

Prenylindole Alkaloids from *Raputia praetermissa* (Rutaceae) and their Chemosystematic Significance

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O extrato diclorometano do caule de *Raputia praetermissa* levou ao isolamento de quatro compostos novos, 4-desóxi-raputindol C (**1**), raputimonindol A-B (**2, 3**) e hexadecanil 2-hidróxi-4-metóxi-cinnamato (**5**), juntamente com os alcalóides 5-(4-metóxi-metilfuran-2-il)-1*H*-indol (raputimonindol C), furoquinolinos maculosidine, robustine, evolitrine e dictamine. O estudo do extrato hexano levou ao isolamento de *N*-metil-4-metóxi-quinolin-2(1*H*)-ona, skimmianina, cicloartenona, sitosterol, stigmasterol e sitostenona. Os alcalóides antranfílicos isolados indicam que o gênero possui afinidade química relevante com aqueles da tribo Cusparieae, mas difere de *Neoraputia* devido à ausência de alcalóides prenilindoís neste último, cujas espécies foram anteriormente incluídas em *Raputia*.

The dichloromethane extract from the stems of *Raputia praetermissa* afforded four new compounds, 4-deoxyraputindole C (**1**), raputimonindole A-B (**2, 3**), and hexadecanyl 2-hydroxy-4-methoxy-cinnamate (**5**), besides the alkaloids 5-(4-methoxymethylfuran-2-yl)-1*H*-indole (raputimonindole C), furoquinolines maculosidine, robustine, evolitrine and dictamine. The hexane extract yielded *N*-methyl-4-methoxyquinolin-2(1*H*)-one, skimmianine, cycloartenone, sitosterol, stigmasterol and sitostenone. The anthranilate alkaloid content indicates that the genus is strongly related to those included in Cusparieae tribe, but differs from *Neoraputia* by the absence of prenylindole alkaloids in the late, whose species have previously been placed in *Raputia*.

Keywords: *Raputia praetermissa*, *Neoraputia*, Rutaceae, prenylindole alkaloids, chemosystematics

Introduction

The *Raputia* genus was established by Aublet in 1775,¹ and based on morphological characteristics of *R. aromatica* Aubl. Emmerich later dismembered this genus,¹ placing most of the species into *Neoraputia* Emmerich, *Sigmatanthus* Huber ex Emmerich, and *Raputiarana* Emmerich. Following the research of Kallunki and Pirani,^{2,5} a total of eleven species have now been included in the *Raputia* genus: *R. aromatica* Aubl., *R. maroana* (R. S. Cowan) Kallunki, *R. neblinensis* (R. S. Cowan) Kallunki, *R. ulei* (K. Krause) Kallunki, *R. brevipedunculata* Kallunki, *R. megalantha* Kallunki, *R. simulans* Kallunki, *R. amazonica* (Huber) Kallunki (synonym: *Ravenia amazonica* Huber), *R. szczyrbanii* (Steyerm.) Kallunki (synonym: *Lubaria szczyrbanii* Steyerm.), *R. hirsuta* (Gereau) Kallunki and

R. praetermissa Pirani & Kallunki. *Raputia* and *Neoraputia* are assigned to the tribe Cusparieae and are distributed from Venezuela and French Guiana to Amazonian Colombia, Peru and Brazil.⁵

Previous investigations of *Neoraputia* reported the presence of eleven polymethoxylated flavonoids, six flavones, three 5,6-(2'',2'')-dimethylpyrano)flavones, one 6,7-(2'',2'')-dimethylpyrano)flavone and one flavanone from *N. alba* (Engler) Emmerich;^{6,7} five polymethoxylated flavones and two flavanones, 2'-hydroxy-3,4,4',5,6'-pentamethoxychalcone, three 5',6'-(2'',2'')-dimethylpyrano)-polymethoxylated chalcones from *N. magnifica* var. *magnifica* (Engler) Emmerich;^{8,9} ten polymethoxylated flavonoids, six flavones, three 6,7-(2'',2'')-dimethylpyrano) flavones and one 6-(3''-hydroxy,3''-methyl-*trans*-but-1''-enyl)flavone from *N. paraensis*.^{10,11} A reinvestigation of *N. paraensis* searching for alkaloids afforded flindersine, skimmianine, 8-methoxyflindersine and dictamine.¹²

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C-glycosylflavones were also reported from *N. paraensis*,¹³ which was cited as *Raputia paraensis*, but this species was transferred to *Neoraputia* by Emmerich.¹

The first investigation about the chemistry of *Raputia* reported the presence of cyclopentyl bisindole alkaloids raputiindoles A-D from *R. simulans* Kllunki.¹⁴

In this paper we report a phytochemical study on *R. praetermissa* Pirani & Kallunki, and the chemosystematic significance of isolated compounds is discussed in order to clarify the relationships between *Raputia* and *Neoraputia*.

Results and Discussion

The dichloromethane extract from the stems of *Raputia praetermissa* afforded four prenylindole alkaloids (**1-4**), a cinnamic acid derivative (**5**) (Figure 1), and furoquinoline alkaloids maculosidine, robustine, evolitrine¹⁵ and dictamnine.¹⁶ The hexane extract yielded *N*-methyl-4-methoxyquinolin-2(1*H*)-one,¹⁷ skimmianine,¹⁷ cycloartenone,¹⁸ sitosterol, stigmasterol and sitostenone.

Compound **1**, C₂₆H₂₆N₂ (HREIMS), was identified as a bisindole alkaloid. The presence of two indole nucleus was suggested by an UV absorption maximum at 328 nm, an IR band at 3426 cm⁻¹ (NH), and ¹H NMR signals for N-H protons at δ 8.00 and 8.04 (brs, no correlation in the HSQC spectrum) (Table 1), which in the COSY experiments showed cross peaks with the ¹H signals at δ _H 7.14 (dd, 3.2, 2.5 Hz), 6.50 (ddd, 3.2, 2.5, 1.0 Hz), 7.11 (dd, 3.0, 2.5 Hz) and 6.44 (ddd, 3.0, 2.5, 1.0 Hz), respectively. These signals were then assigned to 2N-H (δ 8.00 and 8.04), 2H- α (δ _H 7.14 and 7.11) and 2H- β (δ _H 6.50 and 6.44) of the indole rings, respectively. HMBC cross peaks (Figure 2) between the signals H-4' (δ _H 7.30)/C-3' (δ _C 102.3), C-6' (δ _C 144.3),

7'a (δ _C 135.5) and 40.7 (CH); H-7' (δ _H 7.18)/C-3'a (δ _C 127.3), C-5' (δ _C 139.4), and δ _C 48.5 (quaternary carbon) led to the assignment of a 5',6'-dialkylindole system. Moreover, the observed cross peaks between the ¹H signals at δ _H 4.07 (H-6), 2.43 (H-5a) and 1.81 (H-5b), and the ¹³C signals for C-5', C-6' and 48.5 (quaternary carbon, C-3) suggested a 3,3,6-trisubstituted cyclopentyl fused to the indole ring at C-5' and C-6'. An isobutene group was identified from the ¹H NMR signals at δ _H 5.22 (dsep, 9.0, 1.0), 1.82 (br s, 3H), and 1.79 (br s, 3H), assigned to olefinic proton and methyl groups, respectively. This was supported by the HMBC experiments which showed correlations from the olefinic proton at δ _H 5.22 to the methyl carbons at δ _C 25.9 and 18.3. The isobutene group was connected at C-6 due to coupling of H-6 to the olefinic proton at δ _H 5.22. A methyl group must be connected at C-3 due to the observed HMBC cross peaks between its ¹H signal at δ _{3H} 1.59 and the ¹³C signals for C-6' and C-3. This structural unit was corroborated by the MS base-peak at *m/z* 223.13387 (100.0%) resulting from C-3-C-2 cleavage.

The ¹H NMR spectrum also showed signals for three hydrogens with *ortho* and *meta* coupling constants, δ _H 7.50 (d, 0.7 Hz), 7.23 (dd, 8.4, 0.7 Hz) and 7.22 (d, 8.4 Hz), suggesting the second indole nucleus to be monosubstituted. In addition, the presence of a *trans*-disubstituted double bond was evidenced by two vinylic protons at δ _H 6.13 (δ _C 126.8) and 6.40 (d, 16.0 Hz; δ _C 136.0) with a vicinal coupling constant of 16.0 Hz. From HMBC experiments the observed cross peak between the signal with *meta* coupling constant at δ _H 7.50 and the ¹³C signal for C-3'' (δ _C 102.7), permitted the assignment of the signal at δ _H 7.50 to H-4'' (C-4'', δ _C 118.7). The *meta* coupling constant for H-4'' indicated that C-5'' was substituted. The

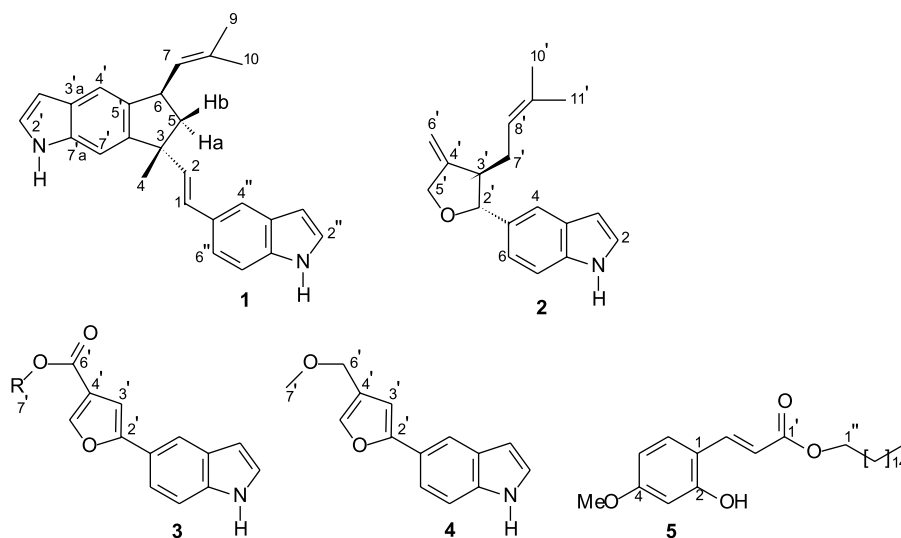


Figure 1. Compounds isolated from *Raputia praetermissa*.

Table 1. ^1H NMR data for **1-4**

H	1	H	2	3	4
1'	8.00 br s				
2'	7.14 dd (3.2, 2.5)				
3'	6.50 ddd (3.3, 2.5, 1.0)				
4'	7.30 s				
7'	7.18 s	2'	4.64 d (5.6)		
1	6.13 d (16.0)	3'	2.73 td (7.0, 5.6)	6.88 s	6.65 s
2	6.40 d (16.0)	5'a	4.44 dd (13.1, 2.3)	8.04 s	7.46 s
4	1.59 s	5'b	4.65 dd (13.1, 2.3)		
5a	2.43 dd (12.1, 6.8)	6'a	4.99 d (2.3)		4.11 s
5b	1.81 dm (12.1)	6'b	4.97 d (2.3)		
6	4.07 dm (9.0)	7'	2.28 br t (7.0)	3.87 s	3.44 s
7	5.22 dsep (9.0, 1.0)	8'	5.12 tsep (7.0, 1.2)		
9	1.82 br s	10'	1.64 br s		
10	1.79 br s	11'	1.60 br s		
1''	8.04 br s	1	8.15 br s	9.85 br s	8.35 br s
2''	7.11 dd (3.0, 2.5)	2	7.19 d (3.1, 1.6)	7.25 dd (2.0, 1.2)	7.14 t (2.4)
3''	6.44 ddd (3.0, 2.5, 1.0)	3	6.53 m	6.55 br t (2.0)	6.58 m
4''	7.50 d (0.7)	4	7.60 d (0.8)	7.97 br s	8.00 br s
6''	7.23 dd (8.4, 0.7)	6	7.20 dd (8.5, 2.0)	7.50 dd (8.5, 1.6)	7.53 dd (8.4, 1.8)
7''	7.22 d (8.4)	7	7.37 d (8.5)	7.43 d (8.5)	7.33 d (8.4)

^1H NMR spectrum was acquired in CDCl_3 at 400 MHz. TMS was used as internal standard. Chemical shifts are shown in the δ scale with J values (Hz) in parentheses.

attachment of the ethylenyl bridge to C-3 and C-5'' was evidenced from HMBC correlations H-1 (δ_{H} 6.13)/C-3, C-4'' and H-2 (δ_{H} 6.40)/C-3, C-6'. These spectral characteristics are in agreement with those published for raputindole C (**6**, Figure 3), isolated from *Raputia simulans*.¹⁴ The main difference observed in the ^1H and ^{13}C NMR spectra (Tables 1 and 2) of compound **1**, when compared to those of **6**, was the replacement of the signal for an oxymethylene by a methyl singlet (C-4, δ_{3H} 1.59, δ_{C} 27.1). The relative stereochemistry of compound **1**, named as 4-deoxyraputindole C, was determined from gNOESY experiments. The nOes of H-2, H-1, and H-5a, coming from H-6, indicated that H-6 and the ethylenylindole system are on the same side of the five-membered ring, whereas nOe between H-5b and CH_3 -4, required the methyl group to be *anti* (β) to H-6, and *syn* to the isobutene chain.

Compounds **2-4** showed the spectral characteristic of a 5-substituted-1*H*-indole alkaloid (Tables 1, 2). The ^{13}C NMR spectrum (Table 2) revealed resonances for C-2 to C-7a in close agreement with those for the corresponding carbons in the structural unit 5-substituted-1*H*-indole of compound **1**, except for C-5 whose chemical shift was affected by a different substituent. Elemental analysis and

MS indicated the molecular formula to be $\text{C}_{18}\text{H}_{21}\text{NO}$ for compound **2**, requiring the presence of an aliphatic chain of 10 carbons and one oxygen. A heterocyclic oxolane was identified from the ^1H NMR spectrum, which showed a proton at δ_{H} 2.73 (td, 7.0, 5.6, H-3'; δ_{C} 51.6) coupled to a proton attached to carbon adjacent to an oxygen atom (δ_{H} 4.64, d, 5.6, H-2'; δ_{C} 86.6), and two doublet of doublets for an oxymethylene (δ_{H} 4.44, 4.65, J 13.1, 2.3, 2H-5'; δ_{C} 71.5). Observed HMBC cross peaks between the ^1H signals at δ_{H} 2.73 (H-3') and 4.64 (H-2') with the ^{13}C signals at δ_{C} 132.7 (C-5) and 119.1 (C-4), respectively, as well as those of H-5'a and H-5'b (δ_{H} 4.44 and 4.65) with C-3' (δ_{C} 51.6), suggested the attachment of the heterocyclic oxolane to C-5 of the indole nucleus. The presence of an isopentene group was deduced from the proton resonances at δ_{H} 5.12 (tsep, 7.0, 1.2, H-8'), 1.64 (br s, 3H-10'), 1.60 (br s, 3H-11'), 2.28 (br t, 7.0, 2H-7'), and corroborated by the HMBC correlation between the methyl and methylene protons with the olefinic carbons at δ_{C} 132.7 (C-9') and 121.7 (C-8'). This was supported by the mass spectrum which showed fragment at m/z 211 [$\text{C}_{18}\text{H}_{21}\text{NO} - \text{C}_4\text{H}_8$]⁺. The ^1H - ^1H coupling between the methylene at δ_{H} 2.28 and H-3' (δ_{H} 2.73) indicated the linkage of the isopentene chain to C-3'. An oxymethylene (δ_{H} 4.99, 4.97, J 2.3, 2H-6';

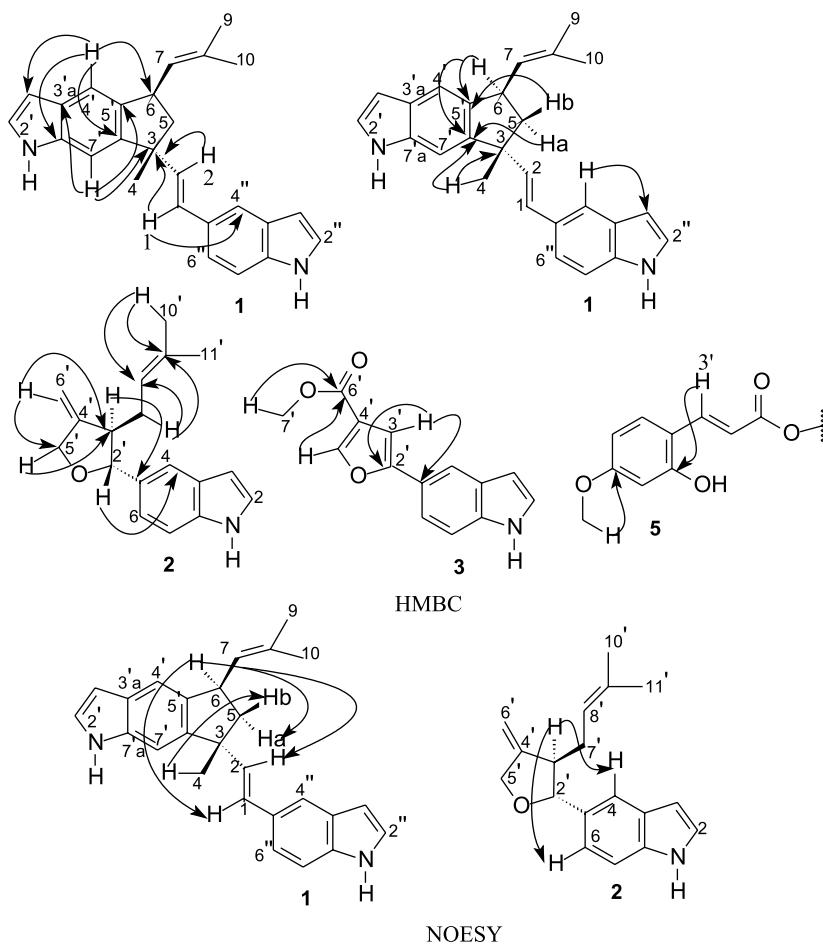


Figure 2. Relevant HMBC and NOESY interactions observed for compounds **1-3** and **5**.

