

Research Article

Eucalyptus ESTs corresponding to the enzyme glutamine synthetase and the protein D1, sites of action of herbicides that cause oxidative stress

Edivaldo Domingues Velini¹, Maria Lúcia Bueno Trindade¹, Elza Alves¹, Ana Catarina Catâneo², Celso Luis Marino³, Ivan de Godoy Maia³, Edson Seizo Mori⁴, Edson Luiz Furtado⁵, Iraê Amaral Guerrini^{6#} and Carlos Frederico Wilcken⁷

Abstract

This work was aimed at locating *Eucalyptus* ESTs corresponding to the GS enzyme (Glutamine Synthetase, EC = 6.3.1.2) and to the D1 protein, which are directly related to resistance to herbicides that promote oxidative stress. Glutamine Synthetase corresponds to the site of action of the herbicide glufosinate. Herbicides that belong to groups such as ureas, uracils, triazines and triazinones act on the D1-Qb complex (receptor of electrons from the Photosystem II) by inactivating it. The clusters EGEQRT3302E01.g, EGEQRT3001F12.b; EGEZLV1203B04.g; EGBGFB1211H06.g and EGEZLV1205F09.g enclosed complete sequences (with 356 amino acids) of the Glutamine Synthetase enzyme. The cluster EGEQSL1054G06.g is a consensus of four reads and enclosed a complete sequence of D1 Protein (with 353 amino acids). The comparison of the sequences of Protein D1 from different species showed that the substitutions of serine (S) by glycine (G) or serine (S) by threonine (T) at the position 264 could produce plants resistant to herbicides that act on electron flow on Photosystem II. The sequence of amino acids corresponding to the cluster EGEQSL1054G06.g had a serine in position 264 indicating sensitivity of the *Eucalyptus* plants to herbicides that act on this site.

Key words: Eucalyptus, glutamine synthetase, D1 protein, herbicide.

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Introduction

Four herbicide action mechanisms are known to cause plant death by promoting oxidative stress: interference with the flow of electrons of Photosystem I, or Photosystem II, and action on the enzymes protoporphyrinogen

Send correspondence to Edivaldo Domingues Velini. Universidade Estadual Paulista, Faculdade de Ciências Agrárias, Fazenda Experimental Lageado, Laboratório de Matologia, 18603-970 Botucatu, SP, Brazil. E-mail: velini@fca.unesp.br. #Supported by CNPq.

IX oxidase (Protox or PPO, E.C. 1.3.3.4), or Glutamine Synthetase (GS, EC = 6.3.1.2). Oxidative stress is associated with the production of highly reactive free radicals. These radicals are very effective in oxidizing lipids from cell membranes.

This work aimed at studying *Eucalyptus* ESTs related to the action of herbicides on the flow of electrons in Photosystem II and the synthesis of glutamine. The action of diquat and paraquat is related to the electron flow deviation in photosystem I but their effects are not directly asso-

¹Universidade Estadual Paulista "Júlio de Mesquita Filho", Faculdade de Ciências Agrárias, Fazenda Experimental Lageado, Laboratório de Matologia, Botucatu, SP, Brazil.

²Universidade Estadual Paulista "Júlio de Mesquita Filho", Instituto de Biociências, Laboratório de Xenobióticos, Botucatu, SP, Brazil.

³Universidade Estadual Paulista "Júlio de Mesquita Filho", Instituto de Biociências, Laboratório de Genética, Botucatu, SP, Brazil.

⁴Universidade Estadual Paulista "Júlio de Mesquita Filho", Faculdade de Ciências Agrárias, Fazenda Experimental Lageado, Laboratório de Melhoramento Vegetal, Botucatu, SP, Brazil. ⁵Universidade Estadual Paulista "Júlio de Mesquita Filho", Faculdade de Ciências Agrárias, Fazenda Experimental Lageado, Laboratório de Fitopatologia, Botucatu, SP, Brazil.

⁶Universidade Estadual Paulista "Júlio de Mesquita Filho", Faculdade de Ciências Agrárias, Fazenda Experimental Lageado, Laboratório de Solos, Botucatu, SP, Brazil.

⁷Universidade Estadual Paulista "Júlio de Mesquita Filho", Faculdade de Ciências Agrárias, Fazenda Experimental Lageado, Laboratório de Entomologia, Botucatu, SP, Brazil.

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ciated to the interaction with enzymes or proteins; for this reason, they should not be included in this work. The enzyme Protox, highly relevant for herbicide action and physiological modifications in plants, has been discussed in a separate paper.

