potencial dos fungos amazônicos como fornecedores de lipases para biocatálise.

In this study, 212 fungi were isolated from Amazon region plants, aiming to obtain mycelium bound-lipase-producing biocatalysts. These isolates were submitted to hydrolytic and synthetic activity assays. When submitted to the tributyrine substrate test, 87% of the isolates showed hydrolytic activity. Of these, 30% showed good growth in lipase inducing liquid media and were submitted to evaluation of synthetic activity in esterification and transesterification reactions in organic solvents. The nine fungi which had the best synthetic activity were evaluated in the (*R,S*)-2-octanol resolution reaction, in order to verify the enantioselectivity of mycelium-bound lipases. The isolate UEA_115 was the most versatile biocatalyst, showing good performance in esterification reactions (conversion > 90%) and good ability for the resolution of (*R,S*)-2-octanol (ee_s 29%; ee_p 99%; *c* 22%; *E* > 200). Thus, this study has demonstrated the great potential of the Amazonian fungi as lipase suppliers for biocatalysts.

Keywords: fungi, Amazon region, mycelium-bound lipase, biocatalyst screening, biocatalysis

Introduction

In recent years, global concern regarding the impact of human actions on the environment has been growing. The implementation of the concept of sustainable development in several productive activities has become an urgent issue. Fortunately, biotechnology also offers an increasing potential to meet the global demands for processes and products in greater harmony with the sustainability concept.^{1,2} In this regard, one of the most promising tools in biotechnology is the field of applied biocatalysis,

also referred to by some authors as biotransformation or enzymatic technology.^{2,3}

The advantages of using enzymes as biocatalysts in industrial processes have been shown to be ever more evident. Great versatility of catalyzed reactions, mild reaction conditions, and regio, chemo and enantioselective natures are those most frequently mentioned in the literature.⁴ Such characteristics allow the enzymes to promote reactions that would be difficult to stimulate through conventional organic synthesis techniques.⁵ From the ecological point of view, the most important factor is that the high selectivity and efficiency of biological catalysts lead to reactions with few byproducts and less energy

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consumption, making enzymes a more environmentally 'clean' alternative than traditional chemical catalysis.⁶ Many industrial sectors are strong candidates to make use of these advantageous aspects of biocatalysts on a large scale, and many are already using.⁷

The finding that enzymes can be active in organic media has opened an immense field of applications, particularly for the fine chemicals pharmaceutical industry. The obtainment of enantiomerically pure drugs, with potentializing action, which also avoid the risks of using racemic mixtures, tragically illustrated by the case of thalidomide, is of great interest to the pharmaceutical industry.⁸ The use of enzymes (in particular lipases) for the synthesis of chiral drugs, has been well established for years, and biotransformations are now accepted as common methodologies in the preparation of this type of medicine.⁹ In fact, in recent years the application of biological systems (enzymes or whole cells) is cited as the most significant development in the field of synthetic chemistry.¹⁰ Optically active alcohols are extremely useful starting materials and intermediates in synthetic organic chemistry and in the pharmaceutical industry. The obtainment of enantiopure alcohols by biocatalytic reactions has attracted considerable attention.¹¹⁻¹³

Many other fields in which biotransformation processes are already in use or, at least, in an advanced state of development can be cited: treatment of residual waters, bioremediation, biopolymers, tests for clinical diagnosis and biofuel production are among the most important.^{2,7} In some industrial sectors, it is possible to detect an incipient change in paradigm; the traditional chemical catalytic processes tend now to be gradually replaced by biocatalytic processes. This change meets not only the demands for taking advantage of the beneficial characteristics of the enzymes, but also the need to satisfy the growing desire for sustainable technology.¹⁴

The processes which use lipases are particularly attractive due to the versatility of the application of these enzymes.¹⁵ The various technological segments which employ them include the food industry (development of aromas and maturation of cheeses), detergents, and oil chemistry (hydrolysis of oils and fats, synthesis of surfactants), as well as applications in the treatment of oily residues originating from the leather and paper industries.

It is worth noting the use of lipases for obtaining medicines in their enantiomerically active forms.^{16,17} In fact, lipases have been widely recognized as the most important group of biocatalysts for biotechnological applications.¹⁸⁻²⁰ They have been used to catalyze hydrolysis, esterification, transesterification and interesterification reactions, with

extreme process simplicity, higher quality of the final product and excellent yield.¹⁵ They have become so attractive for employment in biotransformation because: (i) they are stable in organic solvents; (ii) they do not require co-factors; (iii) they have wide specificity for substrates; and (iv) they exhibit high enantioselectivity.^{18,21,22}

Although higher animals and plants also produce them, most lipases currently employed in biotechnological applications have been isolated from microorganisms, mainly fungi and bacteria.^{21,23} From the economic and industrial point of view, microorganisms present advantages over higher animals and plants as sources of lipases, due to the high cost of enzyme isolation from higher organisms²⁴ since microorganisms grow fast and are easily handled. Commercial lipases of fungal origin are generally extracellular enzymes, although there is considerable evidence for the activity of mycelium-bound lipases and their study has received attention in recent years. Lipases bound to the cell, both in fungi and in bacteria, also have great stability in organic solvents.^{23,25-27}

The industrial demand for new sources of lipases has continued to stimulate the isolation and screening of new lipolytic microorganisms.²³ The immense biological diversity in the Amazon region and the importance of fungi as enzyme sources, has led to the search for new biocatalysts with special characteristics and possibilities for application in biotechnological processes.^{16,28} The access to a wide range of lipase types has made them even more desirable for two additional reasons: (i) from the large number of lipases cited in the literature, only the enzymes of a few species have been shown to be stable and have biosynthetic capacities suitable for routine use in organic reactions; and (ii) many important problems remain unsolved in the application of lipases in certain industrial processes.²⁹

Considering the need for production processes which are more committed to the ideal of sustainability, this study aims to contribute to the broadening of knowledge regarding the biotechnological potential of fungi from the Amazon region, particularly those isolated from flora, as potential mycelium bound-lipase sources for application in the field of biocatalysis. In this study, lipase-producing Amazonian fungi were tested for both hydrolytic and synthetic activities. Fungi performance on tributyrin and olive oil hydrolysis, as well as on synthetic reactions in organic solvent were examined. The synthesis of pentyl oleate and dodecyl linoleate by esterification and the formation of ethyl palmitate by transesterification were evaluated and the strains with higher synthetic activity were used for the resolution of (*R,S*)-2-octanol.

Experimental

Isolation and maintenance of fungi

The plants used in this study were acquired from markets and fairs in Manaus, capital city of the state of Amazonas, Brazil. All solvents and reagents used in this work were of analytical grade, are commercially available and were used without further purification.

For the isolation of the endophytic microorganisms different plant species from the Amazon region were used: spiked pepper (*Piper aduncum*), manioc (*Manihot esculenta*), tucumã palm (*Astrocaryum aculeatum*), Brazil nut (*Bertholletia excelsa*), uxi (*Endopleura uchi*) and cocoa (*Theobroma cacao*). In the isolation of the endophytic microorganisms, different plant material (leaves, stems, roots, seeds and fruits) were washed in running water and then dried on paper. They were then submitted to surface disinfection by way of immersion in 70% alcohol, 2% hypochlorite, 70% alcohol and again in sterile distilled water.³⁰ Fragments of the different plant tissues were transferred to Petri dishes containing potato-dextrose-agar (PDA) culture medium supplemented with the antibiotic tetracycline (100 mg L⁻¹) and incubated at 28 °C for approximately four days. Following this period, growth of the endophytic microorganisms was verified on each of the plant materials analyzed and the fungi obtained were purified by way of cross-breeding in Petri dishes to obtain monospore cultures. This procedure was repeated twice in order to ensure the purity of the isolates. When necessary, dilution and seeding in culture medium were used to obtain isolated colonies. After purification, the isolates were stored in test tubes containing the culture medium (PDA) and kept in a cold chamber at 5 °C until later use.

The isolation of the phytopathogenic fungi was carried out from the leaves and fruits of different plants: peach palm (*Bactris gasipaes*), tucumã palm (*Astrocaryum aculeatum*), Brazil nut (*Bertholletia excelsa*), aloe vera (*Aloe vera*), cocoa (*Theobroma cacao*), banana (*Musa paradisiaca*) and sugar cane (*Saccharum* sp.) that had lesions due to the presence of fungi. The material was washed in running water (to remove excess impurities) and then tissue fragments were removed from the transition region between the lesion and the healthy area. The explants were submitted to surface disinfection in 70% alcohol and 2% hypochlorite, followed by two washings in autoclaved distilled water.^{28,31} The fragments were placed on filter paper to remove excess water and transferred to Petri dishes containing the culture medium (PDA) supplemented with chloranphenicol (250 mg L⁻¹). The dishes were then incubated at 28 °C until the appearance of fungal hyphae (24-48 h). The fungi were purified as previously described,

and stored in test tubes containing the culture medium PDA, and kept in a cold chamber at 5 °C until later use.

The wood degrading fungi were kindly provided by professor Ademir Castro e Silva of UEA. The fungi were grown on Petri dishes containing malt extract agar media, and kept in a cold chamber.

Hydrolytic assays

The preliminary lipolytic screening was carried out according to the methodology described by Torres *et al.*²³ with modifications. The medium used was composed of mycological peptone (5.0 g L⁻¹), yeast extract (3.0 g L⁻¹), agar (10.0 g L⁻¹), and tributyrine (Sigma) added to give a final concentration of 0.1% (v/v). After autoclaving the medium (121 °C, 20 min) the mixture was emulsified by vigorous shaking for 15 min, and after cooling to approximately 60 °C, the medium was transferred to Petri dishes and rapidly cooled. The agar concentration used for the tributyrine assay is in the range suggested by Lawrence (under 1.5%),²⁹ assuring adequate protein diffusion into the solid medium.

The inoculant to be tested was previously cultivated in PDA medium for 5-7 days at 28 °C. A disc of the solid medium containing the inoculant was placed in the center of a Petri dish with agar containing the medium with tributyrine. The dishes were incubated at 25 °C for 14 days and the presence of a translucent halo around the disc was used to confirm the hydrolytic activity of the isolates tested. All assays were carried out in triplicate.