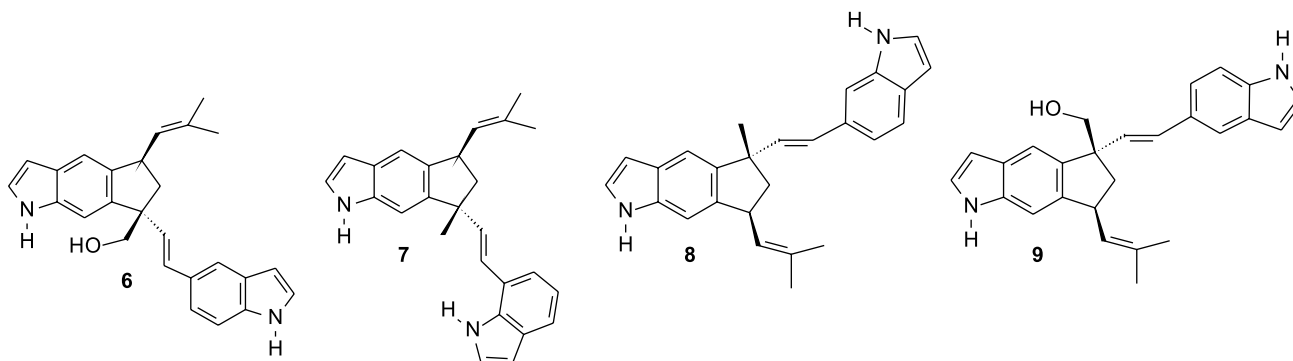


Figure 3. Raputindoles **A** (**8**), **B** (**7**), **C** (**6**) and **D** (**9**) isolated from *Raputia simulans*.

δ_C 103.6) must be connected at C-4' of oxolane ring on the basis of the observed cross peaks between the ^1H signals at $\delta_{2\text{H}}$ 4.99/4.97 and the ^{13}C signals for C-3' (δ_C 51.6) and C-5' (δ_C 71.5). As for compound **1**, the relative stereochemistry of **2** was deduced from gNOESY experiments. The nOes of the H-4 and H-6, coming from H-3', indicated that H-3' and the indole system must be on the same side of the oxolane ring. The above data confirmed the structure of **2**, here named as raputimonindole A.

As commented above compounds **2-4** showed the spectral characteristic of a 5-substituted-1*H*-indole alkaloid. Elemental analysis and MS indicated the molecular formula to be $\text{C}_{14}\text{H}_{11}\text{NO}_3$ and $\text{C}_{14}\text{H}_{13}\text{NO}_2$ for compounds **3** and **4**, respectively, requiring the presence of an aliphatic chain of 6 carbons ($\text{C}_6\text{H}_5\text{O}_3$ and $\text{C}_6\text{H}_7\text{O}_2$, respectively) and indole nucleus ($\text{C}_8\text{H}_6\text{N}$). Their ^1H NMR resonances, when compared to those of **2**, showed low field shifts for the disubstituted furan ring protons. In

Table 2. ^{13}C NMR data for **1-4**, and **6**

C	1	6 ¹⁴	C	2	3	4
2'	123.8	124.1				
3'	102.3	102.4				
3'a	127.3	128.0				
4'	115.4	116.0				
5'	139.4	140.3				
6'	144.3	139.8				
7'	105.8	106.2				
7'a	135.5	135.5				
1	126.8	131.1	2'	86.6	156.0	156.1
2	136.0	131.2	3'	51.6	102.3	103.7
3	48.5	55.1	4'	152.1	121.0	123.8
4	27.1	68.9	5'	71.5	146.3	139.2
5	50.0	43.9	6'	103.6	165.0	66.0
6	40.7	40.6	7'	29.0	51.8	57.6
7	128.5	128.3	8'	121.7		
8	132.4	132.7	9'	132.7		
9	25.9	25.9	10'	18.1		
10	18.3	18.3	11'	25.7		
2''	124.4	123.9	2	124.4	125.5	124.9
3''	102.7	102.9	3	102.8	102.4	102.8
3''a	127.3	128.1	3a	127.7	128.0	127.9
4''	118.7	119.0	4	119.1	116.6	116.1
5''	129.8	129.3	5	132.7	123.0	123.0
6''	120.4	120.5	6	121.0	118.6	118.7
7''	110.9	111.0	7	110.9	111.7	111.2
7''a	135.1	135.4	7a	135.5	136.0	135.3

^{13}C NMR spectrum was acquired in CDCl_3 at 100 MHz. Assignments based on HSQC and HMBC experiments.

compound **3**, the existence of a cross peak between the ^1H signal at δ_{H} 6.88 assigned to H-3', and the ^{13}C signal at δ_{C} 123.0 assigned to C-5, determined the position of the furan ring at C-5 of the indole nucleus. The presence of a carbomethoxy group linked at C-4' was indicated by HMBC cross peaks between the ^1H signals at δ_{H} 8.04 (H-5', δ_{C} 146.3) and 3.87 (methyl protons) with the ^{13}C signal at δ_{C} 165.0 (C-6'). The structure of the new natural product is therefore 5-(4-carbomethoxyfuran-2-yl)-1*H*-indole, here named as raputimonindole B (**3**). However, compound **3** was purified by column chromatography on Sephadex and eluted with $\text{MeOH-CH}_2\text{Cl}_2$, hence, **3** could be an artifact.

The ^1H NMR resonances for compound **4**, when compared to those of **2**, showed low field shifts for the disubstituted furan ring protons. In compound **4**, the main difference observed in the ^1H NMR, when compared with

3, was the replacement of the resonance for a carbomethoxy group by two ^1H singlets at $\delta_{2\text{H}}$ 4.11 (δ_{C} 66.0) and $\delta_{3\text{H}}$ 3.44 (δ_{C} 57.6) from a methoxymethylene group. This was supported by the mass spectrum which showed a fragment at m/z 197 [$\text{C}_{14}\text{H}_{13}\text{NO}_2 - \text{H}_2\text{CO}$]⁺. These signals together with the mass and ^{13}C NMR spectral data are consistent with **4** being 5-(4-methoxymethylfuran-2-yl)-1*H*-indole, which has previously been isolated from *Raputia simulans* Kallunki.¹⁹ However, the isolation of 5-(4-methoxymethylfuran-2-yl)-1*H*-indole was cited without spectroscopic data in an congress whose abstracts were published in *Planta Medica Proceedings*.¹⁹ Thus, its spectroscopic data are cite here for the first time, and it was named raputimonindole C.

The cinnamic acid (**5**) derivative showed the spectral characteristic of a *trans* α,β -unsaturated carboxyl functional group ($\delta_{\text{H}\beta}$ 7.61, d, *J* 15.9, $\delta_{\text{C}\beta}$ 144.5; $\delta_{\text{H}\alpha}$ 6.29, d, *J* 15.9, $\delta_{\text{C}\alpha}$ 115.7; COOR 167.3). In addition, the ^1H NMR showed signals for one methoxyl group at δ_{H} 3.92 (s, 3H; δ_{C} 55.9), three aromatic hydrogens at δ_{H} 7.07 (dd, 8.1, 1.7 Hz), 7.03 (d, 1.7 Hz) and 6.91 (d, 8.1 Hz), clearly indicating the aromatic ring to be 1,2,4-trisubstituted. From the HMBC experiments, the cross peaks observed between the signal of methoxyl group at $\delta_{3\text{H}}$ 3.92 with δ_{C} 146.7, and the ^1H signal at δ_{H} 7.61 (H-3') with δ_{C} 147.9 but not with δ_{C} 146.7, indicated the presence of a 2-hydroxy-4-methoxy-cinnamic acid derivative. The ^{13}C NMR spectrum revealed resonances for an aliphatic chain of sixteen carbons, one being attached to carboxylate as indicated by the HMBC cross peak between the ^1H signal at δ_{H} 4.18 and the ^{13}C signal at δ 167.3 (C-1'). The presence of a hexadecanyl chain was corroborated by the MS spectrum, which showed an ion at m/z 279 [$\text{HC}\equiv\text{C-COO}-(\text{CH}_2)_{14}-\text{CH}_2$]⁺. The new compound was therefore identified as hexadecanyl 2-hydroxy-4-methoxy-cinnamate (**5**). The structural assignment was also supported by comparison of its ^{13}C NMR spectrum with that of 4-hydroxy-2-methoxy-cinnamic acid.²⁰

A number of 3,5- and 3,6-diprenylated indoles have been reported for the Annonaceae genera *Isolona*,²¹⁻²³ *Uvaria*,²² *Annonidium*,²⁴ *Monodora*,²⁵ *Hexalobus*,²⁶ *Asteranthe*,²⁷ *Greenwayodendron*,²⁸ and *Polyalthia*.²⁹ However, there are only a few examples of 3-, 5-, 6- and 7- and 3,7-prenylated indoles reported in the Rutaceae genera *Raputia*,¹⁴ *Esenbeckia*,^{30,31} *Murraya*,³²⁻³⁴ *Merrillia*³⁵ and *Glycosmis*.^{36,37} Four bisindoles (**6-9**; Figure 3) similar to compound **1** have been isolated from *Raputia simulans*.¹⁴ One bisindole, yuehchukene, which may be regarded as the product of Diels-Alder-type condensation of two 3-isopentenylindoles, occurs in *Murraya* species.³² While several bisindoles derived from 2-prenyltryptamine have been isolated from *Flindersia* species (Rutaceae),³⁸

pyrano[3,2-b]indole skeleton (koniamborine), a novel type of alkaloid was isolated from *Boronella koniambiensis* (Rutaceae).³⁹

The anthranilate alkaloid content found in *R. praetermissa* indicates that the genus is strongly related to those included in Cusparieae tribe. As mentioned in the introduction, the polymethoxylated flavonoids form an extremely good marker for the *Neoraputia*. Their use in this context shows that *R. praetermissa* differs substantially from *Neoraputia* species, and reinforce its inclusion in *Raputia* genus. Furthermore, the prenylindole alkaloids have been reported only from *Esenbeckia* and *Raputia* in Cusparieae, thus suggesting an affinity of this tribe with subfamily Aurantioideae, where similar prenylindoles occur in *Murraya*,³³⁻³⁵ *Merrillia*³⁶ and *Glycosmis*.^{37,38} It is noteworthy that *Neoraputia* shares with *Murraya* and *Citrus* a propensity for producing polymethoxylated flavonoids,^{9,12,40,41} showing also chemical affinity with Aurantioideae.

Experimental

General experimental procedures

Optical rotations were measured by using a Perkin Elmer 241 spectropolarimeter; NMR: Bruker DRX 400, with TMS as internal standard; high resolution EI-MS: Fisons VG Autospec; GC-MS: low resolution on a HP-2576 instrument; IR: Bomem-FT/IR; UV: Hewlett Packard/8452A; Elemental analyses: on a EA 1108, CHNS-O (Fisons).

Plant material

Raputia praetermissa was collected in the Forest Reserve Adolpho Ducke, Amazonas, Brazil, and identified by J. R. Pirani (Department of Botany, University of São Paulo). A voucher specimen (189865) is deposited in the Herbarium of the Instituto Nacional de Pesquisa da Amazônia (INPA), Manaus, AM (Brazil).

Extraction and isolation

Ground stems (4.4 kg) were extracted 3 times at room temperature using hexane, followed by CH₂Cl₂ and MeOH. The concentrated hexane extract (13.3 g) was subjected to silica gel (230-400 mesh) column chromatography with successive elution with hexane, CH₂Cl₂, EtOAc and MeOH, yielding 6 fractions. Fraction 2 was flash rechromatographed twice on silica gel with successive elution with hexane, CH₂Cl₂, EtOAc and MeOH, and then by preparative TLC

(silica gel; hexane-acetone 9:1), yielding cycloartenone (10 mg). Fraction 3 was flash rechromatographed twice as above, and then by gel permeation column chromatography (Sephadex LH 20, CH₂Cl₂-MeOH 2:8) affording *N*-methyl-4-methoxyquinolin-2(1*H*)-one (30 mg). Fraction 4 was chromatographed on silica gel and Florisil (1:1) with hexane-EtOAc-MeOH gradient elution to give two fractions (A and B). Fraction A was subjected to column chromatography over silica gel and eluted with hexane-acetone gradient, yielding a mixture of sitosterol and stigmasterol. Fraction B was purified by preparative TLC (silica gel; hexane-acetone 9:1), yielding sitostenone (60 mg). Fraction 5 was chromatographed on silica gel and Florisil (1:1) with hexane-EtOAc-MeOH gradient elution to give skimmianine (50 mg).

The concentrated dichloromethane extract (30.0 g) was subjected to column chromatography over silica gel (70-230 mesh) under vacuum. Elution with hexane, CH₂Cl₂, EtOAc and MeOH yielded 4 fractions. Fraction 1 was flash rechromatographed on silica gel with hexane-EtOAc-MeOH gradient, yielding compound **1** (700 mg) and a new fraction C. Fraction C was flash rechromatographed twice as above, and then by preparative TLC (silica gel; hexane-acetone 6:1), yielding compound **5**. Fraction 2 was chromatographed on silica gel and Florisil (1:1) with hexane-acetone-MeOH gradient elution to give new fractions D, E and F. Fractions D and E were rechromatographed over Sephadex LH 20 (MeOH) to give compounds **2** and **4**, respectively. Fraction F was flash rechromatographed on silica gel with hexane-acetone-MeOH gradient elution, yielding robustine (50 mg). Fraction 3 was rechromatographed as above using hexane-CH₂Cl₂-MeOH gradient to yield two fractions. Both fractions were rechromatographed over Sephadex LH 20 (MeOH-CH₂Cl₂ 2:8) affording maculosidine (80 mg), and a new fraction containing compound **3** which was purified by preparative TLC (silica gel; hexane-acetone 5:1). Fraction 4 was rechromatographed on silica gel and Florisil (1:1) with hexane-acetone-MeOH gradient elution to give evolitrine (50 mg) and dictamnine 120 mg).

