

## Copper Improves the Production of Laccase by the White-Rot Fungus *Pleurotus pulmonarius* in Solid State Fermentation

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### ABSTRACT

*Pleurotus pulmonarius* (Fr) Quélet, produced laccase as the main ligninolytic enzyme when cultivated on solid-state cultures using corn cob as substrate. The addition of copper greatly increased the production of enzyme. The addition of 25.0 mM CuSO<sub>4</sub> increased the level of laccase from 270 to 1,420 U.L-1 and the fungus showed high resistance to copper under the conditions used in this work.

**Key words:** Copper, laccase, *Pleurotus pulmonarius*, phenoloxidase, solid state fermentation

### INTRODUCTION

Laccases (benzenediol:oxygen oxidoreductases, EC 1.10.3.2) are glycosylated polyphenol oxidases that contain four copper ions per molecule. These enzymes catalyse the one-electron oxidation of a wide variety of organic and inorganic substrates, including mono-, di- and polyphenols, methoxyphenols, aromatic amines, and ascorbate, with concomitant four-electron reduction of oxygen to water (Leonowicz et al., 2001). Laccases are produced by the majority of white-rot fungi described to-date as well as by other types of fungi and plants, insects and some bacteria. Fungal laccases are believed to be involved in the degradation of lignin, the removal of potentially toxic phenols arising during lignin degradation, the fruit body development, pigment production and antimicrobial activity (Eggert, 1997).

Laccases have become important industrially relevant enzymes because of a number of diverse applications, e.g. for biocatalytic purposes such as delignification of lignocellulosics and cross-linking of polysaccharides, for bioremediation applications such as waste detoxification and textile dye transformation, for food technological uses, and for biosensor and analytical applications (Mayer and Staples, 2002).

In several organisms, laccases are constitutively produced in small amounts. However, their production can be considerably enhanced by a wide variety of substances, including aromatic or phenolic compounds, metal ions, alcohol, and detergents (Leonowicz et al., 2001). The increase in laccase activity by white-rot fungi in response to aromatic and phenolic substances such as ferulic acid, 2,5 xylidine, p-anisidine and veratryl alcohol is well-documented, the effect of metal ions, especially copper has only recently been

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addressed. The effect of copper on laccase synthesis was studied in *Trametes versicolor* (Collins and Dobson, 1997), *Pleurotus ostreatus* (Palmieri et al., 2000) among several other white-rot fungi.

Recently, the capability of the white-rot fungus *Pleurotus pulmonarius* to produce high titres of laccase and efficiently to decolourise industrial dyes on submerged and solid-state systems was described (Zilly et al., 2002). The production of laccase was greatly affected by the presence of soluble phenolic compounds in the medium (Tychanowicz et al., 2004). The aim of this work was to study the effect of copper on the production of laccase by *P. pulmonarius* cultivated in solid-state cultures.

## MATERIALS AND METHODS

### Organism and culture conditions

*Pleurotus pulmonarius* CCB-19 was obtained from the Culture Collection of the São Paulo Botany Institute. It was cultured on potato dextrose agar slants for 2 weeks at 28° C. When the slant was fully covered with the mycelia, mycelial plugs measuring 10 mm in diameter were made and used as inoculum for solid state cultures. For production of enzymes on solid-state system, four mycelial plugs were transferred to 250 ml Erlenmeyer flasks containing 5 g of corn cob (approximate size 3 x 3 x 2 mm<sup>3</sup>) enriched with 0.15 g of glucose and 0.014 g ammonium tartrate. Approximately 15 mL of mineral solution (Herpöel et al., 2000) was used to adjust the moisture content to 75%. This medium was designated in this work as basal medium. The basal medium was supplemented with different amounts of CuSO<sub>4</sub>·5H<sub>2</sub>O, MnSO<sub>4</sub>·H<sub>2</sub>O, ZnSO<sub>4</sub>·7H<sub>2</sub>O, AgNO<sub>3</sub>, CdSO<sub>4</sub>·8H<sub>2</sub>O, HgCl<sub>2</sub> and MgCl<sub>2</sub>·6H<sub>2</sub>O ranging from 0 to 15.0 mM. Dry weight of the substrate and moisture content were determined gravimetrically, after drying samples at 105° C. Incubation was carried out at 30° C. At periodic intervals, 50 ml of cold water was added to the cultures and the mixtures were shaken for 1 h at 4° C. The mixtures were filtered, and the filtrates were used as enzymatic extract. Results were expressed as the mean of at least three different cultures.