For the olive oil test the methodology described by Colen was adopted, with modifications.³⁰ The solid medium was prepared with yeast extract (0.5 g L⁻¹), mycological agar (25 g L⁻¹), (NH₄)₂SO₄ (5.0 g L⁻¹), (NH₂)₂CO (2.0 g L⁻¹), MgSO₄·7H₂O (1.0 g L⁻¹), NaCl (1.0 g L⁻¹), olive oil (10 mL L⁻¹, Sigma) and bile salts (3.0 g L⁻¹, Difco). The medium was heated and then homogenized in a blender for 1 min, to obtain a stable emulsion, followed by autoclaving at 121 °C for 15 min. After cooling to approximately 60 °C the medium was transferred to Petri dishes for rapid cooling. The dishes containing the medium with olive oil were inoculated with a disc of around 7.0 mm diameter cut from the PDA medium culture. The dishes were incubated at 25 °C for 10 days and the formation of a translucent halo around the fungi was used to confirm the hydrolytic activity of the isolates.

Obtainment of fungal mycelia

The fungi which gave positive results in the test with tributyrine were cultivated in synthetic liquid

medium consisting of asparagine (2.0 g L^{-1}), glucose (2.0 g L^{-1}), K_2HPO_4 (1.0 g L^{-1}), MgSO_4 (0.5 g L^{-1}), thiamine hydrochloride (5.0 mg L^{-1} ; Sigma), $\text{Fe}(\text{NO}_3)_3 \cdot 7\text{H}_2\text{O}$ (1.45 mg L^{-1}), $\text{MnSO}_4 \cdot \text{H}_2\text{O}$ (0.235 mg L^{-1}) and sunflower oil (2.0% v/v, Sigma), used to induce the production of lipases.²⁰ The medium was inoculated with four discs of approximately 7.0 mm diameter of culture in PDA medium and incubated at $28 \text{ }^\circ\text{C}$ for 9 days under orbital shaking (200 rpm). The fungal mycelium was collected from the culture medium by vacuum filtration, washed with distilled water and acetone,²² dried for 48 h in a desiccator until constant weight and ground to a powder.

Synthetic assays

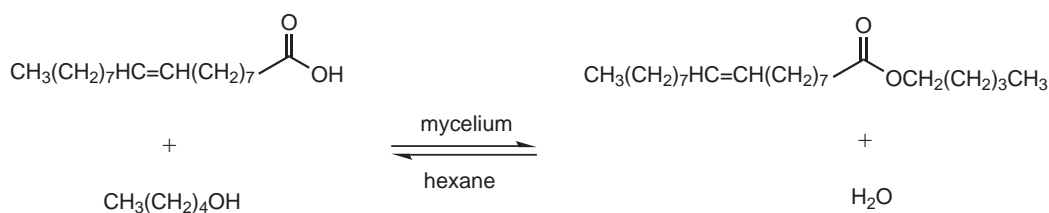
After the selection of the fungal isolates which produced hydrolase in solid media, the synthetic activity of lipases produced by the selected fungi was evaluated through the monitoring of the synthesis of pentyl oleate and dodecyl linoleate by esterification, and the synthesis of ethyl palmitate by transesterification. All assays were carried out in duplicate.

The fungi which showed good growth in synthetic liquid media, *i.e.*, which produced above 500 mg of mycelium

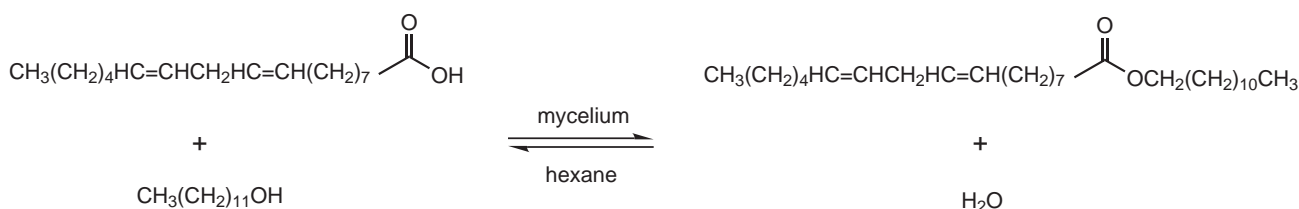
(dry weight) after nine cultivation days were selected for the synthetic screening. All synthetic reactions were carried out in organic media. Reactions performed in organic media eliminate microbiological contamination, since organic solvents are hostile environments for living cells.²⁰

In order to study the synthetic activity of mycelium-bound lipases, 50 mg of mycelium were suspended in *n*-hexane (J. T. Backer, HPLC grade) containing equimolar amounts of fatty acid and alcohol for esterification reactions, and with an excess of one of the reagents for transesterification reactions. The reactions were incubated for 72 h under orbital shaking at 200 rpm and $28 \text{ }^\circ\text{C}$. In the synthesis of pentyl oleate (Scheme 1), 6.66 mmoles of oleic acid and *n*-pentanol in 30 mL of solvent were used. To obtain the dodecyl linoleate (Scheme 2), 0.5 mmoles of linoleic acid and dodecanol in 10 mL of *n*-hexane were used. The transesterification reaction (Scheme 3) was carried out with 1.25×10^{-2} mmoles of *p*-nitrophenyl palmitate (Sigma) and 5×10^{-2} mmoles of ethanol in 10 mL of *n*-hexane.

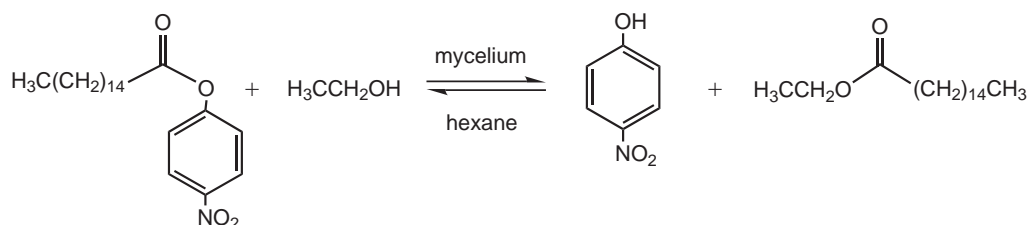
The isolates which provided the greatest conversions to product in the synthetic reactions were evaluated in relation to their enantioselective activity. To this end, the (*R,S*)-2-octanol resolution reaction *via* transesterification



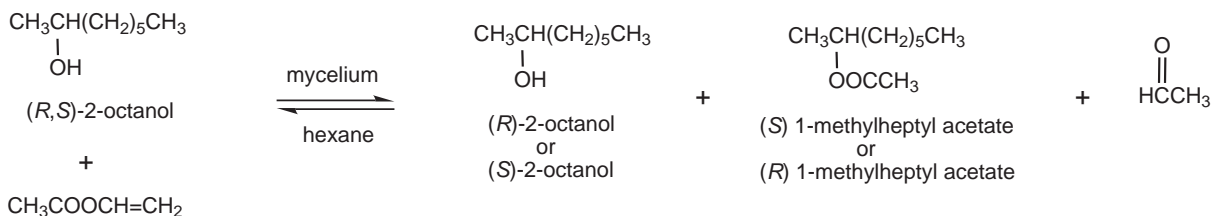
Scheme 1. Biocatalytic esterification reaction of oleic acid with *n*-pentanol for synthetic screening.



Scheme 2. Biocatalytic esterification reaction of linoleic acid with dodecanol for synthetic screening.



Scheme 3. Biocatalytic transesterification reaction of *p*-nitrophenyl palmitate with ethanol for synthetic screening.



Scheme 4. Reaction of biocatalytic resolution of (*R,S*)-2-octanol with vinyl acetate for enantioselective evaluation.

with vinyl acetate (Scheme 4) was carried out. 50 mg of mycelium in a solution containing 2.0 mmoles of (*R,S*)-2-octanol and 6.0 mmoles of vinyl acetate in 10 mL of *n*-hexane were used.

In all biocatalytic reactions two control reactions were carried out: one containing only the reaction solution, without addition of mycelium (blank, B); and one where 50 mg of the commercial enzyme Novozym 435 (lipase B of *Candida antarctica*, CAL-B) was added to the reaction medium (biocatalytic standard, S). Reactions containing only the fungal mycelium and organic solvent were also carried out in order to verify the occurrence of soluble by-products or cellular components in the reaction environment. These reactions were followed by thin layer chromatography (TLC, Merck) using hexane:ethyl acetate 9:1 as eluent. No spots appeared after 72 h of reaction.

Analytical methodology

Aliquots of the biocatalytic reactions were removed periodically and used for the determination of the percent conversion to product (%c) and of the enantiomeric ratio (E). The conversions to pentyl oleate were obtained by nuclear magnetic resonance of hydrogen (¹H NMR) (Varian Unity Inova AC 500 MHz) through the comparison of the integrals of the hydrogen singlets of the alcohol and the hydrogen atoms of the ester.³⁴ On analyzing the ¹H NMR spectra for the alcohol and the ester a difference can be noted between the frequencies of the hydrogen peaks of the methylene group (-CH₂) bound to the oxygen atom, present in both molecules (RCH₂OH; R'COOCH₂R). The hydrogen atoms of the methylene group of the alcohol gave peaks at a lower frequency (more shielded) than those of the ester (less shielded) in the ¹H NMR spectra. Due to this feature, through the integration of the areas of the spectra, the conversion of the esterification reaction can be calculated.

The conversions to dodecyl linoleate and ethyl palmitate were obtained by high performance liquid chromatography (HPLC, Varian ProStar 310, UV detector at 210 nm, column Microsorb 300-5 C4 - 25 cm, acetonitrile/water gradient) through comparison of the peak area of the ester produced

by the fungi with the peak area of the ester produced by the commercial enzyme Novozym 435 (100% conversion).