4-Deoxyraputindole C (1)

Brown solid; $[\alpha]_D^{25} + 94$ (CHCl₃; *c* 0.0012); UV (acetone) λ_{max}/nm : 230; IR (KBr) ν_{max}/cm^{-1} : 3425.7; ¹H NMR (400 MHz, CDCl₃), see Table 1; ¹³C NMR (100 MHz, CDCl₃), see Table 2; HREI-MS, 366.20560 (37.5; calc. for C₂₆H₂₆N₂), 294.11201(10.0), 223.13387 (100), 167.07123 (20), 130.06388 (30).

Raputimonindole A (2)

Yellow solid; $[\alpha]_D^{25} - 41$ (CHCl₃; *c* 0.003); UV (acetone) λ_{max}/nm : 228; IR (KBr) ν_{max}/cm^{-1} : 3411.5; ¹H NMR

(400 MHz, CDCl₃), see Table 1; ¹³C NMR/DEPT (100 MHz, CDCl₃), see Table 2. Anal. found C 80.28%, H 7.80%, N 5.20%; calc. for C₁₈H₂₁NO, C 80.86%, H 7.92%, N 5.24%, O 5.98 %; MS *m/z* 267 [M]⁺ (10), 252 (5), 211 (50), 144 (100), 107 (80), 79 (70).

Raputimonoindeole B (3)

Yellow solid; UV (acetone) λ_{max}/nm: 230; IR (KBr) ν_{max}/cm⁻¹: 3310.2, 1770.1; ¹H NMR (400 MHz, CDCl₃), see Table 1; ¹³C NMR/DEPT (100 MHz, CDCl₃), see Table 2. Anal. found C 69.76%, H 4.58%, N 5.80%; calc. for C₁₄H₁₁NO₃, C 69.70%, H 4.60%, N 5.81%, O 19.90 %; MS *m/z* 241 [M]⁺ (100), 226 [C₁₄H₁₁NO₃ – Me]⁺ (15), 210 [C₁₄H₁₁NO₃ – OMe]⁺ (10), 198 (10), 154 (30), 105 (40), 77 (50).

Raputimonoindeole C (4)

Yellow solid; UV (acetone) λ_{max}/nm: 232; IR (KBr) ν_{max}/cm⁻¹: 3315.4; ¹H NMR (400 MHz, CDCl₃), see Table 1; ¹³C NMR/DEPT (100 MHz, CDCl₃), see Table 2. Anal. found C 73.90%, H 5.80%, N 6.15%; calc. for C₁₄H₁₃NO₂, C 73.99%, H 5.77%, N 6.16%, O 14.08 %; MS *m/z* 227 [M]⁺ (100), 197 (80), 168 (70), 98 (30).

Hexadecanyl 2-hydroxy-4-methoxy-cinnamate (5)

Amorphous white solid; ¹H NMR (400 MHz, CDCl₃): δ 7.61 (d, *J* 15.9 Hz, H-3'), 7.07 (dd, *J* 8.1, 1.7 Hz, H-5), 7.03 (d, *J* 1.7 Hz, H-3), 6.91 (d, *J* 8.1 Hz, H-6), 6.29 (d, *J* 15.9, H-2'), 3.92 (s, OMe), 4.18 (t, *J* 6.8 Hz, H-1''), 1.69 (quint, *J* 6.8 Hz, H-2''), 1.25 (br s, 3''-15'') and 0.88 (t, *J* 6.6 Hz, H-16); ¹³C NMR (100 MHz, CDCl₃): δ 167.3 (C-1'), 147.9 (C-2), 146.7 (C-4), 144.5 (C-3'), 127.0 (C-1), 123.0 (C-5), 115.7 (C-2'), 114.6 (C-6), 109.3 (C-3), 64.5 (C-1''), 55.9 (OMe), 29.6-28.7 (C-3''-C-15''), 14.0 (C-16''); MS *m/z* 418 [M]⁺ (5), 279 (10), 207 [279 – C₅H₁₂]⁺ (10), 167 (50), 149 (100), 71 (40), 57 (45).

Note from the Editor

During the edition of the present paper, the spectroscopic data of compound **4** have been published online on March 7, 2011 by *Planta Medica*, in the entitled Letter "Simple Indole Alkaloids from the Neotropical Rutaceous Tree *Raputia simulans*" by K. Vougiannopoulou, N. Fokialakis, N. Aliagiannis, C. Cantrel, A-L Skaltsounis.

Supplementary Information

¹H and ¹³C NMR spectra of compounds **1-5** are available free of charge at <http://jbcs.sbq.org.br> as PDF file.

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References

1. Aublet, J. B. C. F.; *Histoire des Plantes de la Guiane Française*, Vol. 2; P. E. Didot: London and Paris, 1775; Emmerich, M.; *Rodriguesia* **1978**, *30*, 223.
2. Kallunki, J. A.; *Brittonia* **1990**, *42*, 175.
3. Kallunki, J. A.; *Brittonia* **1994**, *46*, 279.
4. Kallunki, J. A.; *Brittonia* **2009**, *61*, 28.
5. Pirani, J. R.; *Rodriguesia* **2005**, *56*, 189.
6. Arruda, A. C.; Vieira, P. C.; Fernandes, J. B.; da Silva, M. F. G. F.; Francisco, R. H. P.; Rodrigues, A. M. G. D.; Lechat, J. R.; *Phytochemistry* **1991**, *30*, 3157.
7. Arruda, A. C.; Vieira, P. C.; Fernandes, J. B.; da Silva, M. F. G. F.; *J. Braz. Chem. Soc.* **1993**, *4*, 80.
8. Passador, E. A. P.; da Silva, M. F. G. F.; Rodrigues-Fo, E.; Fernandes, J. B.; Vieira, P. C.; Pirani, J. R.; *Phytochemistry* **1997**, *45*, 1533.
9. Tomazela, D. M.; Pupo, M. T.; Passador, E. A. P.; da Silva, M. F. G. F.; Vieira, P. C.; Fernandes, J. B.; Rodrigues-Fo, E.; Oliva, G.; Pirani, J. R.; *Phytochemistry* **2000**, *55*, 643.
10. Souza, J. P. I.; Arruda, A. C.; Arruda, M. S. P.; *Fitoterapia* **1995**, *66*, 465.
11. Souza, J. P. I.; Arruda, A. C.; Muñoz, G. D.; Arruda, M. S. P.; Müller, A. H.; *Phytochemistry* **1999**, *52*, 1705.
12. Moraes, V. R. S.; Tomazela, D. M.; Ferracin, R. J.; Garcia, C. F.; Sannomiya, M.; Soriano, M. P. C.; da Silva, M. F. G. F.; Vieira, P. C.; Fernandes, J. B.; Rodrigues-Fo, E.; Magalhães, E. G.; Magalhães, A. F.; Pimenta, E. F.; de Souza, D. H. F.; Oliva, G.; *J. Braz. Chem. Soc.* **2003**, *14*, 380.
13. Bakhtiar, A.; Gleye, J.; Moulis, C.; Fouraste, I.; *Phytochemistry* **1991**, *30*, 3840.
14. Vougiannopoulou, K.; Fokialakis, N.; Aliagiannis, N.; Cantrell, C.; Skaltsounis, A.-L.; *Org. Lett.* **2010**, *12*, 1908.
15. Robertson, A. V.; *Aust. J. Chem.* **1963**, *16*, 451.
16. Cortez, L. E. R.; Cortez, D. A. G.; Ferreira, A. G.; Vieira, P. C.; da Silva, M. F. G. F.; Fernandes, J. B.; *Braz. J. Pharmacogn.* **2006**, *16*, 164.
17. Nayar, M. N. S.; Sutar, C. V.; Bhan, M. K.; *Phytochemistry* **1971**, *10*, 2843.
18. Khuong-Huu, F.; Sangare, M.; Chari, V. M.; Bekaert, A.; Devys, M.; Barbier, M.; Lukacs, G.; *Tetrahedron Lett.* **1975**, 1787.

19. Vougiopoulou, K.; Fokialakis, N.; Aligiannis, N.; Cantrell, C.; Skaltsounis, A.-L.; *Planta Med.* **2008**, *74*, 189.
20. Chen, H.; Jiang, H.; Morgan, J. A.; *Phytochemistry* **2007**, *68*, 306.
21. Makangara, J. J.; Henry, L.; Jonker, S. A.; Nkunya, M. H. H.; *Phytochemistry* **2004**, *65*, 227.
22. Achenbach, H.; Raffelsberger, B.; *Tetrahedron Lett.* **1979**, *28*, 2571.
23. Achenbach, H.; Löwel, M.; *Phytochemistry* **1995**, *40*, 967.
24. Achenbach, H.; Renner, C.; *Heterocycles* **1985**, *23*, 2075.
25. Adeoye, A. O.; Oguntimein, B. O.; Clark, A. M.; Hufford, C. D.; *J. Nat. Prod.* **1986**, *49*, 534.
26. Achenbach, H.; Renner, C.; Addae-Mensah, I.; *Heterocycles* **1984**, *22*, 2501.
27. Nkunya, M. H. H.; Jonker, S. A.; Mdee, L. K.; Waibel, R.; Achenbach, H.; *Nat. Prod. Lett.* **1996**, *9*, 71.
28. Yoo, H.-D.; Cremin, P. A.; Zeng, L.; Garo, E.; Williams, C. T.; Lee, C. M.; Goering, M. G.; O'Neil-Johnson, M.; Eldridge, G. R.; Hu, J.-F.; *J. Nat. Prod.* **2005**, *68*, 122.
29. Kunesch, N.; Cave, A.; Leboeuf, M.; Hocquemiller, R.; Dubois, G.; Guittet, E.; Lallemand, J. Y.; *Tetrahedron Lett.* **1985**, *26*, 4937.
30. Monache, F. D.; Monache, G. D.; Souza, M. A. M.; Cavalcanti, M. S.; Chiappeta, A.; *Gazz. Chim. Ital.* **1989**, *119*, 435.
31. Monache, F. D.; Benedito R. D.; *Gazz. Chim. Ital.* **1990**, *120*, 387.
32. Kinoshita, T.; Tatara, S.; Ho, F.-G.; Sankawa, U.; *Phytochemistry* **1989**, *28*, 147.
33. Wu, T.-S.; Liou, M.-J.; Jong, T.-T.; Chen, Y.-J.; Lai, J.-S.; *Phytochemistry* **1989**, *28*, 2873.
34. Wu, T.-S.; Liou, M.-J.; Lee, C.-J.; Jong, T.-T.; McPhail, A. T.; McPhail, D. R.; Lee, K.-H.; *Tetrahedron Lett.* **1989**, *30*, 6649.
35. Kong, Y.-C.; But, P. P.-H.; NG, K.-H.; Cheng, K.-F.; Chang, K.-L.; Wong, K. M.; Gray, A. I.; Waterman, P. G.; *Biochem. Syst. Ecol.* **1988**, *16*, 47.
36. Vajrodaya, S.; Bacher, M.; Greger, H.; Hofer, O.; *Phytochemistry* **1998**, *48*, 897.
37. Wang, J.; Zheng, Y.; Efferth, T.; Wang, R.; Shen, Y.; Hao, X.; *Phytochemistry* **2005**, *66*, 697.
38. Fernandez, L. S.; Buchanan, M. S.; Carroll, A. R.; Feng, Y. J.; Quinn, R. J.; Avery, V. M.; *Org. Lett.* **2009**, *11*, 329.
39. Grougnet, R.; Magiatis, P.; Fokialakis, N.; Mitaku, S.; Skaltsounis, A.-L.; Tillequin F.; Sevenet, T.; Litaudon, M.; *J. Nat. Prod.* **2005**, *68*, 1083.
40. Ferracin, R. J.; da Silva, M. F. G. F.; Fernandes, J. B.; Vieira, P. C.; *Phytochemistry* **1998**, *47*, 393.
41. Ribeiro, A. B.; Abdelnur, P. V.; Garcia, C. F.; Belini, A.; Severino, V. G. P.; da Silva, M. F. G. F.; Fernandes, J. B.; Vieira, P. C.; de Carvalho, S. A.; de Souza, A. A.; Machado, M. A.; *J. Agric. Food Chem.* **2008**, *56*, 7815.

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Supplementary Information

Prenylindole Alkaloids from *Raputia praetermissa* (Rutaceae) and their Chemosystematic Significance

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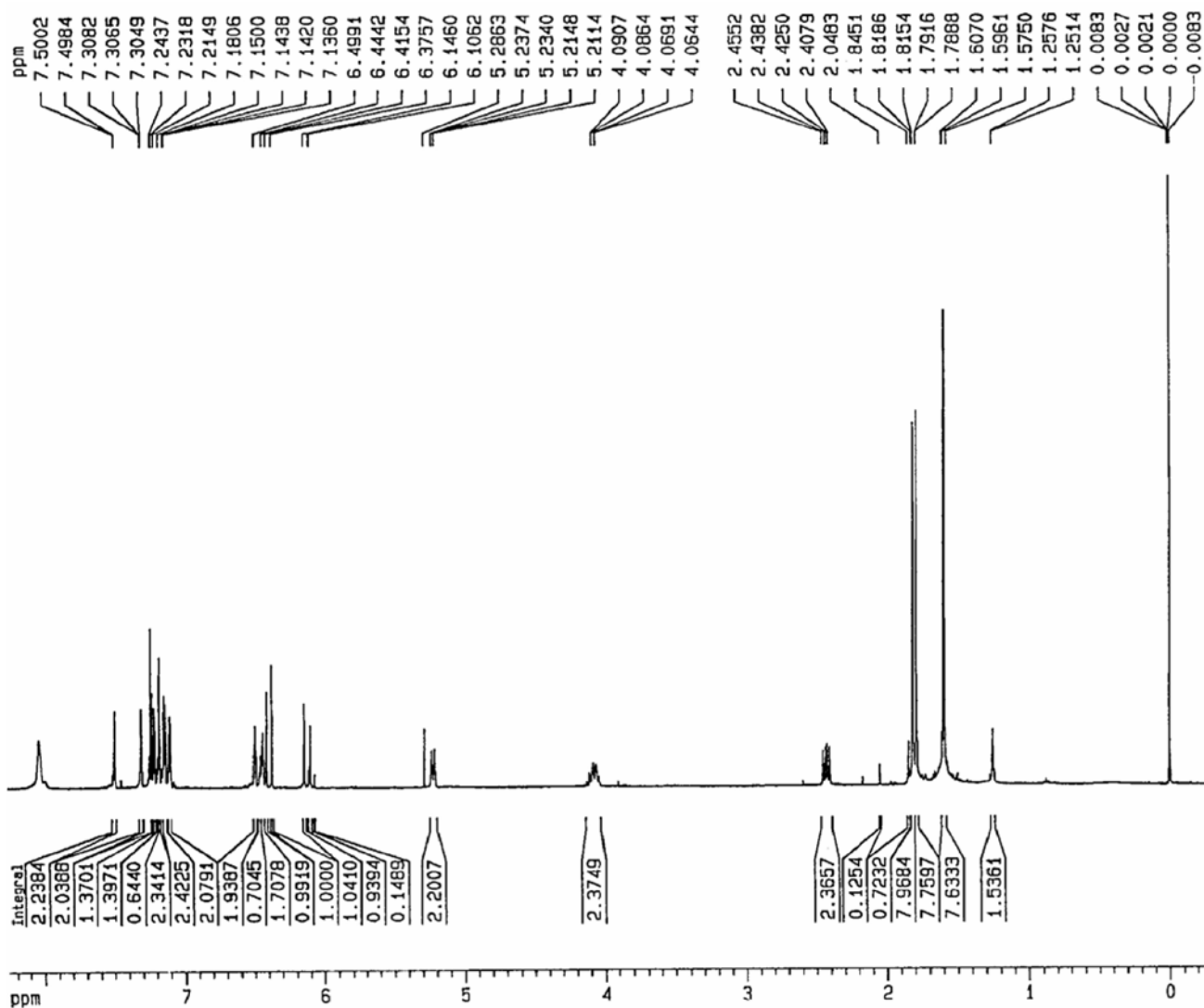


Figure S1. ¹H NMR spectrum of 1.