Variability in the levels of action of the herbicides that act upon the sites addressed in this paper depends on characteristics of the site itself (amino acids sequence in the proteins and expression levels), as well as the plants ability to inactivate the free radicals produced, which actually generate plant intoxication. Variability in absorption and translocation capacity (even at small distances) is also an important factor that conditions the effectiveness of any compound with herbicidal action. The study of *Eucalyptus* ESTs related to detoxifying systems, which are important in defining the levels of action of herbicides that cause oxidative stress, was already carried out by Alves *et al.* (2005).

GS takes part in the glutamine synthesis and glufosinate is the only commercially available herbicide that acts on this enzyme. The death of plants intoxicated by glufosinate is not caused by a lack of glutamine or glutamate, but by the excess of non-incorporated ammonia. Working with GS is quite interesting, since glufosinate is applied in postemergence (compatible with reduced tillage in foresting areas), acting on grasses and broad-leaf plants. In addition, this compound is produced by fermentation and shows low toxicity and high safety to environment. The mechanism of action and the genes involved in resistance to this herbicide have been described by Kishi and Ishisuka (1992), Baron *et al.* (1994), Baron *et al.* (1994a), Downs *et al.* (1994), Altenburger *et al.* (1995), Bridges and Hess (2002) and Thill (2002).

White *et al.* (1990), Schwartz *et al.* (1996) and Bridges and Hess (2002) presented two sequences that code for the production of the enzyme phosphinothricin-N-acetyltransferase, which has the ability to inactivate glufosinate: the PAT gene from *Streptomyces hygroscopicus* and the BAR gene from *Streptomyces viridochromogenes*. However, no references have been found on the variability of the GS enzyme that could be related to the glufosinate herbicide selectivity. In addition, the information presented indicate the GS potential for changing the production of glutamate and, consequently, of porphyrins in plants.

According to Thill (2002), the sequences that code for the production of the enzyme phosphinothricin-N-acetyltransferase have been introduced or are in the process of being introduced into a total of 22 crops, among which are soybean, beet, canola, rice, and corn. No references have been found on *Eucalyptus* species that would have received the sequence. The plants or crops into which the enzyme production has been incorporated become resistant to the herbicide glufosinate.

With regard to interference with the electron flow in the Photosystem II, according to van Rensen (1982), Itoh and Iwaki (1989), Judson and Rowson (1989), Fuerst *et al.* (1991), Giardi et al. (1992), Arnaud et al. (1994), Egner et al. (1996), Weller (1997), Huppatz (1996), Hess (2000) and Weller (2002) the action results from binding the herbicides to the D1 protein, preventing the formation of the D1-QB complex, a receptor of electrons from this photosystem. According to those authors, the D1 protein, which is the main component that determines the functionality of the complex, is genetically coded in the chloroplast. The change of a single amino acid in the protein's sequence could modify the photosynthetic efficiency and confer resistance to several herbicides that inhibit the electron flow in Photosystem II (Examples: ureas, uracils, triazines and triazinones). Some of these herbicides show a high potential for use in *Eucalyptus* because of their control spectrum, low cost, long residual effect, preemergence and early postemergence action, low toxicity, low environmental risk, and easy transportation through the litter fall, in rain water. The herbicides diuron and ametryn are examples of compounds that present the above-mentioned characteristics, and belong to the urea and triazine groups, respectively.

This work was aimed at locating *Eucalyptus* ESTs corresponding to GS enzymes (Glutamine Synthetase, EC = 6.3.1.2) and to the D1 protein, which are directly related to the action or resistance to herbicides, synthesis of porphyrins and photosynthetic activity in plants.

Material and Methods

This work resulted from an analysis of the information bank produced in the first stage of the *Eucalyptus* Genome Project (Projeto Genoma do *Eucalyptus* - FORESTs), jointly developed by FAPESP and a consortium of four companies in the forestry industry (Duratex, Ripasa, Suzano, and VCP) and executed with the participation of 20 laboratories from the State of São Paulo associated with the AEG network (https:forests.esalq.usp.br). In all, 123,889 reads constructed from expressed sequence tags (ESTs) of cDNA libraries, mainly derived from *Eucalyptus grandis* tissues, were obtained. The tissues were removed from different organs of plants submitted to different growing conditions. The makeup and coding of the libraries are described in Table 1.

The search for enzyme sequences corresponding to GS enzyme and D1 protein was performed using the BLAST tool (Altschud *et al.*, 1997). The amino acid sequences for the GS enzyme and the D1 protein, described for different plant species, were compared with the information from the FORESTs project database using the "tBLASTn" option, allowing the identification of Clusters associated with them. Only Clusters effectively aligned with the amino acid sequences were selected, using an e-value < e-70 as a selection criterion.

