Enhanced Detection of Ponceau 4R Food Dye by Glassy Carbon Electrode Modified with Reduced Graphene Oxide

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This paper reports the development of a glassy carbon electrode modified with a reduced graphene oxide (r-GO/GCE) for the determination of the food dye named Ponceau 4R (PNC). The modified sensor presented excellent signal gain in relation to the GCE and GCE modified with graphene oxide besides exhibiting well defined oxidation peaks for PNC at potential of 0.61 V. Through the application of square wave voltammetry using the medium of 0.1 mol L⁻¹ Britton-Robinson (BR) buffer (pH 5.0), analytical curves were constructed under optimized conditions, where linear regions were found within the range of 0.200 to 20.0 μ mol L⁻¹. The limits of detection and quantification were 2.84 × 10⁻⁸ and 9.46 × 10⁻⁸ mol L⁻¹, respectively. The sensor was successfully applied towards the determination of PNC in instant juice sample. A comparison made between the result obtained through the application of this sensor and via the high-performance liquid chromatography (HPLC) technique showed no significant difference between the two methods.

Keywords: voltammetric sensor, dye, voltammetric determination, electrochemical sensor, electroanalysis

Introduction

Food colorings are dye additives defined as any substance that confers, enhances or restores food color. They are widely used in the food industry for the enhancement of the aesthetic appeal of processed food. Food dyes are controlled by restrictive legislation worldwide.¹⁻³ In Brazil, the current legislation on food dyes is stipulated under Resolutions 382 to 388 of ANVISA (National Agency of Sanitary Surveillance).⁴ The legislation limits the use of only eleven synthetic dyes in foods and beverages, with acceptable daily intake in the range of 0.10 to 15 mg kg⁻¹. Ponceau 4R (PNC, E124) is an intense red synthetic food colorant found in sweets, powder juices, milk derivatives, among other products. Studies have shown that this food additive is associated with cases of allergies and increasing rates of hyperactivity in children.^{5,6} As a result, its daily intake level allowed by international regulatory agencies is likely to be scaled down.7 Considering that great part of these dyes is released from industrial effluents and domestic sewage, these substances are regarded as contaminants of surface waters and drinking water processing plants.8 In this unpleasant scenario, analytical methods capable of detecting dyes are conceivably highly required.

The literature reports the use of spectrophotometry,^{9,10} high-performance liquid chromatography (HPLC),¹¹ spectrometry,¹² differential pulse polarography (DPP)¹³ and voltammetry^{14,15} for the determination of Ponceau 4R. Among the aforementioned mechanisms, electrochemical techniques are known to be endowed with crucial properties, such as high sensitivity, selectivity, low cost, simplicity, in addition to speed and capacity of miniaturization. These essential properties render them suitable for the development of new, efficient methods for dye determination.^{14,16} Apart from the interesting and inherent characteristics of voltammetric techniques, one can think of the possibility of modifying the surface of electrodes using specific materials, thus resulting in improvements in the electroactive area, sensitivity, selectivity and catalytic activity.17-21

Graphene is a widely explored material useful for developing electrochemical sensors by virtue of its special physicochemical properties including high surface area, excellent conductivity and high mechanical strength.²² It is formed by single sheets of carbon bonded together by sp² bonds. Graphene displays superior performance as electrocatalyst, in addition to better conductivity and

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large surface area compared to other carbon nanotubebased materials.^{17,23-26} Although graphene has demonstrated great applicability in electroanalysis, Jampasa *et al.*²⁷ demonstrated that screen printed carbon electrode modified with reduced graphene oxide can be used for determining Tartrazine (TZ) and Sunset Yellow (SY) dyes in beverages. They obtained linear relationships in the concentration range of 0.01 to 20 mmol L⁻¹ for SY and 0.02 to 20 mmol L⁻¹ for TZ. They reported to have found limits of detection (LOD) of 0.50 and 4.50 nmol L⁻¹ for SY and TZ, respectively.