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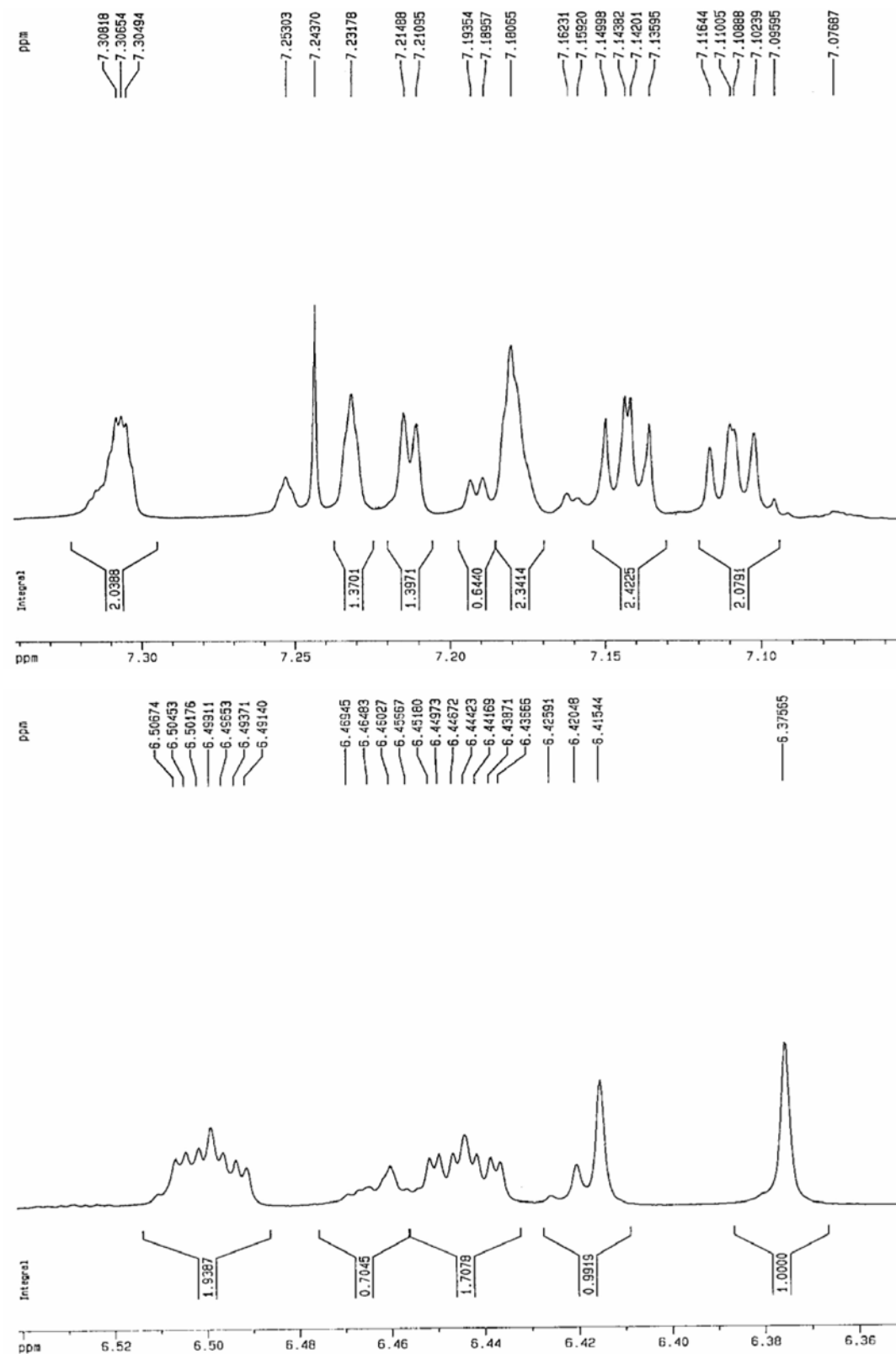
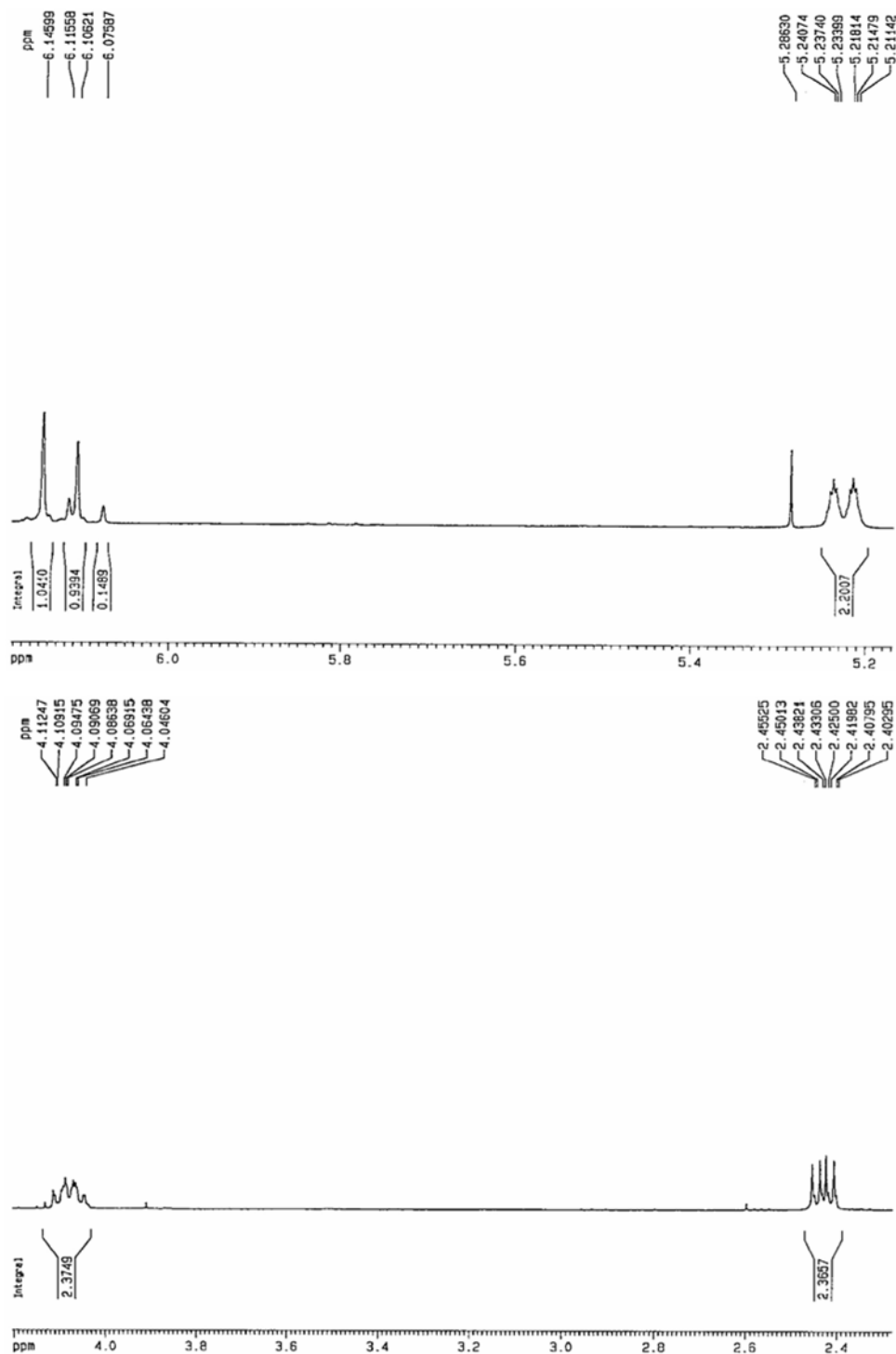


Figure S2. Expanded ¹H NMR spectrum of 1.

**Figure S3.** Expanded ¹H NMR spectrum of **1**.

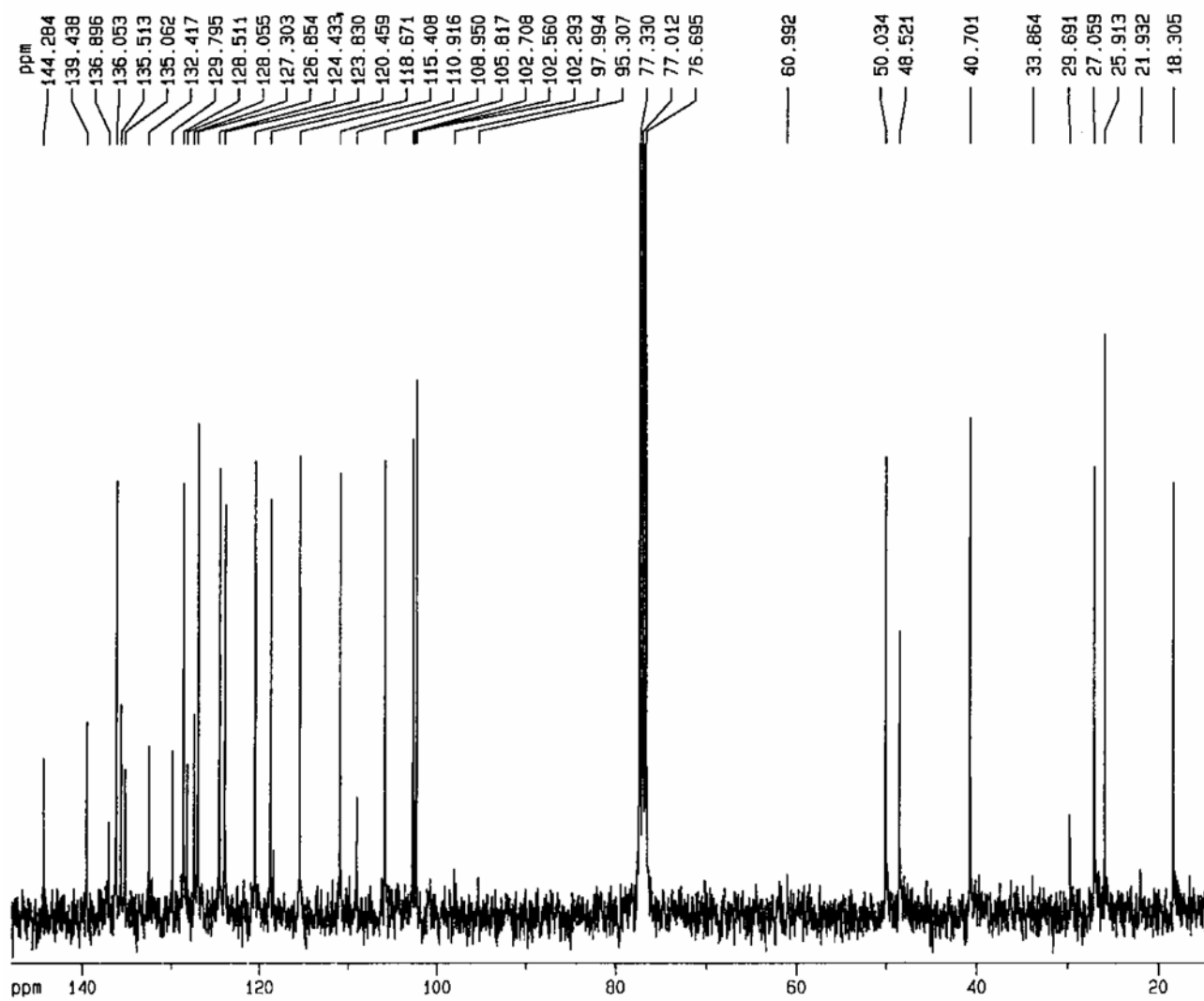
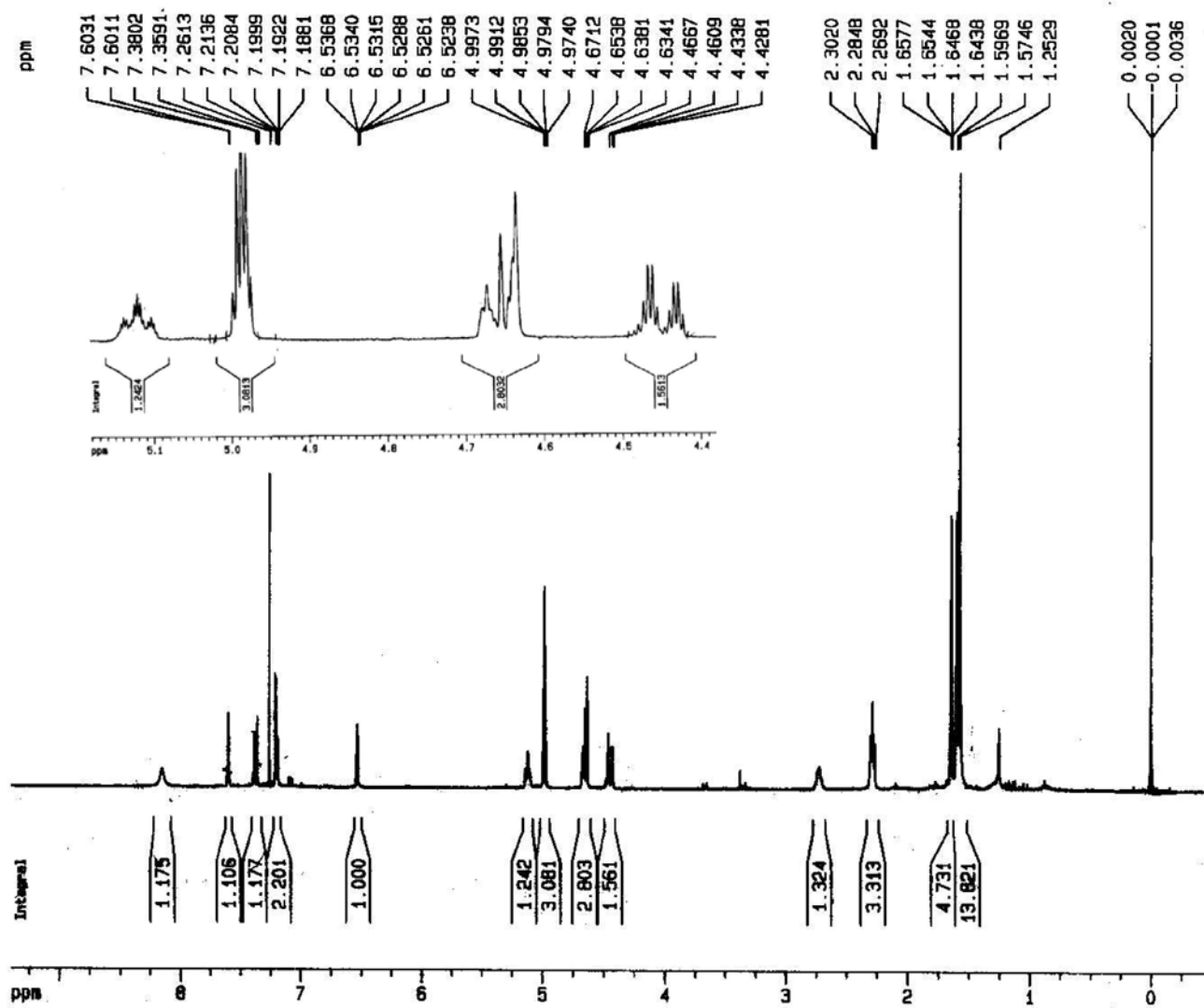
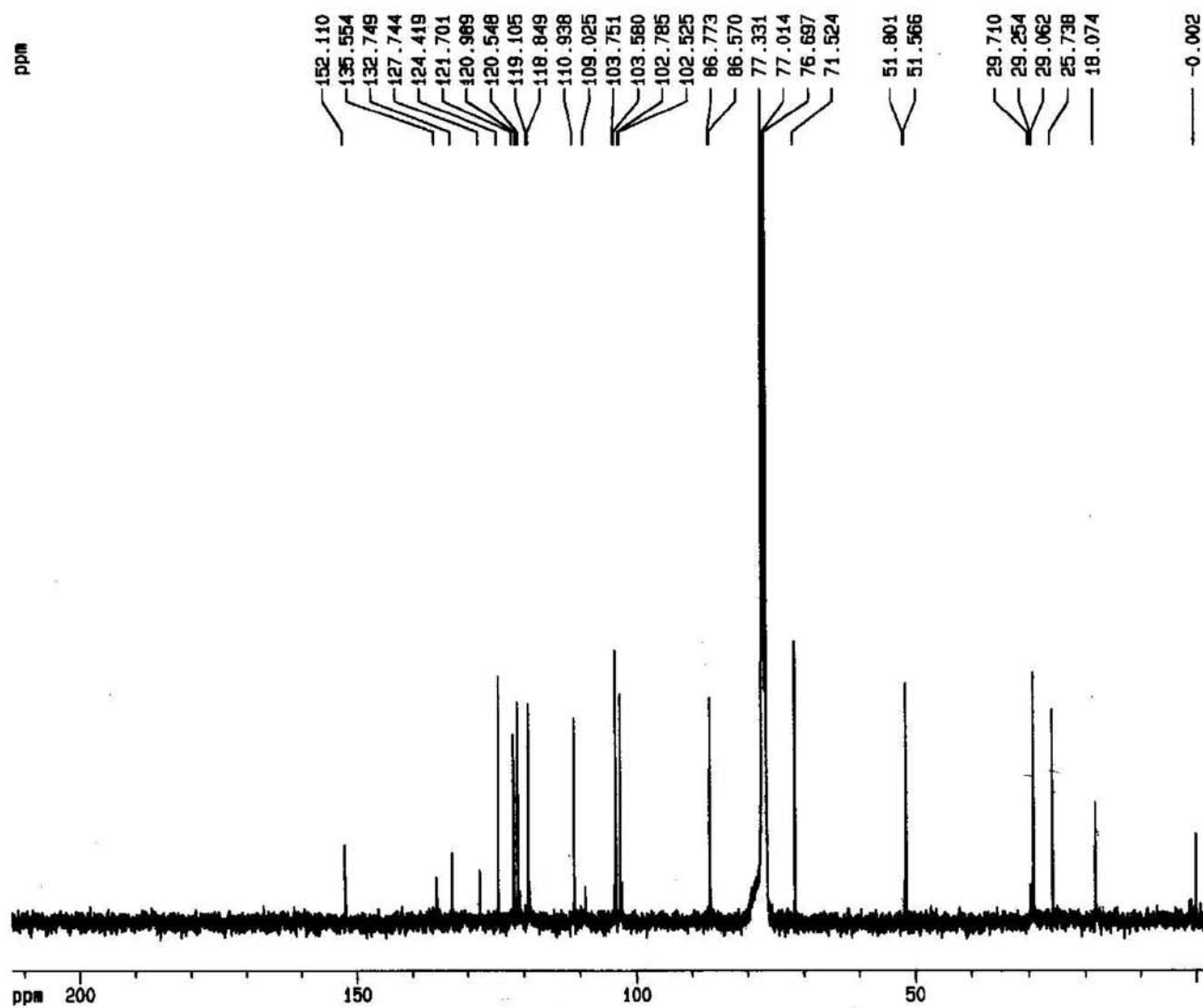
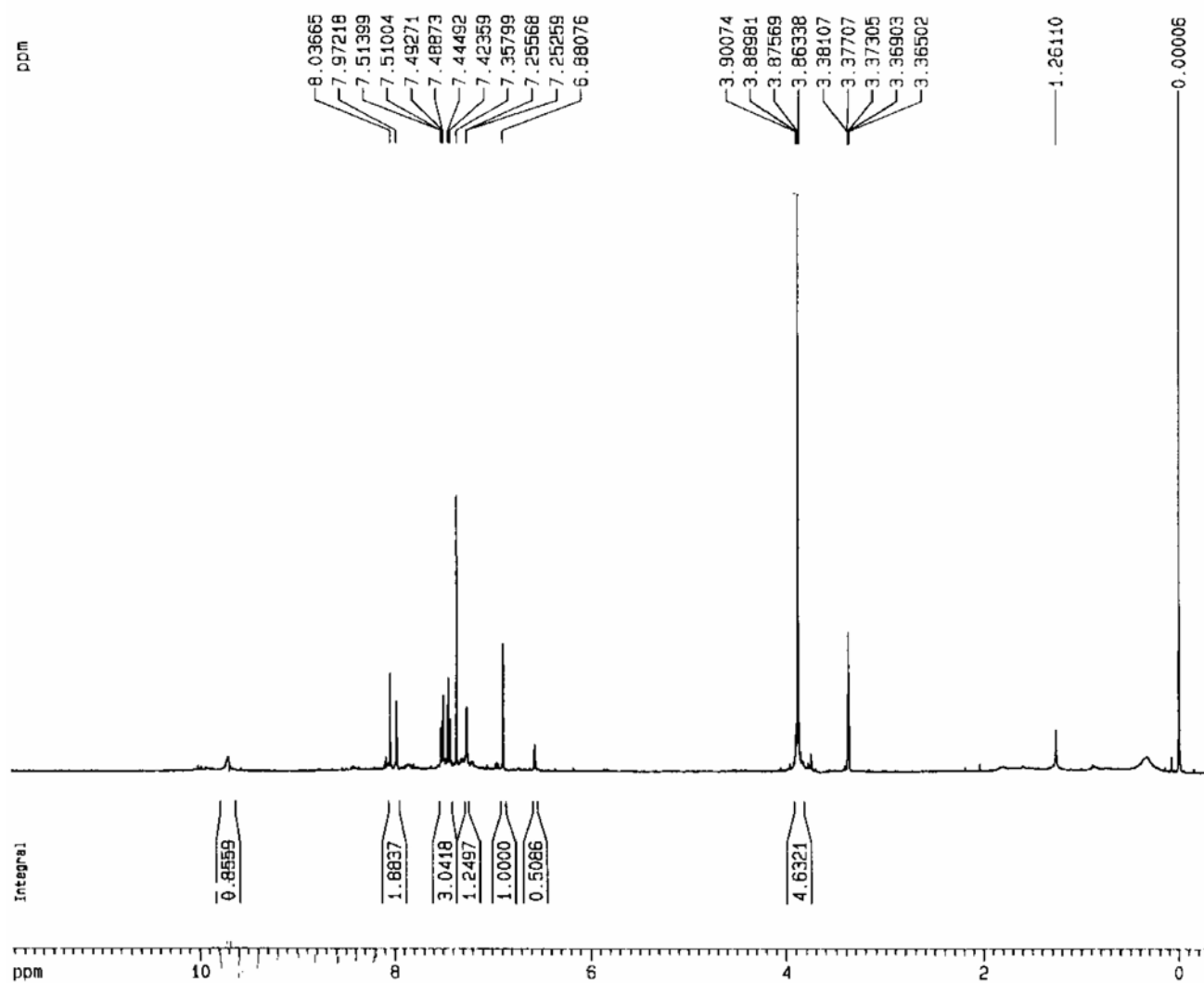