The nucleotide sequences of the selected Clusters were compared with the NCBI (National Center Biotechnology Information) and the geneBank amino acid se-

quence databases after translation in all possible frames. The procedure allowed to confirm the alignment with sequences of the GS enzyme or the D1 protein from different plant species, to find the translation frame for the cluster

Table 1 - Codes and source tissues of cDNA libraries approved by the FORESTS project.

Code	Tissues / growing condition
BK1	Bark, sapwood, heartwood, and pith of 8-year old <i>E. grandis</i> trees
CL1	E. grandis calluses formed in the dark
CL2	E. grandis calluses formed in the light
FB1	Buds, flowers and fruits
LV1	Seedling leaves
LV2	Leaves from trees efficient and poorly efficient in phosphorus and boron utilization
LV3	Leaves colonized with the caterpillar <i>Thyrinteina</i> sp. for 7 days
RT3	Nursery seedling roots
RT6	Roots of trees resistant and susceptible to frost
SL1	E. grandis seedlings grown in the dark and exposed to light for 3 h prior to RNA extraction
SL4	E. globulus seedlings grown in the dark
SL5	E. saligna seedlings grown in the dark
SL6	E. urophylla seedlings grown in the dark
SL7	E. grandis seedlings grown in the dark
SL8	E. camaldulensis seedlings grown in the dark
ST2	Stems of six-month old seedlings susceptible to water deficit
ST6	Stems of seedlings susceptible to water deficit
ST7	Stems of trees resistant and susceptible to frost
WD2	E. grandis wood

and to obtain values of identity percents and similarity probability values (e-value) for the alignments.

Based on the translation frame that produced the best alignments and using the software GENERUNR, the nucleotide sequence corresponding to the clusters was translated into amino acids for the identification and analysis of Open Read Frames. The software CLUSTAL was used to align the amino acid sequences corresponding to the identified ORFs and the amino acid sequences of different plant species (with e-value < e-70 as previously described).

Results and Discussion

The sequences corresponding to the BAR and PAT genes presented by White *et al.* (1990) and Schwartz *et al.* (1996) were not found in the ESTs database of the FOR-ESTs project, indicating that their incorporation to genotypes of commercial interest should be done from external gene sources. The development of glufosinate-tolerant clones is an excellent opportunity for simplifying and reducing weed control costs in *Eucalyptus*, but the incorporation of BAR or PAT genes can only be done by producing transgenic clones.

With regard to the GS (Glutamine Synthetase), twelve clusters were identified showing high similarity with a sequence corresponding to the enzyme, obtained from *Arabidopsis thaliana*. It were observed e-values ranging from 0.0 to 4e-75 (Table 2). Nine of the clusters corresponded to consensual sequences from 2 to 106 reads. Three of the clusters were single reads. The nucleotide sequences of the clusters were compared with the NCBI (National Center Biotechnology Information) and the geneBank amino acid sequence databases after translation in all possible frames and the results are presented in Table 3.

Table 2 - Clusters from *Eucalyptus*, identified through the tBlastn tool, which showed homology with the GS enzyme (Glutamine Synthetase, EC = 6.3.1.2) and with the Protein D1.

Enzyme protein	Species	Accession	Cluster	e-value	N. of reads	N. of nucleotides
GS	Arabidopsis thaliana	AAM67495	EGEZLV1205F09.g	e-177	49	1787
	Arabidopsis thaliana	AAM67495	EGEQRT3302E01.g	0.0	45	1290
	Arabidopsis thaliana	AAM67495	EGEZLV1203B04.g	0.0	38	1433
	Arabidopsis thaliana	AAM67495	EGEQRT3001F12.b	0.0	106	2376
	Arabidopsis thaliana	AAM67495	EGBFB1211H06.g	0.0	5	1383
	Arabidopsis thaliana	AAM67495	EGEZSL8047G03.g	e-128	7	836
	Arabidopsis thaliana	AAM67495	EGUTST6223H02.g	e-117	3	834
	Arabidopsis thaliana	AAM67495	EGCCFB1220G03.g	e-100	1	404
	Arabidopsis thaliana	AAM67495	EGJFST2005F09.g	6e-99	9	1413
	Arabidopsis thaliana	AAM67495	EGCECL2011D08.g	2e-97	1	789
	Arabidopsis thaliana	AAM67495	EGEPST2215E04.g	8e-80	1	788
	Arabidopsis thaliana	AAM67495	EGEQRT3104B08.g	4e-75	2	1204
D1	Arabidopsis thaliana	NP_051039	EGEQSL1054G06.g	0.0	4	1156
	Lactuca sativa	AAL28074	EGEQSL1054G06.g	0.0	4	1156

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Only values of e-value superior to e-70 were observed by aligning the amino acid sequence of the cluster "EGE PST2215E04.g" with sequences of GS from literature. The results observed for all the other eleven clusters are presented in Table 3. Due to the high number alignments with e-values smaller than e-70, they were counted and the numbers are presented in the fourth column of Table 3. For five clusters it was possible to identify ORFs lengthening 356 amino acids and corresponding to GS sequences from literature.

