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Updated and annotated checklist of recent mammals from Brazil

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Abstract: An updated and annotated checklist of mammals occurring in Brazil is presented. A total of 751 native species, distributed in 249 genera, 51 families and 11 orders were recorded to the country. The Brazilian mammalian fauna shows an elevated rate of endemism (30%; 223 species). Among the species evaluated by IUCN (668 species; 90%), a total of 80 (10.6% of total mammalian fauna) are Threatened, 28 (3.9%) are considered as Near Threatened, two species (0.3%) are presumable Extinct, 96 (12.8%) are considered with Deficient Data for conservation and 462 (61.6%) are considered as Least Concern. Fifteen new species were described since the last national compilation (published in 2017), which associated to new records to the country and synonimizations resulted in an increment of 30 species. Eight non-native species were introduced to the country, including the recently established Asiatic cervids *Rusa unicolor* (sambar) and *Axis axis* (chital). Seven native species (five primates and two hystricomorph rodents) have been translocated from their areas of natural occurrence to other areas inside the country.

Key words: biodiversity, conservation, Mammalia, Neotropical region, taxonomic list.

INTRODUCTION

Currently, more than 6,400 mammal species are recognized worldwide. The Neotropics is the most diverse biogeographic realm, comprising around 25% of the mammalian diversity (Burgin et al. 2018). Within the Neotropical realm, Brazil is the largest country, covering an area nearly half of South America and housing two of the *hotspots* for the world conservation of biodiversity (Cerrado and Atlantic Forest) (Myers et al. 2000), as well as a great part of the Amazon Forest. Some authors (Mittermeier et al. 1997, Costa et al. 2005) have considered Brazil as the country with the highest mammalian diversity in the world.

Considerable attempts have been made to dimension the Brazilian mammalian diversity (Fonseca et al. 1996, Vivo 1996, Costa et al. 2005, Reis et al. 2006, 2011, Paglia et al. 2012, Percequillo & Gregorin 2017) and each new list adds new species as the result of a crescent effort on field sampling and taxonomic revisions (Fonseca et al. 1996, Paglia et al. 2012). The last compilation of mammals from Brazil (Percequillo & Gregorin 2017) listed a total of 721 species (or 778 valid nominal taxa, including subspecies). This list was produced as part of a joint effort of the Brazilian Society of Zoology aiming to produce an online database of the Brazilian fauna. It brought an accretion of 20 species in relation to the previous list of mammals occurring in Brazil (Paglia et al. 2012) and represents a very useful reference for the diversity of the group in the country. However, several new taxa described posteriorly to Paglia et al. (2012) were omitted by Percequillo & Gregorin (2017) (e.g. Hrbek et al. 2014, Feijó et al. 2015, Moratelli &

Dias 2015, Christoff et al. 2016, Pavan et al. 2017). Moreover, some taxonomic revisions resulting in nomenclatural changes conducted posteriorly to Paglia et al. (2012) annotated list (e.g. Berta & Churchill 2012, Feijó & Cordeiro-Estrela 2016, Nascimento & Feijó 2017) were not considered by Percequillo & Gregorin (2017). Still, progress in alpha-taxonomy has already been made after the compilation of Percequillo & Gregorin (2017) (e.g. Percequillo et al. 2017, Boubli et al. 2018, Garbino et al. 2019, Miranda et al. 2018, Feijó et al. 2018, Serrano-Villavicencio et al. 2019, Pavan 2019, Feijó et al. 2019). These three factors drew attention to the need for a reassessment of the mammalian diversity occurring in Brazilian territory.

Information on alpha-taxonomy is crucial in this current period of accelerated loss of biodiversity (Grieneisen et al. 2014). In this view, lists of species are of great importance for knowledge on regional diversity and the development of conservation plans. Herein, we compile all the information available about the valid mammalian species occurring in Brazil to the present date, aiming to produce an updated and annotated list for the country. This new compilation presents all new taxa described since the publication of Paglia et al. (2012) and Percequillo & Gregorin's (2017) lists as well as taxonomic modifications occurred since then, accompanied by justifications based on recent findings. We also included available and unpublished data on the status of exotic and invasive mammalian species in Brazil.