Figure S4. ^{13}C NMR spectrum of 1.

Figure S5. ¹H NMR spectrum of 2.

Figure S6. ^{13}C NMR spectrum of 2.

**Figure S7.** ^1H NMR spectrum of 3.

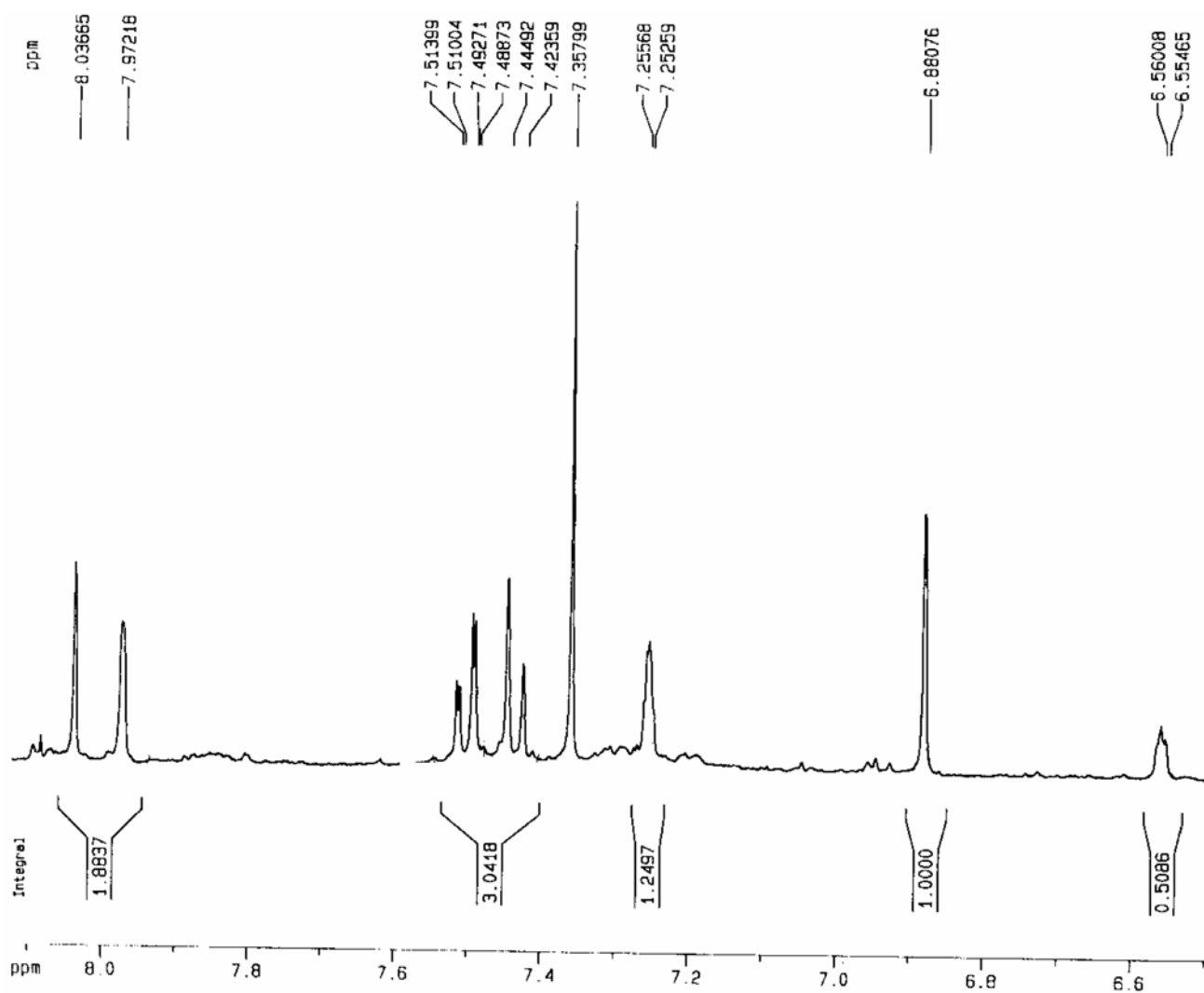
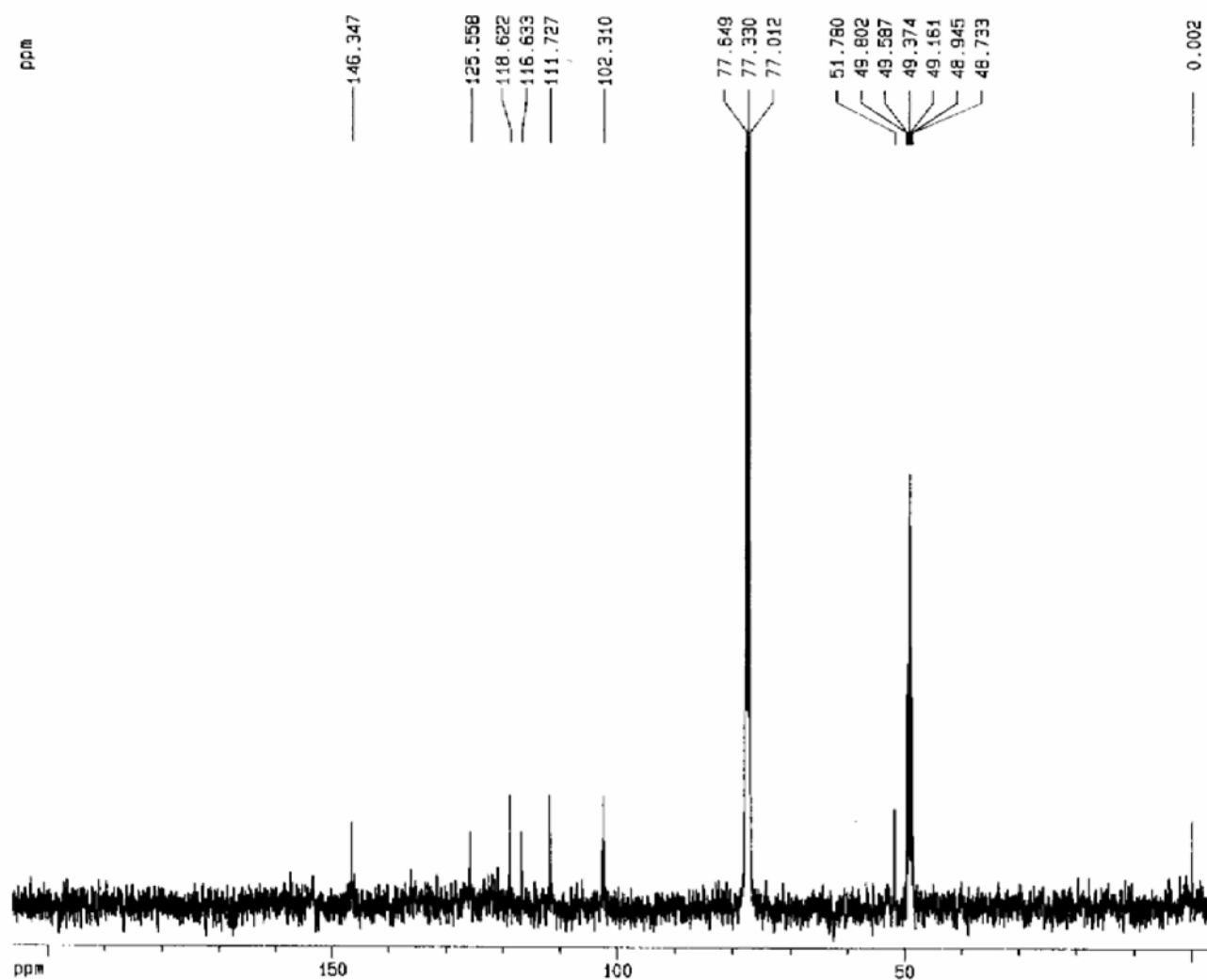


Figure S8. Expanded ¹H NMR spectrum of 3.

