

CHRONIC MYOCARDIAL DAMAGE IN EXPERIMENTAL *T. cruzi* INFECTION OF A NEW WORLD PRIMATE, *CEBUS* sp. MONKEY

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SUMMARY

Eighteen *Cebus apella* monkeys, (juvenile and adult of both sexes) were inoculated five years ago, with three *Trypanosoma cruzi* strains (CA1, n = 10; Colombian, n = 4 and Tulahuén, n = 4), either by conjunctival or intraperitoneal route, once or repeatedly. Parasitological, hematological, serological, enzymatic, radiographic, electro and echocardiographic findings have been previously published¹⁵ and they are similar to those observed in human pathology. The most frequent electrocardiographic alteration was right branch bundle block.

Six animals, chosen at random, were sacrificed. Those sacrificed 20 to 25 months post-first inoculation showed focal accumuli of leukocytes with myocytolysis. Foci of diffuse interstitial fibrosis with mild infiltrate of leukocytes among fibers were observed in the animals sacrificed 36 to 47 months post-inoculation. No parasites were seen. The lesions were more prominent in the ventricular walls and the septum. The fact that the infiltrates were predominant in the animals sacrificed at a shorter time after first inoculation and that fibrosis was more severe in those sacrificed at a longer time suggests that there is a progression of the infiltrative lesions to fibrosis, with a leukocytic activity indicative of a chronic phase.

These lesions are similar to those described in human chronic Chagas' disease. This would demonstrate that this model is useful in evaluating a progress in the knowledge of the pathogenesis which is still a controversial issue, immunology, immunogenesis and chemotherapeutic agents of the chronic and indeterminate phases of this disease.

KEY WORDS: Cebus monkey; Chagas' disease; Pathology.

INTRODUCTION

Chagas' disease or American trypanosomiasis is a zoonosis, restricted to the American continent, caused by transmission of the protozoon *Trypanosoma cruzi* by triatomine bugs. It can

also be transmitted by alternative mechanisms such as blood transfusion, congenital transmission, laboratory accidental infection, organ transplantation and the oral route⁶.

This investigation received support from the UNDP/WORLD Bank/WHO Special Programme for Research and Training in Tropical Disease (ID 790122), and by the Universidad del Salvador, Buenos Aires, Argentina.

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An efficient study of the pathogenesis of the indeterminate and chronic phases becomes difficult due to the slow evolution of Chagas' disease¹¹.

These difficulties have been accentuated "due to the lack of suitable animal models for chronic Chagas' disease, which would produce lesions resembling those found in human"¹⁷.

The diversification of the research objectives and of the animal models chosen by the different research groups, makes it difficult to develop and reproduce, the electrocardiographic and pathological patterns of this disease. The different animal models under study are: rat²⁸, mouse²², dog²³ and rabbit¹⁶, in search of the ideal experimental model¹.

Chagas' disease pathogenesis remains without a clear explanation although different mechanisms: immunological^{12, 27, 32}, neurogenic²¹, hypoxemic^{20, 34} have been proposed.

As experimental Chagas' disease has been studied in *Cebus sp.* monkeys by other authors without producing clear results^{4, 5, 13, 24, 35}, the objective of the present work is to describe myocardial damage observed, after a five-year follow-up, in the *Cebus apella* monkey, a New World prima-

te, native to the Paraguayan Chaco, experimentally infected with different *T. cruzi* strains.

Partial parasitological, immunological, electrocardiographic and echocardiographic studies have been previously reported¹⁴⁻¹⁵.

The results show that this monkey is a suitable model of chronic Chagas' disease.

MATERIAL AND METHODS

Fifty-three *Cebus apella* monkeys, with normal electrocardiograms, echocardiograms and specific serology for Chagas' disease, were selected from a breeding and rearing outdoor colony. They were kept in captivity in the indoor colony in individual cages with free water provision and feeding was based on a standard pellet diet (25% protein, 3% lipids) prepared by Cargill (Buenos Aires, Argentina), supplemented with fresh fruit twice a week.

The animals were divided into four groups, one control and three infected. The age, sex, weight, parasite strain, route, and dose of inoculum and number of inoculations carried out are detailed in the summary of the experimental design (Table 1).

EXPERIMENTAL DESIGN

STRAIN MATERIAL AND METHODS	CA 1	COLOMBIAN	TULAHUEN	CONTROL
Number of animals	10	4	4	35
Sex	male	2 male - 2 female	male	male
Estimated age at first inoculation (years)	6-10	1,2-3	4-4,5	5-9
Weight (g)	2110 - 3320	940 - 1800	1660 - 1950	2250-2570
Date of first inoculation	06-80, 07-81	09-82, 10-82	11-82, 12-82	—
Number of inoculations at 06-84	1 / 2	17 / 18	10 / 11	—
Number of <i>T. cruzi</i> (each inoculation)	4 X 10 ⁴ to 1 X 10 ⁶	3 X 10 ⁶	3 X 10 ⁶	—
Route	conjunctival	i.p.	i.p.	—

The 10 adult monkeys receiving the CA1 strain were inoculated with the parasite's metacyclic forms by the conjunctival route. Three received one inoculation of 4×10^4 parasites; four received one inoculation of 1×10^6 parasites; and the last three received two inoculations, the first of 4×10^4 parasites and the second of 1×10^6 parasites one year later.

The two other inoculated groups (receiving the Colombian and Tulahuen strains) were repeatedly inoculated with 3×10^6 blood forms of the parasite by the intraperitoneal route — a route intended to produce better absorption of the parasite. The four young males receiving the Tulahuen strain were inoculated 10 or 11 times at intervals ranging from few days to 30 weeks and the four juvenile inoculated with the Colombian strain were inoculated 18 or 19 times at intervals ranging from 3 to 24 weeks. The periodic reinoculations were performed in order to approximate conditions found by people living in endemic areas, where the periods of natural reinfection vary.