### Enzyme assays

Laccase activity was performed spectrophotometrically at 525 nm in a reaction medium containing 0.5 mM syringaldazine ( $\epsilon_{525}=65 \text{ mM}^{-1}\text{cm}^{-1}$ ) in 0.1 M phosphate buffer (Leonowicz and Grzywnowicz, 1981). The aryl alcohol oxidase (AAO) activity was estimated by the oxidation of 5 mM veratryl alcohol (3,4 dimethoxybenzyl alcohol) to veratraldehyde in 0.1 M sodium phosphate buffer, pH 6.0 ( $\epsilon_{310}=9.30 \text{ mM}^{-1}\text{cm}^{-1}$ ) (Muñoz et al., 1997). The manganese peroxidase (MnP) activity was assayed by the oxidation of 1 mM MnSO<sub>4</sub> in 0.05 M sodium malonate, pH 4.5, in the presence of 0.1 mM H<sub>2</sub>O<sub>2</sub>. Manganic ions, Mn<sup>+3</sup>, form a complex with malonate, which absorbs at 270 nm ( $\epsilon_{270}=11.59 \text{ mM}^{-1}\text{cm}^{-1}$ ) (Wariishi et al., 1992). The lignin peroxidase activity was determined as the oxidation rate of 4 mM veratryl alcohol to veratraldehyde in 0.1 M tartrate buffer pH 3.0 in the presence of 0.2 mM of H<sub>2</sub>O<sub>2</sub> (Tien and Kirk, 1983). The enzymatic activity was expressed as international units (U) defined as the amount of enzyme required to produce 1  $\mu\text{mol product}\cdot\text{min}^{-1}$  and expressed as U.L<sup>-1</sup>. The differences in laccase production (P<0.01) among the treatments tested were assessed using one-way ANOVA with Tukey's honestly significant difference contrasts.

### Determination of residual glucose and ammonium nitrogen

Residual glucose was determined by using the glucose oxidase kit from *Labtest*® (Brazil). Residual NH<sub>4</sub><sup>+</sup>-N was determined by the salicylate-hypochlorite method (Forster, 1995).

### Chemicals

The enzymatic substrates, syringaldazine and veratryl alcohol were obtained from Sigma Chemical Corp. (St. Louis, Mo). All other reagents were of analytical grade.