The results of the alignment of the five longest sequences from Eucalyptus and sequences from eight other species are presented in Figure 1. If the same amino acid was observed in a certain position for all studied sequences, the column was marked with an asterisk enabling the identification of several conserved regions with lengths ranging from one to twelve amino acids. Results indicated that it is viable develop specific primers to be used in additional sequencing of the enzyme Glutamine Synthetase aiming to find promoters and SNPs to induce different levels of expression of the gene or activity of GS and, as a result, plants of Eucalyptus exhibiting higher or lower levels of sensitivity to glufosinate. The results also showed that the clusters EGEQRT3302E01.g, EGEQRT3001F12.b; EGEZLV12 03B04.g; EGBGFB1211H06.g and EGEZLV1205F09.g enclosed complete sequences of the Glutamine Synthetase enzyme. It must be pointed out that the 267 reads related to GS were present in almost all libraries, indicating the expression of this gene in different tissues and conditions.

Information relative to the cluster EGEQSL1054 G06.g with a high homology with amino acid sequences corresponding to the D1 protein obtained from the species *Arabidopsis thaliana* and *Lactuca sativa* are presented in Table 2. The cluster is a consensus of four reads from calluses and seedlings libraries. The nucleotide sequences of the clusters were compared with the NCBI (National Cen-

ter Biotechnology Information) and the geneBank amino acid sequence databases after translation in all possible frames and it was obtained a total of 582 alignments with e-value smaller than e-70. The results of the alignment of the *Eucalyptus* sequence with sequences from fourteen other species are presented in Table 4. For three of these fourteen species, it was possible to find sequences corresponding to Protein D1 from plants susceptible or resistant to herbicides acting on the flow of electrons in Photosystem II. The translation of the nucleotide sequence of the cluster EGEQSL1054G06.g using the frame "+2" allowed to identify an ORF lengthening 353 amino acids and corresponding to Protein D1 sequences from literature.

The alignment of the sequences using CLUSTALX indicated an almost complete homology among the sequences from different species. It was possible to identify conserved regions lengthening from eleven to 83 amino acids. In effect, differences among the amino acid sequences were observed only at the positions 11, 24, 78, 155, 238, 264, 281 and 346-353.

The comparison of the sequences of Protein D1 showed that the position 264 is the only one in which substitutions of amino acids produced plants resistant to herbicides (Figure 2). The substitutions of serine (S) by glycine (G) produced resistant plants of *Amaranthus powelli* and *Bromus tectorum*. In *Nicotiana tabacum*, resistance to herbicides resulted from the substitution of serine (S) by threonine (T). It must be highlighted that the sequence of amino acids corresponding to the cluster EGEQSL1054G06.g had a serine in position 264 indicating sensitivity of the *Eucalyptus* plants to herbicides that act on this site.

It is important to point out that the Protein D1 was completely sequenced and presents a high level of preservation. Considering that D1 Protein is encoded in chloroplasts and that changes in the amino acid sequences can

Table 3 - Translation frames for the nucleotide sequence of the clusters, minimum and maximum valures of e-value and sizes of the ORFs corresponding to the GS enzyme (Glutamine Synthetase, EC = 6.3.1.2).

Cluster	Frame	Size of the ORF	Number of	e-value	
		correspondisng to GS	sequences ⁽¹⁾	Minimum	Maximum
EGEQRT3302E01.g	+1	356	282	0.0	e-70
EGEQRT3001F12.b	+3	356	288	e-177	e-70
EGEZLV1203B04.g	+3	356	296	0.0	e-70
EGBGFB1211H06.g	+1	356	295	0.0	e-70
EGEZLV1205F09.g	+2	356	288	0.0	e-70
EGJFST2005F09.g	+3	303	135	e-105	e-70
EGEZSL8047G03.g	+2	258	140	e-110	e-70
EGUTST6223H02.g	+1	240	142	e-104	e-70
EGEQRT3104B08.g	+2	193	76	9e-80	e-70
EGCCFB1220G03.g	+1	163	151	e-104	e-70
EGCECL2011D08.g	+2	148	74	3e-78	e-70

⁽¹⁾ Number of amino acid sequences from literature aligning with sequences corresponding to the clusters from Eucalyptus.