The present work was conducted under two-fold objectives. First, it used the excellent electrical and electrocatalytic properties of reduced graphene oxide to modify glassy carbon electrode (GCE). Second, it sought to develop a voltammetric method for determining PNC. The method developed was then applied towards the direct determination of PNC in artificial beverage sample without any pretreatment. Our method demonstrated to be in considerable agreement with high-performance liquid chromatography analysis which was used for comparison.

Experimental

Reagents and equipment

All reagents used were of analytical grade. 4 mg mL⁻¹ of graphene oxide and Ponceau 4R dye ($\geq 99.0\%$) were supplied by Sigma-Aldrich. Sodium hydroxide was obtained from Synth. The commercial strawberry-flavored artificial powder juice was purchased from a nearby supermarket. The solutions were prepared using ultrapure water (with resistivity above $18 \text{ M}\Omega \text{ cm}$) provided by Milli-Q System (Millipore). The Britton-Robinson (BR) buffer solution was prepared by mixing 0.1 mol L⁻¹ of boric acid (Merck), acetic acid (Merck), o-phosphoric acid (Merck) and sodium hydroxide (Merck). Standard solution of 0.01 mol L⁻¹ of PNC dye was prepared in aqueous medium. Voltammetric measurements were recorded on an Autolab PGSTAT 302N galvanostat/potentiostat controlled by NOVA software with a conventional three electrode system. The pH measurements were conducted in a TECNOPON mPA 210 pH meter. The morphological and surface characterization of the electrode material was carried out by scanning electron microscopy (SEM) using FEG-SEM of the model JEOL 7500F. Chromatographic analysis was performed using a high-performance liquid chromatography-diode array detector (HPLC-DAD) (Shimadzu, model LC10ATVp) controlled by software CLASS VP.

Preparation of the electrode modified with r-GO

Initially, an aqueous suspension of graphene oxide (GO) of 1.0 mg mL⁻¹ was placed in an ultrasonic bath for 4 h aiming at promoting exfoliation. Afterwards, 0.2 mol L⁻¹ of sodium sulfate solution was added, yielding a suspension of 0.5 mg mL⁻¹ GO in 0.1 mol L⁻¹ of sodium sulfate. The GCE was properly cleaned by sanding various grain sizes prior to being modified. For the GO modification, a drop containing 10 μ L of the GO suspension was poured onto the GCE surface, and the electrode was in turn carefully placed in an oven at 50 °C for 15 min aiming at drying the modifier. The r-GO/GCE was prepared through the electrodeposition of the previously prepared graphene oxide suspension in a glassy carbon electrode applying a potential of -1.4 V for 500 s, forming a stable and reproducible film.^{28,29}

After each modification, the sensor was subjected to successive voltammetric cycles (15 cycles) in the range of 0.40 to 1.1 V in 0.1 mol L^{-1} BR buffer solution at a scan rate of 75 mV s⁻¹ for stabilization of the modified electrode surface.

Analytical procedure

Measurements were recorded in a 10 mL electrochemical cell using a conventional system with three electrodes: Ag/AgCl (3 mol L⁻¹) being the reference electrode, a platinum wire as auxiliary electrode, and GCE with and without modifications as working electrodes. All the voltammograms were obtained transferring 100 μ mol L⁻¹ of the selected dye in BR buffer under pH of 7.0. Cyclic voltammograms were obtained using a scan rate of 75 mV s⁻¹. Square wave voltammograms (SWV) were recorded using optimized conditions of accumulation time (30 s), frequency (25 Hz), step potential (10 mV) and pulse amplitude (70 mV).

The adherence of Ponceau 4R to the r-GO/GCE film was investigated by the immersion of the modified electrode in a solution (pH 5.0) containing Ponceau 4R dye, for 30 s under stirring. Subsequently, the electrode was washed with pure water and transferred to a voltammetric cell containing 10 mL of supporting electrolyte (pH = 5.0) and square wave voltammograms were recorded in the range of 0.45 to 0.90 V.