MATERIALS AND METHODS

The present annotated checklist was produced based on the two previous published lists of Brazilian mammals (Paglia et al. 2012, Percequillo & Gregorin 2017). An extensive research on all information (scientific articles, books and book chapters) published from 2012 to August 2019 was made on platforms Google, Google Scholar, Scopus and Web of Science, using as search terms the name of each genus and family presented on each previous lists (Paglia et al. 2012, Percequillo & Gregorin 2017) associated to "Brazil" and "New" (examples: "Didelphidae Brazil", "new Didelphidae", "*Monodelphis* Brazil", "new *Monodelphis*"). The search terms "Sigmodontinae Brazil", "new Sigmodontinae", "new genus mammal Brazil" and "new genus Mammalia Brazil" were also applied. The contents of each genus were also compared with the Mammal Diversity Database (www.mammaldiversity.org) (ASM 2019). We also checked the Brazilian institutional platform of academic productivity (Plataforma Lattes, accessed on: lattes.cnpq.br) for the latest scientific articles published by the main mammalian taxonomists in activity in Brazil. All the new taxa described after the publication of the latest list (Percequillo & Gregorin 2017) were added to this new list together with information on type locality and distribution. The taxonomic modifications as well as new evidences for taxa revalidation or refutation occurred after the publication of Percequillo & Gregorin's (2017) list were included as 'Remarks' containing justifications based on the respective researches. We have also included as 'Remarks' observations on forms with unstable taxonomy investigated in the last two years, even when in nomenclatural concordance with Percequillo & Gregorin (2017). Information on status of conservation of all species was given based on the latest evaluation of the International Union for the Conservation of Nature (IUCN) and on the Brazilian Red List (ICMBio 2018). The species endemic to the country were also indicated.

Finally, we present a section including the list of all allochthonous species so far recorded

living in the wild in natural environments in Brazil, as well as the native displaced species. That information was obtained by published data (articles, books, and book chapters), unpublished data (doctoral thesis, master's dissertations, meeting communications) and personal communications of researchers. We included only wild non-native species, excluding feral and free-roaming domestic species (*Canis familiaris*, *Felis catus*, *Bos* sp., *Equus* sp., *Capra* sp.). To know more about these domestic species, see Rosa et al. (2017a).

RESULTS

A total of 751 autochthonous mammalian species, distributed in 249 genera, 51 families and 11 orders were assigned to Brazil. The most speciose order is Rodentia with 258 species, followed by Chiroptera (182 spp.), Primates (126 spp.), Didelphimorphia (62 spp.), Artiodactyla (57 spp.), Carnivora (36 spp.), Cingulata (12 spp.), Pilosa (12 spp.), Lagomorpha (2 spp.), Perissodactyla (2 spp.), and Sirenia (2 spp.). A total of 80 species (10.6%) are included in some global threat category (12 Critically Endangered, 31 Endangered and 37 Vulnerable), 28 (3.7%) are considered as Near Threatened, two species (0.3%) are presumable Extinct, 96 (12.8%) are considered with Deficient Data for conservation and 462 (61.6%) are including in Least Concern (non-threatened) category (IUCN 2019). Eighty-three species (11%) are pending (re)evaluation due to recent taxonomic rearrangements or new species descriptions. According to the Brazilian red list (ICMBio 2018), 110 taxa (15%) are listed under some threat category (10 species are Critically Endangered, 41 Endangered, and 52 Vulnerable). A total of 223 species (30%) are endemic to Brazil (Appendix).