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                              ..*.*************
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EGEQRT3302E01.g MSLLSDLLNLNLSDTTEKVIAEYIWIGGSGMDLRSKARTLNGPVSDPSKLPKWNYDGSSTNQAPGEDSEVILYPQ
                                                                                                75
EGEQRT3001F12.b MSLLSDLLNLNLSDTTEKIIAEYIWIGGSGMDLRSKARTLNGPVSDPAKLPRWNYDGSSTGQAPGEDSEVILYPQ
                                                                                                75
EGEZLV1203B04.g MSLLSDLVNLNLSDCTEKIIAEYIWIGGSGMDMRSKARTFPGPVNDPSKLPKWNYDGSSTNQAPGEDSEVILYPQ
EGBGFB1211H06.g MSALNDLLNFDLFDYTDKIIAEYIWIGGSGIDMRSKARTLRGPVTDPAKLPKWNYDGSSTDQAPGHDSEVILYPQ
EGEZLV1205F09.g INRLENLLNLDVTPYTDKIIAEYIWIGGSGIDLRSKSRTISRPVEHPSELPKWNYDGSSTGQAPGEDSEVILYPQ
                                                                                                75
    A.thaliana MSLLADLVNLDISDNSEKIIAEYIWVGGSGMDMRSKARTLPGPVTDPSKLPKWNYDGSSTGQAPGQDSEVILYPQ
                                                                                                75
         G.max MSLLSDLINLNLSDTTEKVIAEYIWIGGSGMDLRSKARTLPGPVSDPSELPKWNYDGSSTGQAPGEDSEVILYPQ
                                                                                                75
75
75
75
75
      L.sativa MALLSDLVNLDLSSITDKIIAEYIWIGGSGMDLRSKARTLSGPVSDPSELPKWNYDGSSTGOAPGEDSEVIIYPO
     N.tabacum MAHLSDLVNLNLSDSTOKIIAEYIWIGGSGMDVRSKARTLSGPVDDPSKLPKWNYDGSSTGOAPGEDSEEILYPO
      R.sativus MSLLTDLINLNLSETTDKIIAEYIWVGGSGMDMRSKARTLPGPVSDPSELPKWNYDGSSTGOAPGEDSEVILYPO
      O.sativa MANLTDLVNLNLSDCSDKIIAEYIWVGGSGIDLRSKARTVKGPITDVSQLPKWNYDGSSTGQAPGEDSEVILYPQ
         Z.mays MACLTDLVNLNLSDTTEKIIAEYIWIGGSGMDLRSKARTLPGPVTDPSKLPKWNYDGSSTGQAPGEDSEVILYPQ
          ruler 1.....10......20.....30.....40.....50.....60.....70....
                *:*:*** ..:**:** *** *:***** **::*:
                                                             * .***:*****: . .**:***:**:
EGEQRT3302E01.q AIFKDPFRRGNNILVMCDAYTPAGEPIPTNKRHAAAKVFSHPDVLAEEPWYGIEQEYTLLQRDVKWPIGWPLGGY
                                                                                               150
EGEQRT3001F12.b AIFKDPFRRGNNILVMCDAYTPAGEPIPTNKRCDAAKIFSQPDVVAEVPWYGIEQEYTLLQKNVRWPLGWPVGGY
                                                                                               150
EGEŽLV1203B04.g AIFKDPFRGGNNILVMCDAYTPGGQPIPTNKRCAAAKIFSĤPNVVVEEPWYGLEQEYTLLQKDVKWPLGWPVGGF
                                                                                               150
EGBGFB1211H06.g AVFRDPFRRGSNILVMCDAYTPAGEPIPTNKRFNAAKVFSDPHVVAEEPWYGIEQEYTLLQKDVQWPIGWPVGGF
                                                                                               150
EGEZLV1205F09.g AIFKDPFRGGNNILVICDTYTPAGEPIPTNKRARAAEIFSNQKVINEVPWYGIEQEYTLMQTNVQWPLGWPVGGY
                                                                                               150
    A.thaliana AIFKDPFRRGNNILVMCDAYTPAGEPIPTNKRHAAAEIFANPDVIAEVPWYGIEQEYTLLQKDVNWPLGWPIGGF
                                                                                               150
          G.max AIFRDPFRRGNNILVICDAYTPAGEPIPTNKRHAAAKVFSHPDVVAEVPWYGIEQEYTLLQKDIQWPLGWPVGGF
                                                                                               150
      L.sativa AIFKDPFRRGNHLLVMCDAYTPAGEPIPTNKRAAAAKIFSNPEVEKEVTWYGIEQEYTLLQKDTNWPLGWPLGGF
                                                                                               150