In order to compare the conversion values that were determined by different analytical methods (¹H NMR and HPLC), samples from the esterification reaction of linoleic acid with dodecanol were analyzed by the two techniques. Both analytical methods were shown to be adequate to determine biocatalytic reaction conversions, since a good correlation between the observed values (R² = 0.976) was observed.³⁵

The conversions to (*R*)- or (*S*)-2-octanol were obtained by gas chromatography coupled to mass spectrometry (Shimadzu GCMS-QP2010) using a chiral stationary phase (Chirasil-Dex CB 25 m × 0.25 mm ID × 0.25 mm). Column temperature was 80 °C. The He pressure was 56.9 kPa; and the temperatures of the injector and the detector were 220 °C and 275 °C, respectively. The calculation of the enantiomeric ratio (E) was carried out according to the formula described by Faber.⁴ The values of the enantiomeric excess of the substrate (*ee*_s) and of the product (*ee*_p) were obtained through the peak areas of the enantiomers.

Results and Discussion

Isolation of Amazonian fungi

A total of 77 isolates were obtained from the endophytic fungi of fragments of six different plant species; 45 were isolated from phytopathogenic fungi growing on fragments of seven different plant species; and 90 wood degrading fungi were isolated from the stems of different species (Figure 1a).

As can be seen in Figure 1b, a greater number of phytopathogenic isolates from Brazil nut, tucumã palm and peach palm were obtained. In the case of endophytic fungi (Figure 1c), spiked pepper allowed the obtainment of the greatest number of isolates. Endophytic microorganisms (fungi and bacteria) are those that inhabit inside a plant for at least a period of its vital cycle, and are found in tissues such as leaves, branches and roots. Apparently, they do not cause any damage to the host, which distinguishes them from the phytopatogen microorganisms. Their

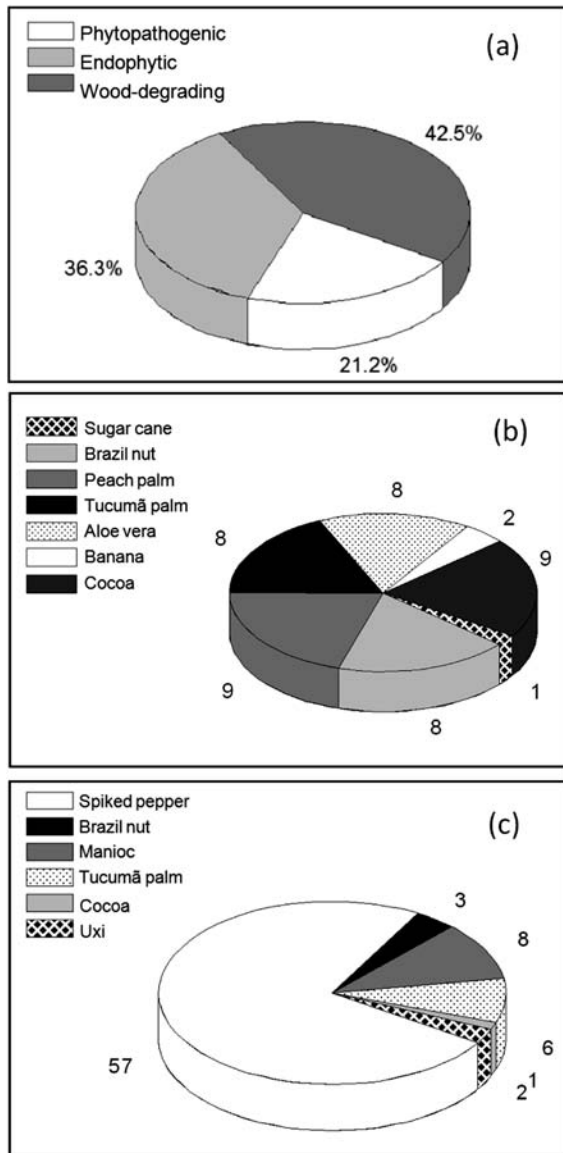


Figure 1. (a) Fungal isolates obtained from Amazon region plants; (b) Number of phytopathogenic isolates and their respective hosts; (c) Number of endophytic isolates and their respective hosts.

presence implies a symbiotic interaction in all the plants investigated until now.³³ These microorganisms gained interest in the 1970's due to the possibility of producing pharmacologically active substances with biotechnological potential such as antitumor agents (*Pestalotiopsis microspora*, taxol) and antifungal agents (*Cryptosporiopsis cryptocandina*, quercine), besides producing plant growth factors, toxins and enzymes.³⁷⁻⁴⁰

The 212 fungal isolates were stored in triplicate and transferred every three months to a new dish containing PDA medium in order to maintain the fungal cellular viability. The isolates were coded and the information relating to the origin of the organisms was grouped (Table 1).

Hydrolytic screening

The 212 fungal isolates were submitted to preliminary tests with tributyrine in order to evaluate the hydrolytic capacity. Of these, the isolates from 74 endophytic, 41 phytopathogenic and 70 wood degrading fungi showed the formation of a halo, indicating the production of hydrolases, that is, 87% of the Amazonian fungi evaluated in this study produced enzymes with hydrolytic activity. This result indicates the great potential of these species as a source of microorganisms which produce hydrolytic enzymes.

Of the species, spiked pepper, Brazil nut, uxi and cocoa, 100% of their endophytic isolates produced halos. Of the endophytic isolates of tucumã palm and manioc, 83 and 75%, respectively, showed a positive result. Of the total of 77 endophytic isolates, 96% produced halos in the presence of tributyrine.

Among the phytopathogenic fungi evaluated, peach palm, the fruit of which had around 20% oil in its composition,⁴¹ provided isolates with 100% halo appearances, as much as the isolates from aloe vera. Of the phytopathogenic fungi isolated from Brazil nuts (which have good proportions of unsaturated fatty acids, 75.6%)⁴² and tucumã palm, 87.5% showed hydrolytic activity and 71% of the isolates from cocoa presented positive result. Of the 45 phytopathogenic fungi evaluated, 89% produced halo when cultivated on tributyrine.

Within the 90 wood degrading fungi evaluated, 70 showed the production of hydrolases, which represents 78% of these isolates.

Studies involving the screening of microorganisms as sources of lipolytic enzymes frequently describe carrying out a preliminary selection of the hydrolytic activity associated with the strains.^{23,26,29,32} The initial selection for the detection of a particular activity has a significant weighting in the screening process, since it aims to eliminate the worst candidates and, in general, is carried out through the cultivation of microorganisms in a solid culture medium, in Petri dishes.⁴⁴

Triacylglycerides such as triolein or -butyrin are used as standard substrates for the determination of lipase activity.⁴ Tributyrine is the substrate which has been most used for detection of lipolytic activity.^{23,29,32,45} On being hydrolyzed, tributyrine generates dibutyryne, monobutyryne, glycerol and butyric acid, all of which are soluble in water, facilitating the visualization of the translucent halo.³² Despite the drawbacks associated with the use of tributyrine (since it is not a specific substrate for lipases as it can also be hydrolyzed by esterases), this is a convenient substrate, since it can be dispersed in water by agitation, without the need to add emulsifiers.⁴⁶ Some authors have also recommended the use

Table 1. Identification code for isolated Amazonian fungi and their respective origin