The follow-up of the animals of the control group and the infected ones, along the course of the natural evolution, was performed by means of the control of parasitemia (fresh-drop, Strouts method²¹ and/or xenodiagnosis⁸ specific serology (indirect hemagglutination — IHB (Celloghost - Chagas Behringwerke) and enzyme immunoassay - ELISA 37), hematological parameters, plasmatic proteins and electrocardiogram. Xenodiagnosis was carried out with third nymphal stage *Triatoma infestans*, using 4 boxes containing 10 bugs each, that were examined 30 and 60 days afterwards.

The electrocardiograms were recorded using a Fukuda FSC-7100 monitor at a speed of 25 a 50 mm/sec. The animals were handled under 10 mg/kg/w ketaminehydrochloride anesthesia (Ketalar, Parke Davis, Buenos Aires, Argentina). The precordial leads used were V1 to V6 as in human, and V1R and V3R in order to obtain a better evaluation of the right cavities.

These studies were carried out once a week during the first three months of the infection and then twice a month during the first two years. At present, they are being performed once a month.

In order to carry out the objective of the present work, six animal chosen at random, from those inoculated that showed electrocardiographic disturbances were sacrificed¹⁷. Four animals of the control group were sacrificed with the same purpose.

The whole heart and samples of intestine, liver, spleen, skeletal muscle and kidneys were collected. The heart was immediately removed, washed in 0.9% saline and fixed in Zamboni's liquid²⁰.

The heart was cut according to Olsen's technique²⁶. The material, 3 tissue samples of each ventricle and auricle, as embedded in paraffin after previous dehydration increasing alcohols and cleared in benzene; the sections were stained with hematoxylin-eosin, Masson's trichromic and Movat's pentachrome and examined under light microscope.

By means of serial sections, the following areas were studied: the union of a superior cava vein (SCV) and the right auricle (RA); a search of the sinus node, and the portion of the interventricular septum corresponding to the septal flap of the tricuspid, a search of the atrioventricular node and the His bundle, were attempted.

RESULTS

General Comments

The follow-up of the control group showed no alterations in the different parameters studied. Neither gross nor microscopic cardiac lesions were observed in the control animals sacrificed.

During the acute phase, positive parasitemia was detected in all the infected animals either by the fresh drop, the Strout's method or xenodiagnosis. The period in which parasitemia started to become negative ranged from 5 to 60 weeks in the animals infected with CA1 strain, 18 to 54 weeks in those with the Colombian strain and 46 to 49 in those with the Tulahuen strain.

The highest titers for chagasic specific serology were 1/128 and 1/256 according to the *T. cruzi* strain utilized¹⁵. They started to decrease

two and a half years after infection. At the moment of the sacrifice, all the animals had electrocardiographic patterns compatible with Chagas' disease.

Neither electrocardiographic alterations nor spontaneous deaths were recorded during acute phase, or immediately to inoculations.

At present, all the infected monkeys show electrocardiographic (Table 2)¹⁷, and echocardiographic¹⁸ disturbances, which remained unmodified during the follow-up.

The electrocardiographic patterns in the group of infected animals resemble those described in human chagasic cardiomyopathy in the chronic phase^{3, 19}. In the six sacrificed monkeys a right bundle branch block was detected in the three that had been inoculated with the CA1 strain. Left anterior hemiblock was observed in one monkey inoculated with CA1 strain, and in one with Colombian strain^{1/2}. In the one with Tulahuen strain an intermittent right bundle branch block was detected. The other monkey inoculated with the Colombian strain exhibited repolarization disturbances.

Morphological finding:

Control group:

From the anatomical point of view, the "in situ" gross study of the heart of the primate, showed a vertical localization in relation to the longitudinal axis, similar to that of the slender human.

From the microscopical point of view the cardiac morphology was similar to that described in humans, both in the contractile tissue and the conduction system.

Infected animals:

Gross anatomy

In the animals sacrificed the heart was slightly enlarged and flabby with dilatation of the right side chambers (Fig. 1).

Thinning of the apical region of the left ventricle (apical aneurysm) was not found. Neither macroscopic nor microscopic signs of chronic cardiac failure were found.

Table 2
Electrocardiographic Disturbances in Monkeys Inoculated with Different *T. cruzi* Strains and Examined During the Chronic Phase

Strain	Monkey	RD	LVO	RVH	LAH	IIRBB	ASF	RBBB
CA 1 (n=10)	36	×						
	68	×				×		
	76	●			×			×
	82	●						×
	84		×	×				
	104	●						×
	116				×			
	125		×	×				
	173		×					
	313		×					
Colombian (n=4)	1	●			×			
	2		×		×			
	308	●	×					
	399					×		
Tulahuen (n=4)	190			×			×	
	233		×					
	388	●				×		
	391		×				×	

● *Sacrificed*

RD = Repolarization disturbances

LVO = Left ventricular overload

RVH = Right ventricle hypertrophy

LAH = Left anterior hemiblock

IIRBB = Intermittent right bundle branch block

ASF = Anteroseptal fibrosis

RBBB = Right bundle branch block



Fig. 1 — Macroscopy of a heart with right side chamber dilatation.

Light microscopy

A diffuse focal myocarditis with scattered fibrosis proportional to the time of infection was observed in all the layers of the heart and particularly in the ventricles. The coronary arteries showed no alterations.