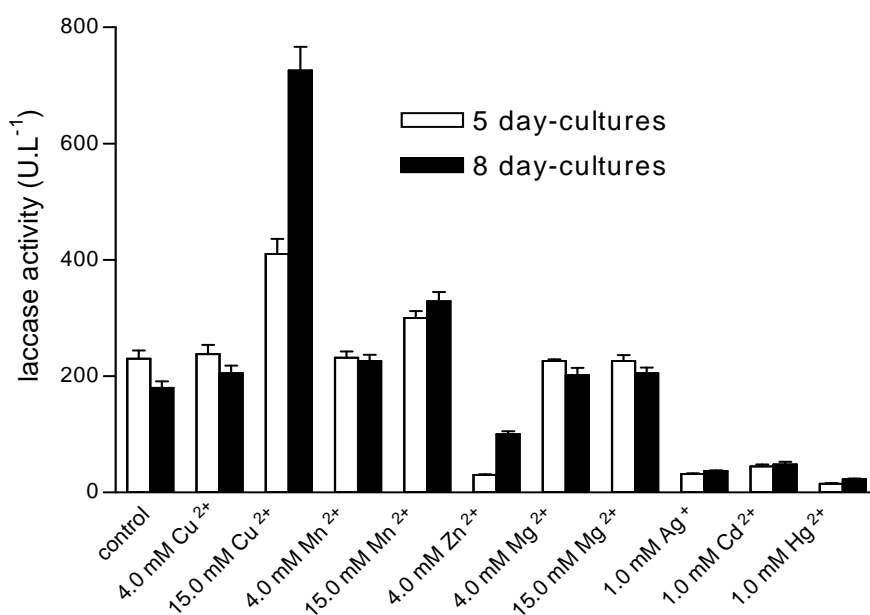
## RESULTS AND DISCUSSION

### Effect of copper and other metal ions on laccase production by *P. pulmonarius* on solid state cultures

To study the effect of metallic ions on the growth and on laccase formation, the basal medium (glucose-ammonium tartrate-corn cob powder) was supplemented with Cu<sup>2+</sup>, Ag<sup>+</sup>, Cd<sup>2+</sup>, Cu<sup>2+</sup>,

$\text{Hg}^{2+}$ ,  $\text{Mn}^{2+}$  and  $\text{Zn}^{2+}$  (Fig. 1).  $\text{Ag}^+$ ,  $\text{Cd}^{2+}$  and  $\text{Hg}^{2+}$  were highly toxic for the organism and very low growth was observed in these cultures. When 15.0 mM of  $\text{Cu}^{2+}$  or  $\text{Mn}^{2+}$  were added to the cultures, a stimulatory effect in the laccase activity, more evident to copper, was observed. These results are in agreement with the data of literature where copper has been reported as a strong laccase

inducer in white-rot fungi (Collins and Dobson 1997).  $\text{Mg}^{2+}$  did not produce any effect in both growth and laccase production. No visual alteration on the growth and on the levels of laccase activity was observed by the addition of 4.0 mM of  $\text{Cu}^{2+}$ . In the cultures supplemented with 15.0 mM of  $\text{Cu}^{2+}$ , an evident visual inhibition of growth was observed (data not shown).



**Figure 1** - Effect of metallic ions in the production of laccase by *P. pulmonarius* in solid-state fermentation.

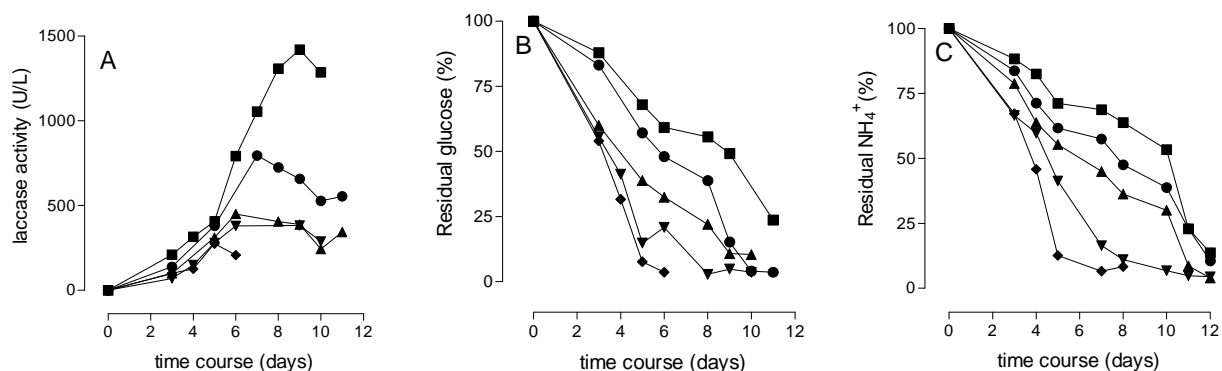
The time course of solid-state cultures supplemented with different amounts of copper is shown in Fig. 2. When copper was added to the cultures up to 10.0 mM, no inhibition in the consumption of substrates glucose and ammonium, and no alteration in the levels of laccase activity were observed in the cultures. In the culture where copper at 10.0-25.0 mM was present, the decrease in the glucose and ammonium consumption by the fungus suggested inhibition of growth (Fig. 2B-2C). However, in these cultures, laccase activity increased up to 8-fold when compared with the levels of enzyme obtained in the control cultures (270 U.L<sup>-1</sup> in the basal medium) to 1,420 U.L<sup>-1</sup> in the medium supplemented with 25.0 mM  $\text{CuSO}_4$ . No growth was observed when more than 40 mM of copper was added in the cultures, even after 20 days of cultivation. Very low AAO, Mn peroxidase and lignin peroxidase activities were detected in all