Application of the proposed method

Analysis of PNC on r-GO/GCE

0.1 g of strawberry flavor artificial powder juice sample was dissolved in 10 mL of ultra-pure water. 1 mL of the sample was then transferred to an electrochemical cell containing 9 mL of 0.10 mol L^{-1} BR buffer (pH 5.0). Following the immersion of the electrode in the aforementioned solution for 10 s under stirring, the electrode was subsequently transferred to a voltammetric cell containing 10 mL of 0.10 mol L^{-1} BR buffer (pH 5.0) and analyzed as described in the previous section.

Analysis of PNC by HPLC-DAD

For HPLC-DAD analysis, 0.5 g of powder juice sample was dissolved in 50 mL of ultra-pure water. Quantification of the PNC dye was performed by the standard addition method. The chromatographic measurements were recorded using C18 column (Phenomenex) with mobile phase consisting of 0.1 mol L⁻¹ of (A) ammonia acetate solution (pH 8.0) and (B) acetonitrile in gradient mode under the following conditions: 0-4 min 5% B; 4-10 min 95% B; 10-15 min 100% B.³⁰ The flow rate of 1 mL min⁻¹ and sample injection volume of 20 μ L were employed.

Results and Discussion

Characterization of r-GO/GCE

Figures 1a and 1b compare the scanning electron microscopy (SEM) of glassy carbon electrode modified with graphene oxide (GO/GCE) and reduced graphene oxide (r-GO/GCE). While agglomerated blocks are observed in Figure 1a, the surface of the r-GO/GCE (Figure 1b) illustrates a structure of wrinkled sheets owing to the modification that took place after the reduction of graphene oxide.^{27,31}

In order to investigate the mechanism by which this chemical structure changes the electroactive area of the electrode, a comparison was made in relation to the electrode active area by recording cyclic voltammograms for 1.00 mmol L⁻¹ potassium hexacyanoferrate(III) solution as model compound (diffusion coefficient = 7.6 μ cm² s⁻¹) in 0.1 mol L⁻¹ KCl solution for both GCE modified with GO and r-GO. The following Randles-Sevcik equation was applied:³²

$$i_{pa} = 2.69 \times 10^5 n^{\frac{3}{2}} A D_0^{\frac{1}{2}} C_0 v^{\frac{1}{2}}$$
 (1)

where i_{pa} is the anodic peak current (µA), n is the electron number, A is the electrode active area (cm²), D₀ is the diffusion coefficient (cm² s⁻¹), v^{1/2} is the scan rate square root (V s⁻¹) and C₀ being the analyte concentration (mol cm⁻³). The electroactive areas for GCE modified with GO and r-GO/GCE correspond to 1.96 × 10⁻³ and 7.61 × 10⁻³ cm², respectively. These results suggest that the simple electrochemical procedure used to produce



Figure 1. SEM of (a) GO/GCE and (b) r-GO/GCE for 50,000-fold amplification.

the r-GO electrode yielded an increase in surface area of approximately four times bigger compared to the GO/GCE electrode.

Electrochemical oxidation of PNC

Figure 2 shows cyclic voltammograms recorded for 100 μ mol L⁻¹ PNC dye in 0.1 mol L⁻¹ BR buffer (pH 7.0) on GCE (Figure 2a), GO/GCE (Figure 2b) and r-GO/GCE (Figure 2c). For the GCE (Figure 2a), the voltammograms present an oxidation peak at 0.64 V and a small peak at 0.58 V in the reverse scan, where the ratio of $i_{pc}/i_{pa} = 0.36$ $(i_{nc}$ is the cathodic peak current). One will note that the peak displays characteristics of reversible systems with subsequent chemical reactions since the reverse peak undergoes a slight increase at higher scan rates.³² When the GO/GCE (Figure 2b) is used, electrochemical behavior similar to that of GCE is observed albeit with a decrease in magnitude of the peak. This result can be attributed to the low conductivity of GO, a material that has many sp³ carbons due to the presence of oxide groups (for example, -COOH and -OH). Interestingly, an excellent performance is observed on the r-GO/GCE. The oxidation peak is shifted to less negative potential (0.61 V), and a 20-fold increase is noted in the anodic peak current. In addition, the electrodic process is found to be closer to the reversible process,

where $i_{pc}/i_{pa} = 0.98$ and the $E_{pc} - E_{pa} = 30.8$ mV (E_{pc} and E_{pa} are the cathodic and anodic peak voltages, respectively).³² This behavior suggests that fast electron transfer can be occurring as a result of the reconstitution of a large part of the sp² bonds between carbons, increasing the conductivity of the material while facilitating the electron transfer.³³ This behavior indicates that probably the hydroxyl group in the dye molecule undergoes oxidation involving two electrons in a reversible system.³²



