

HERBASPIRILLUM SEROPEDICAE AND SUGARCANE ENDOPHYTIC INTERACTION INVESTIGATED BY USING HIGH PRESSURE FREEZING ELECTRON MICROSCOPY

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ABSTRACT

The interaction between sugar cane plantlets and *H. seropedicae* was investigated using High Pressure Freezing followed by Freeze Substitution. Microscopical observation showed consistent differences between this approaches when compared with the conventional preparation, specially related to appearance of the bacteria cell and the endophytic attachment to the host cell wall.

Key words: *H. seropedicae*, high pressure freezing, endophytic interaction.

INTRODUCTION

Herbaspirillum seropedicae are Gram-negative bacteria capable of fixing nitrogen and promoting plant growth in different grasses (1,4,8). Ecological studies have shown that these bacteria can be isolated from the interior of roots, stems and leaves of many grasses, but never from soil samples (7). In addition, studies involving light and transmission electron microscopy have clearly demonstrated the endophytic nature of the plant-bacteria interaction (5,9). At structural level, the interactions between *Herbaspirillum seropedicae* and sugar cane plants have been receiving attention over the last 13 years. Studies based on chemically fixed samples of roots and shoots have demonstrated that the endophytic bacteria colonize randomly as single cells or micro colonies in the apoplast (intercellular spaces, cell wall and xylem lumen) as cited by James and Olivares (5). In the present study, we used High Pressure Freezing Technique (HPF) followed by Freeze Substitution to investigate the sugar cane / *H. seropedicae* interaction at microscopic level.

MATERIALS AND METHODS

Bacterial strains and growth conditions

The bacterial strain used in this study was *Herbaspirillum seropedicae* (BR11175), grown overnight at 30°C and 120 rpm in Dyg's medium.

Plant Material

Micro-propagated plantlets of sugar cane (3) var. RB72454 were grown in a modified 2% sucrose MS medium (6) and inoculated with 0.1 mL of the suspension containing 10⁸ cell.mL⁻¹.

Cryo-technique preparation and microscopy evaluation

Plantlets were harvested one week after inoculation, and small pieces of roots and leaves (0.1 – 0.6 mm) were carefully collected in filter paper and inserted into the support of aluminium covered with hexadecane (1-hexadecene, Fluka, Buchs, Switzerland). The freezing process was performed at -196°C and at 2,100 Bar (HPM 010 - High Pressure Freezing Machine, Baltec). The frozen samples were transferred to the Freezer Substitution

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Apparatus (Baltec) and, then fixed with 2% osmium tetroxide in 100% acetone solution for 24h (-85°C/8h, -60°C/8h e -20°C/8h). Samples were washed twice (acetone) and infiltrated with Spurr resin. Polymerised blocks were sectioned, stained and viewed using a transmission electron microscope (TEM) (Zeiss EM 900) operating at 80 KV under standard conditions.

RESULTS AND DISCUSSION

Microscopic observations showed significant differences between the cryo-technique approaches as compared with the conventional preparation, specially in relation to the appearance of the bacteria cell (Figs. 1 and 2) and the type of the endophytic attachment to the host cell wall (Figs. 1c, 1d, 2d, 2e and 2f). *H.*

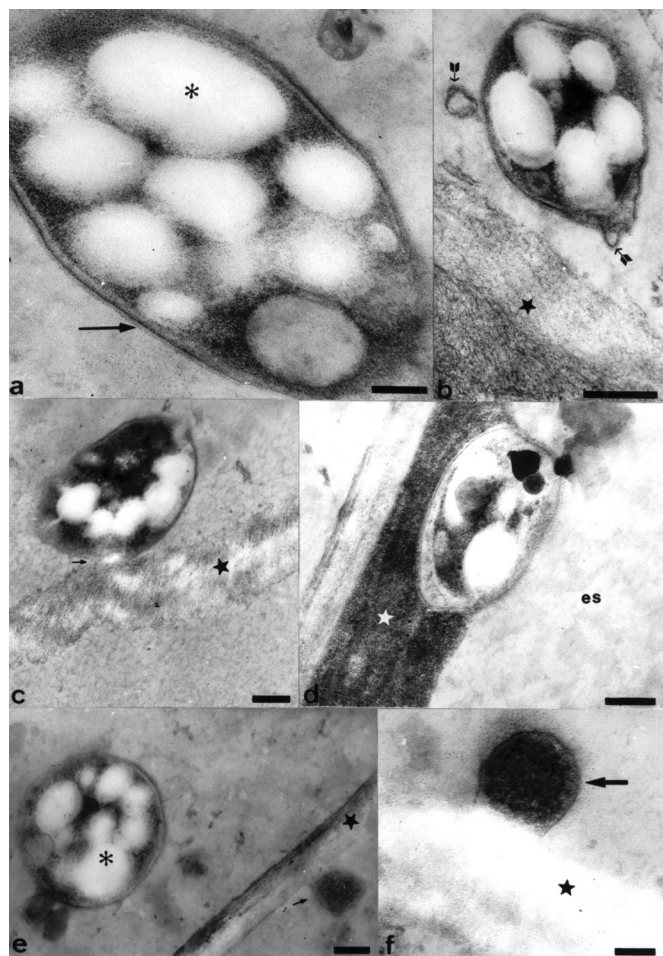


Figure 1. *H. seropedicae* strain BR11175 colonizing the apoplast close to vascular parenchyma cells of the leaf tissue of the sugarcane plants (var. RB72454). **Figure a:** Note various and large PHB granules (asterisk) in the cytoplasm of non-adhered bacteria. Bacteria cell wall (arrow). **Figure b:** Bacteria close to the plant cell wall (star); cell wall structures (arrows). **Figure c:** Bacteria adhered to plant cell wall. Note that at the adhesion site both cell walls limits can not be identified (arrow). **Figure d:** Bacteria immersed in plant cell wall (star). **Figure e:** Bacteria close to the plant cell wall (star). Multivesicular body in the periphery of plant cytoplasm (arrow). **Figure f:** Fusion of the multivesicular body with plant cell membrane. Legend: asterisk = poly- β -hydroxybutyrate; es = extracellular space. Magnifications: Bars = a, b – 100 nm; c, d, e – 150 nm; f – 120 nm.