     N.tabacum AIFKDPFRRGNNILVICDCYTPAGEPIPTNKRHSAAKIFSHPDVVVEEPWYGLEOEYTLLOKDINWPLGWPLGGF
                                                                                              150
      R.sativus AIFKDPFRRANNILVMCDAYTPAGEPIPTNKRHAAAKVFSOPDVVAEVPWYGIEOEYTLLOKDVKWPVGWPIGGF
                                                                                               150
       O.sativa AIFKDPFRRGDNILVMCDCYTPQGEPIPTNKRHSAAKIFSHPDVVAEVPWYGIEQEYTLLQKDVNWPLGWPVGGF
                                                                                               150
         Z.mays AIFKDPFRRGNNILVMCDCYTPAGEPIPTNKRYSAAKIFSSPEVAAEEPWYGIEQEYTLLQKDTNWPLGWPIGGF
                                                                                               150
          ruler ...80.....90.....100.....110.....120.....130.....140......150
                {\tt EGEQRT3302E01.g} \  \  {\tt PGPQGPYYCGIGGDKAFGRDVVDSHYKACLYAGINISGINGEVMPGQWEFQVGPAVGISAGDELWVARYILERIT}
                                                                                              225
EGEORT3001F12.b PGPOGPYYCGVGADKALGRDIVDSHYKACLYAGINISGINGEVMPGOWEFOVGPAVGISAGDELWVARYILERIT
                                                                                               225
EGEZLV1203B04.g PGPQGPYYCGAGADKAFGRDIVDAHYKACLYAGINVSGINGEVMPGQWEFQVGPSVGISASDELWVARYILERIT
                                                                                               225
EGBGFB1211H06.g PGPQGPYYCGVGADKAFGRDIVDAHYKACLYAGINISGINGEVMPGQWEFQVGPAVGISAADQVWVARYILERIT
                                                                                               225
EGEZLV1205F09.g PGPQGPYYCGVGADKSFGRDISDAHYKACLYAGINISGTNGEVMPGQWEYQVGPSVGIEAGDHIWCSRYLLERIT
A.thaliana PGPQGPYYCSIGADKSFGRDIVDAHYKASLYAGINISGINGEVMPGQWEFQVGPSVGISAADEIWIARYILERIT
                                                                                               225
                                                                                               225
          G.max PGPQGPYYCGVGADKAFGRDIVDAHYKACIYAGINISGINGEVMPGQWEFQVGPSVGISAGDEIWAARYILERIT
      L.sativa PGPQGPYYCGIGADKAFGRDIVDAHYKACLYAGVNISGINGEVMPGQWEFQVGPSVGIAAADQIWVARYILERIT
                                                                                               225
      N.tabacum PGPQGPYYCGIGAGKVFGRDIVDSHYKACLYAGINISGINGEVMPGQWEFQVGPSVGISAADELWAARYILERIT
                                                                                               225
      R.sativus PGPQGPYYCGVGADKSFGRDVVDAHYKACLYAGINISGINGEVMPGQWEFQVGSAVGISAGDEIWVARYILERIT
                                                                                               225
       O.sativa PGPQGPYYCAAGAEKAFGRDIVDAHYKACIYAGINISGINGEVMPGQWEFQVGPSVGIAAADQVWVARYILERVT
                                                                                               225
         Z.mays PGPQGPYYCGIGAEKSFGRDIVDAHYKACLYAGINISGINGEVMPGQWEFQVGPSVGISSGDQVWVARYILERIT
          ruler ......160......170......180......190......200......210......220.....
                   {\tt EGEQRT3302E01.g} \ {\tt EIAGIVLSFDPKPIQGDWNGAGAHTNYSTKSMRSDGGYEVIKKAIEKLGLKHKEHIAAYGEGNERRLTGRHETAD}
                                                                                              300
EGEQRT3001F12.b EIADVVLSFDPKPIQGDWNGAGAHTNYSTKSMRNEGGYEIIKKAIEKLGLRHKEHIAAYGEGNERRLTGRHETAD
                                                                                               300