Strain Code	Type ^a	Host	Tissue	Strain Code	Type	Host	Tissue
UEA_001	PT	<i>B. excelsa</i>	seed	UEA_107	EF	<i>M. esculenta</i>	stem
UEA_002	PT	<i>A. aculeatum</i>	fruit	UEA_108	FT	<i>A. vera</i>	leaf
UEA_003	WD	Amazon wood	stem	UEA_109	EF	<i>M. esculenta</i>	leaf
UEA_004	WD	Amazon wood	stem	UEA_110	WD	Amazon wood	stem
UEA_005	WD	Amazon wood	stem	UEA_111	WD	Amazon wood	stem
UEA_006	PT	<i>T. cacao</i>	fruit	UEA_112	WD	Amazon wood	stem
UEA_007	PT	<i>T. cacao</i>	fruit	UEA_113	WD	Amazon wood	stem
UEA_008	PT	<i>A. aculeatum</i>	fruit	UEA_114	WD	Amazon wood	stem
UEA_009	WD	Amazon wood	stem	UEA_115	WD	Amazon wood	stem
UEA_010	PT	<i>B. gasipaes</i>	fruit	UEA_116	WD	Amazon wood	stem
UEA_011	PT	<i>A. vera</i>	leaf	UEA_117	WD	Amazon wood	stem
UEA_012	PT	<i>A. vera</i>	leaf	UEA_118	WD	Amazon wood	stem
UEA_013	PT	<i>T. cacao</i>	fruit	UEA_119	WD	Amazon wood	stem
UEA_014	PT	<i>B. excelsa</i>	seed	UEA_120	WD	Amazon wood	stem
UEA_015	PT	<i>A. aculeatum</i>	fruit	UEA_121	WD	Amazon wood	stem
UEA_016	PT	<i>B. gasipaes</i>	fruit	UEA_122	WD	Amazon wood	stem
UEA_017	PT	<i>B. gasipaes</i>	fruit	UEA_123	WD	Amazon wood	stem
UEA_018	PT	<i>A. vera</i>	leaf	UEA_124	WD	Amazon wood	stem
UEA_019	PT	<i>A. vera</i>	leaf	UEA_125	WD	Amazon wood	stem
UEA_020	PT	<i>B. excelsa</i>	seed	UEA_126	WD	Amazon wood	stem
UEA_021	PT	<i>B. excelsa</i>	seed	UEA_127	WD	Amazon wood	stem
UEA_022	PT	<i>B. gasipaes</i>	fruit	UEA_128	WD	Amazon wood	stem
UEA_023	PT	<i>T. cacao</i>	fruit	UEA_129	WD	Amazon wood	stem
UEA_024	PT	<i>T. cacao</i>	fruit	UEA_130	WD	Amazon wood	stem
UEA_025	PT	<i>A. vera</i>	leaf	UEA_131	EP	<i>M. esculenta</i>	stem
UEA_026	PT	<i>T. cacao</i>	fruit	UEA_132	EP	<i>M. esculenta</i>	root
UEA_027	PT	<i>B. excelsa</i>	seed	UEA_133	EP	<i>M. esculenta</i>	root
UEA_028	PT	<i>A. aculeatum</i>	fruit	UEA_134	EP	<i>M. esculenta</i>	stem
UEA_029	PT	<i>B. excelsa</i>	seed	UEA_135	PT	<i>B. gasipaes</i>	fruit
UEA_030	PT	<i>A. aculeatum</i>	fruit	UEA_136	PT	<i>A. aculeatum</i>	fruit
UEA_031	PT	<i>B. excelsa</i>	seed	UEA_137	PT	<i>A. vera</i>	leaf
UEA_032	PT	<i>A. aculeatum</i>	fruit	UEA_138	PT	<i>T. cacao</i>	fruit
UEA_033	PT	<i>B. gasipaes</i>	fruit	UEA_139	PT	<i>B. excelsa</i>	seed
UEA_034	PT	<i>B. gasipaes</i>	fruit	UEA_140	WD	Amazon wood	stem
UEA_035	PT	<i>B. gasipaes</i>	fruit	UEA_141	WD	Amazon wood	stem
UEA_036	PT	<i>A. aculeatum</i>	fruit	UEA_142	WD	Amazon wood	stem
UEA_037	PT	<i>B. gasipaes</i>	fruit	UEA_143	WD	Amazon wood	stem
UEA_038	EP	<i>M. esculenta</i>	leaf	UEA_144	WD	Amazon wood	stem
UEA_039	EP	<i>P. aduncum</i>	leaf	UEA_145	EP	<i>P. aduncum</i>	root
UEA_040	EP	<i>P. aduncum</i>	root	UEA_146	EP	<i>P. aduncum</i>	stem
UEA_041	EP	<i>P. aduncum</i>	leaf	UEA_147	EP	<i>P. aduncum</i>	root
UEA_042	EP	<i>A. aculeatum</i>	seed	UEA_148	EP	<i>P. aduncum</i>	root
UEA_043	EP	<i>P. aduncum</i>	root	UEA_149	EP	<i>P. aduncum</i>	leaf
UEA_044	EP	<i>P. aduncum</i>	stem	UEA_150	EP	<i>P. aduncum</i>	leaf
UEA_045	EP	<i>P. aduncum</i>	root	UEA_151	EP	<i>P. aduncum</i>	leaf
UEA_046	EP	<i>P. aduncum</i>	leaf	UEA_152	EP	<i>P. aduncum</i>	leaf
UEA_047	EP	<i>B. excelsa</i>	leaf	UEA_153	EP	<i>P. aduncum</i>	stem
UEA_048	EP	<i>P. aduncum</i>	root	UEA_154	EP	<i>P. aduncum</i>	root
UEA_049	EP	<i>P. aduncum</i>	root	UEA_155	EP	<i>P. aduncum</i>	stem
UEA_050	EP	<i>B. excelsa</i>	stem	UEA_156	EP	<i>P. aduncum</i>	root
UEA_051	EP	<i>B. excelsa</i>	seed	UEA_157	EP	<i>P. aduncum</i>	root
UEA_052	EP	<i>P. aduncum</i>	stem	UEA_158	EP	<i>P. aduncum</i>	root
UEA_053	EP	<i>A. aculeatum</i>	fruit	UEA_159	EP	<i>P. aduncum</i>	root

Table 1. continuation

Strain Code	Type ^a	Host	Tissue	Strain Code	Type	Host	Tissue
UEA_054	EP	<i>P. aduncum</i>	stem	UEA_160	EP	<i>P. aduncum</i>	leaf
UEA_055	EP	<i>P. aduncum</i>	stem	UEA_161	EP	<i>P. aduncum</i>	root
UEA_056	EP	<i>P. aduncum</i>	leaf	UEA_162	EP	<i>P. aduncum</i>	stem
UEA_057	EP	<i>P. aduncum</i>	root	UEA_163	EP	<i>P. aduncum</i>	stem
UEA_058	EP	<i>P. aduncum</i>	stem	UEA_164	EP	<i>P. aduncum</i>	stem
UEA_059	EP	<i>P. aduncum</i>	root	UEA_165	EP	<i>P. aduncum</i>	stem
UEA_060	EP	<i>P. aduncum</i>	root	UEA_166	EP	<i>P. aduncum</i>	leaf
UEA_061	EP	<i>A. aculeatum</i>	seed	UEA_167	EP	<i>P. aduncum</i>	root
UEA_062	EP	<i>P. aduncum</i>	stem	UEA_168	EP	<i>P. aduncum</i>	leaf
UEA_063	EP	<i>A. aculeatum</i>	seed	UEA_169	EP	<i>P. aduncum</i>	leaf
UEA_064	EP	<i>P. aduncum</i>	root	UEA_200	WD	Amazon wood	stem
UEA_065	EP	<i>A. aculeatum</i>	fruit	UEA_201	WD	Amazon wood	stem
UEA_066	EP	<i>P. aduncum</i>	leaf	UEA_202	WD	Amazon wood	stem
UEA_067	EP	<i>P. aduncum</i>	root	UEA_203	WD	Amazon wood	stem
UEA_068	EP	<i>P. aduncum</i>	leaf	UEA_204	WD	Amazon wood	stem
UEA_069	EP	<i>P. aduncum</i>	leaf	UEA_205	WD	Amazon wood	stem
UEA_070	EP	<i>P. aduncum</i>	stem	UEA_206	WD	Amazon wood	stem
UEA_071	EP	<i>T. cacao</i>	seed	UEA_207	WD	Amazon wood	stem
UEA_072	EP	<i>A. aculeatum</i>	fruit	UEA_208	WD	Amazon wood	stem
UEA_073	EP	<i>P. aduncum</i>	root	UEA_209	WD	Amazon wood	stem
UEA_074	EP	<i>E. uchi</i>	fruit	UEA_210	WD	Amazon wood	stem
UEA_075	EP	<i>P. aduncum</i>	root	UEA_211	WD	Amazon wood	stem
UEA_076	EP	<i>E. uchi</i>	fruit	UEA_212	WD	Amazon wood	stem
UEA_077	EP	<i>P. aduncum</i>	leaf	UEA_213	WD	Amazon wood	stem
UEA_078	EP	<i>P. aduncum</i>	leaf	UEA_214	WD	Amazon wood	stem
UEA_079	EP	<i>P. aduncum</i>	leaf	UEA_215	WD	Amazon wood	stem
UEA_080	EP	<i>P. aduncum</i>	leaf	UEA_216	WD	Amazon wood	stem
UEA_081	EP	<i>P. aduncum</i>	leaf	UEA_217	WD	Amazon wood	stem
UEA_082	EP	<i>P. aduncum</i>	leaf	UEA_218	WD	Amazon wood	stem
UEA_083	PT	<i>A. vera</i>	leaf	UEA_219	WD	Amazon wood	stem
UEA_084	PT	<i>Saccharum sp.</i>	fruit	UEA_220	WD	Amazon wood	stem
UEA_085	PT	<i>Banana</i>	fruit	UEA_221	WD	Amazon wood	stem
UEA_086	PT	<i>Banana</i>	fruit	UEA_222	WD	Amazon wood	stem
UEA_087	PT	<i>T. cacao</i>	fruit	UEA_223	WD	Amazon wood	stem
UEA_088	PT	<i>T. cacao</i>	fruit	UEA_224	WD	Amazon wood	stem
UEA_089	WD	Amazon wood	stem	UEA_225	WD	Amazon wood	stem
UEA_090	WD	Amazon wood	stem	UEA_226	WD	Amazon wood	stem
UEA_091	WD	Amazon wood	stem	UEA_227	WD	Amazon wood	stem
UEA_092	WD	Amazon wood	stem	UEA_228	WD	Amazon wood	stem
UEA_093	WD	Amazon wood	stem	UEA_229	WD	Amazon wood	stem
UEA_094	WD	Amazon wood	stem	UEA_230	WD	Amazon wood	stem
UEA_095	WD	Amazon wood	stem	UEA_231	WD	Amazon wood	stem
UEA_096	WD	Amazon wood	stem	UEA_232	WD	Amazon wood	stem
UEA_097	WD	Amazon wood	stem	UEA_233	WD	Amazon wood	stem
UEA_098	WD	Amazon wood	stem	UEA_234	WD	Amazon wood	stem
UEA_099	WD	Amazon wood	stem	UEA_235	WD	Amazon wood	stem
UEA_100	WD	Amazon wood	stem	UEA_236	WD	Amazon wood	stem
UEA_101	WD	Amazon wood	stem	UEA_237	WD	Amazon wood	stem
UEA_102	WD	Amazon wood	stem	UEA_238	WD	Amazon wood	stem
UEA_103	WD	Amazon wood	stem	UEA_239	WD	Amazon wood	stem
UEA_104	WD	Amazon wood	stem	UEA_240	WD	Amazon wood	stem
UEA_105	WD	Amazon wood	stem	UEA_241	WD	Amazon wood	stem
UEA_106	EP	<i>M. esculenta</i>	leaf	UEA_242	WD	Amazon wood	stem

^aEP = Endophytic; PT = Phytopathogenic; WD = Wood-degrading.

of tributyrine in the initial selection of microorganisms with lipolytic enzymes, but with later confirmation with other lipid substrates of greater specificity for lipases.³³

Recent studies have adopted as a criterion the size of the acyl chain resulting from the hydrolysis of acylglycerol to distinguish between lipases and esterases.⁴⁸ Lipases (EC 3.1.1.3) have been characterized as carboxylesterases which hydrolyze long chain acylglycerides (with more than 10 carbon atoms) and the enzymes which hydrolyze acylglycerides with a chain size of less than 10 carbon atoms are considered esterases (EC 3.1.1.1).⁴⁹ Esterases and lipases differ in the size of this hydrophobic interaction area as well as in the shape, deepness and physico-chemical properties of their substrate binding pocket.⁵⁰ However, according to Faber, the main difference between lipases and esterases is their physico-chemical interaction with their substrates, since these enzymes have distinct kinetic behaviors.⁴

Most lipases show activity at the interface of an aqueous and non-aqueous solvent, known as interfacial enzyme activation, the activity of which is associated with conformational changes in the protein domain called 'lid', which gives the substrate access to the active site.^{48,51} However, some lipases have little or no interfacial activation and, thus, show both the activities of lipases and esterases.⁵² Consequently, hydrolysis catalyzed by lipases must be preferentially carried out in biphasic medium, the use of a substrate in a concentration which constitutes an organic phase in an aqueous medium or solubilization of the substrate in a hydrophobic solvent ($\log P > 1.5$) being sufficient.⁴

In order to verify the hydrolytic capacity of the lipases produced by Amazonian fungi, the test was carried out in the presence of olive oil, a substrate considered more specific for lipases than tributyrine,³³ since olive oil is composed of 71% oleic acid.⁵³ The test with olive oil was applied to 22 fungi which tested positive in the tributyrine assay, selected randomly.

