

Original Article

Cytotoxicity of iodine-131 radiopharmaceutical in tumor and non-tumor human cells and radioprotection by integral juices of *Vitis labrusca* L.

Citotoxicidade do radiofarmacêutico iodo-131 em células humanas tumorais e não tumorais e radioproteção por sucos integrais de *Vitis labrusca* L.

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Abstract

Iodine-131 (I-131) radioisotope it causes the formation of free radicals, which lead to the formation of cell lesions and the reduction of cell viability. Thus, the use of radioprotectors, especially those from natural sources, which reduce the effects of radiation to healthy tissues, while maintaining the sensitivity of tumor cells, stands out. The objective of the present study was to evaluate the cytoprotective/radioprotective effects of whole grape juices manufactured from the conventional or organic production systems, whether or not exposed to ultraviolet (UV-C) light irradiation. The results showed that I-131 presented a cytotoxic effect on human hepatocellular cells (HepG2/C3A) at concentrations above 1.85 MBq/mL, after 24 and 48 hours of treatment, though all concentrations (0.0037 to 7.40 MBq/mL) were cytotoxic to non-tumor human lung fibroblast (MCR-5) cells, after 48 hours. However, grape juices (10 and 20 µL/mL) did not interfere with the cytotoxic effect of the therapeutic dose of I-131 on tumor cells within 48 hours of treatment, while protecting the non-tumor cells, probably due to its high antioxidant activity. In accordance with their nutraceutical potential, antioxidant and radioprotective activity, these data stimulate *in vivo* studies on the use of natural products as radioprotectants, such as grape juice, in order to confirm the positive beneficial potential in living organisms.

Keywords: human lung fibroblast cells (MRC-5), human liver tumor cells (HepG2/C3A), radioprotection, MTT test, whole grape juice.

Resumo

O radioisótopo iodo-131 (I-131) causa a formação de radicais livres, que levam à formação de lesões celulares e redução da viabilidade celular. Assim, destaca-se a utilização de radioprotetores, principalmente de origem natural, que reduzem os efeitos da radiação nos tecidos saudáveis, mantendo a sensibilidade das células tumorais. O objetivo do presente estudo foi avaliar os efeitos citoprotetores/radioprotetores de sucos de uva integral fabricados em sistemas de produção convencional ou orgânico, expostos ou não à radiação ultravioleta (UV-C). Os resultados mostraram que o I-131 apresentou efeito citotóxico nas células hepatocelulares humanas (HepG2/C3A) em concentrações acima de 1,85 MBq/mL, após 24 e 48 horas de tratamento, embora todas as concentrações (0,0037 a 7,40 MBq/mL) fossem citotóxicas para células de fibroblasto de pulmão humano não tumoral (MCR-5), após 48 horas. No entanto, os sucos de uva (10 e 20 µL/mL) não interferiram no efeito citotóxico da dose terapêutica de I-131 nas células tumorais em 48 horas de tratamento, protegendo as células não tumorais, provavelmente devido ao seu alto poder antioxidante. atividade. De acordo com seu potencial nutraceutico, atividade antioxidante e radioprotetora, esses dados estimulam estudos *in vivo* sobre o uso de produtos naturais como radioprotetores, como o suco de uva, a fim de confirmar o potencial benéfico positivo em organismos vivos.

Palavras-chave: células de fibroblastos de pulmão humano (MRC-5), células tumorais de fígado humano (HepG2/C3A), radioproteção, teste MTT, suco de uva integral.

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1. Introduction

Iodine-131 (I-131) is a radioisotope used mainly in the diagnosis and treatment of the thyroid gland (Béron and Wémeau, 2020; Hailan and Yassin, 2021), but also has diagnostic applications in the labeling of antibodies and the adrenal gland, in the treatment of ganglion, lung or bone metastases (Sapienza et al., 2005) and the treatment of glioblastoma tumors (Koosha et al., 2021) and hepatocellular carcinoma (Boucher et al., 2007; Trall and Ziessman, 2003; Costa and Sapienza, 2012).

Hepatocellular carcinoma or primary liver cancer is a malignant epithelial tumor that arises from parenchymal liver cells and is one of the malignant diseases that affect a large number of individuals worldwide (Ahmadzadehfar et al., 2011). Its treatment is performed with I-131-lipiodol, which is prepared by replacing the iodine atom of the lipiodol molecule with its radioactive isotope I-131, which has a half-life of 8.06 days and releases beta particles and gamma radiation during its decay (Trall and Ziessman, 2003; Ahmadzadehfar et al., 2011; Costa and Sapienza, 2012).

The use of radiopharmaceuticals in carcinoma treatments is based on the fact that when they are present, in sufficient quantity in the tissue or organ, they can cause cell destruction and prevent the formation of new tissues, mainly by excitation and ionization, which may alter cell molecules chemically and result in their death (Fischer et al., 2018; Asadian et al., 2020; Nouailhetas et al., 2021). Besides, ionizing radiation may cause swelling or damage of the nucleus and cell structure as a whole, increased cell media viscosity, increased permeability, cell membrane damage, or cell cycle arrest (Fischer et al., 2018), induction of apoptosis, necrosis and autophagy, as well as organ fibrosis, atrophy and inflammation (Yun and Wang, 2017). However, these effects may also affect healthy cells (Murayama et al., 2021) and pulmonary complications have been evidenced in patients undergoing I-131-lipiodol due to tumoral arteriovenous permeation resulting in migration of the molecule to the pulmonary circulation (Ahmadzadehfar et al., 2011; Costa and Sapienza, 2012).

Consequently, the use of radioprotectants stands out. These compounds are administered together with irradiation and reduce their effects on healthy tissues, maintaining the sensitivity to radiation damage to tumor cells (Painuli and Kumar, 2016; Kuruba and Gollapalli, 2018). Radioprotectors, due to their antioxidant activities and the reduction of oxidation, prevent radiation-generated free radicals from attacking cell biomolecules, preventing injury formation and loss of cell integrity (Kunwar et al., 2012; Kumar et al., 2021; Rahgoshai et al., 2021) and promoting DNA repair (Kitabatake et al., 2020).

The evaluation of natural compounds as radioprotective agents is relevant due to their non-toxic essence, their easy availability (Yun and Wang, 2017; Kuruba and Gollapalli, 2018; Dowlath et al., 2021; Kumar et al., 2021), their antioxidant activity and the possibility that they may act effectively in reducing cancer incidence without adverse side effects (Fischer et al., 2018; Kuruba and Gollapalli, 2018; Dowlath et al., 2021). Plants possess a renewable source of metabolites with enormous chemical structural

diversity, which may have potential therapeutic relevance (Bomfim et al., 2022).