EGEZLV1203B04.g EIAGVVLSFDPKPIQGDWNGAGAHTNYSTKSMRNEGGYEIIKKAIEKLGLRHKEHIAAYGEGNERRLTGRHETAD
                                                                                               300
EGBGFB1211H06.g EIAGVVLTFDPKPIKGDWNGAGAHTNYSTKSMREHGGLNVIKKAIEKLGLRHKHHIAAYGEGNERRLTGRHETAD
                                                                                               300
EGEZLV1205F09.g EQTGVVLTLDPKPIEGDWNGAGCHTNYSTKSMREEGGFEVIKKAILNLSLRHKEHISAYGEGNERRLTGKHETAS
A.thaliana EIAGVVVSFDPKPIPGDWNGAGAHTNYSTKSMREEGGYEIIKKAIEKLGLRHKEHISAYGEGNERRLTGHHETAD
                                                                                               300
                                                                                               300
          G.max EIAGVVVSFDPKPIKGDWNGAGAHTNYSTKSMREDGGYEVIKAAIDKLGKKHKEHIAAYGEGNERRLTGRHETAD
                                                                                               300
       L.sativa EIYGVVVSFDPKPIPGDWNGAGAHTNYSTKTMREEGGYEVIKKAIEKLGLRHKEHIAAYGEGNERRLTGRHETAD
                                                                                               300
      N.tabacum EIAGVVVSFDPKPIPGDWNGAGAHTNYSTKSMRNEGGYEVIKKAIENLGLRHKEHIAAYGEGNERRLTGRHETAD
                                                                                               300
      R.sativus EIAGVVVSFDPKPIPGDWNGAGAHCNYSTKSMREDGGYEIIKKAIDKLGLRHKEHIAAYGEGNERRLTGHHETAD
                                                                                               300
       O.sativa EVAGVVLSLDPKPIPGDWNGAGAHTNFSTKSMREPGGYEVIKKAIDKLALRHKEHIAAYGEGNERRLTGRHETAD
                                                                                               300
         Z.mays EIAGVVVTFDPKPIPGDWNGAGAHTNYSTESMRKEGGYEVIKAAIEKLKLRHKEHIAAYGEGNERRLTGRHETAD
                                                                                               300
          ruler ..230......240......250......260......270......280.....290......300
                **** ******.*:***: ****:********:*::**.::*::
EGEQRT3302E01.g INTFLWGVANRGASIRVGRDTEKAGKGYFEDRRPASNMDPYVVTSMIAETTIIWKP--
                                                                             356
EGEORT3001F12.b INTFKWGVANRGASIRVGRDTEREGKGYFEDRRPASNMDPYVVTSMIAETTILWNP--
                                                                             356
EGEZLV1203B04.g INTFKWGVANRGASIRVGRDTEKDGKGYFEDRRPASNMDPYVVTSMIAETTILWKP--
EGBGFB1211H06.g INTFSWGVANRGASIRVGRDTEKDGKGYFEDRRPASNMDPYVVTSMIAETTILLKP--
EGEZLV1205F09.g INTFSWGVANRGCSIRVGRDTEKQGKGYLEDRRPASNMDPYIVTSLLAETTLLWEP--
                                                                             356
    A.thaliana INTFLWGVANRGASIRVGRDTEKEGKGYFEDRRPASNMDPYVVTSMIAETTLLWNP--
                                                                             356
          G.max INTFLWGVANRGASVRVGRDTEKAGKGYFEDRRPASNMDPYVVTSMIADTTILWKP--
                                                                             356
      L.sativa INTFLWGVANRGASIRVGRDTEKEGKGYFEDRRPASNMDPYVVTSMIAETTILWDNKS
                                                                             358
     N.tabacum INTFKWGVANRGASIRVGRDTEREGKGYFEDRRPASNMDPFVVTSMIAETTILSEP --
                                                                             356
      R.sativus INTFLWGVANRGASIRVGRDTEKEGKGYFEDRRPASNMEPYIVTSMIAETTILWNP--
                                                                             356
       O.sativa INTFKWGVANRGASIRVGRDTEKEGKGYFEDRRPASNMDPYVVTGMIAETTLLWKON-