Of the 22 isolates evaluated, seven fungi (UEA_006, UEA_041, UEA_053, UEA_055, UEA_057, UEA_064 and UEA_015) gave a negative result in the test with olive oil, suggesting that the positive results in the test with tributyrine of these strains was due to the action of esterases.

The 185 Amazon fungi which showed the formation of halos in the tests with tributyrine were cultivated in a lipase production inductor liquid medium. The fungi which showed good growth were then selected for the synthetic screening.

Synthetic screening

The 64 isolates which had the best growth in the lipase production inductor medium, reaching greater mycelium

masses after nine days of cultivation, were selected to carry out the biocatalytic reactions for synthetic screening using hexane as the solvent.

Considering that lipases are employed in both hydrolytic and synthetic reactions, good results in hydrolytic reactions do not guarantee good performances in synthetic reactions in organic solvents. Thus, synthetic screening is fundamental to evaluate the potential application of mycelium-bound lipases.^{23,29} Purified commercial enzymes, although widely available, are not selected specifically for catalytic reactions in non-aqueous environments. Thus, synthetic screening in organic media is an alternative which allows the selection of enzymes with specific characteristics for the reaction of interest.⁵⁴

The results of the synthetic screening are shown in Figures 2, 3 and 4, which give only the results of the reactions mediated by the Amazonian fungi with a conversion above 10%.

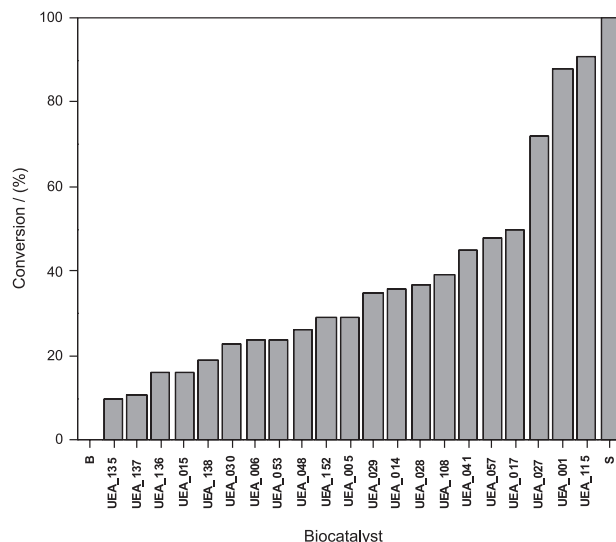


Figure 2. Conversions to pentyl oleate in biocatalytic reactions mediated by Amazonian fungi at 28 °C and 72 h. B and S stand for blank and standard control reactions, respectively.

In the synthesis of pentyl oleate, 21 fungi showed conversions above 10%, with four being above 50%. The isolates UEA_115, UEA_001 and UEA_027 had the most notable performance with 91%, 88% and 72% conversion, respectively (Figure 2). In the synthesis of dodecyl linoleate, 15 fungi had conversions above 10% with six above 50%. The isolates UEA_007, UEA_115 and UEA_041 were the most efficient in this reaction with 98%, 94% and 78% conversion, respectively (Figure 3). In the synthesis of ethyl palmitate, 14 fungi had conversions above 10% with five above 50%. The fungi UEA_007 and UEA_001 had the best results in this reaction, with 87% and 79% of conversion, respectively (Figure 4). It can be

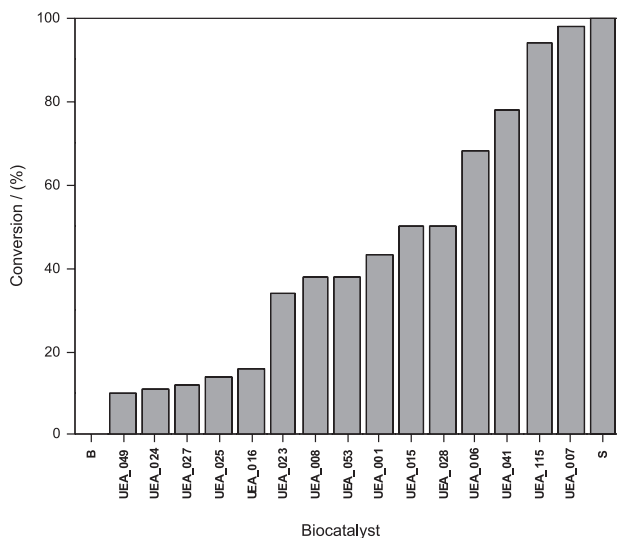


Figure 3. Conversions to dodecyl linoleate in biocatalytic reactions mediated by Amazonian fungi at 28 °C and 72 h. B and S stand for blank and standard control reactions, respectively.

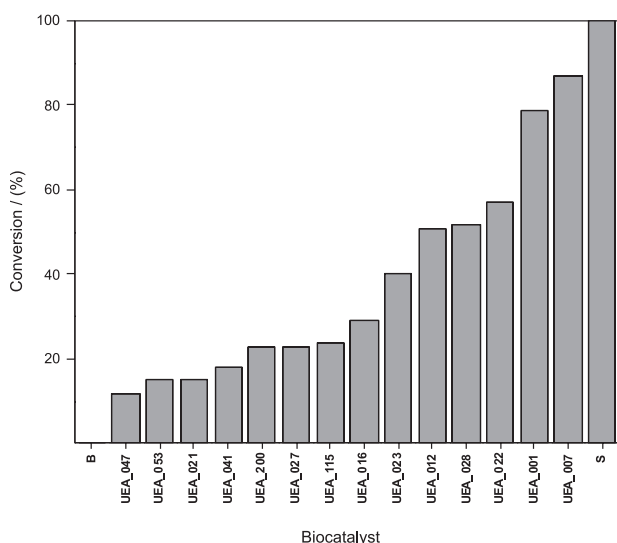


Figure 4. Conversions to ethyl palmitate in biocatalytic reactions mediated by Amazonian fungi at 28 °C and 72 h. B and S stand for blank and standard control reactions, respectively.

seen from these results that the Amazonian fungi evaluated produced lipases with pronounced synthetic activity, being able to act efficiently on the substrates used.

Some strains had a good biocatalytic activity for more than one type of substrate. The strain UEA_001, for example, obtained 88% of conversion to pentyl oleate and 79% of conversion to ethyl palmitate. This behavior was similar to that observed for the fungus UEA_007 which showed 98% of conversion to dodecyl linoleate and 87% of conversion to ethyl palmitate. These two isolates had excellent performance for both the esterification and transesterification reactions, obtaining conversion percentages comparable with the commercial enzyme

Novozym 435 (100% conversion). These results suggest two possibilities: the fungi UEA_001 and UEA_007 have mycelium-bound lipases able to promote excellent conversions to the product, acting on substrates with different chemical structures; or these strains produce different lipases, specific to each of the two substrates.

Using the isolate UEA_115 excellent conversions to product were observed in the two esterification reactions evaluated. This strain showed 91% conversion to pentyl oleate and 94% conversion to dodecyl linoleate. In this case, a preference of the lipases produced by this fungus for the esterification reaction was noted, since in the transesterification reaction the conversion to ethyl palmitate was 24%. The small structural differences in the substrates studied appear not to limit the action of mycelium-bound lipases produced by this isolate in esterification reactions.

It is known that lipases can convert a large variety of non-natural substrates, indicating that they are not completely ‘rigid’ enzymes, but undergo small conformational changes during the formation of the enzyme-substrate complex. This characteristic gives lipases a great versatility for the catalysis of different reactions. Furthermore, the ‘induced fit’ mechanism developed by Koshland at the end of the 1960s⁵⁵ indicated that the enzyme can change its conformation under the influence of the substrate structure, and according to Faber, “The most typical *induced-fit* enzymes are the lipases”.⁴

On the other hand, reactions carried out in organic solvents, in general, result in enzymes with greater rigidity, since in aqueous medium the breaking and formation of hydrogen bonds involved in the formation of the enzyme-substrate complex are assisted by water, which does not occur in hydrophobic solvents.⁵⁶ Another factor that may influence the selectivity of biocatalysts is immobilization.⁴ Nascimento *et al.*⁵⁷ reported that the use of immobilized lipase increased the enantioselectivity and the conversion percentage in the resolution of α -methylene- β -hydroxy esters, when compared to the reaction with the free enzyme.

In the enzymes adhered to the mycelium, the structure formed by the mass of hyphae acts as a natural matrix and produces an effect similar to that of supports used in the immobilization of biocatalysts,²⁷ that is, the mycelium-bound lipases may have a less flexible three-dimensional conformation, and thus only more specific substrates will be transformed.

The mycelium-bound lipases of the isolates UEA_027 and UEA_041 showed less flexibility in relation to the different substrates used in the synthetic screening reactions, since in contrast to the isolates UEA_001,

UEA_007 and UEA_115, only one of the substrates was successfully biotransformed. With the isolate UEA_027 a good percentage of conversion to pentyl oleate (72%) was obtained, with the isolate UEA_041 showing 78% conversion to dodecyl linoleate. These two isolates did not show good performances in the other biocatalytic reactions (conversions below 45%) under the experimental conditions employed. However, they were able to use the substrates, suggesting that, after optimization of the reaction conditions, the performance of these enzymes can be improved.

In order to select the fungi with the best performance for biocatalysis application, a comparison between the results obtained in the hydrolysis and synthesis reactions was considered (Table 2).