The mononuclear infiltrates of lymphocytes, plasmocytes and macrophages formed perivascular or interstitial accumuli around the fibers which showed myocytolysis (fig. 2).

These infiltrates were also observed in neurons and fibers of the parasympathetic nervous plexus of the interauricular septum. In some cases the nerves were completely infiltrated (fig. 3).

Another conspicuous feature was the diffuse focal fibrosis that was interstitial, perivascular or of substitution. This lesion was predominantly observed in the interventricular septum and the ventricular walls (Fig. 4).

In the conduction system, the specialized cells showed foci of myocytolysis surrounded by mild infiltrates. In the perinodal zone, a more severe infiltration was observed especially in the pericardium around the sinus node (Fig. 5 to 7). A mild fibrosis was detected in the atrioventricular node and in the His bundle (fig. 8).

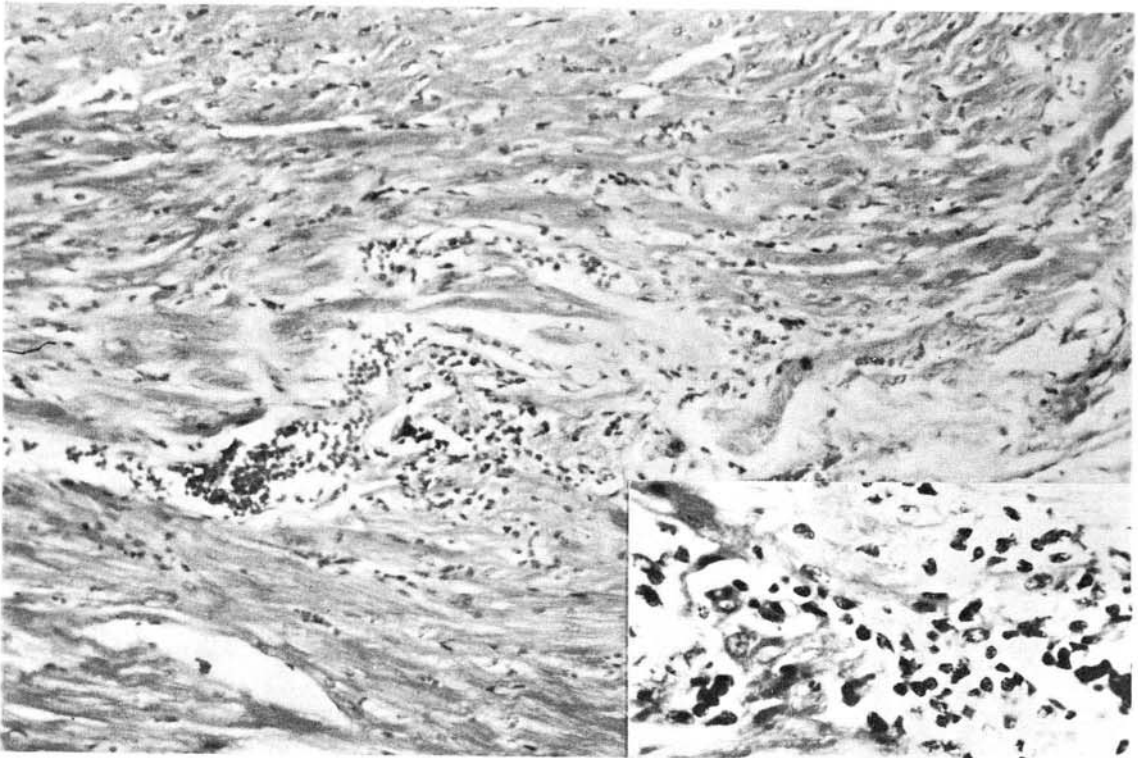


Fig. 2 — Area of free wall of the left ventricle of a monkey, 21 months after first inoculation with Colombian strain. Focal infiltrates among muscular fibers are seen (HE 40x).

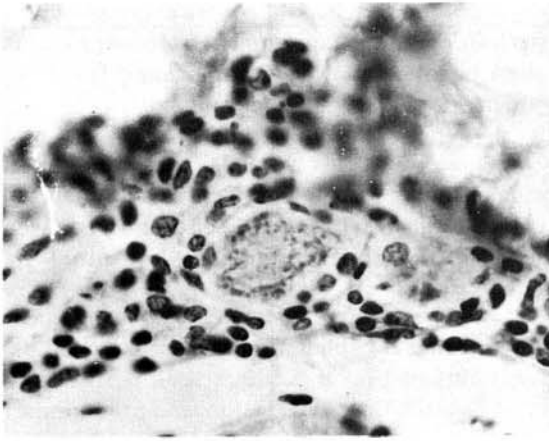


Fig. 3 — Neuron cells of the ganglia of the interauricular wall surrounded by infiltrates of lymphocytes in a monkey 21 months after inoculation with Colombian strain (HE-400x).

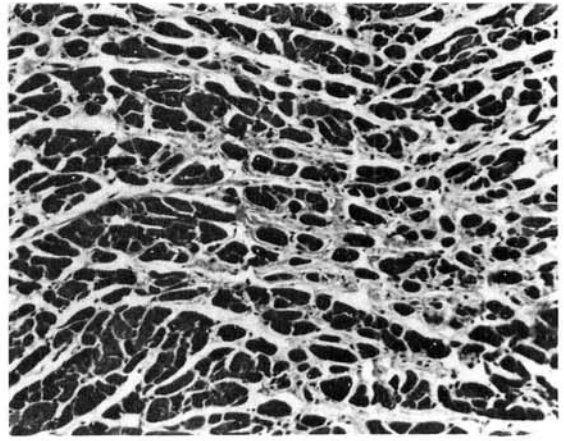


Fig. 4 — Septum in a monkey 58 months after first inoculation with CA1 strain in which a marked interstitial fibrosis and scarce mononuclear elements in the fibrotic tissue are observed (Masson's trichromic-100x).