In this sense, grapes (*Vitis labrusca* L.) and their derivatives stand out, which have numerous nutritional, pharmacological and therapeutic benefits (Apostolou et al., 2013; Kim et al., 2013; Huang et al., 2015; Devi and Singh, 2017; Lopes et al., 2019), mainly due to the presence of phenolic compounds and their antioxidant activity (Fernandes et al., 2013; Maestre et al., 2013; Pinto et al., 2022).

Therefore, the present study aimed to verify the cytotoxic activities of diagnostic and therapeutic doses of I-131 in human hepatocellular carcinoma (HepG2/C3A) and non-tumoral human lung fibroblast (MRC-5) cells. Also, to evaluate the radioprotective effects of whole grape juices produced from the conventional or organic production system, whether or not exposed to type C ultraviolet light irradiation (UV-C), which has been shown to increase the synthesis of flavonoids, anthocyanins and aromatic compounds (Cantos et al., 2001; Maurer et al., 2017; Pinto et al., 2022), without producing cytotoxic or mutagenic substances (Düsmen et al., 2014).

2. Materials and Methods

2.1. Cell cultures

Considering that I-131 can be used for the treatment of hepatocellular carcinoma (Boucher et al., 2007; Trall and Ziessman, 2003; Costa and Sapienza, 2012) and that due to pulmonary circulation of I-131, pulmonary complications of patients have already been identified (Ahmadzadehfar et al., 2011; Costa and Sapienza, 2012), we used in the present study, as a test system, the human hepatocarcinoma cells, HepG2/C3A (Cat. 0291) and non-tumor human lung fibroblast, MRC-5 (Cat. 0180), obtained from the Rio de Janeiro Cell Bank. The cells were cultivated in 25cm² culture flasks containing complete culture medium (DMEM supplemented with 10% of fetal bovine serum and 1 mL/L antibiotic/antimycotic solution) and kept in a CO₂ incubator (5%, 95% humidity) at 37° C. Under these conditions, the cell cycle of both cells is approximately 24 hours. Cells in the logarithmic growth phase were used in these experiments.

2.2. Treatment solution

2.2.1. Grape juice

Vitis labrusca L. variety Concord grapes harvested in 2012 were cultivated in the municipality of Verê, belonging to the Francisco Beltrão microregion, in southwestern Paraná, Brazil. The grapes were obtained from two nearby properties (5.3km), one using conventional cultivation practice (altitude 564 m, latitude 25° 54' 01" S and longitude 52° 53' 51" W) and one with organic cultivation practice (altitude 492 m, latitude 25° 51' 21" S and longitude 52° 55' 06" W) under similar climatic and soil conditions.

The selection and treatment of grapes with UV-C irradiation was described by Cantos et al. (2001) and Düsmen et al. (2014). The following parameters were

used: radiation fluence rate of 65.6 J/m² at a distance of 30cm from the light source. The grapes were arranged in a single layer in trays that were irradiated in a cabin equipped with three UV-C lamps (Philips® 90 W) for 5 minutes. The bunches of grapes were rotated 180° and the light source remained for another 5 minutes, totaling 10 minutes of irradiation time. The irradiated material was stored for three days at 25 ± 5° C in the absence of light to promote the biosynthesis of bioactive compounds.

Organic and conventional grapes, with or without UV-C postharvest treatment, were used to obtain whole juices. The whole juice was obtained in a steam puller at 90° C and bottled in glass. Sterile samples of the four conventional, UV-C-treated and conventional UV-C-treated organic grape juices were stored in a -4° C freezer and thawed at the time of experimentation.

2.2.2. I-131 radioisotope

The I-131 radioisotope was obtained from the Institute of Nuclear Energy Research, São Paulo, Brazil. On the day of the experiment, doses of I-131 were prepared using a CRC®-25R Dose Calibrator (Capintec Inc., USA) in order to determine the activity of the radiopharmaceutical. As I-131 was diluted in saline solution, the calculation was made of the amount of µL needed to obtain each concentration that was evaluated. The cells were directly exposed to radioactive iodine, which was diluted in the culture medium, and remained so throughout the experimental period.

2.2.3. Chemical agents

Methyl methane sulfonate (MMS - 99%, CAS 66-27-3), Dimethyl sulfoxide (DMSO - 99.7%, CAS 67-68-5) and 2,2-diphenyl-1-picrylhydrazil (DPPH, CAS 1898-66-4) were purchased from Sigma. Dulbecco's Modified Eagle Medium (DMEM), Fetal Bovine Serum, Antibiotic/Antimycotic and MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide] (98%, CAS 298-93-1) were purchased from Gibco® Life Technologies (Carlsbad, CA, USA).

2.3. MTT cytotoxicity test

For the cytotoxicity test, the MTT assay was performed as described by Lopes et al. (2019). The 96-well cell culture plates were used, wherein each well 10⁴ HepG2/C3A or MRC-5 cells were seeded. After stabilization, the cultures were treated as follows: control group (CO), cytotoxic agent methyl methane sulfonate (MMS 50µM) and I-131 concentrations, defined after pilot tests and spanning a wide range of equivalent diagnostic and therapeutic doses, with activities based clinical extrapolations (0.0037; 0.0185; 0.037; 0.185; 0.37; 0.74; 1.85; 3.70; 7.40 MBq/mL). The cytoprotection/radioprotection experiment was performed with the following treatments: control group (CO); organic, conventional, organic UV-C post-treated and conventional UV-C post-treated grape juices (10 and 20 µL/mL); I-131 (1.85 MBq/mL); and cytoprotective treatments with organic grape juice (10 and 20 µL/mL) + I-131 (1.85 MBq/mL), conventional grape juice (10 and 20 µL/mL) + I-131 (1.85 MBq/mL), organic UV-C post-treated grape juice (10 and 20 µL/mL) + I-131 (1.85 MBq/mL),

and conventional UV-C post-treated grape juice (10 and 20 µL/mL) + I-131 (1.85 MBq/mL).

After 24 or 48 hours of incubation, the culture medium was replaced with MTT (0.2 mg/mL) and the absorbance readings were performed in a 550nm microplate reader (Lopes et al., 2019).

Cell viability was estimated based on the mean absorbance (A) of the control by the following formula: $A_{\text{treatment}}/A_{\text{control}} \times 100$. The experiments were performed in three independent repetitions.