REMARKS

Order Didelphimorphia

- 1. *Gracilinanus peruanus* (Tate, 1931) was resurrected from the synonym of *G. agilis* (originally *Marmosa gracilis peruana* Tate, 1931) based on phylogenetic analyses of cytochrome b (Cyt*b*) sequences and external and craniodental morphology (Semedo et al. 2015). The occurrence of this taxon in Brazil was assigned to the states of Rondônia and Mato Grosso (Semedo et al. 2015).
- 2. *Marmosa (Micoureus) phaea* Thomas, 1899 was recorded for the right bank of Rio Tiquié, Comunidade Colina, Amazonas state (0.12° N, 69.01° W) (Voss et al. 2020).
- 3. Voss et al. (2020) based on molecular and morphological evidence resurrected *Marmosa (Micoureus) rapposa* Thomas, 1899 (type locality: Huadquiña, Cuzco, Peru). This species was previously treated a junior synonym or subspecies of *Marmosa (Micoureus) regina* Thomas, 1898. In Brazil, *M. rapposa* is recorded in Mato Grosso and Mato Grosso do Sul states (Voss et al. 2020).
- 4. *Marmosa (Micoureus) rutteri* Thomas, 1924 (type locality: "Tushemo, near Masisea", Ucayali, Peru) is recognized as a valid species by Voss et al. (2020) apart from *Marmosa (Micoureus) regina* Thomas, 1898. *Marmosa (M.) rutteri* is distributed in the lowland Amazon forest of southeastern Colombia, eastern Ecuador, eastern Peru and western Brazil. Records in Brazil include Acre and Amazonas states.
- 5. In the subgeneric classification of *Marmosa* of Voss et al. (2014a), *Marmosa lepida* Thomas, 1888 is placed within *Stegomarmosa* Pine, 1972.

- 6. Díaz-Nieto et al. (2016) performed phylogenetic analysis on Cyt*b* sequences of all recognized *Marmosops* species and found an internal dichotomy, leading to a subgeneric classification.
- 7. *Marmosops impavidus* (Tschudi, 1845) was considered a *nomen dubium* by Díaz-Nieto et al. (2016: 931) and therefore it was not included herein.
- 8. Voss et al. (2019) recognized *Metachirus myosuros* (Temminck, 1824) (type locality: Ipanema, São Paulo, Brazil) as a species apart from *M. nudicaudatus* based on cranial traits and phylogenetic analyses of the Cyt*b*. *Metachirus nudicaudatus* is now restricted to the Guiana Shield region with confirmed records from Amapá state in Brazil. Voss et al. (2019) reported specimens of *M. myosuros* from the Brazilian states of Acre, Amazonas, Bahia, Espírito Santo, Minas Gerais, Rio de Janeiro, São Paulo, and Rondônia.
- 9. Pavan et al. (2016), analyzing sequences of Cyt*b* and four nuclear genes, recovered monophyletic species groups within *Monodelphis*. Subsequently, Pavan & Voss (2016) formally organized such groups into subgenera, showing diagnostic morphological characters.
- 10. *Monodelphis (Microdelphys) gardneri* Solari, Pacheco, Vivar & Emmons, 2012 was described from specimens collected in the montane forests of central Peru and there is no confirmed record for Brazil. The species is not included in the present list.
- 11. *Monodelphis (Monodelphis) vossi* was described from three specimens collected in two savanna localities in Roraima state (Pavan 2019). Referred as *Monodelphis* "species 3" in Pavan et al. (2016) and Pavan & Voss (2016).
- 12. *Monodelphis (Mygalodelphys) handleyi* Solari, 2007 was reported from Brazil based on one individual collected in an Amazonian savanna area, Humaitá, Amazonas (Bezerra et al. 2019).
- 13. *Monodelphis (Mygalodelphys) saci* was described from 18 specimens collected in Brazilian Amazon (states of Pará, Mato Grosso, Rondônia and Acre) (Pavan et al. 2017). Referred as *Monodelphis* "species 2" in Pavan et al. (2016) and Pavan & Voss (2016).
- 14. Voss et al. (2018) revised *Philander* through mitochondrial and nuclear genes phylogeny and morphological analyses and considered eight valid species. According to these authors, *Philander canus* (Osgood, 1913) (type locality: Moyobamba, San Martín, Peru) is the name applicable to the form occurring in western Brazil (Cerrado biome).
- 15. According to Voss et al. (2018), *Philander quica* (Temminck, 1824) is the name applicable to the Brazilian Atlantic Forest form, considering that the holotype of *P. frenatus* (Olfers, 1818) was collected in eastern Amazonia. Voss et al. (2018) considered *P. frenatus* (Olfers, 1818) a junior synonym of *P. opossum* (Linnaeus, 1758).
- 16. *Philander pebas* was described from specimens collected on the left bank of the Rio Juruá, state of Amazonas. In Brazil, it is referred to Amazon Forest habitats of Amazonas and Acre states (Voss et al. 2018).