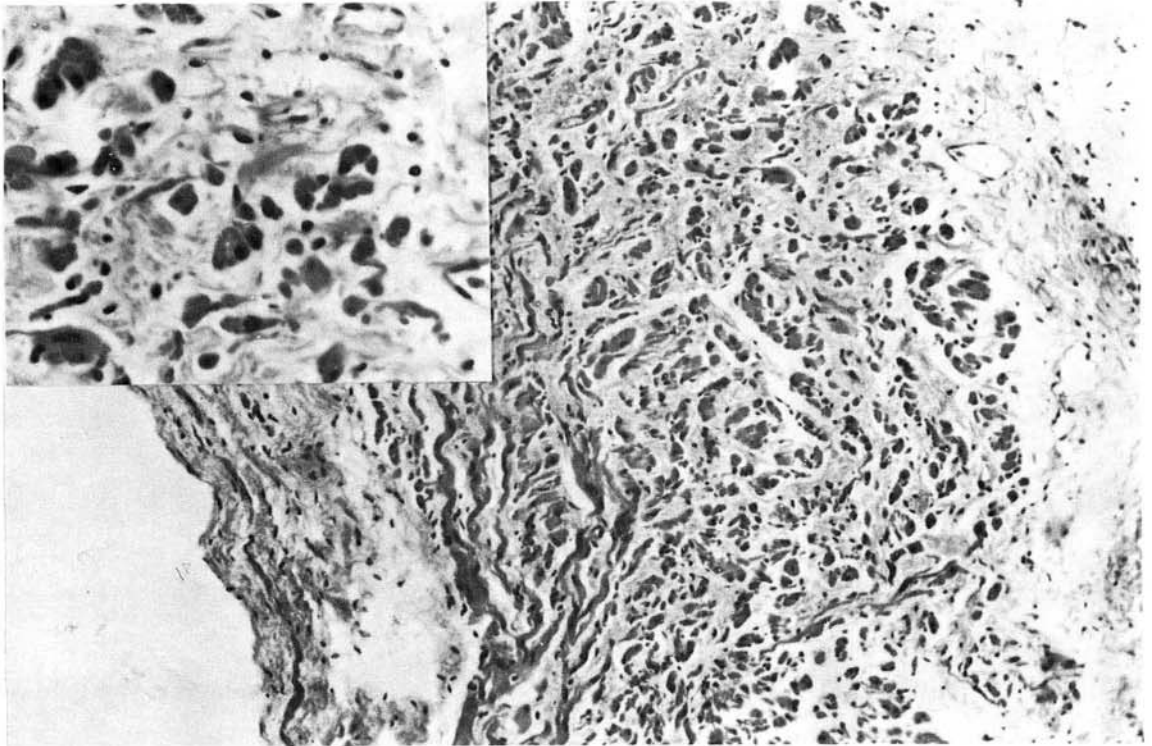


Fig. 5 — Sinus node of a monkey 53 months after first inoculation with CA1 strain presenting slight infiltrates among the specialized fibers (HE-40x).

Foci of severe myocardial fibrosis, either interstitial or of substitution, were the image more commonly found in the animals sacrificed 36 to 47 months post-infection whereas the principal

characteristic observed in those sacrificed 21 to 25 months after first inoculation was the presence of interfibrillar and perivascular infiltrates with foci of myocytolysis.