cultures, supplemented or not with copper. Copper is an essential micro-nutrient for most living organisms, and copper requirements by micro-organisms are usually satisfied by very low concentrations of the metal, in order of 1-10  $\mu\text{M}$ . However, copper present in higher concentration is extremely toxic to microbial cells (Labbé and Thiele 1997), although some copper-tolerant fungi had already been described (De Groot and Woodward 1999). The results suggested that in solid-state cultures, *P. pulmonarius* presented high resistance to copper.

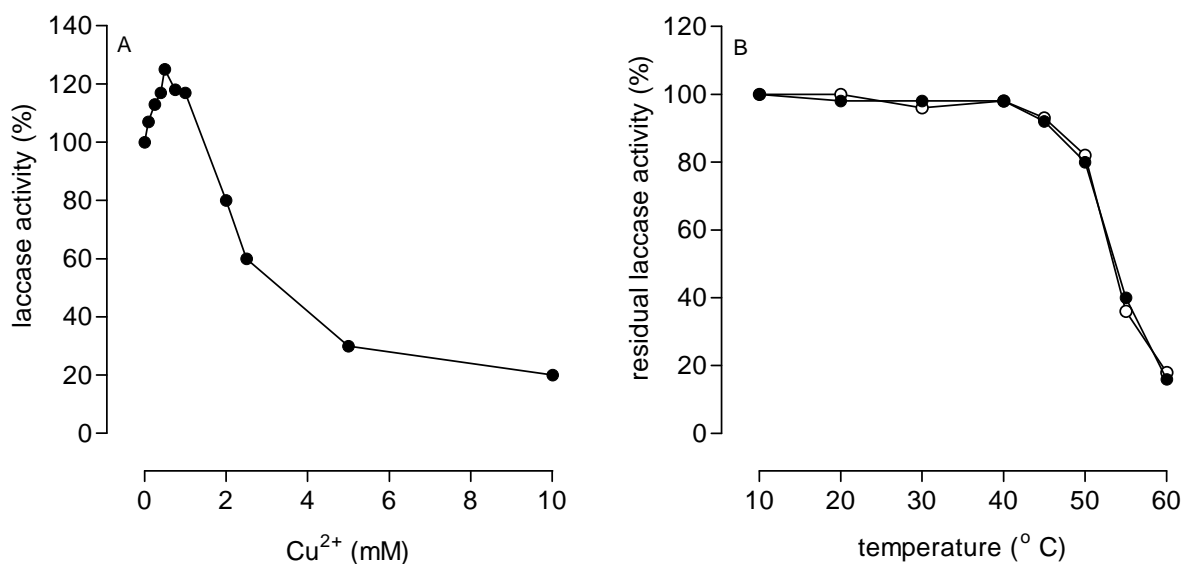
In the ascomycete *Podospora anserina*, in which laccase mRNA, amongst others, increased in response to copper and aromatic compounds, it was postulated that laccase acts as a defense mechanism against oxidative stress (Fernandez-Larrea and Stahl, 1996). This protective function was partly attributed to the chelation of copper ions during synthesis of the laccase enzyme. It is

well known that copper can cause oxidative damage of proteins by the induction of oxidative stress associated with the production of reactive oxygen species like hydroxyl or superoxide radicals (Stoys and Bagchi 1995). In *Pleurotus ostreatus* cultures, the presence of copper decreased the activity of an extracellular protease

(Palmieri et al., 2001). This might explain the positive of copper on enzyme stabilization in the crude extracts. Metal responsive elements (MRE) were identified in the promoters of *P. ostreatus* laccase genes *poxc* and *poxa 1b* (Baldrian, 2003). These MERs interact with copper-responsive transcription factors.