Order Sirenia

- 1. *Trichechus pygmaeus* Van Roosmalen et Van der Vlist, 2015 was described based on the holotype (CCM181) and an alive individual kept in semi-captivity.

According to Van Roosmalen (2015), *T. pygmaeus* differs from *T. inunguis* by overall smaller size, darker coloration, skull morphology, and fewer cheek teeth. In addition, Van Roosmalen (2015) obtained a fragment of 410 bp of the left domain of the mitochondrial control region (D-loop) from the living specimen and recovered a haplotype found by Garcia-Rodriguez et al. (1998) for *T. inunguis*. Still, Van Roosmalen (2015) interpreted this result as a possible consequence of the slow mutation rate of the control region in manatees, or even a past event of isolation of *T. pygmaeus* followed by the hybridization with *T. inunguis*. The specific status of *T. pygmaeus* was contested by authorities who claimed that specimens and individuals attributed to this taxon are actually juvenile *T. inunguis* (see Jefferson et al. 2015). Therefore, herein we did not consider *T. pygmaeus* as valid species.

Order Cingulata

- 1. Feijó et al. (2019) performed phylogenetic analyses using two mitochondrial markers (Cyt*b* and cytochrome oxidase subunit I [COI]) and one nuclear marker (exon 28 of the von Willebrand factor [vWF]) and recovered three clades within *Dasypus*, which were treated as subgenera (*Dasypus*, *Hyperoambon* and *Muletia*).
- 2. Feijó & Cordeiro-Estrela (2016) revised *Dasypus kappleri* using morphological and morphometric (linear and 2D geometric) analyses and considered it a species complex, which was further supported by phylogenetic analyses (Feijó et al. 2019). *Dasypus kappleri beniensis* Lönnberg, 1942 was elevated to full species. In Brazil, this taxon is known from the states of Pará, Rondônia, and Mato Grosso (Amazon biome).
- 3. Feijó & Cordeiro-Estrela (2016) restricted *Dasypus kappleri* Krauss, 1862 to the form occurring to the north of Amazon River (Guiana shield) in French Guiana, Suriname, Guyana, eastern Venezuela and Brazilian states of Amapá, Amazonas and Pará.
- 4. Feijó & Cordeiro-Estrela (2016) resurrected *Dasypus pastase* (Thomas, 1901) (type locality: "Sarayacu, upper Pastasa River", Pastaza, Ecuador) to the form occurring on the foothills of the eastern Andes in Peru, Ecuador, Colombia, Venezuela (south of the Orinoco River) and Brazilian state of Amazonas (south of Javari river).
- 5. Feijó et al. (2018) revised the genus *Dasypus* through qualitative morphological analysis and linear and 2D geometric morphometrics, recognizing *Dasypus hybridus* (Desmarest, 1804) as a subspecies of *Dasypus septemcinctus* Linnaeus, 1758. Molecular phylogeny and species delimitation analyses also supported this arrangement (Feijó et al. 2019).
- 6. Gibb et al. (2015) erected family Chlamyphoridae Bonaparte, 1850 to include subfamilies Euphractinae, Chlamyphorinae and Tolypeutinae, a clade highly divergent from *Dasypus* clade in the mitogenomic phylogenetic analysis performed by the authors. Dasypodidae was restricted to *Dasypus* genus.
- 7. The occurrence of *Cabassous chacoensis* Wetzel, 1980 in Brazil is uncertain. In its description, Wetzel (1980) referred to one alleged specimen from Brazil obtained from the Buenos Aires Zoo in 1904. Besides that, no other individual of this

species was ever recorded in the country. Therefore, considering there is no reliable evidence of this species in Brazil and following recent reviews (Wetzel et al. 2008, Hayssen 2014, Brandão et al. 2019), we did not consider *C. chacoensis* as part of the Brazilian mammalian fauna.