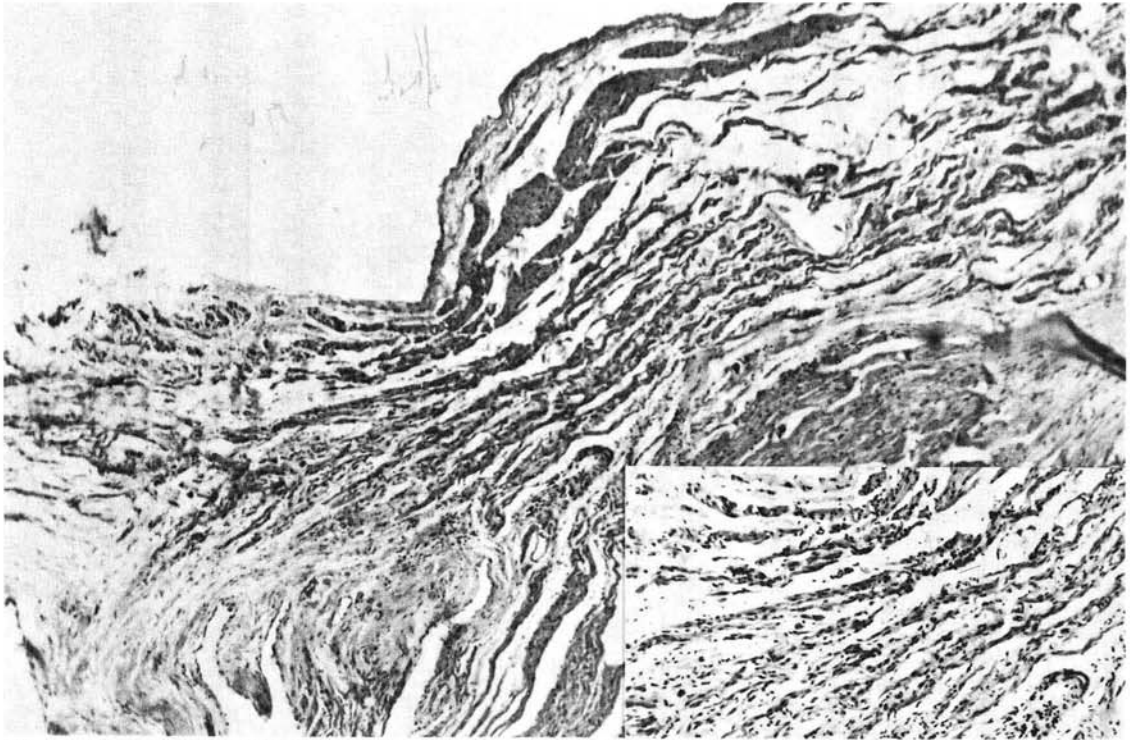


Fig. 6 — Atrioventricular node of a monkey 67 months after first inoculation with CA1 strain (HE-40x).
Insert: mononuclear elements are more concentrated in the perinodal zone (HE-100x).

This fact would suggest that there is a progression to fibrosis of the infiltrative lesions observed in the animals sacrificed sooner after first inoculation. On the other hand, mild mononuclear infiltrates among the myocardial fibers and foci of myocytolysis could be detected together with an important septal and ventricular fibrosis.

The ventricular myocardial cells showed enlarged nuclei and fibers, a sign of myocardial hyperthrophy.

DISCUSSION

The development of an animal model that resemble human Chagas' disease has been a priority of the research plan devised by the RSG-SWG of the TDR of WHO.

The main objectives in this area, are to improve understanding of the immunopathogenesis of chronic lesions, the trial of new drugs and to develop vaccines against this disease.

Most experimental infections have been carried out in mice²², rats²⁸ and dogs²³, which are susceptible to *T. cruzi* infection³. However the observations in susceptible hosts are usually limited to the early acute phase of the infection, probably due to the high mortality rate ensured by parasite inoculation. On the other hand, the *T. cruzi* experimental infection in mice does not reproduce either the distribution or the extent of the panmyocardites observed in man²⁵. In contrast to mice, rats and dogs, rabbits and primates are apparently more resistant to *T. cruzi* infection^{33, 35}. The course of experimental Chagas' disease in adult *Cebus sp.* monkeys has been previously studied but with not very clear results^{4, 5}.

These experimental studies have aimed at different aspects (clinical, parasitological, epidemiological, immunological) and have been carried out using different animal species, routes of inoculation, strain of parasites, length of infection, follow-up and method of evaluation complicating the interpretation of them.

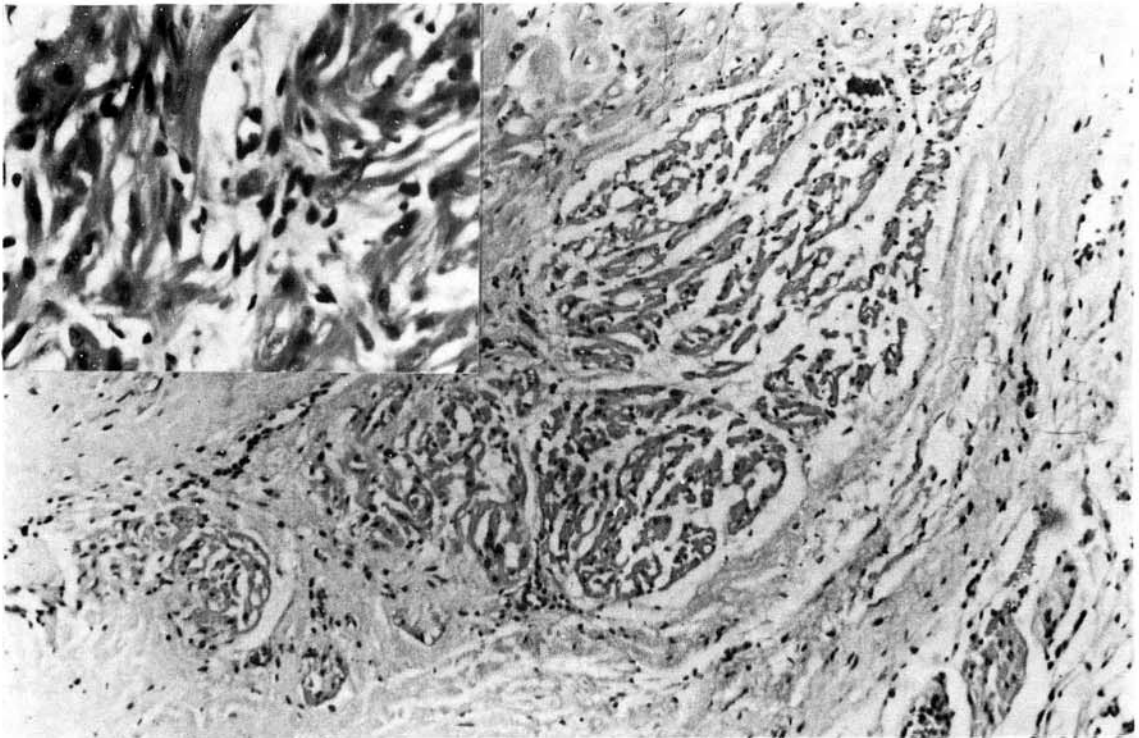


Fig. 7 — Bundle of His of a monkey 48 months after first inoculation with CA1 strain showing infiltrates of lymphocytes around it (HE-400x).

Insert: A low number of lymphocytes are seen among the specialized fibers (HE-400x).