Hydrolytic activity screening methods in solid medium can be easily carried out for large numbers of organisms, but they are not suitable to evaluate the true potential for the production of enzymes of synthetic interest.²⁶ The differences between the levels of synthetic and hydrolytic activity may be accentuated by the fact that tests such as that with tributyrine analyze the activity of both extracellular and mycelium-bound hydrolases in hydrophilic medium, whereas the synthesis of esters indicates the presence of mycelium-bound lipase activity in hydrophobic medium.^{23,26}

The isolates UEA_001, UEA_007, UEA_028, UEA_041 and UEA_115, gave positive results in the hydrolytic tests and presented at least one high conversion percentage (> 52%) in one of the synthetic reactions (Table 2).

These isolates can be considered the most versatile within those studied and, consequently, the most promising to be employed in biocatalysis.

The isolates UEA_006, UEA_012, UEA_015, UEA_016, UEA_018, UEA_041 and UEA_053 showed positive results in the hydrolytic test with the tributyrine substrate and negative results in the test with olive oil. Although these strains did not hydrolyze this substrate, many showed significant synthetic activity (Table 2). Thus, the results suggest that the hydrolytic test with the substrate olive oil, for these experimental conditions, is not a suitable method for the screening of lipases with application in organic synthesis, since these seven fungi would be eliminated during the hydrolytic selection. Therefore, these results confirm the data in the literature,^{23,29} which state that there is no correlation between the hydrolytic and synthetic activity of lipases.

One of the main applications of lipases in biocatalysis is the obtainment of enantiomerically pure compounds.²⁰ Therefore, after the screening tests, nine strains of fungi (*Aspergillus* and *Trichoderma* genus) isolated from Amazon region plants were selected (UEA_001, UEA_006, UEA_007, UEA_014, UEA_023, UEA_027, UEA_041, UEA_053 and UEA_115). These had the highest conversion percentages in synthetic activity screening. These nine fungi are being identified through molecular biology techniques. For the preliminary evaluation of the enantioselective potential of the nine strains selected, the resolution of (*R,S*)-2-octanol reaction was carried out.

Table 2. Performance of the Amazonian fungi in the different screening tests

Isolate	Tributyrine ^a	Olive oil ^a	Pentyl oleate ^b / (%)	Dodecyl linoleate ^c / (%)	Ethyl palmitate ^c / (%)
UEA_027	+	+	72	12	23
UEA_001	+	+	88	43	79
UEA_017	+	+	50	1	5
UEA_115	+	+	91	94	24
UEA_041	+	-	45	78	18
UEA_053	+	-	24	38	15
UEA_016	+	-	2	16	29
UEA_028	+	+	37	50	52
UEA_023	+	+	6	34	40
UEA_006	+	-	24	68	0
UEA_007	+	+	8	98	87
UEA_012	+	-	1	8	51
UEA_015	+	-	16	50	4
UEA_018	+	-	0	0	18

^aHydrolytic assays. Positive tests indicate halo formation. ^bDetermined by ¹H NMR. ^cDetermined by HPLC with column Microsorb 300-5 C4 column, acetonitrile/water gradient.

Resolution of (*R,S*)-2-octanol

Lipases have been widely applied as versatile biocatalysts for the kinetic resolution of various racemates.^{16,17,55-57} Our research group has initiated the screening of enantioselective lipase from Amazonian fungi with the resolution of (*R,S*)-2-hexanol. A good potential of these microorganisms for producing chiral compounds was observed.⁵⁸ Thus, following these studies on enantioselective screening, the isolates selected in the synthetic reactions were used to mediate the resolution of the (*R,S*)-2-octanol reaction via transesterification with vinyl acetate (acyl donor). The values for the conversion percentages (%c), substrate and product enantiomeric excess (ee_s and ee_p), and enantiomeric ratio (E) can be seen in Table 3.

Table 3. Conversion and enantioselectivity of the (*R,S*)-2-octanol resolution reaction mediated by Amazonian fungi after 120 h reaction

Biocatalyst	Percentage Conversion ^a	% ee_s ^b	% ee_p ^b	E ^d
UEA_001	0	0	0	0
UEA_006	5	6	99	> 200
UEA_007	8	9	99	> 200
UEA_014	5	6	99	> 200
UEA_023	2	3	99	> 200
UEA_027	7	8	99	> 200
UEA_041	8	9	99	> 200
UEA_053	18	22	99	> 200
UEA_115	22	29	99	> 200
Novozym 435	69	99	44	11

^aReaction with 50 mg of mycelium in a solution containing 2.0 mmoles of (*R,S*)-2-octanol and 6.0 mmoles of vinyl acetate in 10 mL of *n*-hexane.

^bDetermined by GC-MS with Chirasil-Dex CB column. ^dCalculated according to the formula described by Faber (2004).

The values for the parameters used to evaluate the enantioselective activity given in Table 3 show that the fungal strains displayed a resolution capacity lower than Novozym 435, since the biocatalytic reactions mediated by Amazonian fungi presented lower ee_s than the commercial enzyme.

According to Faber, E values lower than 15 are unacceptable for practical purposes. E values between 15 and 30 are considered moderate to good, and above these values are excellent, a value of 200 being the limit of accuracy of the technique, since, above this value, even an extremely small variation in ee_s or ee_p causes a significant change in the numerical value of E.⁴

The reactions mediated by the Amazonian fungi gave values for the conversion of methyl heptyl acetate lower than the reaction using Novozym 435. However, two fungal

isolates (UEA_053 and UEA_115) showed promising enantioselective capacity, since they had high values of ee_p and E. The strain UEA_053 presented ee_s 22%; ee_p 99%; c 18% and E > 200, whereas the isolate UEA_115 showed ee_s 29%; ee_p 99%; c 22% and E > 200. These results indicate the potential of these two strains for the obtainment of optically active alcohols. Consequently, the optimization of the reaction conditions will be of fundamental importance to increase conversion to the product.

Conclusions

The fungi isolated from plants of the Amazon region showed a great potential as providers of mycelium-bound lipases for use in biotransformations. Of the 212 fungi evaluated in this study, the mycelium-bound lipases of the isolates UEA_001, UEA_007 and UEA_115 showed properties desirable for use in biocatalysis: good hydrolysis activity, and good esterification and transesterification capacity in organic media. The isolate UEA_115 was the most versatile biocatalyst, since it presented good performance in the esterification reactions (conversions > 90%) and also showed potential for resolution of racemic 2-octanol (ee_s 29%; ee_p 99%; c 22%; E > 200).

These results confirm the difficulty in obtaining a universal method for the selection of microorganisms which produce lipases, since for each specific substrate and type of reaction employed a suitable biocatalysts can be found from the biodiversity available.

On comparing the action of the Amazonian fungi selected with the commercial enzyme Novozym 435, it can be observed that the non-purified mycelium-bound lipases showed excellent biocatalytic potential. It is important to note that most of the enzymes used in organic synthesis are employed in the form of a crude extract, with low cost. However, the preparations contain only around 1 to 30% of the enzyme of interest, the rest being inactive proteins, stabilizers, buffers or carbohydrates originating from the culture broth from which they were obtained.⁴ Also, it should be considered that crude preparations are frequently more stable than purified enzymes, since they constitute more robust catalytic systems, as can be observed in the results obtained in this study.

The studies in the area of biocatalysis have contributed to strengthening the relationship between chemistry and biotechnology, employing knowledge of chemistry in biological sciences, and encouraging scientists from both fields of knowledge to work in collaboration.⁶² Furthermore, biocatalytic screening is a tool which effectively contributes to knowledge of the biodiversity of the Amazon region,

since the species still not accessed may provide products of great utility to mankind, which represents a key argument in favor of its conservation.⁶³

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Supplementary Information

Chromatograms used for the determination of conversion, enantiomeric excess and enantiomeric ratio of (*R,S*)-2-octanol resolution reactions are available free of charge at <http://jbcs.sbq.org.br>, as PDF file.

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Potential Application in Biocatalysis of Mycelium-Bound Lipases from Amazonian Fungi

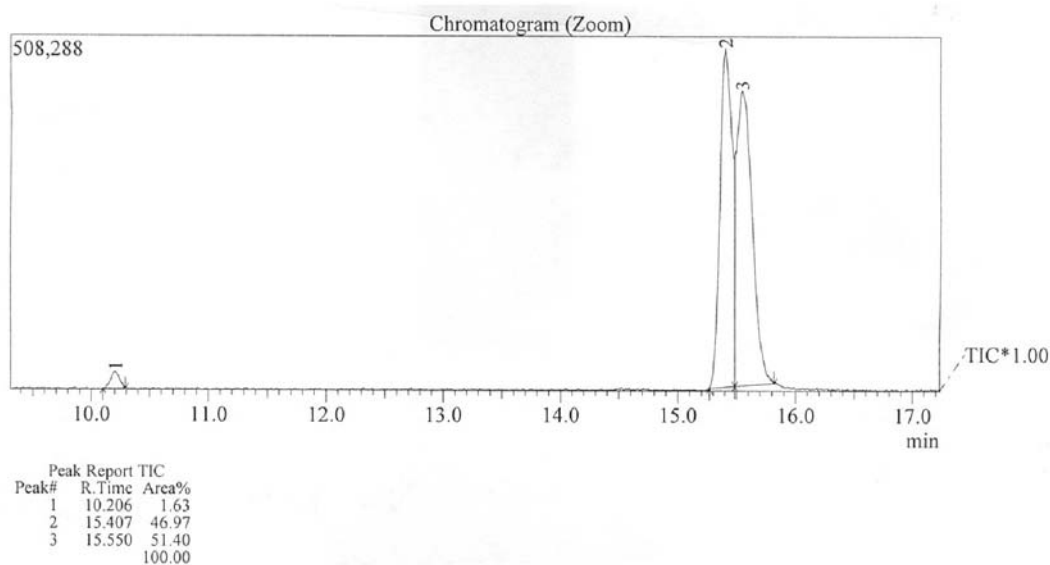
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Chromatograms used for the determination of conversion, enantiomeric excess and enantiomeric ratio of (*R,S*)-2-octanol resolution reactions are presented. Analyses were performed by gas chromatography coupled to mass spectrometry (Shimadzu GCMS-QP2010) using a chiral

stationary phase (Chirasil-Dex CB 25 m × 0.25 mm ID × 0.25 mm). Column temperature was 80 °C. The He pressure was 56.9 kPa and the temperatures of the injector and the detector were 220 °C and 275 °C, respectively.