**Figure S9.** ¹³C NMR spectrum of 3.

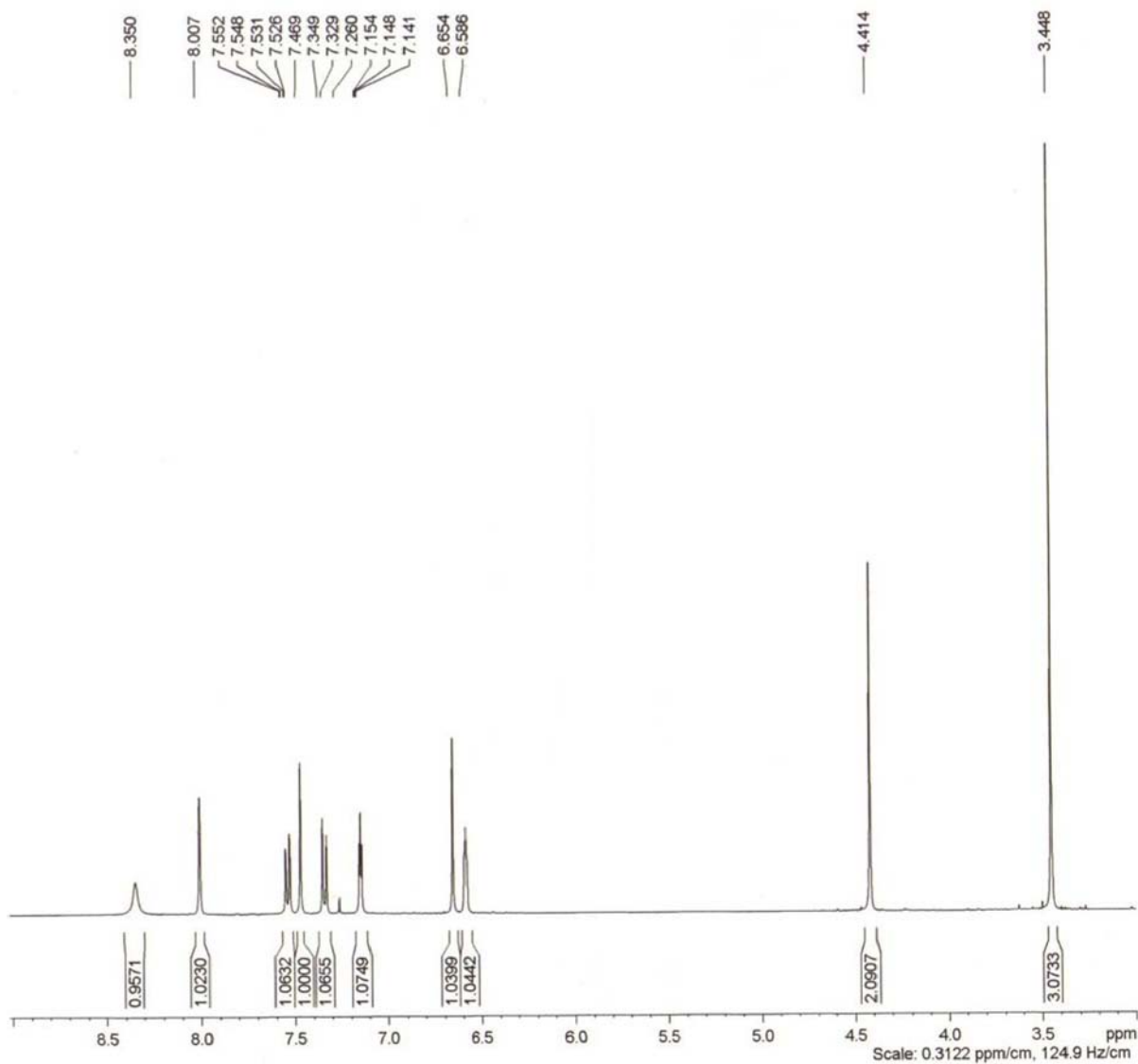


Figure S10. ¹H NMR spectrum of 4.

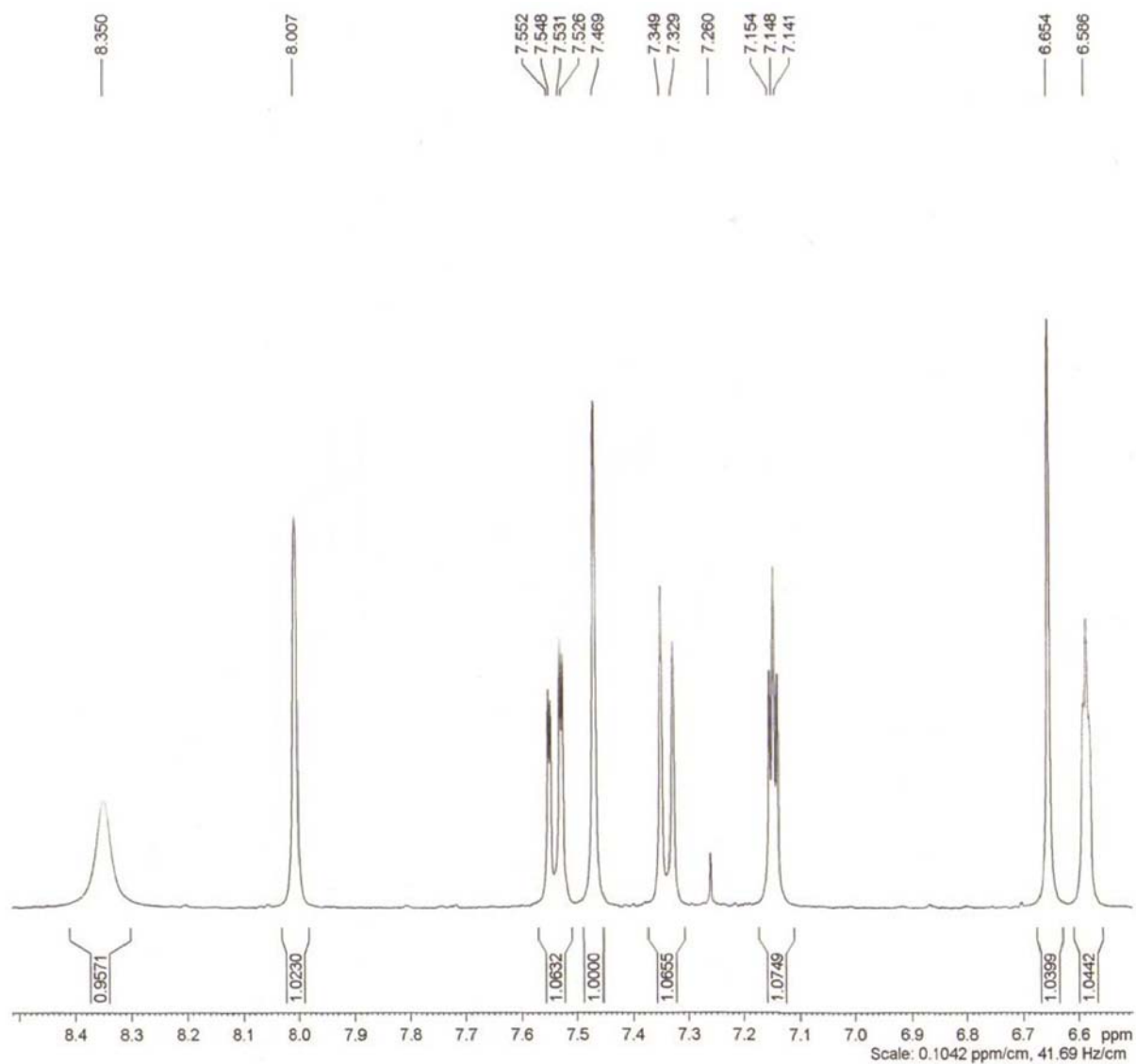


Figure S11. Expanded ^1H NMR spectrum of **4**.

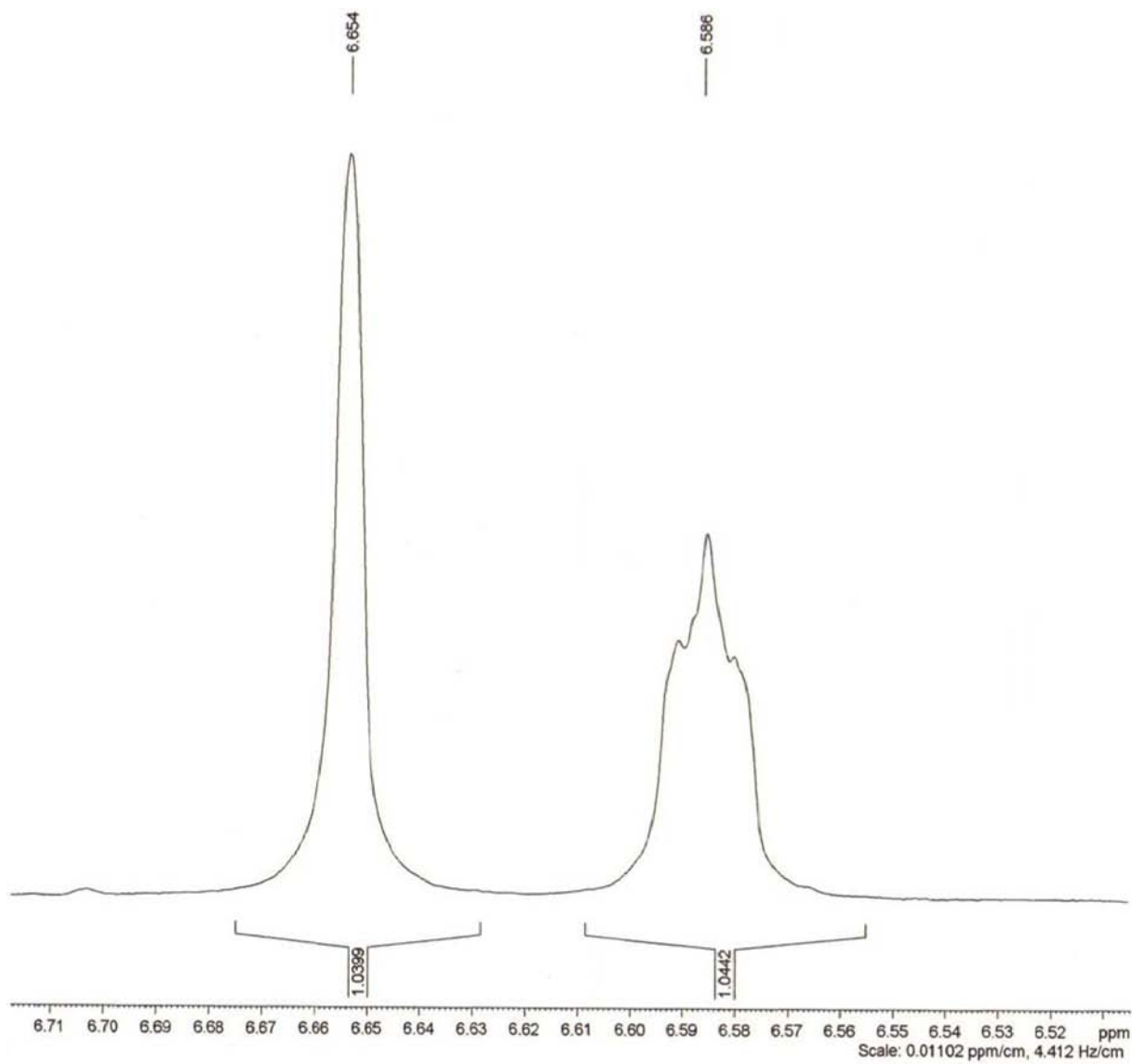
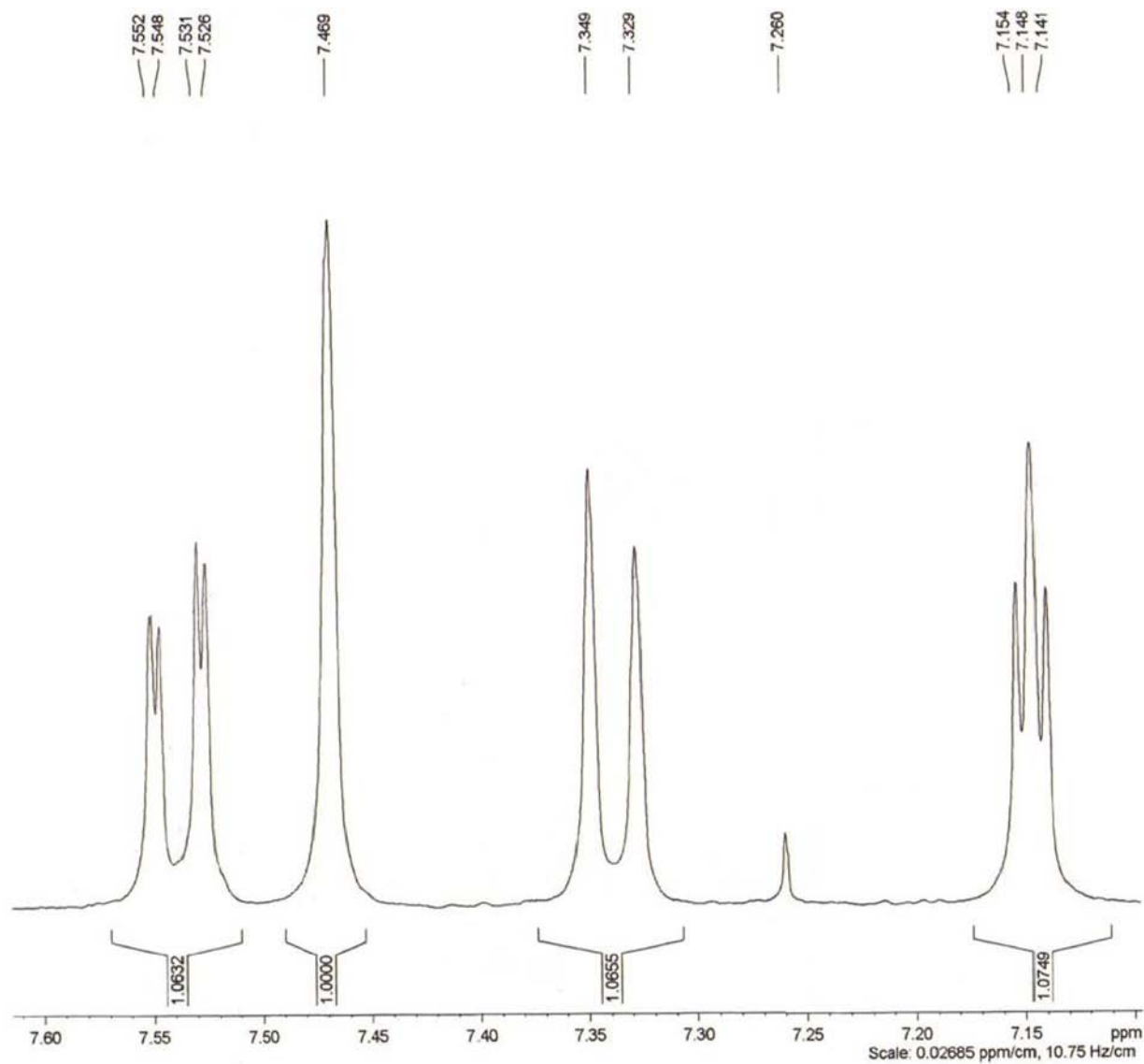


Figure S12. Expanded ¹H NMR spectrum of **4**.

**Figure S13.** Expanded ^1H NMR spectrum of **4**.

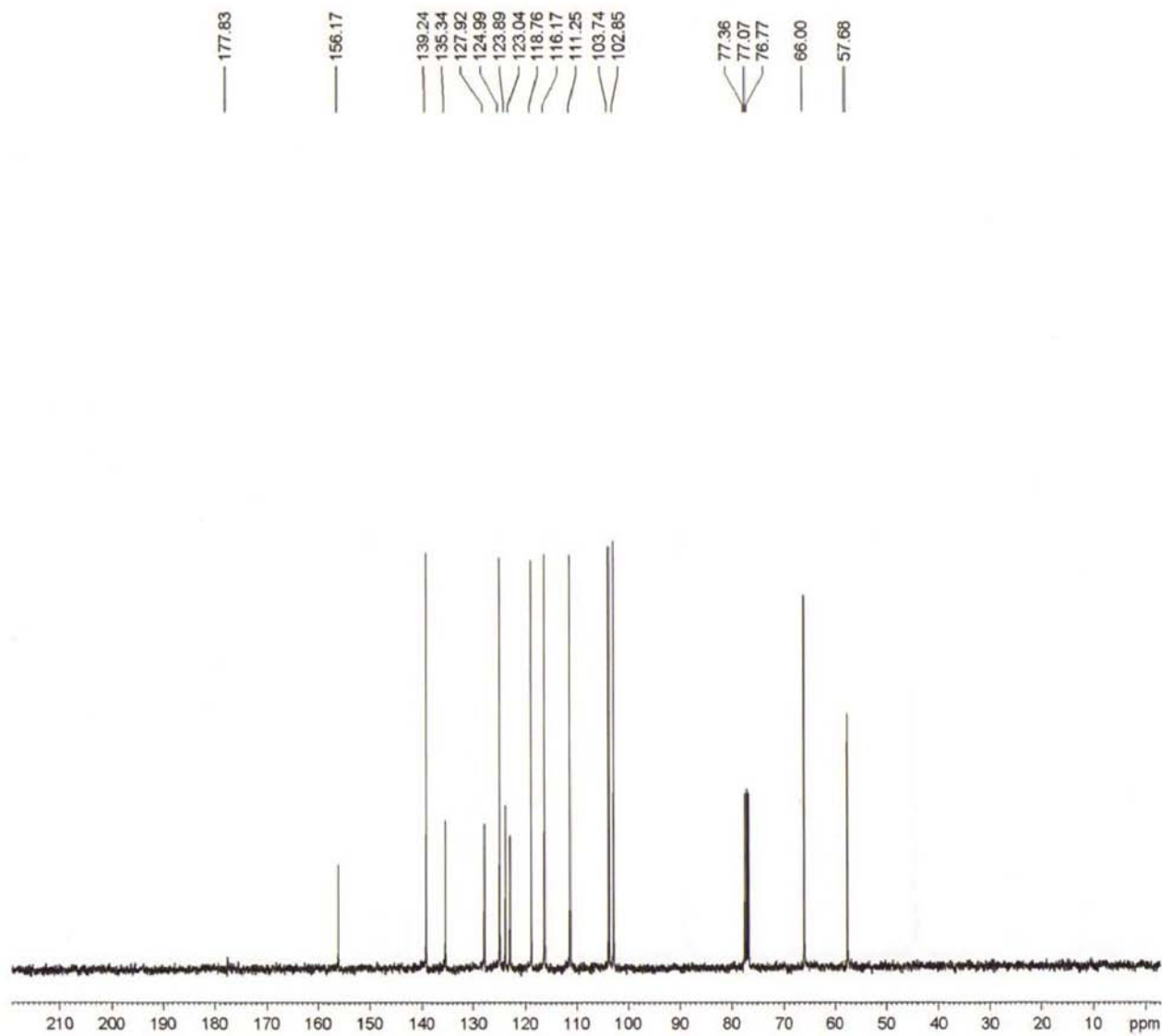


Figure S14. ^{13}C NMR spectrum of 4.