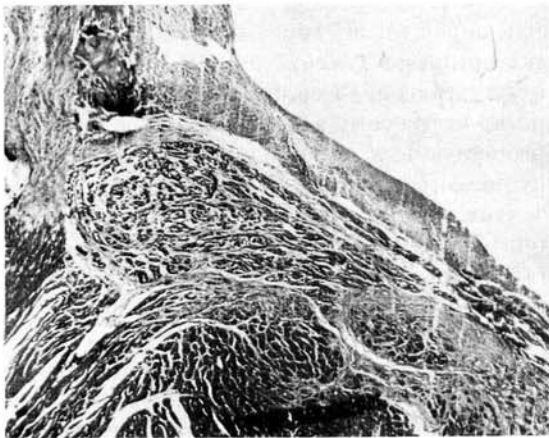


Fig. 8 — Bundle of His giving rise to the left branch in a monkey 48 months after first inoculation with Colombian strain. In the upper part of the muscular septum large fibrotic foci that seem to compromise the left branch of the bundle of His, are observed (HE-40x).

An animal model of human disease is a "living organism with an inherited, naturally acquired or induced by pathological process that

in one or more respects, closely resembles the same phenomenon in man"³⁸. That is, it should accurately reproduce the disease or lesion under study¹⁹.

The results obtained in this work, utilizing a different methodology to that used by other authors when experimentally infecting the *Cebus* monkey^{4, 5} are very interesting specially those concerning the histopathological and clinical aspects since they resemble human chronic chagasic pathology².

During the acute phase, monkeys inoculated with Tulahuen strain showed positive parasitemia detected either by the fresh drop or the Strout method until week 8-30. In those infected with the Colombian or CA1 strain, the parasitemia could be detected only by xenodiagnosis and until weeks 39-54, respectively. This agrees with findings in man in the acute phase. Positive serology was observed from week 3 on. Neither electrocardiographic alterations nor sponta-

neous deaths were recorded during this phase. These results would suggest that the morbidity observed in the course of the chronic phase in this model, as in humans, is not related to parasitemia. The electrocardiographic findings were similar to those described in humans during the chronic phase of the disease.

The histological sections of the heart showed focal myocarditis distributed in all the cavities, specially in the septum and free ventricular wall.

Mild to moderate focal infiltrates, formed by histiocytes, plasmocytes and lymphocytes, as well as myocytolysis of the myocardial fibers, were evident. Focal and diffuse myocardial fibrosis, progressively increased, predominantly in septum and ventricles, as the time of infection augmented (36 and 46 months post-first inoculation) and evident sign of lesional evolution and chronicity, supported by the presence of activated fibroblast among the myocardial fibers.

It was possible to detect an hypertrophic response of the cardiac muscular fiber and substitution and interfibrillar fibrosis.

The focal nature of the myocardial lesion would be an indirect evidence of the microcirculatory compromise in this experimental model²⁰.

The inflammatory lesions were not related to the presence of the parasite, a fact reported by VIANNA in 1911³⁶, and that represents one of the characteristics of the disease. No amastigotes were found in the serial section study. Although the bulk of evidence indicates that intracellular parasites play a negligible role in the pathogenesis of Chagas' disease, recent papers suggest that intracellular parasitism might be more frequent than previously thought⁶.

The lesions previously described, are similar to those usually found in human chronic Chagas' disease and that were described by ZILTON A. ANDRADE in humans³ and that are worthy to remember: "1) the cellular infiltrate composed of macrophages and lymphoid cells, tends to accumulate focally in areas where the local myocardial fibers show varying degrees of degenerative changes; 2) fibrosis appears as an outstanding feature, not only in focally dense areas but

also in delicate and diffuse interstitial areas, sometimes involving each cardiac fiber; 3) congestion and edema are found throughout the myocardium; 4) the parasites are difficult to find in the histological sections".

In this model, the cardiac lesions were independent of the age, sex and weight of the animal and a rather long indetermined phase, as it occurs in man who develops lesions years and even decades after being infected, and a chronic phase starting 2 and 3 years after infection, were observed.

The fact that the 100% of the animals survived the acute phase, and that histopathological lesions were similar to those of man, would demonstrate the extrapolation to the conditions and mechanism of human natural infection.

The animals infected with the Colombian and Tulahuen strains, repeatedly and using the i.p. route and a larger number of parasites than with the CA1 strain, showed histopathological lesions, in a shorter time (11-18 months post-infection, Colombian and Tulahuen strains) than those receiving one or two inoculations (36-47 months post-infection, CA1 strain).

CONCLUSIONS

The *Cebus apella* monkey, native to an endemic area, and reproducing well in captivity, would be an experimental model suitable for the study of chronic Chagas' disease, since it reproduces the human electrocardiographical and histopathological alterations, in a short-time after experimental infection.

The use of this primate will permit us to progress in the knowledge of the pathogenesis and immunopathology of both, the indeterminate and chronic phase of this disease. It will also permit us to evaluate new immunogenic and chemiotherapeutic agents acting in these stages.

This pathology, still unknown and difficult to treat, continues to be, as Chagas said¹⁰, one of the most important and serious medico-social problems in Latinamerican rural areas. Furthermore, at present, it is also a serious problem in

the urban zones since the frequency of this pathology increases, due to the internal migrations and transfusional transmission by non detected chagasic donors.

RESUMO

Lesões miocárdicas crônicas na infecção experimental pelo *T. cruzi* no macaco (*Cebus apella*).

Dezoito macacos *Cebus apella* (jovens e adultos de ambos os sexos) foram inoculados há 5 anos atrás, com 3 cepas de *T. cruzi* (CA1, n=10; Colombiana, n=4 e Tulahuen, n=4) seja por via conjuntival ou intraperitoneal, uma única vez ou repetidamente. Os achados parasitológicos, hematológicos, sorológicos, enzimáticos, radiográficos, eletro e ecocardiográficos foram anteriormente publicados¹⁵ e são semelhantes àqueles vistos no homem. O achado eletrocardiográfico mais freqüente foi o bloqueio do ramo direito.