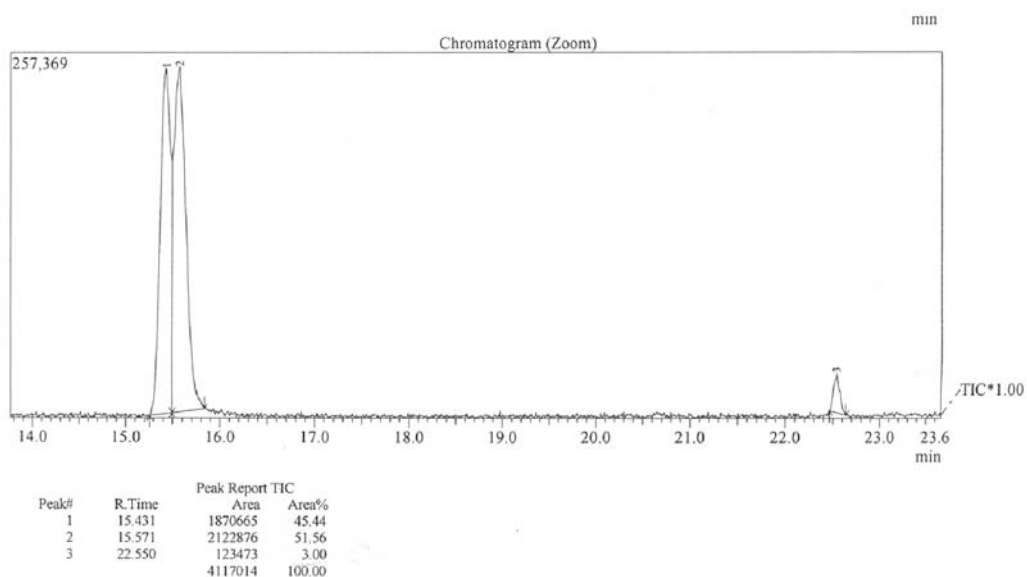


Peak # 1: Reaction By-product

Peak # 2: (*R*) or (*S*)-2-octanol

Peak # 3: (*R*) or (*S*)-2-octanol

Figure S1. GC-MS chromatogram of (*R,S*)-2-octanol resolution mediated by the isolate UEA_001.

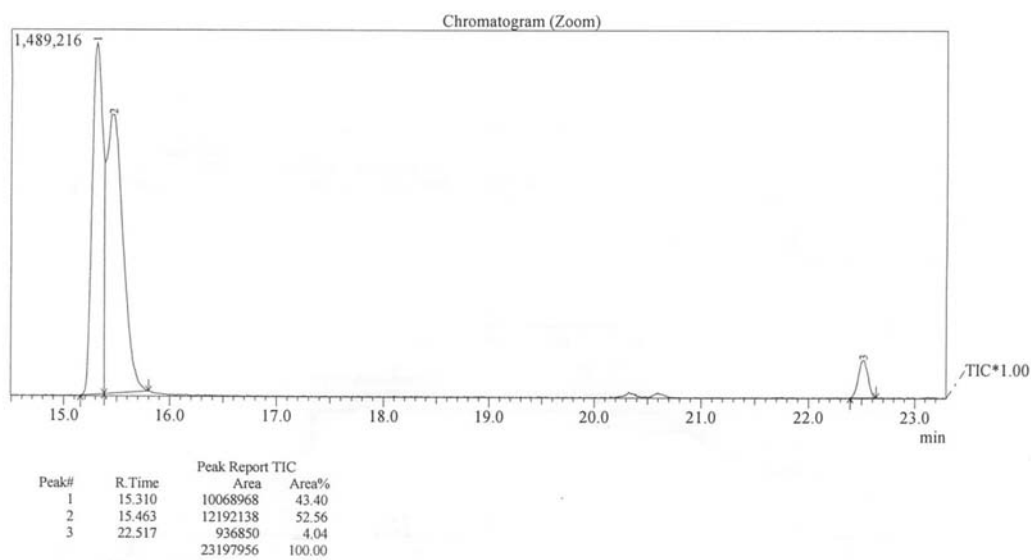


Peak # 1: (*R*) or (*S*)-2-octanol

Peak # 2: (*R*) or (*S*)-2-octanol

Peak # 3: (*R*) or (*S*)-1-methylheptil acetate

Figure S2. GC-MS chromatogram of (*R,S*)-2-octanol resolution mediated by the isolate UEA_006.

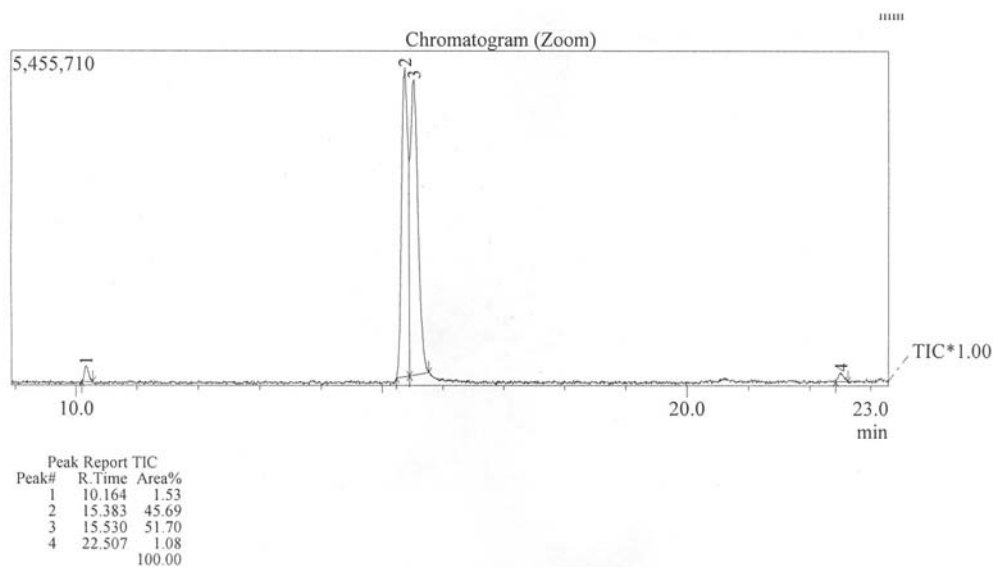


Peak # 1: (*R*) or (*S*)-2-octanol

Peak # 2: (*R*) or (*S*)-2-octanol

Peak # 3: (*R*) or (*S*)-1-methylheptil acetate

Figure S3. GC-MS chromatogram of (*R,S*)-2-octanol resolution mediated by the isolate UEA_007.

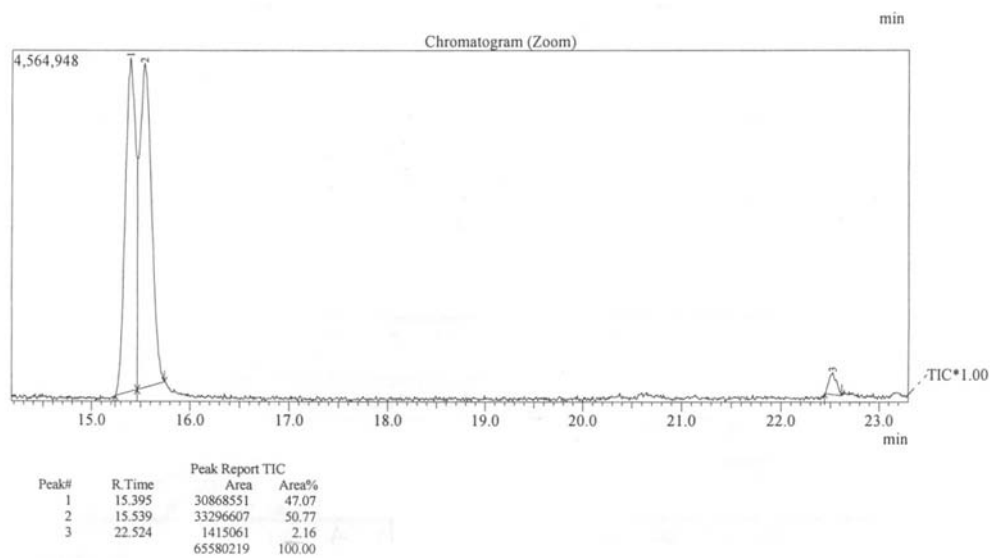


Peak # 1: Reaction By-product

Peak # 2: (*R*) or (*S*)-2-octanol

Peak # 3: (*R*) or (*S*)-2-octanol

Figure S4. GC-MS chromatogram of (*R,S*)-2-octanol resolution mediated by the isolate UEA_014.

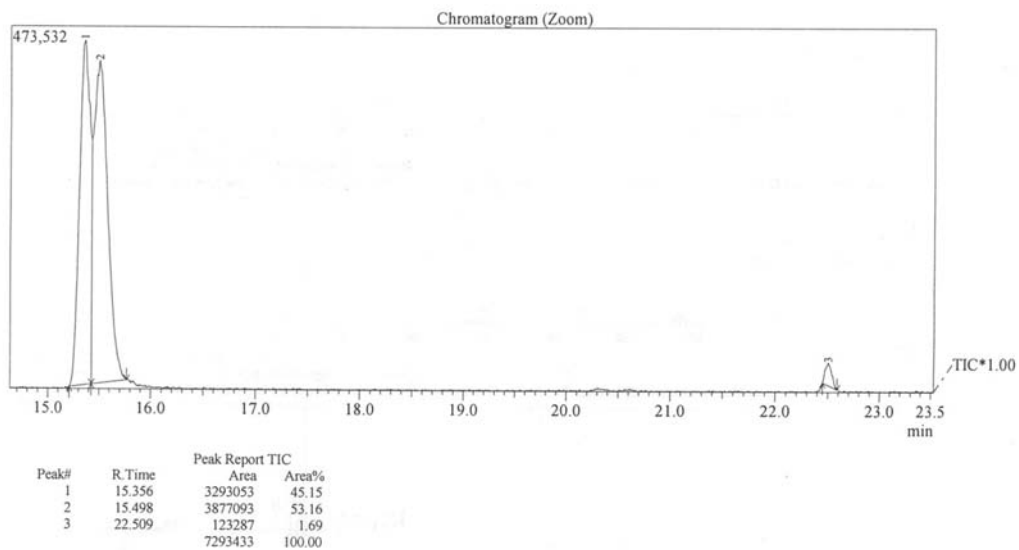


Peak # 1: (*R*) or (*S*)-2-octanol

Peak # 2: (*R*) or (*S*)-2-octanol

Peak # 3: (*R*) or (*S*)-1-methylheptil acetate

Figure S5. GC-MS chromatogram of (*R,S*)-2-octanol resolution mediated by the isolate UEA_023.