All experiments involving the use of radioisotope I-131 were developed at the Mutagenesis Laboratory of the Department of Biotechnology, Genetics and Cell Biology, adequately registered by the Brazilian National Nuclear Energy Commission (CNEN).

2.4. Antioxidant activity

Antioxidant activity was determined by the DPPH (2,2-diphenyl-1-picrylhydrazil) method, as described by Brand-Williams et al. (1995) with modifications by Rufino et al. (2007). Four dilutions of each sample (grape juice) were performed. An aliquot of 0.1 mL of each sample dilution was transferred to test tubes with 3.9 mL of DPPH radical and 0.1 mL of the control solution of methyl alcohol, acetone and water. Then, homogenization was performed in a tube shaker. Spectrophotometer absorbance readings at 515 nm were taken after 30 minutes of reaction. The reaction time was established by previous tests, ranging from 30 minutes to 24 hours. Quantitation was based on the establishment of a standard curve at concentrations 10, 20, 30, 40, 50 and 60 µM from an initial DPPH solution (60 µM). Results were expressed as EC50 (mL juice/g DPPH), e.g., the values obtained correspond to the amount of sample needed to reduce the initial concentration of the DPPH radical by 50%. The experiments were performed in three independent repetitions.

2.5. Statistical analysis

The results were presented as mean and standard deviation, and submitted to one-way analysis of variance (ANOVA), followed by Dunnett's test (juice and radioisotope cytotoxicity experiments) and Tukey's (cytoprotection experiment and antioxidant activity) by the GraffPad® Prism 5 software. Differences were considered to be statistically significant when the p-value was less than 0.05.

3. Results and Discussion

Treatment with the radioisotope I-131 was cytotoxic to HepG2/C3A tumor cells at concentrations above 1.85 MBq/mL after 24 and 48 hours of exposure (Figure 1A). In addition, concentrations above 1.85 MBq/mL showed cell viability below 60% (24 hours) and 70% (48 hours) (Table 1). These data corroborate the commonly used in nuclear medicine, where low doses of I-131 presented a diagnostic effect, while high doses have a therapeutic effect, inducing tissue injury (Trall and Ziessman, 2003).

However, due to radiation migration to the pulmonary circulation, lung cells are affected by the use of

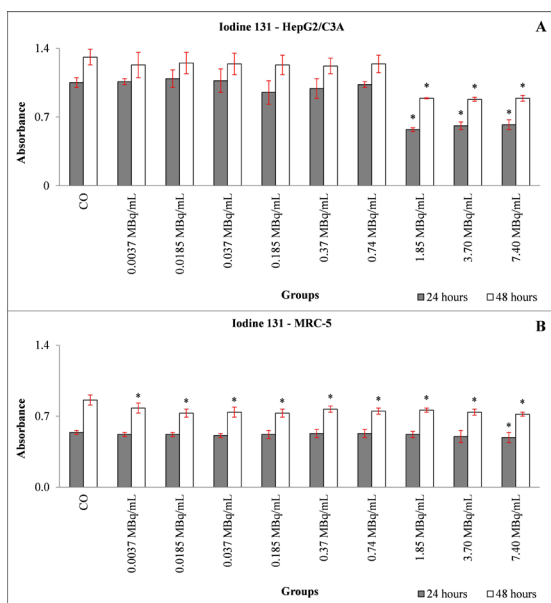


Figure 1. Mean absorbance and standard deviation of HepG2/C3A (A) and MRC-5 (B) cells treated with I-131. CO: Control; 1×10^4 cells per well, incubated for 24 and 48 hours, $n = 3$. *Statistically significant result compared to Control ($p < 0.05$, Dunnett's test).

Table 1. Percentage of viability of HepG2/C3A and MRC-5 cells treated with Iodine-131 for 24 and 48 hours, by the MTT assay.

Groups	HepG2/C3A		MRC-5	
	CV %		CV %	
	24 h	48 h	24 h	48 h
CO	100.0	100.0	100.0	100.0
0.0037 MBq/mL	100.9	93.8	96.2	90.6
0.0185 MBq/mL	103.8	95.4	96.2	84.8
0.037 MBq/mL	101.9	94.6	94.4	86.0
0.185 MBq/mL	90.4	93.8	96.2	84.8
0.37 MBq/mL	94.2	93.1	98.1	89.5
0.74 MBq/mL	98.0	94.6	98.1	87.2
1.85 MBq/mL	54.3	67.9	96.3	88.3
3.70 MBq/mL	58.0	67.1	92.5	86.0
7.40 MBq/mL	59.0	67.9	90.7	83.7

CV = Cell viability. Groups: CO = Control; Treatment with Iodine-131; 1×10^4 cells per well, incubated for 24 and 48 hours, $n=3$.

I-131 (Ahmadzadehfar et al., 2011; Costa and Sapienza, 2012). Data from the present study confirm that for MRC-5 non-tumor lung cells (Figure 1B), all I-131 concentrations (from 0.0037 to 7.40 MBq/mL) were cytotoxic within 48 hours presenting a significant decrease in the absorbances compared to the control. Within 24 hours, only the highest I-131 concentration (7.40 MBq/mL) was cytotoxic. Nevertheless, the minimum cell viability observed was 90.7% for 24 hours and 83.7% for 48 hours, showing a higher resistance of healthy cells

to the harmful effects of radiation when compared to the tumor cells (Table 1), which is essential in terms of diagnostic or therapeutic applications with I-131.

The cytotoxic effect (cell death) induced by the I-131 on HepG2/C3A and MRC-5 cells were possibly due to the induction of oxidative stress generated mainly by beta particles resulting from the radioactive decay of I-131 (Hosseinimehr, 2010; Asadian et al., 2020; Nouailhetas et al., 2021). Reactive oxygen species can interact with cell macromolecules, inducing damage and even cell mortality, and this is one of the main mechanisms of the therapeutic and side effects of I-131 (Hosseinimehr et al., 2013; Nouailhetas et al., 2021).

The simultaneous administration of the radioisotope I-131 and the whole grape juices to HepG2/C3A cells for 24 hours (Figure 2) showed that these treatments, except for the concentration of 20 μ L/mL of UV-C organic grape juice statistically increased the absorbance compared to the treatment with I-131, which was cytotoxic at 24 and 48 hours. Also, simultaneous administration of I-131 and juices presented mean absorbances similar to the treatment with only juice, except for treatment with conventional juice (20 μ L/mL), indicating that even with the administration of I-131, cell divisions were not inhibited, nor cell viability decreased (Table 2).