Figure 2. Cyclic voltammograms for 100 μ mol L⁻¹ of PNC in 0.10 mol L⁻¹ buffer BR, pH 7.0 on (a) GCE; (b) GO/GCE and (c) r-GO/GCE. $\nu = 100 \text{ mV s}^{-1}$.

The effect of scan rate on the oxidation was evaluated by recording cyclic voltammograms for 100 µmol L⁻¹ of PNC in 0.1 mol L⁻¹ BR buffer (pH 5.0) from 5 to 500 mV s⁻¹. The anodic current is found to increase linearly, with the equation $i_{pa} = 7.11 \times 10^{-4} \upsilon + 3.37 \times 10^{-6}$ (coefficient of determination (R²) = 0.98, υ is the scan rate), indicating that the charge transfer is controlled by adsorption process.³² Considering that the process is adsorptive, the surface excess (Γ) was calculated aiming at verifying the electroactive species concentration on the modified electrode surface using the following equation:³²

$$\dot{i}_{pa} = \frac{n^2 F^2 A \Gamma \upsilon}{4 R T}$$
(2)

where i_{pa} is the anodic peak current, n is the electrons number, F is the Faraday constant, v is the scan rate, A is the electrode active area, Γ is the electroactive species surface concentration, R is the gas constant (8.314 J mol⁻¹ K⁻¹) and T is the temperature. By replacing the values in equation 2, the concentration of PNC on the surface of r-GO/GCE is found to be 24.9 nmol cm².

Optimization of parameters

In order to find low levels of PNC dye detection, parameters such as accumulation time, pH and scanning technique were optimized. Considering that the analyte is strongly adsorbed on the electrode surface, the effect of the accumulation time was studied in the range of 5 to 50 s aiming at verifying the analyte response. The measurements were recorded for 30 µmol L⁻¹ of PNC in 0.1 mol L⁻¹ BR buffer (pH 7.0) by cyclic voltammetry ($\upsilon = 100 \text{ mV s}^{-1}$). An increase in the i_{pa} of the dye was observed for up to 30 s of preconcentration (t_{ac} , accumulation time), while a plateau was noted at longer periods beyond 30 s. In view of that, prior to each measurement, the solution was stirred for 30 s.

The pH effect on both the oxidation peak and the i_{pa} of the dye was verified using cyclic voltammetry by varying the pH of the 0.1 mol L⁻¹ BR buffer between 3 to 10. The best i_{pa} responses were verified for pH 5 and 7. However, a better peak resolution and lower deviation between measurements was observed for pH 5, hence, it was chosen for the performance of other experiments. The relationship between E_{pa} *vs.* pH can be described by the equation $E_{pa} = -0.0310$ pH + 0.8368 (R² = 0.990), which indicates that the ratio of H⁺/e⁻ participation in the oxidation reaction of the PNC dye is 1/2, respectively. A proposed PNC oxidation mechanism is shown in Figure 3, which is attributed to the oxidation of phenolic hydroxyl group.³⁴

To obtain better results for PNC, the analytical techniques including linear sweep (LSV), differential pulse (DPV) and square wave voltammetry (SWV) were studied for 1 μ mol L⁻¹ of dye in 0.1 mol L⁻¹ BR buffer (pH 5.0) and t_{ac} = 30s. The best responses were found via SWV, which represented an increase of 369 and 105% in i_{pa} compared to the LSV and DPV techniques, respectively. Hence, SWV parameters such as frequency (10-100 Hz), step potential (2-10 mV) and pulse amplitude (20-80 mV)



Figure 3. Proposed mechanism for the oxidation reaction of PNC against r-GO/GCE.

were optimized, where the best conditions were found to be at 25 Hz, 10 mV and 70 mV, respectively.