**Figure 2** - Effect of  $\text{CuSO}_4$  on laccase production and glucose and ammonium consumption by *P. pulmonarius*. The organism was grown on a glucose-ammonium tartrate-corn cob solid system and several amounts of  $\text{CuSO}_4$  was added: (◆) control culture; (▼) plus 10.0 mM  $\text{CuSO}_4$ ; (▲) plus 15.0 mM  $\text{CuSO}_4$ ; (●) plus 20.0 mM  $\text{CuSO}_4$ ; (■) plus 25.0 mM  $\text{CuSO}_4$ .



**Figure 3** - Effect of copper on the activity (A) and stability (B) of crude *P. pulmonarius* laccase. In A: the laccase activity was assayed using 6 day-culture filtrate without supplementation of copper. In B: The enzyme was incubated during 1 h at several temperatures in the absence (●) or presence of 1 mM  $\text{CuSO}_4$  (○)

### Effect of copper on *in vitro* laccase activity

The activity of laccase *in vitro* was determined under different copper concentrations (Fig. 3). At lowest copper concentrations, the enzyme was slightly activated. At 1 mM CuSO<sub>4</sub>, the enzyme was 20% more active than in the absence of copper. However, highest concentrations of copper inhibited the enzyme (Fig. 3A). The presence of 1 mM copper did not increase the stability of the enzyme when it was maintained 1 h at several temperatures (Fig. 3B). These data suggested that the increase of laccase activity in *P. pulmonarius* cultures after copper addition was due to increased laccase production and not by activation and/or stabilisation of the enzyme in the extracellular environmental, considering that 1 mM of copper slightly activated the enzyme and had no effect in the stabilisation of enzyme and the activity of enzyme was inhibited by copper at concentrations higher than 1 mM. Recently, it has been shown that *Pleurotus ostreatus* laccase was greatly activated by the presence of copper in the reaction medium (Baldrian and Gabriel, 2002).

Several studies showed the regulation of the synthesis of several different laccase isoforms by copper occurs at the level of gene transcription (Collins and Dobson, 1997, Palmieri et al., 2000). The supplementation of *P. ostreatus* cultures with copper increased the production of all isoenzymes of laccase produced by the fungus (Baldrian, 2003).

### CONCLUSION

Until now most studies of effect of copper on the production of laccase have been carried out using liquid cultures. To our knowledge, this is the first report that describes the effect of copper on the production of laccase by a white-rot fungus cultivated in solid-state system, a condition under which the fungus grew closer as found in the nature. The addition of copper in solid-state cultures of *P. pulmonarius* efficiently stimulated laccase formation by the fungus. The reason why copper effectively stimulates *P. pulmonarius* laccase synthesis is at present not clear and new experiments are necessary to understand the induction process. However, the use of copper enriched solid-state cultures can be an attractive way to obtain high titres of laccase which could be useful in several industrial applications.

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### RESUMO

*Pleurotus pulmonarius* (Fr) Quélet, um basidiomiceto causador da podridão branca da madeira produz lacase como principal enzima ligninolítica quando cultivado em meio em estado sólido utilizando sabugo de milho como substrato. O íon cobre tem grande efeito na produção da enzima. Os melhores resultados foram obtidos pela adição de 25.0 mM de CuSO<sub>4</sub> que causou uma elevação dos níveis enzimáticos de 270 U.L<sup>-1</sup> para 1.420 U.L<sup>-1</sup>. O fungo mostrou uma alta resistência ao íon cobre nas condições de cultivo utilizadas neste trabalho.

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