Kim et al. (2013) have shown the protection of HepG2/C3A cellular oxidation pretreated with procyanidins, a grape juice derivative. The authors justify this protection by reducing reactive oxygen species levels and modulating the activity of glutathione, malondialdehyde and antioxidant enzymes, events that may have occurred in the present experiment when HepG2/C3A cells were treated with grape juices and the radioisotope.

However, within 48 hours (Figure 2), this cell division-inducing effect was reversed, as all treatments simultaneously performed with I-131 and juices, except 10 μ L/mL of UV-C organic and conventional grape juices, showed a cytotoxic effect compared to control, reducing cell viability by more than 15% compared to the 24-hour treatment (Table 2). Also, all treatments performed with I-131 and juices presented mean absorbances similar to those obtained with the treatment performed with I-131 alone.

According to Costa and Sapienza (2012), the combined administration of chemotherapies can potentiate the effect of I-131-lipiodol in hepatocellular carcinoma treatments, increasing from 40% to 90% the cases with tumor reduction or stabilization. This effect was not observed in the present study considering the simultaneous administration of grape juices and I-131, which resulted in cytotoxic activity similar to the radioisotope alone within 48 hours.

Nevertheless, it is noteworthy that within 48 hours, UV-C conventional grape juice (20 μ L/mL), organic grape juice (20 μ L/mL) and UV-C organic grape juice (10 and 20 μ L/mL) showed a cytotoxic effect on HepG2/C3A cells. These data corroborate the work of Lopes et al. (2019), in which the same juices evaluated in the present study, at concentrations of 10 to 100 μ L/mL, presented antitumor activity for HepG2/C3A cells, especially for the highest concentrations, in the longest exposure time and with UV-C postharvest treatment. According to El-Din et al. (2019),

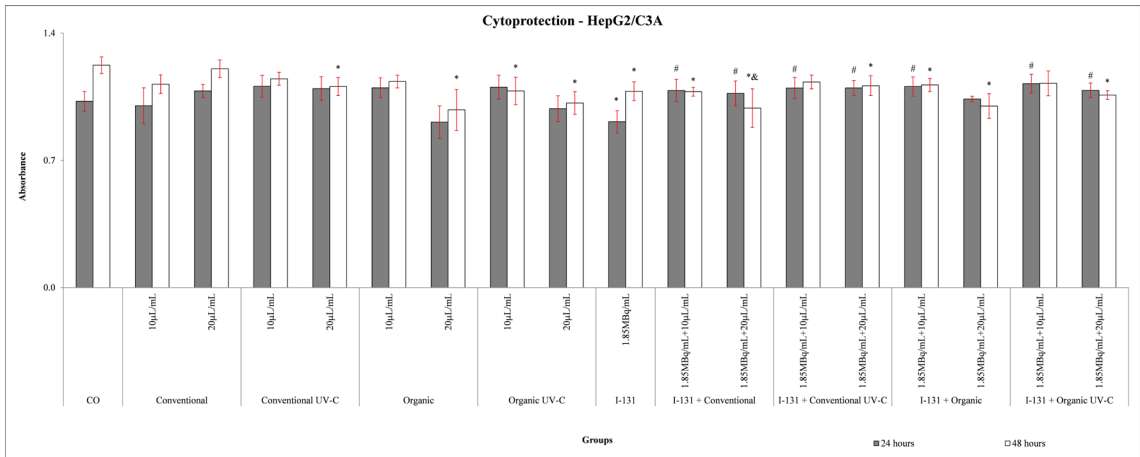


Figure 2. Mean absorbance and standard deviation of HepG2/C3A cells treated with conventional and organic grape juices, exposed or not to UV-C irradiation (10 and 20 µL/mL), treated alone or in cytoprotective tests with I-131 (1.85 MBq/mL). CO: Control; 1×10^4 cells per well, incubated for 24 and 48 hours, n = 3, Tukey test. *Statistically significant result compared to Control; #Statistically significant result compared to treatment with I-131; &Statistically significant result compared to treatment with the same juice concentration, without I-131.

Table 2. Percentage of viability of HepG2/C3A and MRC-5 cells treated with conventional and organic grape juices, exposed or not to UV-C irradiation and Iodine-131, separately and simultaneously, for 24 and 48 hours, by the MIT assay.

Groups	HepG2/C3A		MRC-5			
	CV %		CV %			
	24 h	48 h	24 h	48 h		
Grape Juice	CO		100.0	100.0	100.0	100.0
	Conventional	10 µL/mL	97.6	91.4	101.1	97.2
		20 µL/mL	105.6	98.4	97.7	99.7
	UV-C Conventional	10 µL/mL	108.0	93.8	103.4	116.6
		20 µL/mL	106.8	90.3	101.5	108.6
	Organic	10 µL/mL	107.2	92.6	100.5	105.0
		20 µL/mL	88.8	80.0	97.3	104.4
	UV-C Organic	10 µL/mL	107.6	88.4	100.2	114.3
		20 µL/mL	96.0	83.0	97.3	109.9
	I-131	1.85 MBq/mL	89.0	88.2	71.8	82.9
Cytoprotection: I-131 1.85 MBq/mL + Grape Juice	Conventional	10 µL/mL	105.8	88.0	96.4	94.4
		20 µL/mL	104.2	80.7	87.5	90.9
	UV-C Conventional	10 µL/mL	107.1	92.4	104.4	101.5
		20 µL/mL	107.2	90.7	97.2	94.2
	Organic	10 µL/mL	107.9	91.1	94.6	91.1
		20 µL/mL	101.2	81.6	93.6	99.4
	UV-C Organic	10 µL/mL	109.5	91.8	95.4	93.1
		20 µL/mL	105.8	86.5	90.6	91.7

CV = Cell viability. Groups: CO = Control; conventional and organic grape juices, exposed or not to UV-C irradiation (10 and 20 µL/mL), treated alone or in the cytoprotective tests together with Iodine-131 (1.85 MBq/mL); 1×10^4 cells per well, incubated for 24 and 48 hours, n=3.

grape seed and peel extracts showed antitumor effect by cell cycle arrest, apoptosis induction and inhibition of cell proliferation. Thus, it is suggested the consumption

of grape juices before or after the therapeutic use of I-131, allowing the stimulation of tumor cell elimination by the therapeutic action of these juices. According to Painuli and

Kumar (2016), free radical scavenging is the most common mechanism of antitumor activity, and this antioxidant effect also plays an important role in protecting healthy tissues from the lethal effect of radiation exposure.