Analytical performance

Under optimized conditions of t_{ac} , pH and SWV, voltammograms were recorded for the PNC dye in 0.1 mol L⁻¹ BR buffer (pH 5.0) (Figure 4). By applying a t_{ac} of 30 s, a linear relationship was found in the range of 0.200 to 20.0 µmol L⁻¹ with the following equation: $i_{pa} = 3.65 \times [PNC] + 2.12 \times 10^{-6}$ (R² = 0.998) (inset in Figure 4). The limits of detection (LOD) and quantification (LOQ) were calculated using the following equations: LOD = 3std/m and LOQ = 10std/m, where m and std stand for angular coefficient and standard deviation of the analytical curve, respectively. The values of LOD and LOQ were 2.84 × 10⁻⁸ and 9.46 × 10⁻⁸ mol L⁻¹, respectively.

In Table 1, figures of merit of the proposed method are compared with other methods found in the literature for the determination of PNC. Although the studies reported by Zhang *et al.*,³⁴ Zhang *et al.*,³⁵ Yang *et al.*,³⁶ and Wang and Zhao³⁷ presented lower LOD, these methods have a much lower analytical frequency than the proposed method. In addition, the LOD and LOQ of the proposed method are below the values stipulated by the regulatory agencies (0.10 to 15 mg kg⁻¹).

Selectivity

For the assessment of the degree of interference of other food colorants in the determination of Ponceau 4R, voltammograms were recorded for $100 \,\mu\text{mol}\,\text{L}^{-1}$ of PNC in the presence of Allura Red 40 (V40) and Brilliant Blue FCF (AB) at the same concentration. As depicted in Figure 5,



Figure 4. (a) SWV in 0.1 mol L⁻¹ BR buffer (pH 5.0) at PNC concentrations of (b) 0.200; (c) 0.400; (d) 0.700; (e) 1.98; (f) 4.93; (g) 9.82; (h) 20.0 μ mol L⁻¹. Inserted: relationship between i_{pa} vs. [PNC] in the intervals between 0.200 to 20.0 μ mol L⁻¹.

individual voltammograms for dyes V40 (curve b) and AB (curve c) show anodic peaks at the potentials of 0.75 and 0.91 V, respectively, relative to the dyes oxidation. As can be observed in curve d, the voltammetric profile for the PNC did not show any great difference when analyzed individually (curve a) and when in the presence of the dyes V40 and AB. Clearly, this shows that there is neither overlapping of peaks nor the exertion of influence on the i_{ap} even when PNC is in the presence of the dyes under analysis.

Application of the method

In order to evaluate the applicability of the r-GO/GCE sensor, tap water sample was analyzed. To this end, the tap water sample was fortified with 5.00 μ mol L⁻¹ of PNC and analyzed (n = 3) using the standard addition method. The value found for PNC recovery was 5.20 ± 0.150 μ mol L⁻¹.

Table 1. Comparison of the r-GO/GCE method with other methods reported in the literature regarding the determination of Ponceau

Electrode	Technique	Concentration range / (mol L ⁻¹)	$LOD / (mol L^{-1})$	Reference
MWNT/GCE	DPV	$4.13 \times 10^{-8} - 2.48 \times 10^{-6}$	2.48×10^{-8}	34
IL-EGPE	SWSV	1.00×10^{-8} - 5.00×10^{-6}	1.40×10^{-9}	35
Cu-BTC/CPE	DPV	$2.07 \times 10^{-9} - 8.27 \times 10^{-7}$	1.08×10^{-9}	36
IL-GO-MWCNT/GCE	SWV	8.00×10^{-9} -1.50 × 10 ⁻⁷	6.00×10^{-9}	37
BFE	DPV	$0-9.93 \times 10^{-5}$	1.65×10^{-6}	38
DME	DPP	$0-6.62 \times 10^{-6}$	6.28×10^{-8}	39
DME	DPP	$0-4.13 \times 10^{-6}$	7.28×10^{-8}	39
DME	DPP	$0-5.79 \times 10^{-6}$	7.77×10^{-8}	39
AB-modified GCE	AASV	$8.27 \times 10^{-8} - 2.98 \times 10^{-5}$	1.65×10^{-7}	40
r-GO/GCE	SWV	$2.00 \times 10^{-7} - 9.71 \times 10^{-5}$	2.84×10^{-8}	this work