Seis animais, escolhidos ao acaso, foram sacrificados. Aqueles sacrificados 20 a 25 meses após a primeira inoculação mostraram acúmulos focais de leucócitos com miocitólise. Focos de fibrose intersticial difusa com pequeno infiltrado de leucócitos entre as fibras foram observados em animais sacrificados 36 a 47 meses após a inoculação. Não foram encontrados parasitas. As lesões foram mais proeminentes nas paredes ventriculares e no septo. O achado de infiltrados predominantemente, nos animais sacrificados em tempo mais curto em relação à primeira inoculação e a fibrose mais severa naqueles sacrificados após um tempo maior sugere que existe uma progressão das lesões infiltrativas até a fibrose, com atividade leucocítica indicativa de fase crônica.

Estas lesões são semelhantes àquelas descritas na doença de Chagas crônica humana. Este modelo, portanto, é útil na avaliação do progresso e conhecimento da patogênese da doença, assim como de sua imunologia, imunogenese e da ação da quimioterapia, tanto na sua fase crônica como indeterminada.

ACKNOWLEDGMENT

The authors wish to thank Prof. Dr. Zilton Andrade, Director Centro de Pesquisas Gonçalo

Moniz, Brazil, for reviewing the manuscript and helpful criticism and the technical assistance of Ms. Nery Rolon and Mrs. Claudia Chiesa.

REFERENCES

1. AMORIN, D. — Cardiopatia chagastica. Modelos experimentales. *Arq. bras. Cardiol.*, 42: 243, 1984.
2. ANDRADE, Z. A. & ANDRADE, S. G. — Patologia, In: BRENER, Z. & ANDRADE, Z. A. *Trypanosoma cruzi* e doença de Chagas. Rio de Janeiro, Guanabara Koogan, 1979. p. 199-248.
3. ANDRADE, Z. A. — Mechanisms of myocardial damage in *Trypanosoma cruzi* infection. In: *Cytopathology of parasitic disease*. London, Pitman, 1983. p. 214-233. (Ciba Foundation Symposium, 99).
4. BOLOMO, N. J.; MILEI, J.; COSSIO, P. M.; SEGURA, E.; LAGUENS, R.; FERNANDEZ, L. & ARANA, R. — Experimental Chagas' disease in South American non human primate (*Cebus sp.* monkey). *Medicina (B. Aires)*, 40: 667-672, 1980.
5. BOLOMO, N. J.; MILEI, J.; COSSIO, P.; NAGLE, C.; ARANA, R.; DEL PRADO, C. E. & SEGURA, E. L. — Infección crónica por *Trypanosoma cruzi* en el *Cebus sp.* adulto. *Medicina (B. Aires)*, 41: 678-679, 1981.
6. BRENER, Z. — Pathogenesis and immunopathology of chronic Chagas' disease. *Mem. Inst. Oswaldo Cruz*, 82 (Suppl. 1): 205-213, 1987.
7. BRENER, Z. — Progress recents dans le domaine de la maladie de Chagas. *Bull. Org. mond. Santé*, 60: 845-856, 1982.
8. BRUMPT, E. O. — O xenodiagnóstico. Aplicação do diagnóstico de algumas infecções parasitárias em particular trypanosomose de Chagas. *An. paul. Med. Cirurg.*, 3(5): 97-117, 1914.
9. CAPRIS, T. A. & FERNANDEZ MOORES, A. J. — Alteraciones electrocardiograficas en la cardiopatia chagastica crónica. Estudio de 174 enfermos. *Rev. argent. Cardiol.*, 34: 200-204, 1967.
10. CHAGAS, C. — Nova tripanosomíase humana: estudos sobre a morfologia e o ciclo evolutivo do *Schizotrypanum cruzi*, n.gen.n.sp. agente etiológico de nova entidade morbida do homem. *Mem. inst. Oswaldo Cruz*, 1: 159-219, 1909.
11. COSSIO, P. M.; DIEZ, C.; LAGUENS, R. P. & ARANA, R. M. — Inmunopatología de la enfermedad de Chagas. Hechos y perspectivas. *Medicina (B. Aires)*, 40 (Suppl. 1): 222, 1980.
12. COSSIO, P. M.; LAGUENS, R. P.; DIEZ, C.; SZARFMAN, A.; SEGAL, A. & ARANA, R. M. — Chagasic cardiopathy: antibodies reacting with plasma membrane of striated muscle and endothelial cells. *Circulation*, 50: 1252-1259, 1974.