Studies have shown that the use of antioxidants during radiotherapy does not interfere with the beneficial effects of treatment, and their careful use can improve and reverse the adverse effects of radiotherapy, acting as radioprotective agents and reducing levels of oxidizing agents (Moss, 2007). These effects were observed in the present study since all simultaneous treatments with I-131 and grape juices (organic or conventional, exposed or not to UV-C), were not cytotoxic to MRC- 5 cells (Figure 3). Besides, all treatments performed with I-131 and juices together within 24 hours and the treatment with 10 µL/mL of UV-C conventional grape juice and I-131, within 48 hours, presented statistically higher mean absorbances compared to the treatment with the radioisotope I-131 alone. Furthermore, considering that I-131 was cytotoxic to MRC-5 cells at 24 and 48 hours, with cell viability less than 72% (24 hours) and 83% (48 hours) (Table 2), the data of the present study have shown the radioprotective effect of grape juices.

This effect may have happened because of grape juices are rich in antioxidant substances (Table 3), as shown in other studies with these fruits, their extracts or compounds isolated from them (Trindade et al., 2016), may have reduced the cytotoxicity caused by the radioisotope to MRC-5 cells, preventing free radical formation by I-131 (Alam et al., 2002; Amri et al., 2014) and induced-damage to cells (Andrade et al., 2011). This may have favored the DNA repair and the reconstitution of damaged cell membranes (Picada et al., 2003), and even protecting from radiation-induced apoptosis (Kim et al., 2018).

A similar result was found by Hosseinimehr and Hosseini (2014), who showed that resveratrol, an antioxidant present in grapes, increased I-131-induced cell death in thyroid cancer cells and protected human non-malignant fibroblast

cells (HFFF2) from the toxicity of this radioisotope, also by the MTT test.

Chaturvedi et al. (2021) showed that two fractions of *Cryptosporidium parvum* lysate showed radioprotective activity by normal cells irradiated with 10 Gy, improving cell viability and preventing radiation-induced DNA damage, which could be a candidate for the development of radioprotectant.

In the present study, the radioprotective activities were similar between the different types of grape juices tested (grapes from the organic or conventional production system, post-treated or not with UV-C), and the antioxidant contents of these juices did not differ from each other (Table 3). Data from Kalinova and Vrchotova (2011) and Margraf et al. (2016) corroborate those of the present study since they also did not observe an increase in bioactive compounds in organic foods when compared to conventional ones. Moreover, Cava and Sgroppo (2015) also did not identify the increased antioxidant activity of grape juices treated with UV-C.

So, these data stimulate in vivo studies about the consumption of grape juice as adjuvant in I-131 therapy for human hepatocarcinoma, considering the beneficial

Table 3. Mean and standard deviation of antioxidant activity (EC₅₀) of conventional and organic grape juices, with and without UV-C postharvest treatment.

Grape Juice	EC ₅₀ (mL juice/g DPPH)
Conventional	2,736.63 ± 510.42 a
UV-C Conventional	3,365.13 ± 241.81 a
Organic	2,802.94 ± 175.92 a
UV-C Organic	2,850.53 ± 207.55 a

Means followed by the same letter (a) do not differ statistically from each other at the 5% probability level by Tukey's test, n=3. EC₅₀ = Amount of sample required to reduce initial DPPH concentration by 50%.

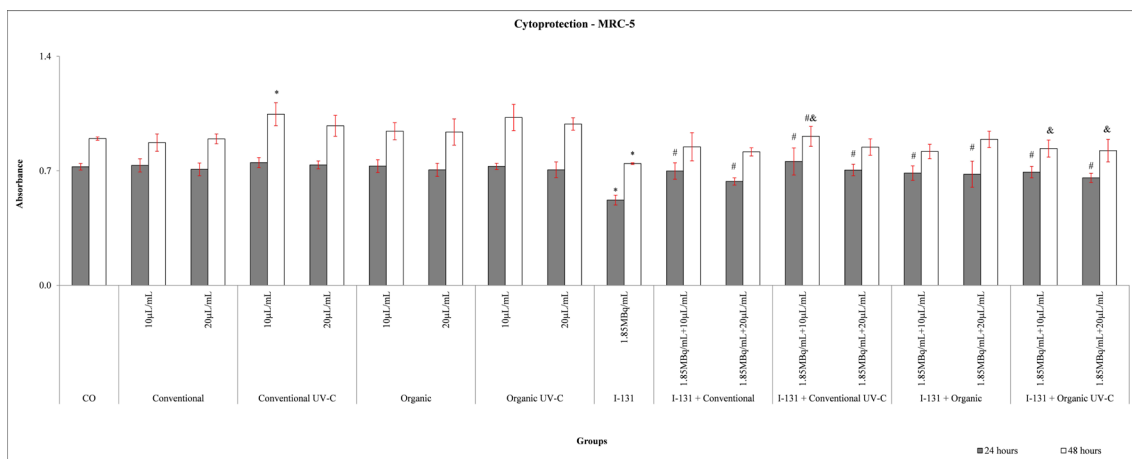


Figure 3. Mean absorbance and standard deviation of MRC-5 cells treated with conventional and organic grape juices, exposed or not to UV-C irradiation (10 and 20 µL/mL), treated alone or in cytoprotective tests with I-131 (1.85 MBq/mL). CO: Control; 1x10⁴ cells per well, incubated for 24 and 48 hours, n = 3, Tukey test. *Statistically significant result compared to Control; #Statistically significant result compared to treatment with I-131; &Statistically significant result compared to treatment with the same juice concentration, without I-131.

potential in vitro of this juice as antitumor, nutraceutical potential, antioxidant activity and radioprotective of stimulated metabolic cells against ionizing damage from radiation.