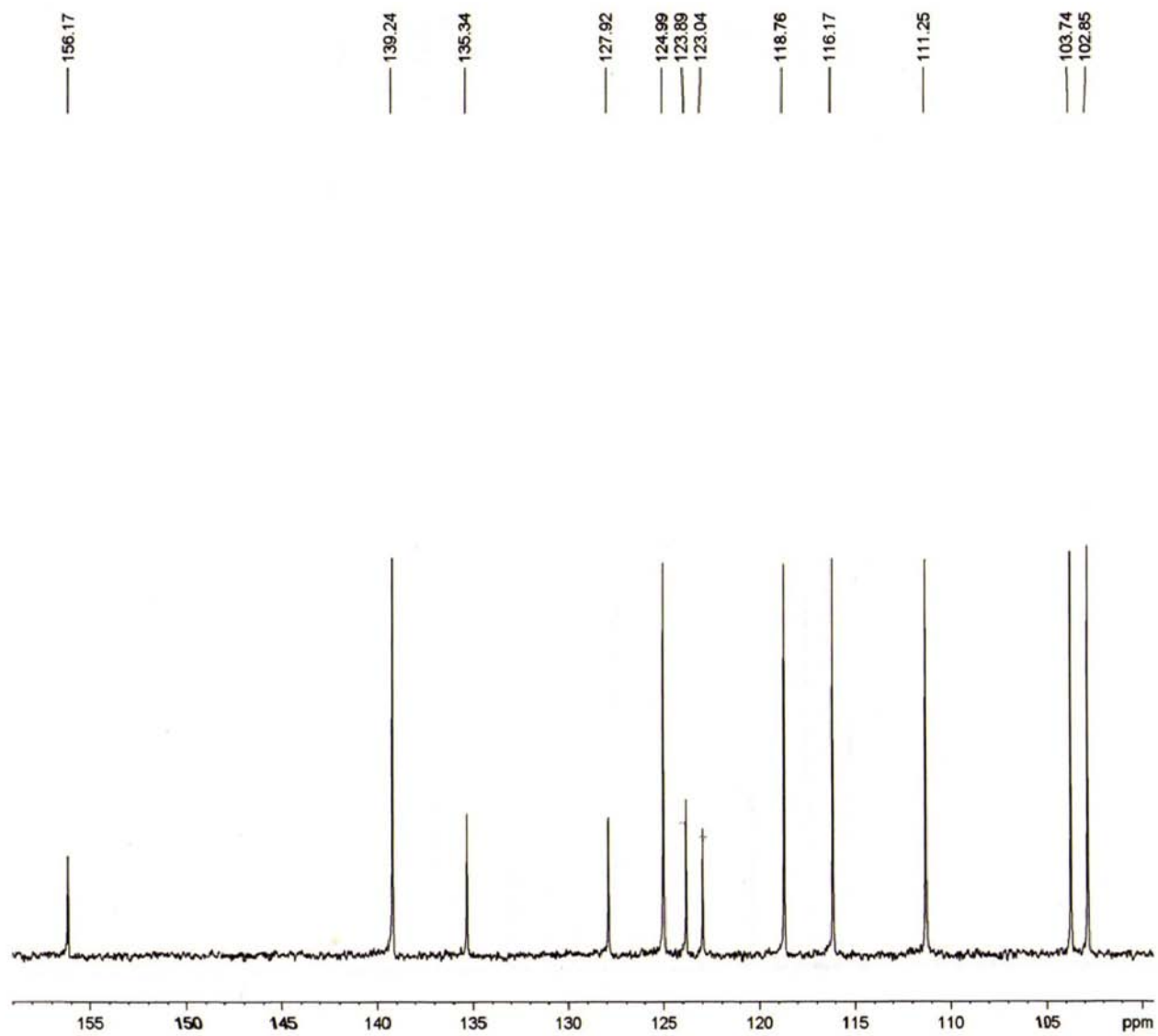


Figure S15. Expanded ^{13}C NMR spectrum of 4.

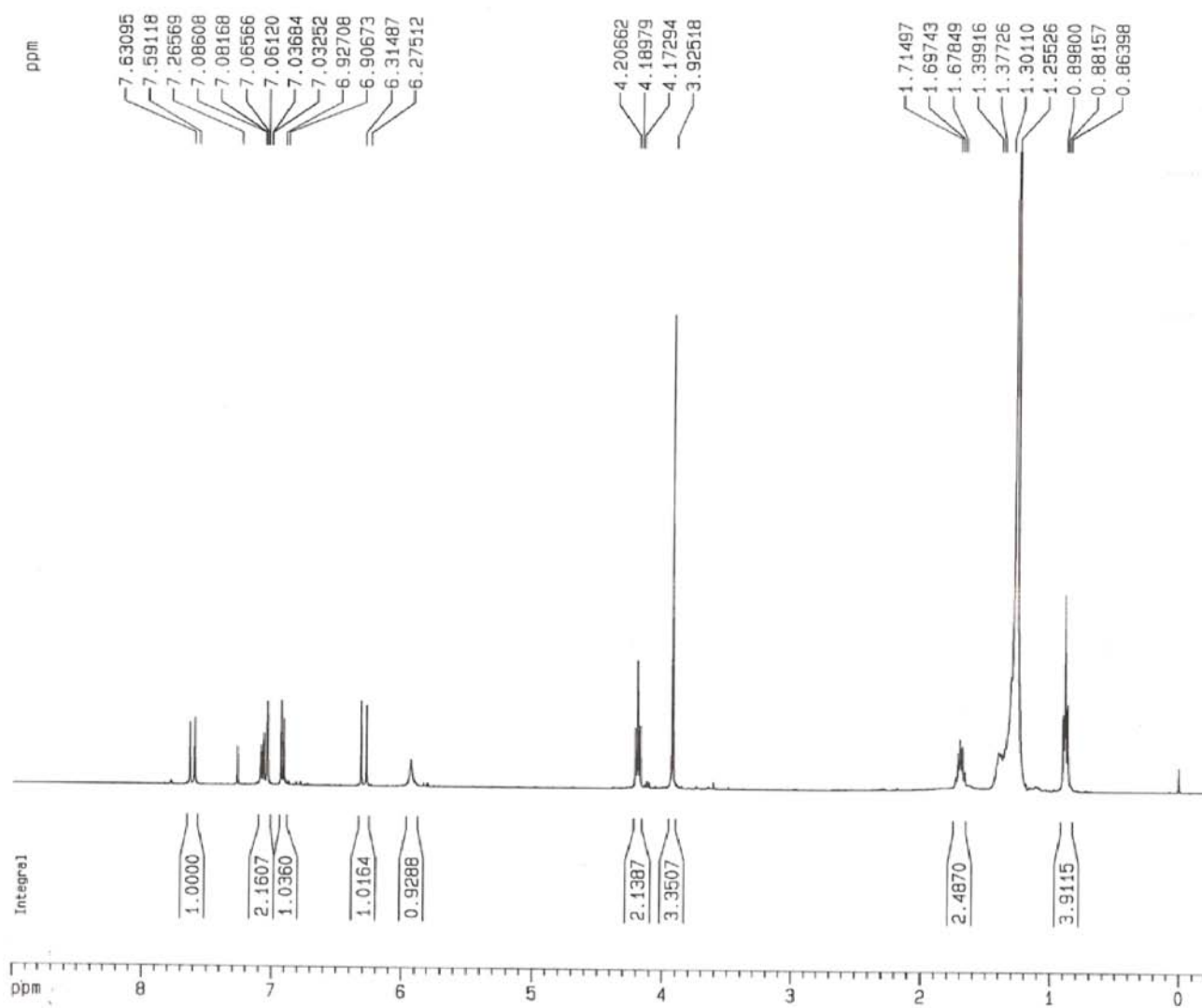
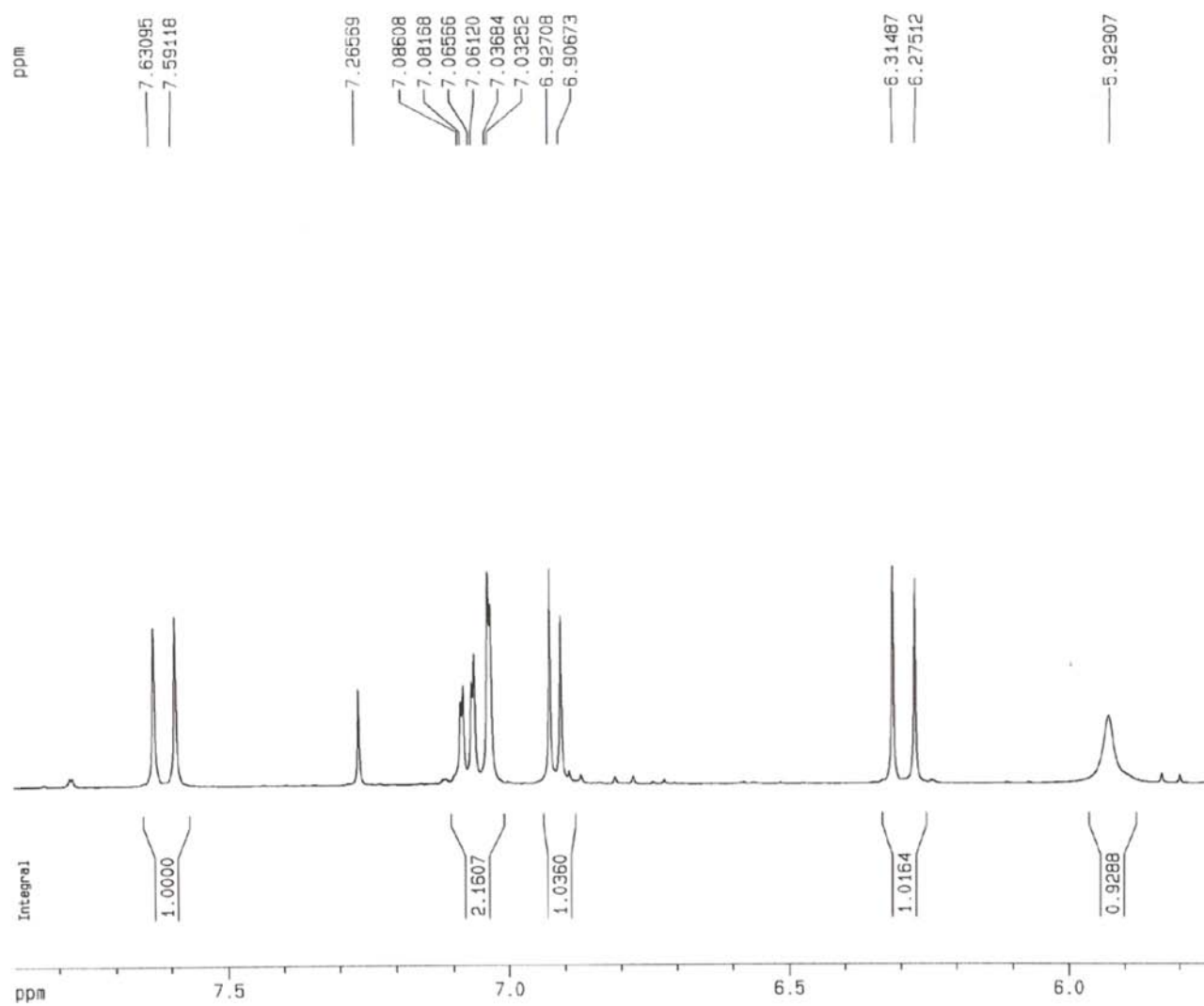
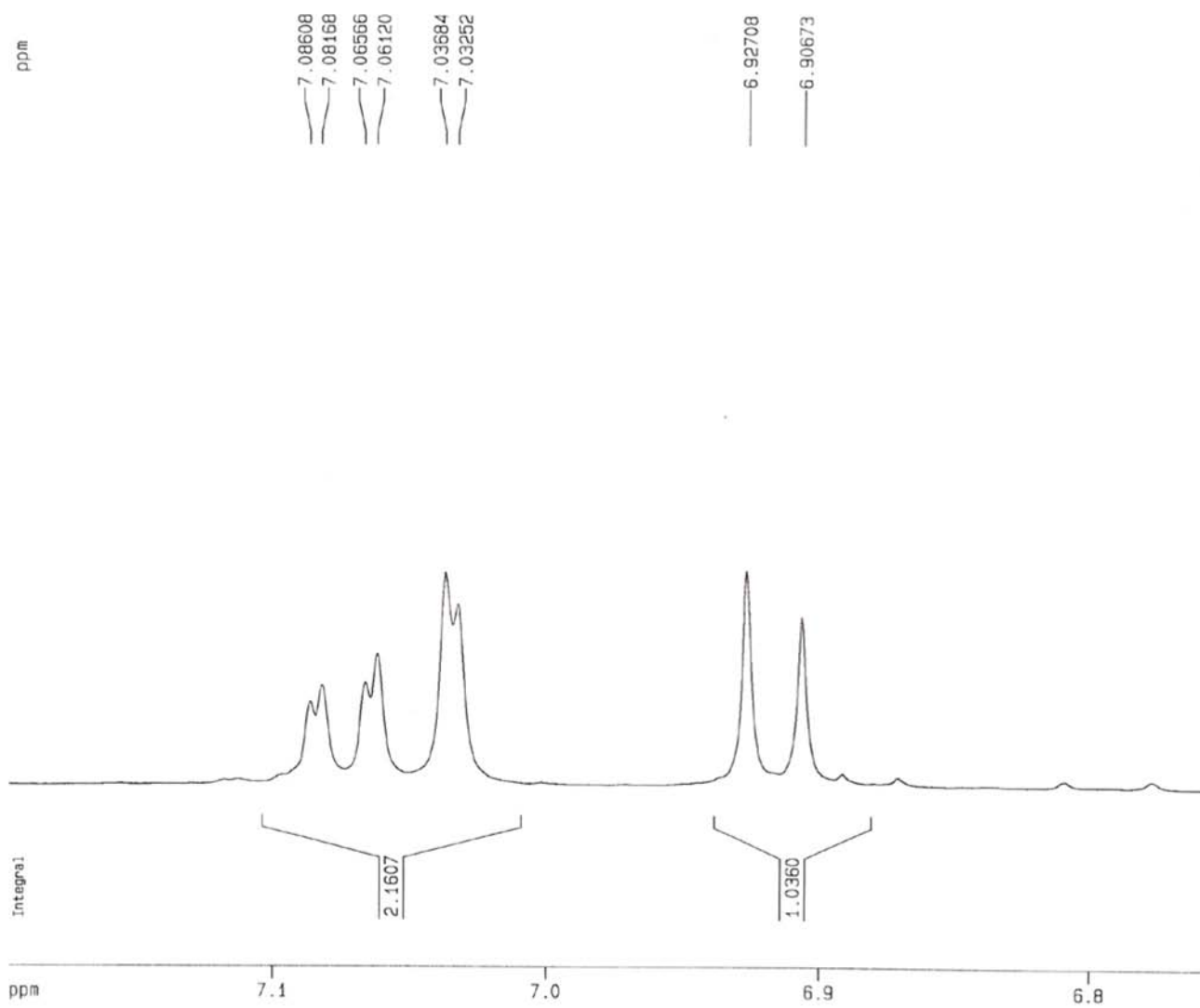


Figure S16. ¹H NMR spectrum of 5.