MWNT/GCE: multi-wall carbon nanotube film-modified glassy carbon electrode; IL-EGPE: ionic liquid modified expanded graphite paste electrode; Cu-BTC/CPE: copper based metal-organic framework modified carbon paste electrode; IL-GO-MWCNT/GCE: ionic liquid-graphene oxides-multi-walled carbon nanotubes modified glassy carbon electrode; BFE: bismuth film electrode; DME: dropping mercury electrode; AB-modified GCE: acetylene black nanoparticle-modified glassy carbon electrode; r-GO/GCE: glassy carbon electrode modified with a reduced graphene oxide; DPV: differential pulse voltammetry; SWSV: square-wave stripping voltammetry; DPP: differential pulse polarography; AASV: adsorptive anodic stripping voltammetry.



Figure 5. Voltammograms obtained for individual oxidation of 100 μ mol L⁻¹ of (a) Ponceau 4R; (b) Allura Red 40, (c) Brilliant Blue FCF and (d) in the presence of the three dyes using r-GO/GCE in BR buffer 0.10 mol L⁻¹, pH 7.0 (ν = 75 mV s⁻¹).

By applying Student's *t*-test, the calculated *t*-value $(t_{calc.} = 2.35)$ was found to be lower compared to the tabulated *t*-value $(t_{tab.} = 4.30)$.⁴¹ This indicates that there was no significant difference at 95% confidence level between the fortified and recovered concentrations.

The potential of the proposed sensor was also evaluated using the instant strawberry flavor juice sample containing the dye under investigation. The sample was diluted as described in Application of the proposed method section, and then inserted into an electrochemical cell containing 0.1 mol L⁻¹ BR buffer (pH 5.0), with no previous treatment. Figure 6A shows the square wave voltammograms and the standard addition curve constructed (inserted) in the range of 2 to 12 µmol L⁻¹ with the aim of determining the sample concentration. The concentration of the PNC dye found in the instant juice sample was 29.8 ± 2.52 µmol L⁻¹ *per* gram of sample.

The method was compared to the HPLC-DAD technique using optimized conditions as described in Analysis of PNC by HPLC-DAD section. As PNC shows maximum absorbance at 509 nm, the chromatographic analysis was carried out monitoring this wavelength. The sample was not subjected to any pre-treatment prior to performing the chromatographic analysis. Figure 6B presents the chromatogram obtained for a sample doped with 4 μ mol L⁻¹ of PNC and under chromatographic conditions optimized in this work. The PNC retention time observed was 7.65 min (Figure 6B).

Table 2 shows the concentrations of PNC in the sample for both methods, where proximity is observed between them.

With the aid of the Student's *t*-test (paired *t*-test), 0.207 was found for the $t_{\text{calc.}}$, which is lower than the $t_{\text{tab.}}$ value (4.30). This result demonstrates that the concentration found in the instant juice sample via the proposed method



Figure 6. (A) Square-wave voltammograms on r-GO/GCE in (a) 0.10 mol L^{-1} of BR buffer (pH 5.0); (b) 1 mL of diluted sample and successive additions of PNC: (c) 2.0; (d) 4.0; (e) 6.0; (f) 8.0 and (g) 12 µmol L^{-1} . Inserted: standard addition curve constructed by the two analytical cells method via adsorption (Analysis of PNC on r-GO/GCE section); (B) chromatogram obtained for a sample doped with 4 µmol L^{-1} of PNC.