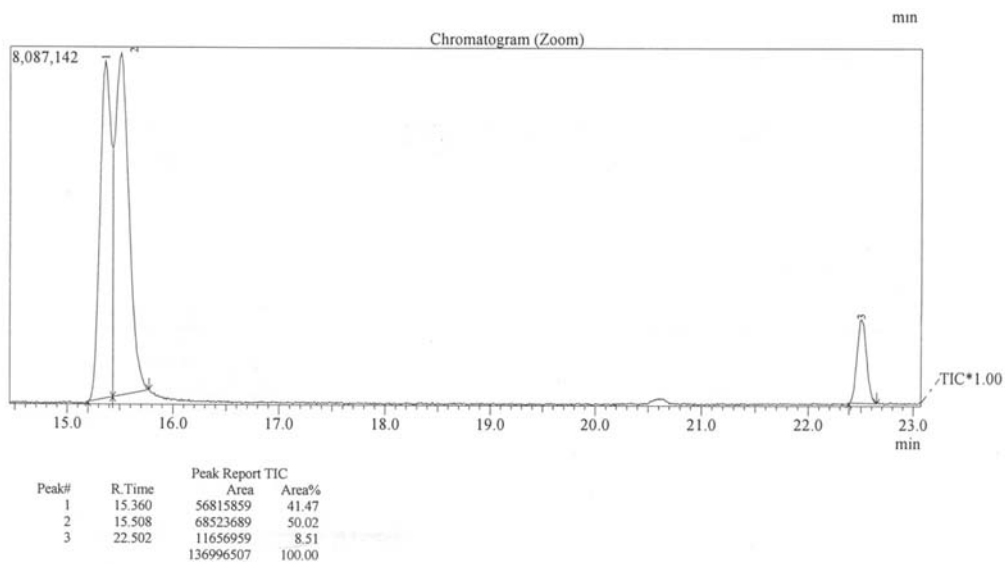


Peak # 1: (*R*) or (*S*)-2-octanol

Peak # 2: (*R*) or (*S*)-2-octanol

Peak # 3: (*R*) or (*S*)-1-methylheptil acetate

Figure S6. GC-MS chromatogram of (*R,S*)-2-octanol resolution mediated by the isolate UEA_027.

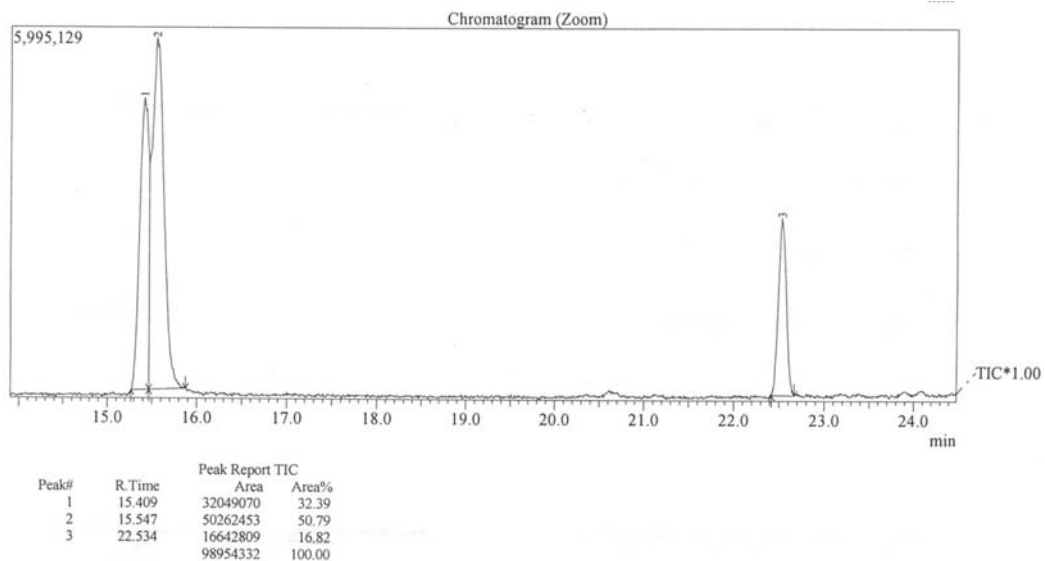


Peak # 1: (*R*) or (*S*)-2-octanol

Peak # 2: (*R*) or (*S*)-2-octanol

Peak # 3: (*R*) or (*S*)-1-methylheptil acetate

Figure S7. GC-MS chromatogram of (*R,S*)-2-octanol resolution mediated by the isolate UEA_041.

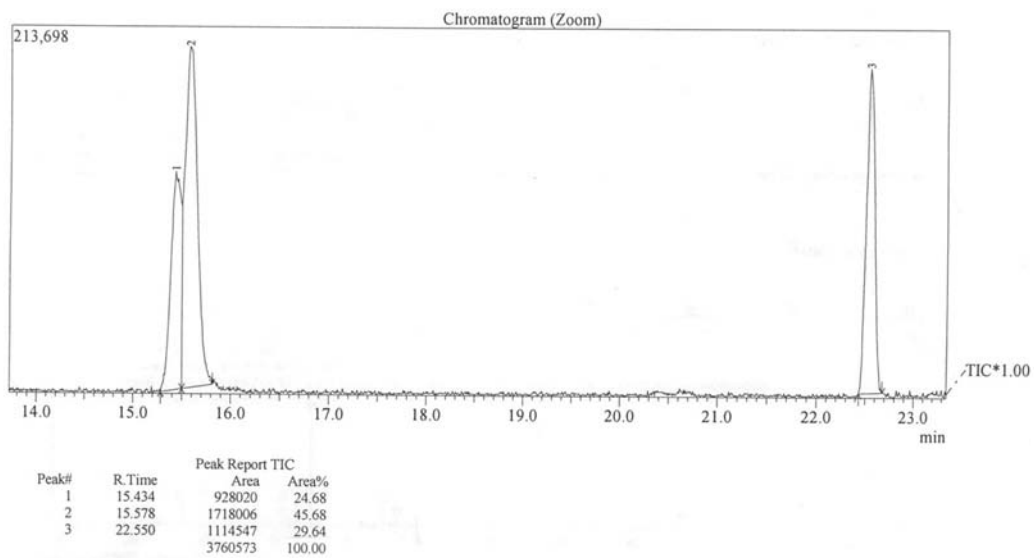


Peak # 1: (*R*) or (*S*)-2-octanol

Peak # 2: (*R*) or (*S*)-2-octanol

Peak # 3: (*R*) or (*S*)-1-methylheptil acetate

Figure S8. GC-MS chromatogram of (*R,S*)-2-octanol resolution mediated by the isolate UEA_053.

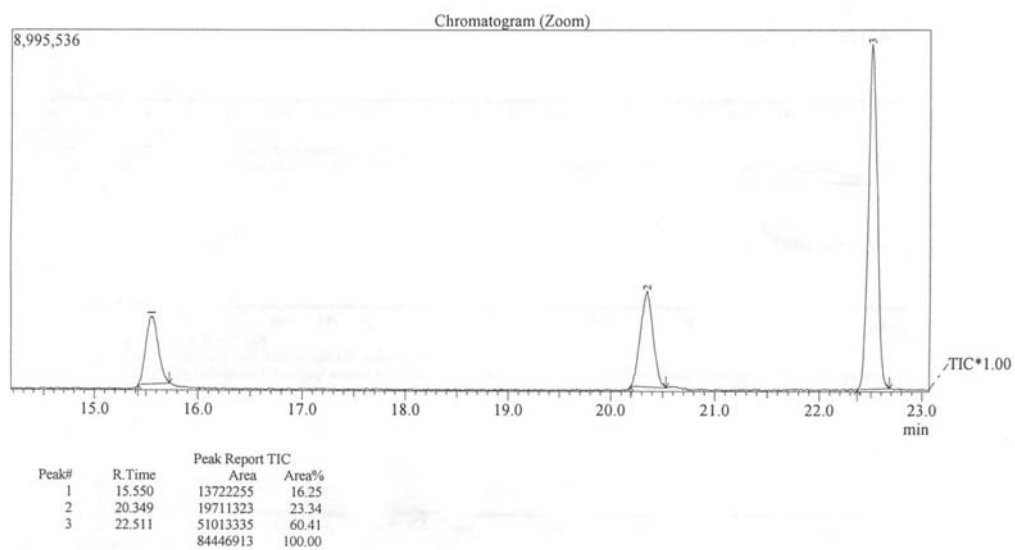


Peak # 1: (*R*) or (*S*)-2-octanol

Peak # 2: (*R*) or (*S*)-2-octanol

Peak # 3: (*R*) or (*S*)-1-methylheptil acetate

Figure S9. GC-MS chromatogram of (*R,S*)-2-octanol resolution mediated by the isolate UEA_115.



Peak # 1: (*R*) or (*S*)-2-octanol

Peak # 2: (*R*) or (*S*)-2-octanol

Peak # 3: (*R*) or (*S*)-1-methylheptil acetate

Figure S10. GC-MS chromatogram of (*R,S*)-2-octanol resolution mediated by the commercial enzyme Novozym 435.