                                                                             357
         Z.mays INTFSWGVANRGASVRVGRETEQNGKGYFEDRRPASNMDPYVVTSMIAETTIVWKP--
                                                                             356
          ruler ......310......320......330......340......350......
```

Figure 1 - Alignment of sequences corresponding to the enzyme Glutamine Synthetase from different higher plant species.

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EGEQSL1054G06.g	IVAAHGYFGRLIFQYASFNNSRSLHFFLAAWPVVGIWFTALGISTMAFNLNGF	300
A.hybridus	IVAAHGYFGRLIFQYASFNNSRSLHFFLAAWPVIGIWFTALGISTMAFNLNGF	300
A.powellii	IVAAHGYFGRLIFQYASFNNSRSLHFFLAAWPVIGIWFTALGISTMAFNLNGF	300
A.thaliana	IVAAHGYFGRLIFQYASFNNSRSLHFFLAAWPVVGIWFTALGISTMAFNLNGF	300
B.tectorum	IVAAHGYFGRLIFQYASFNNSRSLHFFLAAWPVVGIWFTALGISTMAFNLNGF	300
C.reflexa	IVAAHGYFGRLIFQYASFNNSRSLHFFLAAWPVVGIWFTALGISTMAFNLNGF	300
C.sinensis	IVAAHGYFGRLIFQYASFNNSRSLHFFLAAWPVVGIWFTALGISTMAFNLNGF	300
G.max	IVAAHGYFGRLIFQYASFNNSRSLHFFLAAWPVVGIWFTALGISTMAFNLNGF	300
L.sativa	IVAAHGYFGRLIFQYASFNNSRSLHFFLAAWPVVGIWFTALGISTMAFNLNGF	300
N. tabacum	IVAAHGYFGRLIFQYASFNNSRSLHFFLAAWPVVGIWFTALGISTMAFNLNGF	300
O.sativa	IVAAHGYFGRLIFQYASFNNSRSLHFFLAAWPVVGIWFTALGISTMAFNLNGF	300
P.ginseng	IVAAHGYFGRLIFQYASFNNSRSLHFFLAAWPVVGIWFTALGISTMAFNLNGF	300
P.hybrida	IVAAHGYFGRLIFQYASFNNSRSLHFFLAAWPVVGIWFTALGISTMAFNLNGF	300
S.latifolia	IVAAHGYFGRLIFQYASFNNSRSLHFFLAAWPVVGIWFTALGISTMAFNLNGF	300
Z.mays	IVAAHGYFGRLIFQYASFNNSRSLHFFLAAWPVVGIWFTALGISTMAFNLNGF	300
A.powellii R	IVAAHGYFGRLIFQYACFNNSRSLHFFLAAWPVIGIWFTALGISTMAFNLNGF	300
B.tectorum R	IVAAHGYFGRLIFQYAEFNNSRSLHFFLAAWPVVGIWFTALGISTMAFNLNGF	300
	IVAAHGYFGRLIFQYATFNNSRSLHFFLAAWPVVGIWFTALGISTMAFNLNGF	300
ruler	250260270280290300	

Figure 2 - Alignment of sequences of Protein D1 from different higher plant species.

Table 4 - Translation frames for the nucleotide sequence of the cluster EGEQSL1054G06.g, e-values for the alignments, amino acid sequence lengths corresponding to D1 protein of different higher plant species and identity percentages in relation to the sequence of the cluster.

Species	Frame	g.i.	e-value	Length of sequences ⁽¹⁾	Identity (%)
Amaranthus hybridus	+2	131237	0.0	353	95
Amaranthus powellii	+2	16119014	0.0	348	94
Arabidopsis thaliana	+2	515374	0.0	353	95
Bromus tectorum	+2	53828145	0.0	353	95
Cuscuta reflexa	+2	400881	0.0	353	95
Camellia sinensis	+2	51234109	0.0	353	95
Glycine Max	+2	34398494	0.0	353	94
Lactuca sativa	+2	16755684	0.0	97	95
Nicotiana tabacum	+2	11762	0.0	353	95
Oryza sativa	+2	50946439	0.0	353	95
Panax ginseng	+2	52220790	0.0	353	95
Petunia hybrida	+2	131253	0.0	353	95
Silene latifolia	+2	62149327	0.0	353	96
Zea mays	+2	902201	0.0	353	95
Amaranthus powellii ⁽²⁾	+2	16904362	0.0	349	94
Bromus tectorum ⁽²⁾	+2	53828147	0.0	353	94
Nicotiana tabacum ⁽²⁾	+2	4589848	0.0	353	95

⁽¹⁾Number of coded amino acids. ⁽²⁾Resistant to photosystem II inhibiting herbicides.

produce plants with different photosynthetic efficiency and levels of sensitivity to herbicides, it is important to carry out new researches aiming to find *Eucalyptus* plants with different sequences of the protein. The maternal inheritance facilitates the fixation of desirable characteristics and the use of favorable genes in breeding programs. The presence of long conserved regions will be quite helpful to make up the primers required to study specific regions of the Protein D1.

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