Table 2. Determination of strawberry flavor juice sample by the r-GO/GCE method and HPLC-DAD technique (n = 3)

Method	r-GO/GCE	HPLC-DAD
Found values / (µmol L ⁻¹ <i>per</i> gram of sample)	29.8 ± 2.52	29.7 ± 0.643

r-GO/GCE: glassy carbon electrode modified with a reduced graphene oxide; HPLC-DAD: high performance liquid chromatography-diode array detector.

showed no significant difference at 95% confidence level compared to the HPLC-DAD method.

Conclusions

The sensor based on glassy carbon electrode modified with reduced graphene oxide proved to be an excellent alternative for the determination of Ponceau 4R food colorant in instant juice sample. Through the reduction of graphene oxide, it was possible to attain a significant gain in the analyte signal, which paved the way towards the determination of the PNC dye at low levels of concentration. In addition, the sensor did not show interference in the analyte response irrespective of the presence of Allure Red 40 and Brilliant Blue FCF. The proposed method applied for the determination of PNC in juice sample was compared to the HPLC-DAD technique. The results showed no significant difference between the two methods at 95% confidence level.

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References

- Agência Nacional de Vigilância Sanitária (ANVISA), Resolução CNNPA No. 44, de 1977, DOU, seção I, 01/02/78 e 24/04/78, Brazil. Available at http://www.anvisa.gov.br/anvisalegis/ resol/44_77.htm, accessed in November 2017.
- Food and Agriculture Organization of the United Nations (FAO); *Chemical Risks and JECFA*; available at http://www.fao.org/ food/food-safety-quality/scientific-advice/jecfa/, accessed in June 22, 2017.
- 3. http://eur-lex.europa.eu/legal-content/EN/ TXT/?uri=uriserv%3Asa0003, accessed in June 22, 2017.
- 4. Agência Nacional de Vigilância Sanitária (ANVISA); Resolução No. 388, de 05 de agosto de 1999, Aprova o "Regulamento Técnico que Aprova o Uso de Aditivos Alimentares, Estabelecendo suas Funções e seus Limites Máximos para a Categoria de Alimentos 19 - Sobremesas"; D.O.U.; Poder Executivo, de 09 de agosto de 1999. Available at http://portal.anvisa.gov.br/documents/33916/391619/ RESOLUCAO_388_1999.pdf/ac1c03bc-17b8-46a1-b8e5-1003d3a930d8, accessed in June 22, 2017.
- Oliveira, D. F. M.; Batista, P. S.; Muller, P. S.; Velani, V.; França, M. D.; de Souza, D. R.; Machado, A. E. H.; *Dye Pigm.* 2012, 92, 563.
- Thiam, A.; Brillas, E.; Centellas, F.; Cabot, P. L.; Sirés, I.; *Electrochim. Acta* 2015, 173, 523.
- Arnold, L. E.; Lofthouse, N.; Hurt, E.; *Neurotherapeutics* 2012, 9, 599.
- 8. Sarıkaya, R.; Selvi, M.; Erkoç, F.; Chemosphere 2012, 88, 974.
- 9. Altınöz, S.; Toptan, S.; J. Food Compos. Anal. 2003, 16, 517.
- 10. Sayar, S.; Özdemir, Y.; Food Chem. 1998, 61, 367.
- Sadeghi, S.; Hemmati, M.; Garmroodi, A.; *Electroanalysis* 2013, 25, 316.