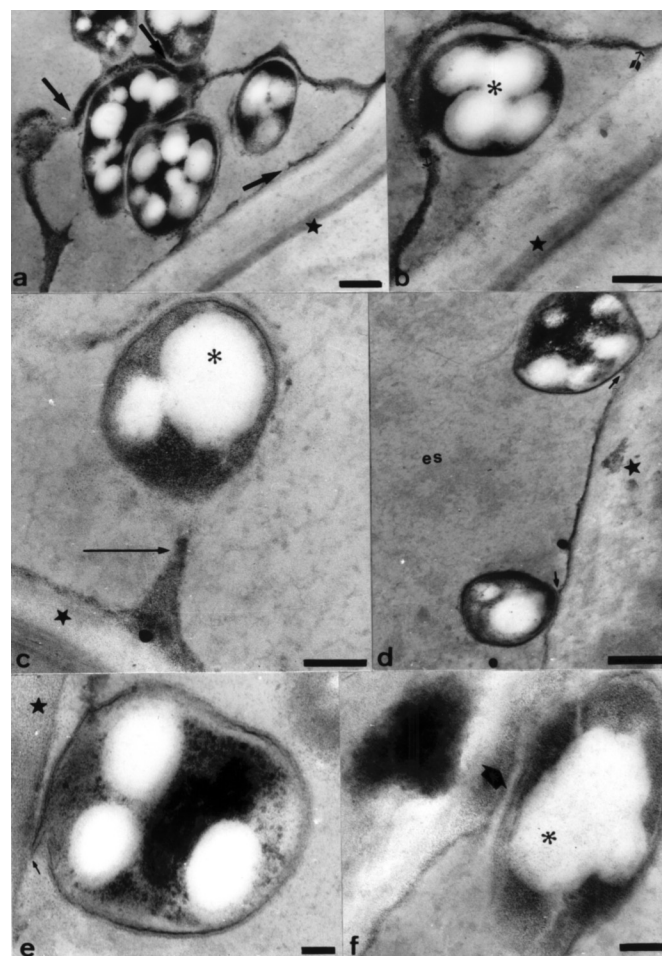


Figure 2. *H. seropedicae* strain BR11175 colonizing the apoplast of cortical cells of the root tissue of the sugarcane plants (var. RB72454). **Figure a and b:** Bacteria near the plant cell wall (star) involved by amorphous material (arrow). **Figure c:** Bacterium attached to the plant cell wall (star) by stalk electron-dense material (arrow). **Figure d:** Bacterium in close contact to the plant cell wall. Note alterations in both cell walls in the adhesion site (arrows). **Figure e:** Bacterium protrusion (arrow) toward to the plant cell wall (star) in the adhesion site. **Figure f:** Bacterium adhered with the plant cell wall (star). Note that the limit of the both cell walls is not clear (arrow). Legend: asterisk = poly- β -hydroxybutyrate; es: extracellular space. Magnifications: Bars = a, d – 0.25 μ m; b – 150 nm; c – 0.75 μ m; e, f – 100 nm.

seropedicae was localized as a single cell or a micro colony inside the apoplast of leaves. Bacteria were also found inside apparently dead vascular parenchyma cells. Root cortex apoplast showed bacteria surrounded by an amorphous matrix of unknown composition (Figs. 2a and 2b), as well as adhered to the plant cell wall by a stalk of electron-dense material (Fig. 2c). Similar results were observed for the first time by Gyaneshwar (2). In both roots and leaves, bacterial cytoplasm has shown numerous poly- β -hydroxybutyrate (PHB) granules. At adhesion sites both bacteria and plant cell walls showed altered aspects (Figs. 1c, 2d, 2e and 2f) and bacterial protrusions could be observed (Fig. 2e). In roots, multivesicular bodies were observed close to the adhesion sites (Figs. 1e and 1f). In conclusion, high-pressure technique was an essential tool to obtain morphological data demonstrating alterations in both bacteria and plant cells during *Herbaspirillum seropedicae* / sugar cane interactions.

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RESUMO

Uso da técnica de congelamento por alta pressão no estudo da interação endofítica de *Herbaspirillum seropedicae* e cana-de-açúcar

A interação entre plântulas de cana-de-açúcar e *H. seropedicae* foi investigada pelo uso da técnica de congelamento por alta pressão seguida de criosubstituição. Observações microscópicas evidenciaram diferenças marcantes entre esta técnica e preparações convencionais, especialmente relacionadas a

ultraestrutura da bactéria e às estruturas envolvidas na adesão à superfície da parede celular da planta hospedeira.

Palavras-chave: *H. seropedicae*, congelamento por alta pressão, interação endofítica.

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