13. DORLAND, J. D. — Infection in monkeys with strains of *Trypanosoma cruzi* isolated in the United States. *Publ. Hlth. Rep.*, 158: 1006-1010, 1943.
14. EIGUCHIDE PALMERO, K.; CARBONETTO, Ch.; MALCHIODI, E. L.; MARGNI, A. & FALASCA, C. A. — Humoral and cellular parameters of the immune system of *Cebus apella* monkeys. Cross reactivity between monkey and human immunoglobulins. *Vet. Immunol. Immunopath.*, 19: 341-349, 1988.
15. FALASCA, C. A.; GRANA, D.; BUCCOLO, J.; GILI, M.; MERLO, A.; ZOPPI, J. & MARESO, E. — Susceptibility of the *Cebus apella* monkey to different strains of *T. cruzi* after single or repeated inoculation. *Bull. Pan Amer. Hlth. Org.*, 20: 117-137, 1986.
16. FIGUEIREDO, F.; MARIN-NETO, J. A. & ROSSI, M. A. — The evolution of experimental *Trypanosoma cruzi* cardiomyopathy in rabbits: further parasitological, morphological and functional studies. *Int. J. Cardiol.*, 10: 277-290, 1986.
17. GILI, M.; GRANA, D.; MARESO, E.; GARCILAZO, E. & FALASCA, C. A. — Electrocardiograma normal y patológico, en el mono *Cebus apella*, un modelo experimental de cardiopatía chagásica crónica. *Rev. argent. Cardiol.*, 55(4), 1987.
18. GILI, M.; GRANA, D.; GARCILAZO, E. & FALASCA, C. A. — Alteraciones electro y ecocardiográficas (Modo M) en un modelo experimental crónico de la enfermedad de Chagas, el primate *Cebus apella*. *Medicina (B. Aires)*, 46: 553, 1986.
19. HELD, J. R. — Appropriate animal models. *Ann. N. Y. Acad. Sci.*, 406: 13-19, 1983.
20. JORG, M. E. — Tripanosomiasis cruzi; anarquía angiopotográfica por descapilarización mesenquimorreactiva, cofactor patogénico de la miocardiopatía crónica. *Pren. méd. argent.*, 61: 94-106, 1974.
21. KÖBERLE, F. — Pathologic anatomy of enteromegaly in Chagas' disease. In: MEETING BOCKUS ALUMNI INTERNATIONAL SOCIETY OF GASTROENTEROLOGY, 2., Rio de Janeiro, 1960. *Proceedings*. Rio de Janeiro, 1961. p. 92-103.
22. LAGUENS, R. P.; MECKERT, P. C. & GELPI, R. J. — Chronic Chagas' disease in the mouse. I. Electrocardiographic and morphological patterns of the cardiopathy. *Medicina (B. Aires)*, 41: 35-39, 1981.
23. MARSDEN, P. & HAGSTROM, J. W. C. — Experimental *Trypanosoma cruzi* infection in beagle puppies. The effect of variations in the dose and source of infecting trypanosomes and the route of inoculations of the course of the infection. *Trans. roy. Soc. trop. Med. Hyg.*, 62: 816-824, 1968.
24. MARSDEN, P.; SEAH, S. K. K.; DRAPER, C. C.; PETTITT, L. E.; MILES, M. A. & VOLLER, A. — Experimental *T. cruzi* infections in Rhesus monkeys. II. The early chronic phase. *Trans. roy Soc. trop. Med. Hyg.*, 70: 247-251, 1976.
25. MOLINA, H. A.; MILEI, J.; RIMOLDI, M. T.; GONZALEZ CAPPA, S. M. & STORINO, R. A. — Histopathology of the heart conducting system in experimental Chagas disease in mice. *Trans. roy. Soc. trop. Med. Hyg.*, 82: 241-246, 1988.
26. OLSEN, E. G. J. — Examination of the heart. In: OLSEN, E. G. J. *The pathology of the heart*. 2nd ed. London, Macmillan Press, 1980. p. 5-10.
27. PAN AMERICAN HEALTH ORGANIZATION. Report of a study group on Chagas' disease. Washington, 1970 (Scientific Publication No. 195).
28. REVELLI, S. S.; AMERIO, N.; MORENO, H. S.; VALENTI, J. L.; BALBARREY, H. & MORINI, J. C. — Enfermedad de Chagas crónica en la rata. Características serológicas, electrocardiográficas e histopatológicas. *Medicina (B. Aires)*, 40 (Suppl. 1): 69-76, 1980.
29. ROSEBAUM, M. B. & ALVAREZ, A. — The electrocardiogram in chronic chagasic myocarditis. *Amer. Heart J.*, 50: 492-527, 1955.
30. STEFANINI, M.; DE MARTINO, C. & ZAMBONI, L. — Fixation of ejaculated spermatozoa for electron microscopy. *Nature*, 216: 173, 1976.
31. STROUT, R. G. — A method for concentrating hemoflagellates. *J. Parasit.*, 48: 100, 1962.
32. TEIXEIRA, A. R. L.; TEIXEIRA, L. & SANTOS-BUSH, C. A. — The immunology of experimental Chagas' disease. IV. The production of lesions in rabbits similar to those of chronic Chagas' disease in man. *Amer. J. Path.*, 80: 163-177, 1975.
33. TEIXEIRA, A. R. L.; FIGUEIREDO, F.; REZENDE, F. J. & MACEDO, V. — Chagas' disease: a clinical, parasitological, immunological and pathological study in rabbits. *Amer. J. trop. Med. Hyg.*, 32: 258-272, 1983.
34. TORRES, C. M. — Miocitólise e fibrose do miocárdio na doença de Chagas. *Mem. Inst. Oswaldo Cruz*, 58: 161-182, 1960.
35. TORRES, C. M. & TAVARES, B. M. — Miocardite no macaco *Cebus* após inoculações repetidas com *Schizotrypanum cruzi*. *Mem. Inst. Oswaldo Cruz*, 5: 85-152, 1958.
36. VIANNA, G. — Contribuição para o estudo da anatomia patológica da moléstia de Carlos Chagas. *Mem. Inst. Oswaldo Cruz*, 3: 276-294, 1911.
37. VOLLER, A.; DRAPER, C. C.; BIDWELL, D. F. & BARTLETT, A. — Microplate enzyme-linked immunosorbent assay for Chagas' disease. *Lancet*, 1: 426-427, 1975.
38. WESSLER, S. — Introduction: What is a model? Animal models of thrombosis and hemorrhagic disease. Bethesda, National Institutes of Health, 1976.

Recebido para publicação em 4/9/1989.