- Tikhomirova, T. I.; Ramazanova, G. R.; Apyari, V. V.; Food Chem. 2017, 221, 351.
- Chanlon, S.; Joly-Pottuz, L.; Chatelut, M.; Vittori, O.; Cretier, J. L.; *J. Food Compos. Anal.* 2005, *18*, 503.
- Zhang, J.; Wang, M.; Shentu, C.; Wang, W.; Chen, Z.; Food Chem. 2014, 160, 11.
- Huang, J.; Zeng, Q.; Wang, L.; *Electrochim. Acta* 2016, 206, 176.
- 16. Chao, M.; Ma, X.; Food Anal. Methods 2015, 8, 130.
- Mazloum-Ardakani, M.; Rajabzadeh, N.; Dehghani-Firouzabadi, A.; Benvidi, A.; Mirjalili, B. B. F.; Zamani, L.; *J. Electroanal. Chem.* 2015, 760, 151.
- Motaharian, A.; Milani Hosseini, M. R.; Lima, J. L. F. C.; Kohlhaas, K. A.; Kleinhammes, A.; Jia, Y.; Wu, Y.; Nguyen, S. T.; Ruoff, R. S.; *Anal. Methods* 2016, *8*, 6305.
- 19. Shang, L.; Zhao, F.; Zeng, B.; Food Chem. 2014, 151, 53.
- Hudari, F. F.; de Almeida, L. C.; da Silva, B. F.; Zanoni, M. V. B.; *Microchem. J.* 2014, *116*, 261.
- Hudari, F. F.; da Silva, B. F.; Pividori, M. I.; Zanoni, M. V. B.; J. Solid State Electrochem. 2016, 20, 2491.
- 22. Shao, Y.; Wang, J.; Wu, H.; Liu, J.; Aksay, I.; Lin, Y.; *Electroanalysis* **2010**, *22*, 1027.
- Wang, Y.; Li, Y.; Tang, L.; Lu, J.; Li, J.; *Electrochem. Commun.* 2009, 11, 889.
- 24. Alwarappan, S.; Erdem, A.; Liu, C.; Li, C.-Z.; *J. Phys. Chem. C* **2009**, *113*, 8853.
- 25. Liu, Y.; Dong, X.; Chen, P.; Chem. Soc. Rev. 2012, 41, 2283.
- Xu, Y.; Gao, H.; Li, M.; Guo, Z.; Chen, H.; Jin, Z.; Yu, B.; Nanotechnology 2011, 22, 365202.
- Jampasa, S.; Siangproh, W.; Duangmal, K.; Chailapakul, O.; *Talanta* 2016, 160, 113.
- Hilder, M.; Winther-Jensen, B.; Li, D.; Forsyth, M.; MacFarlane,
 D. R.; *Phys. Chem. Chem. Phys.* **2011**, *13*, 9187.
- Hudari, F. F.; Zanoni, M. V. B.; *Microchim. Acta* 2017, 184, 4117.
- Zaijun, L.; Xiulan, S.; Qianfang, X.; Ruiyi, L.; Yinjun, F.; Shuping, Y.; Junkang, L.; *Electrochim. Acta* 2012, 85, 42.
- Hudari, F. F.; Costa Ferreira, S. L.; Zanoni, M. V. B.; *Electroanalysis* 2016, 28, 1085.
- Bard, A. J.; Faulkner, L. R. In *Electrochemical Methods: Fundamentals and Applications*; Wiley: New York, 2001, p. 226-260.
- Kucinskis, G.; Bajars, G.; Kleperis, J.; *J. Power Sources* 2013, 240, 66.
- Zhang, Y.; Zhang, X.; Lu, X.; Yang, J.; Wu, K.; Food Chem. 2010, 122, 909.
- Zhang, J.; Wang, X.; Zhang, S.; Wang, W.; Hojo, M.; Chen, Z.; J. Electrochem. Soc. 2014, 161, H453.
- Yang, X.; Sun, D.; Zeng, R.; Guo, L.; Wu, K.; J. Electroanal. Chem. 2017, 794, 229.
- 37. Wang, M.; Zhao, J.; J. Electrochem. Soc. 2015, 162, H321.

- 38. Claux, B.; Vittori, O.; *Electroanalysis* **2007**, *19*, 2243.
- 39. Combeau, S.; Chatelut, M.; Vittori, O.; Talanta 2002, 56, 115.
- 40. Yang, X.; Qin, H.; Gao, M.; Zhang, H.; *J. Sci. Food Agric.* **2011**, *91*, 2821.
- Miller, J. C.; Miller, J. N.; *Statistics for Analytical Chemistry*, 2nd ed.; Ellis Horwood Limited: England, 1988.

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