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References

- AHMADZADEHFAR, H., SABET, A., WILHELM, K., BIRSACK, H.J. and RISSE, J., 2011. Iodine-131-lipiodol therapy in hepatic tumours. *Methods*, vol. 55, no. 3, pp. 246-252. <http://dx.doi.org/10.1016/j.ymeth.2011.05.003>. PMID:21664971.
- ALAM, A., KHAN, N., SHARMA, S., SALEEM, M. and SULTANA, S., 2002. Chemopreventive effect of *Vitis vinifera* extract on 12-*o*-tetradecanoyl-13-phorbol acetate-induced cutaneous oxidative. *Pharmacological Research*, vol. 46, no. 6, pp. 557-564. <http://dx.doi.org/10.1016/S1043661802002268>. PMID:12457631.
- AMRI, A., CLANCHE, S., THÉRON, P., BONNEFONT-ROUSSELOT, D., BORDERIE, D., LAI-KUEN, R., CHAUMEIL, J.C., SFAR, S. and CHARRUEAU, C., 2014. Resveratrol self-emulsifying system increases the uptake by endothelial cells and improves protection against oxidative stress-mediated death. *European Journal of Pharmaceutics and Biopharmaceutics*, vol. 86, no. 3, pp. 418-426. <http://dx.doi.org/10.1016/j.ejpb.2013.10.015>. PMID:24184672.
- ANDRADE, E.R., CRUZ, I.B.M., ANDRADE, V.V.R., PICCOLI, J.C.E., GONZÁLEZ-GALLEGO, J., BARRIO, J.P. and GONZÁLEZ, P., 2011. Evaluation of the potential protective effects of *ad libitum* black grape juice against liver oxidative damage in whole-body acute X-irradiated rats. *Food and Chemical Toxicology*, vol. 49, no. 4, pp. 1026-1032. <http://dx.doi.org/10.1016/j.fct.2011.01.011>. PMID:21266186.
- APOSTOLOU, A., STAGOS, D., GALITSIOU, E., SPYROU, A., HAROUTOUNIAN, S., PORTESIS, N., TRIZOGLOU, I., HAYES, A.W., TSATSAKIS, A.M. and KOURETAS, D., 2013. Assessment of polyphenolic content, antioxidant activity, protection against ROS-induced DNA damage and anticancer activity of *Vitis vinifera* stem extracts. *Food and Chemical Toxicology*, vol. 61, pp. 60-68. <http://dx.doi.org/10.1016/j.fct.2013.01.029>. PMID:23380202.
- ASADIAN, S., MIRZAEI, H., KALANTARI, B.A., DAVARPANAH, M.R., MOHAMADI, M., SHPICHKA, A., NASEHI, L., ES, H.A., TIMASHEV, P., NAJIMI, M., GHEIBI, N., HASSAN, M. and VOSOUGH, M., 2020. β -radiating radionuclides in cancer treatment, novel insight into promising approach. *Pharmacological Research*, vol. 160, p. 105070. <http://dx.doi.org/10.1016/j.phrs.2020.105070>. PMID:32659429.
- BÉRON, A. and WÉMEAU, J.L., 2020. Iodine 131 in the treatment of large goiters. *Médecine Nucléaire*, vol. 44, no. 4, pp. 277-283. <http://dx.doi.org/10.1016/j.mednuc.2020.07.001>.
- BOMFIM, E.M.S., COELHO, A.A.O.P., SILVA, M.C., MARQUES, E.J. and VALE, V.L.C., 2022. Phytochemical composition and biological activities of extracts from ten species of the family Melastomataceae Juss. *Brazilian Journal of Biology = Revista Brasileira de Biologia*, vol. 82, p. e242112. <http://dx.doi.org/10.1590/1519-6984.242112>. PMID:34133563.
- BOUCHER, E., GARIN, E., GUYLLIGOMARC'H, A., OLIVIÉ, D., BOUDJEMA, K. and RAOUL, J.-L., 2007. Intra-arterial injection of iodine-131-labeled lipiodol for treatment of hepatocellular carcinoma. *Radiotherapy and Oncology*, vol. 82, no. 1, pp. 76-82. <http://dx.doi.org/10.1016/j.radonc.2006.11.001>. PMID:17141900.
- BRAND-WILLIAMS, W., CUVELIER, M.E. and BERSET, C., 1995. Use of a free radical method to evaluate antioxidant activity. *Food Science and Technology*, vol. 28, no. 1, pp. 25-30.
- CANTOS, E., ESPÍN, J.C. and TOMÁS-BARBERÁN, F.A., 2001. Postharvest induction modeling method using UV irradiation pulses for obtaining resveratrol-enriched table grapes: a new "functional" fruit? *Journal of Agricultural and Food Chemistry*, vol. 49, no. 10, pp. 5052-5058. <http://dx.doi.org/10.1021/jf010366a>. PMID:11600065.
- CAVA, E.L.M. and SGROPPO, S.C., 2015. Evolution during refrigerated storage of bioactive compounds and quality characteristics of grapefruit [*Citrus paradisi* (Macf.)] juice treated with UV-C light. *Lebensmittel-Wissenschaft + Technologie*, vol. 63, no. 2, pp. 1325-1333. <http://dx.doi.org/10.1016/j.lwt.2015.04.013>.
- CHATURVEDI, P.K., ERDENETUYA, E., PRABAKARAN, D.S., WOO, C.G., KIM, K.H., YU, J.R. and PARK, W.Y., 2021. Radioprotective effects of *Cryptosporidium parvum* lysates on normal cells. *International Journal of Biological Macromolecules*, vol. 178, pp. 121-135. <http://dx.doi.org/10.1016/j.ijbiomac.2021.02.151>. PMID:33636272.
- COSTA, P.L.A. and SAPIENZA, M.T., 2012. Terapia de hepatocarcinoma e outras lesões hepáticas. In: F. HIRONAKA, M.T. SAPIENZA, C.R. ONO, C. BUCHPIGUEL and M.S. LIMA, orgs. *Medicina nuclear: princípios e aplicações*. São Paulo: Atheneu, vol. 1, pp. 473-476.
- DEVI, S. and SINGH, R., 2017. Evaluation of antioxidant and anti-hypercholesterolemic potential of *Vitis vinifera* leaves. *Food Science and Human Wellness*, vol. 6, no. 3, pp. 131-136. <http://dx.doi.org/10.1016/j.fshw.2017.07.002>.
- DOWLATH, M.J.H., KARUPPANNAN, S.K., SINHA, P., DOWLATH, N.S., ARUNACHALAM, K.D., RAVINDRAN, B., CHANG, S.W., NGUYEN-TRI, P. and NGUYEN, D.D., 2021. Effects of radiation and role of plants in radioprotection: a critical review. *The Science of the Total Environment*, vol. 779, p. 146431. <http://dx.doi.org/10.1016/j.scitotenv.2021.146431>. PMID:34030282.
- DÜSMAN, E., ALMEIDA, I.V., LUCCHETTA, L. and VICENTINI, V.E.P., 2014. Effect of processing, post-harvest irradiation, and production system on the cytotoxicity and mutagenicity of *Vitis labrusca* L. juices in HTC cells. *PLoS One*, vol. 9, no. 9, p. e107974. <http://dx.doi.org/10.1371/journal.pone.0107974>. PMID:25244067.
- EL-DIN, N.K.B., ALI, D.A. and ABOU-EL-MAGD, R.F., 2019. Grape seeds and skin induce tumor growth inhibition via G1-phase arrest and apoptosis in mice inoculated with Ehrlich ascites carcinoma. *Nutrition*, vol. 58, pp. 100-109. <http://dx.doi.org/10.1016/j.nut.2018.06.018>. PMID:30391688.
- FERNANDES, F., RAMALHOSA, E., PIRES, P., VERDIAL, J., VALENTÃO, P., ANDRADE, P., BENTO, A. and PEREIRA, J.Á., 2013. *Vitis vinifera* leaves towards bioactivity. *Industrial Crops and Products*, vol. 43, pp. 434-440. <http://dx.doi.org/10.1016/j.indcrop.2012.07.031>.
- FISCHER, N., SEO, E.J. and EFFERTH, T., 2018. Prevention from radiation damage by natural products. *Phytomedicine*, vol. 47, pp. 192-200. <http://dx.doi.org/10.1016/j.phymed.2017.11.005>. PMID:30166104.
- HAILAN, Y. and YASSIN, M., 2021. Secondary chronic myeloid leukemia following radioactive iodine (I131). *Hematology, Transfusion and Cell Therapy*, vol. 43, no. 3, p. S38. <http://dx.doi.org/10.1016/j.htct.2021.10.1024>.
- HOSSEINIMEHR, S.J. and HOSSEINI, S.A.H., 2014. Resveratrol sensitizes selectively thyroid cancer cell to 131-iodine toxicity.