**Figure S17.** Expanded ^1H NMR spectrum of 5.

**Figure S18.** Expanded ^1H NMR spectrum of **5**.

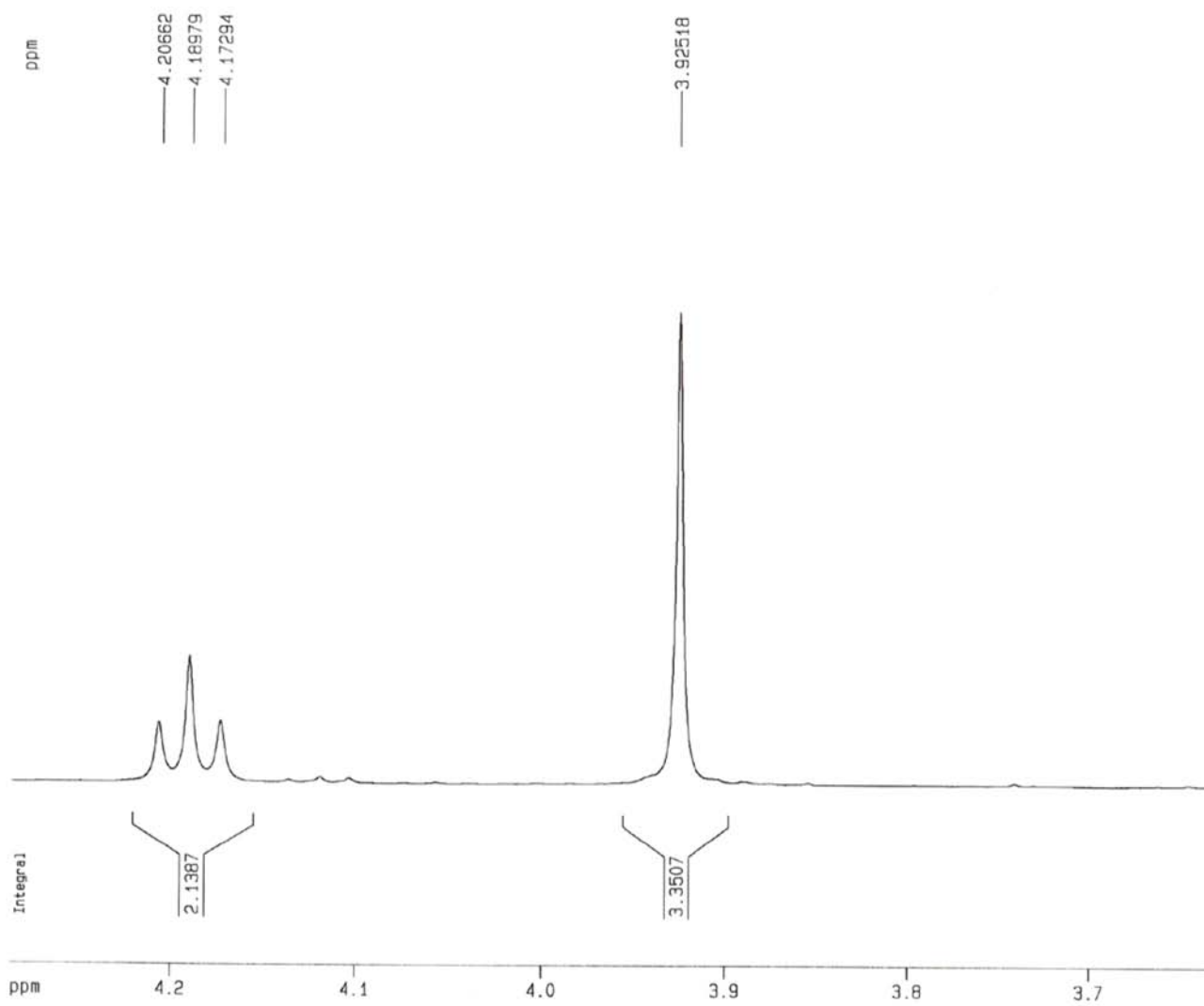


Figure S19. Expanded ^1H NMR spectrum of **5**.

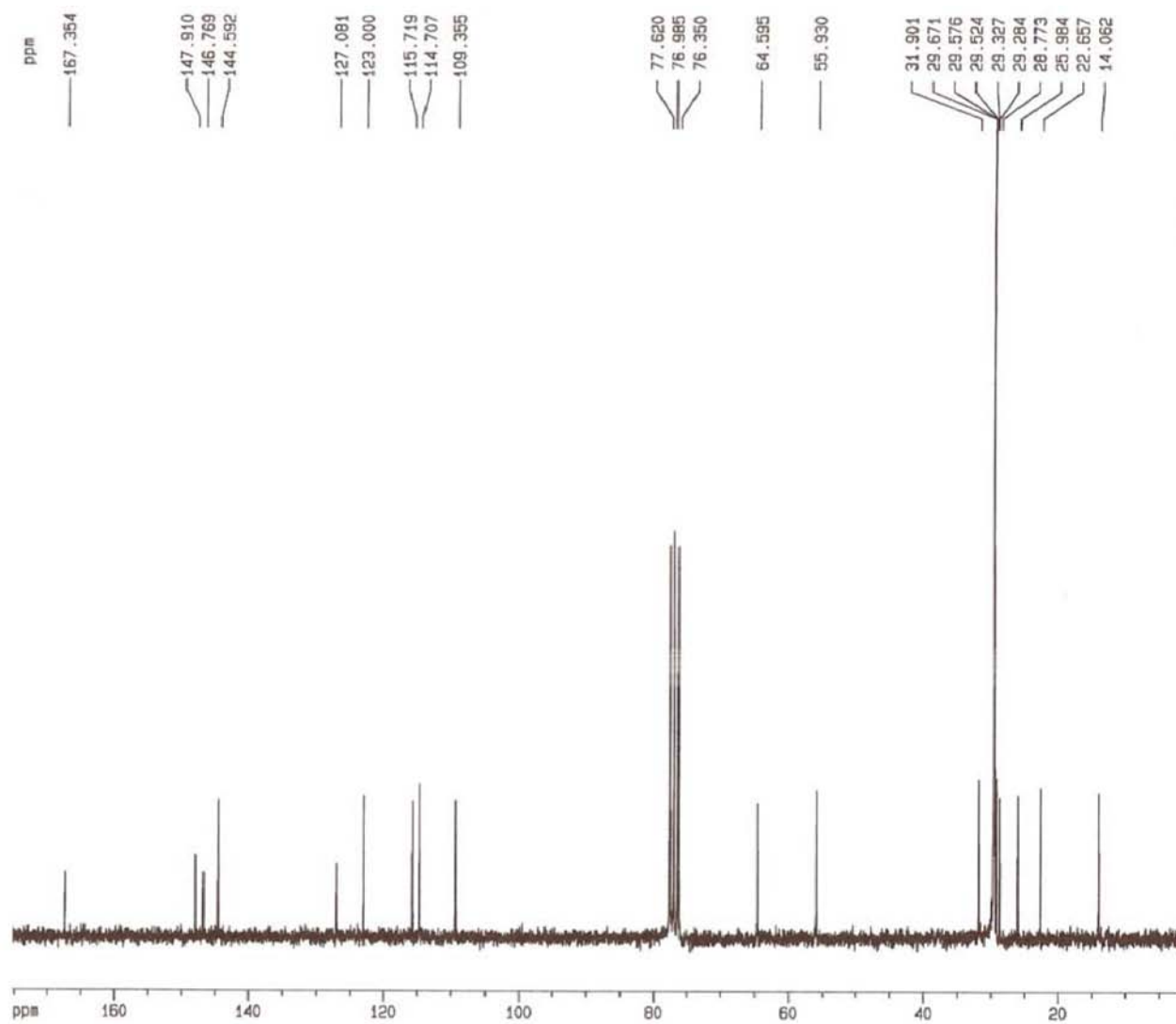
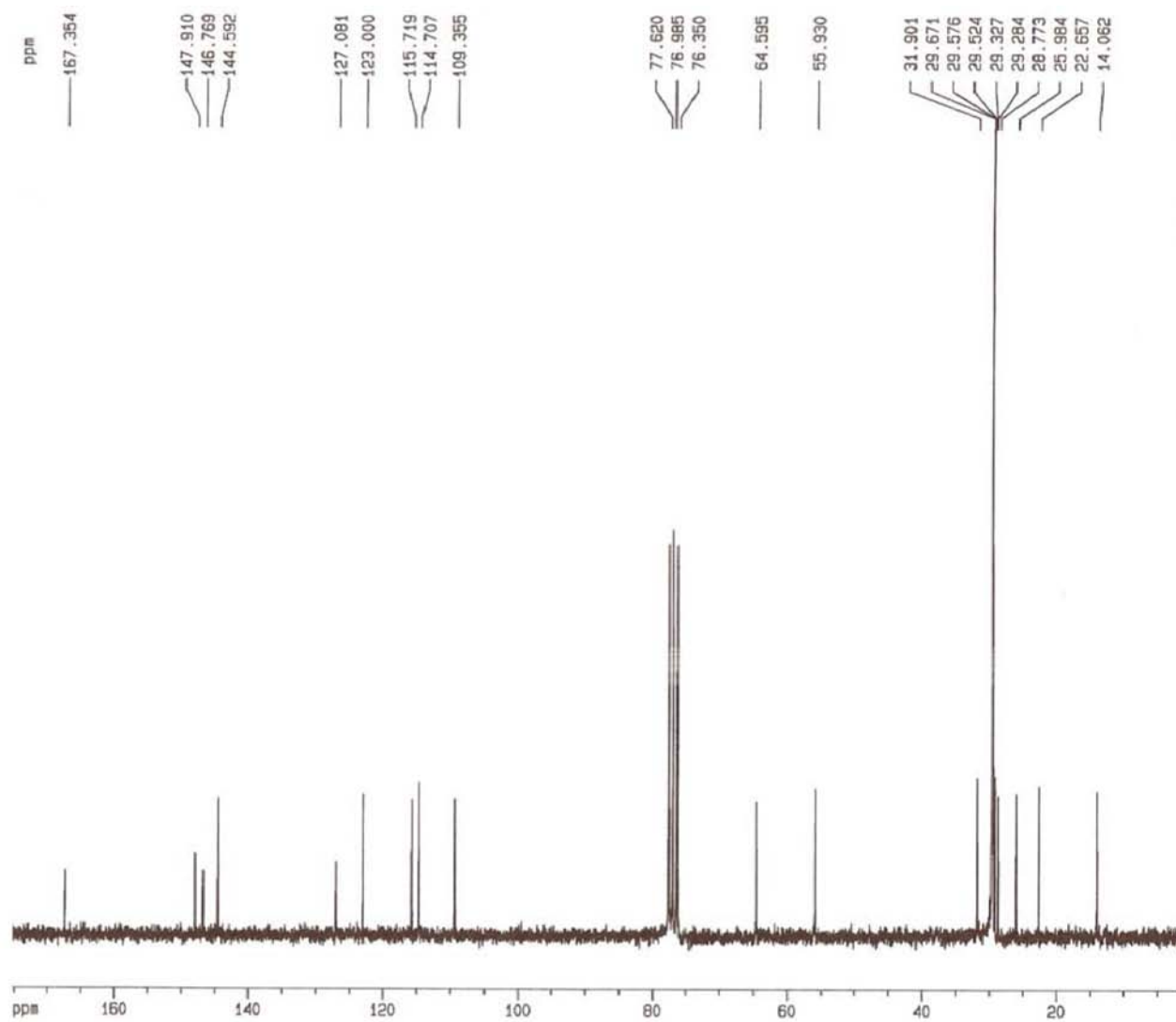


Figure S20. Expanded ^1H NMR spectrum of **5**.

**Figure S21.** ¹³C NMR spectrum of 5.

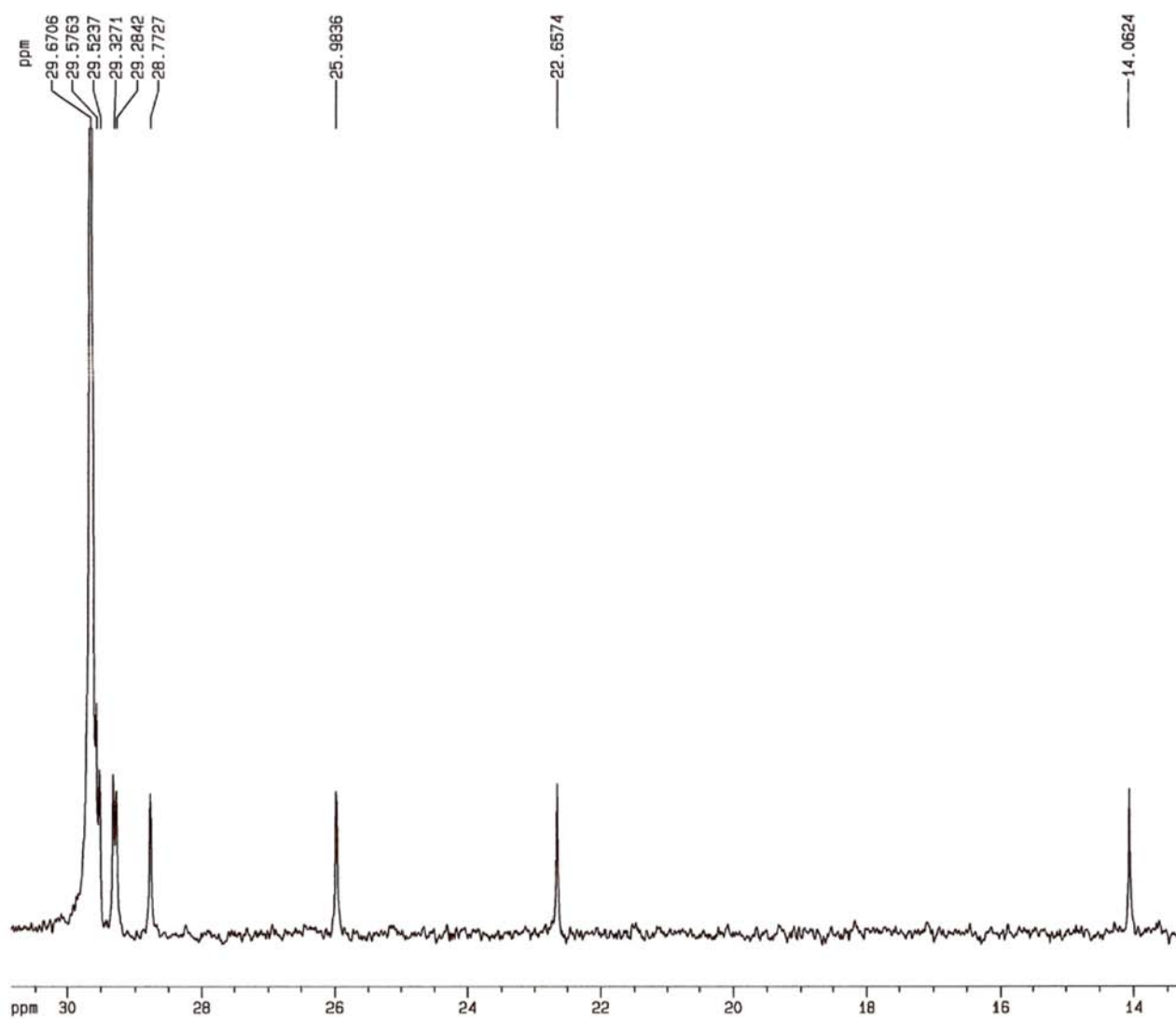


Figure S22. Expanded ^{13}C NMR spectrum of 5.