- Journal of Toxicology*, vol. 2014, p. 839597. <http://dx.doi.org/10.1155/2014/839597>. PMID:25276125.
- HOSSEINIMEHR, S.J., 2010. Flavonoids and genomic instability induced by ionizing radiation. *Drug Discovery Today*, vol. 15, no. 21-22, pp. 907-918. <http://dx.doi.org/10.1016/j.drudis.2010.09.005>. PMID:20933097.
- HOSSEINIMEHR, S.J., SHAFAGHATI, N. and HEDAYATI, M., 2013. Genotoxicity induced by iodine-131 in human cultured lymphocytes. *Interdisciplinary Toxicology*, vol. 6, no. 2, pp. 74-76. <http://dx.doi.org/10.2478/intox-2013-0013>. PMID:24179432.
- HUANG, L.L., PAN, C., WANG, L., DING, L., GUO, K., WANG, H.Z., XU, A.M. and GAO, S., 2015. Protective effects of grape seed proanthocyanidins on cardiovascular remodeling in DOCA-salt hypertension rats. *The Journal of Nutritional Biochemistry*, vol. 26, no. 8, pp. 841-849. <http://dx.doi.org/10.1016/j.jnutbio.2015.03.007>. PMID:25937175.
- KALINOVA, J. and VRCHOTOVA, N., 2011. The influence of organic and conventional crop management, variety and year on the yield and flavonoid level in common buckwheat groats. *Food Chemistry*, vol. 127, no. 2, pp. 602-608. <http://dx.doi.org/10.1016/j.foodchem.2011.01.050>. PMID:23140706.
- KIM, H.M., KIM, S.H. and KANG, B.S., 2018. Radioprotective effects of delphinidin on normal human lung cells against proton beam exposure. *Nutrition Research and Practice*, vol. 12, no. 1, pp. 41-46. <http://dx.doi.org/10.4162/nrp.2018.12.1.41>. PMID:29399295.
- KIM, Y., CHOI, Y., HAM, H., JEONG, H.S. and LEE, J., 2013. Protective effects of oligomeric and polymeric procyanidin fractions from defatted grape seeds on tert-butyl hydroperoxide-induced oxidative damage in HeG2 cells. *Food Chemistry*, vol. 137, no. 1-4, pp. 136-141. <http://dx.doi.org/10.1016/j.foodchem.2012.10.006>. PMID:23200001.
- KITABATAKE, K., KAJI, T. and TSUKIMOTO, M., 2020. ATP and ADP enhance DNA damage repair in γ -irradiated BEAS-2B human bronchial epithelial cells through activation of P2X7 and P2Y12 receptors. *Toxicology and Applied Pharmacology*, vol. 407, p. 115240. <http://dx.doi.org/10.1016/j.taap.2020.115240>. PMID:32941855.
- KOOSHA, F., EYNALI, S., EYVAZZADEH, N. and KAMALABADI, M.A., 2021. The effect of iodine-131 beta-particles in combination with A-966492 and Topotecan on radio-sensitization of glioblastoma: an in-vitro study. *Applied Radiation and Isotopes*, vol. 177, p. 109904. <http://dx.doi.org/10.1016/j.apradiso.2021.109904>. PMID:34454340.
- KUMAR, G.E.N.H., MAURYA, D.K., VISWANATH, B. and BALAJI, M., 2021. Role of phytoconstituents and their mechanism in attenuation of radiation effects: an update. In: B. VISWANATH, ed. *Recent developments in applied microbiology and biochemistry*, vol. 2, pp. 55-76. Amsterdam: Elsevier. <http://dx.doi.org/10.1016/B978-0-12-821406-0.00007-2>.
- KUNWAR, A., ADHIKARY, B., JAYAKUMAR, S., BARIK, A., CHATTOPADHYAY, S., RAGHUKUMAR, S. and PRIYADARSINI, K.I., 2012. Melanin, a promising radioprotector: mechanisms of actions in a mice model. *Toxicology and Applied Pharmacology*, vol. 264, no. 2, pp. 202-211. <http://dx.doi.org/10.1016/j.taap.2012.08.002>. PMID:22968190.
- KURUBA, V. and GOLLAPALLI, P., 2018. Natural radioprotectors and their impact on cancer drug discovery. *Radiation Oncology Journal*, vol. 36, no. 4, pp. 265-275. <http://dx.doi.org/10.3857/roj.2018.00381>. PMID:30630265.
- LOPES, N.B., ALMEIDA, I.V., LUCHETTA, L., DÜSMAN, E. and VICENTINI, V.E.P., 2019. Cytotoxic effects of *Vitis labrusca* (fox grape) whole juices on human tumor and non-tumor cells, *in vitro*. *Genetics and Molecular Research*, vol. 18, no. 2, p. gmr18236. <http://dx.doi.org/10.4238/gmr18236>.
- MAESTRE, R., DOUGLASS, J.D., KODUKULA, S., MEDINA, I. and STORCH, J., 2013. Alterations in the intestinal assimilation of oxidized PUFAs are ameliorated by a polyphenol-rich grape seed extract in an in vitro model and Caco-2 cells. *The Journal of Nutrition*, vol. 143, no. 3, pp. 295-301. <http://dx.doi.org/10.3945/jn.112.160101>. PMID:23325921.
- MARGRAF, T., SANTOS, E.N.T., ANDRADE, E.F., VAN RUTH, S.M. and GRANATO, D., 2016. Effects of geographical origin, variety and farming system on the chemical markers and in vitro antioxidant capacity of Brazilian purple grape juices. *Food Research International*, vol. 82, pp. 145-155. <http://dx.doi.org/10.1016/j.foodres.2016.02.003>.
- MAURER, L.H., BERSCH, A.M., SANTOS, R.O., TRINDADE, S.C., COSTA, E.L., PERES, M.M., MALMANN, C.A., SCHNEIDER, M., BOCHI, V.C., SAUTTER, C.K. and EMANUELLI, T., 2017. Postharvest UV-C irradiation stimulates the non-enzymatic and enzymatic antioxidant system of 'Isabel' hybrid grapes (*Vitis labrusca* × *Vitis vinifera* L.). *Food Research International*, vol. 102, pp. 738-747. <http://dx.doi.org/10.1016/j.foodres.2017.09.053>. PMID:29196007.
- MOSS, R.W., 2007. Do antioxidants interfere with radiation therapy for cancer? *Integrative Cancer Therapies*, vol. 6, no. 3, pp. 281-292. <http://dx.doi.org/10.1177/1534735407305655>. PMID:17761641.
- MURAYAMA, D., YAMAMOTO, Y., MATSUI, A., YASUKAWA, M., TODA, S. and IWASAKI, H., 2021. A case of the accumulation of 131-iodine in the mammary gland after remnant ablation for papillary thyroid carcinoma on lactating period. *Radiology Case Reports*, vol. 16, no. 11, pp. 3442-3444. <http://dx.doi.org/10.1016/j.radcr.2021.08.035>. PMID:34527119.
- NOUAILHETAS, Y., ALMEIDA, C.E.B. and PESTANA, S., 2021 [viewed 20 February 2021]. *Apostila educativa: radiações ionizantes e a vida* [online]. Available from: <https://www.gov.br/cnen/pt-br/material-divulgacao-videos-imagens-publicacoes-publicacoes-1/radiacoesionizantes.pdf>
- PAINULI, S. and KUMAR, N., 2016. Prospects in the development of natural radioprotective therapeutics with anti-cancer properties from the plants of Uttarakhand region of India. *Journal of Ayurveda and Integrative Medicine*, vol. 7, no. 1, pp. 62-68. <http://dx.doi.org/10.1016/j.jaim.2015.09.001>. PMID:27240731.
- PICADA, J., KERN, A., RAMOS, A.L.L.P. and SAFFI, J., 2003. O estresse oxidativo e as defesas antioxidantes. In: J. SILVA, B. ERDTMAN and J.A.P. HENRIQUES, eds. *Genética toxicológica*. Porto Alegre: Alcance, pp. 251-268.
- PINTO, E.P., PERIN, E.C., SCHOTT, I.B., DÜSMAN, E., RODRIGUES, R.S., LUCCHETTA, L., MANFROI, V. and ROMBALDI, C.V., 2022. Phenolic compounds are dependent on cultivation conditions in face of UV-C radiation in 'Concord' grape juices (*Vitis labrusca*). *Lebensmittel-Wissenschaft + Technologie*, vol. 154, p. 112681. <http://dx.doi.org/10.1016/j.lwt.2021.112681>.
- RAHGOSHAI, S., MEHNATI, P., AGHAMIRI, M.R., BORUJEINI, M.H., BANAEI, A., TARIGHATNIA, A., NADER, N.D., KIAPOUR, M. and ABEDI-FIROUZJAH, R., 2021. Evaluating the radioprotective effect of Cimetidine, IMOD, and hybrid radioprotectors agents: an in-vitro study. *Applied Radiation and Isotopes*, vol. 174, p. 109760. <http://dx.doi.org/10.1016/j.apradiso.2021.109760>. PMID:33971548.
- RUFINO, M.S.M., ALVES, R.E., BRITO, E.S., MORAIS, S.M., SAMPAIO, C.G., PÉREZ-JIMÉNEZ, J. and SAURA-CALIXTO, F.D., 2007. Metodologia científica: determinação da atividade antioxidante total em frutas pela captura do radical livre DPPH. *Embrapa*, vol. 127, pp. 1-4. Comunicado técnico.

- SAPIENZA, M.T., ENDO, I.S., CAMPOS NETO, G.C., TAVARES, M.G.M. and MARONE, M.M.S., 2005. Tratamento do carcinoma diferenciado da tireoide com iodo-131: intervenções para aumentar a dose absorvida de radiação. *Arquivos Brasileiros de Endocrinologia & Metabologia*, vol. 49, no. 3, pp. 341-349. <http://dx.doi.org/10.1590/S0004-27302005000300004>. PMID:16543987.
- TRALL, J.H. and ZIESSMAN, H.Á., 2003. *Medicina nuclear*. 2ª ed. Rio de Janeiro: Guanabara Koogan, 423 p.
- TRINDADE, C., BORTOLINI, G.V., COSTA, B.S., ANGHINONI, J.C., GUECHEVA, T.N., ARIAS, X., CÉSIO, M.V., HEINZEN, H., MOURA, D.J., SAFFI, J., SALVADOR, M. and HENRIQUES, J.A.P., 2016. Antimutagenic and antioxidant properties of the aqueous extracts of organic and conventional grapevine *Vitis labrusca* cv. Isabella leaves in V79 cells. *Journal of Toxicology and Environmental Health*, vol. 79, no. 18, pp. 825-836. <http://dx.doi.org/10.1080/15287394.2016.1190675>. PMID:27587288.
- YUN, K.L. and WANG, Z.Y., 2017. Target/signalling pathways of natural plant-derived radioprotective agents from treatment to potential candidates: a reverse thought on anti-tumour drugs. *Biomedicine and Pharmacotherapy*, vol. 91, pp. 1122-1151. <http://dx.doi.org/10.1016/j.biopha.2017.05.001>. PMID:28531942.