- 8. Feijó & Langguth (2013) treated *Cabassous squamicaudis* (Lund, 1845) (type locality: "Rio das Velhas Floddal, Lagoa Santa, Minas Gerais, Brazil") as full species apart from *Cabassous unicinctus* (Linnaeus, 1758) based on phenotypic differences, which has been corroborated by molecular studies (Schetino M.A.A., unpublished data).

Order Pilosa

- 1. Miranda et al. (2018) revised *Cyclopes* through morphological, morphometric and molecular data (Cyt*b*, COI and mitochondrial control region [CR]) and recognized seven valid species.
- 2. Miranda et al. (2018) designated a neotype for *C. didactylus* (Linnaeus, 1758) (FMNH 93175, type locality: Kayser-Gebergte Airstrip, Suriname) and cited the occurrence of this taxon to Suriname, Venezuela, French Guiana, Brazil and Trinidad and Tobago. In Brazil, *C. didactylus* occurs disjointly in Amazon (states of Amazonas, Pará, Maranhão and Piauí) and northeast Atlantic Forest (states of Rio Grande do Norte, Paraíba, Pernambuco and Alagoas).
- 3. *Cyclopes ida* Thomas, 1900 was originally described as a subspecies of *C. didactylus*. Miranda et al. (2018) elevated *ida* to species level and assigned its distribution to Ecuador, Colombia, Peru and Brazilian states of Acre and Amazonas.
- 4. *Cyclopes rufus* (type locality: "Porto Velho, Rondônia, Brazil") was described by Miranda et al. (2018) from specimens collected in the Brazilian state of Rondônia, between Madeira and Aripuanã rivers.
- 5. *Cyclopes thomasi* (type locality: "Porto Walter, Acre, Brazil") was described by Miranda et al. (2018) based on specimens from few localities in Peru and Brazilian states of Acre and Amazonas.
- 6. *Cyclopes xinguensis* (type locality: Vitória do Xingu, Pará, Brazil [Usina Belo Monte]) was described by Miranda et al. (2018) from specimens collected in the Brazilian states of Pará and Amazonas.

Order Primates

- 1. The status of *Cebuella niveiventris* Lönnberg, 1940 as a full species, herein adopted, is resultant from phylogenetic analysis of of *Cytb* sequences and reduced-representation genome sequencing (ddRADseq) (Boubli et al. 2018), which was further supported by molecular species delimitation analysis (Garbino et al. 2019).
- 2. *Mico munduruku* Costa-Araújo, Farias, & Hrbek, 2019 (type locality: Boca do Crepori community (0546'55'S, 5715'14'W), right margin of the mouth of the Crepori River, Itaituba municipality, Pará State, Brazil) was described from specimens collected in seven localities in the Amazon forest, Pará state, between Jamanxim, Novo, Tapajós and Cururú rivers (Costa-Araújo et al. 2019).
- 3. The capuchin monkeys are traditionally classified in two groups: gracile (or untufted) and robust (or tufted) species. Silva-Júnior (2002) treated these two groups as subgenera (*Cebus* for gracile

and *Sapajus* for robust) of *Cebus*. Later, Lynch Alfaro et al. (2012a, b, 2014) advocate for the use of *Cebus* and *Sapajus* as distinct genera, based on morphological, genetic, biogeographic, behavioral, and ecological differences. This new classification has provoked a heated debate (Rosenberger 2012, Feijó & Langguth 2013, Garbino 2015, Gutiérrez & Marinho-Filho 2017). Because there is no objective and universal criteria for defining genera, Garbino (2015) proposed the use of an integrative approach based on multiple sources of evidence to define genera. Herein, we considered *Cebus* Erxeleben, 1777 and *Sapajus* Kerr, 1792 as valid genera.

- 4. Boubli et al. (2008) described *Cacajao hosomi* (type locality: Imeri Mountains, Serra do Xamatá, Amazonas, Brazil) and *C. ayresi* (type locality: right bank of lower Rio Aracáy, Amazonas, Brazil) apart from *C. melanocephalus* (Humboldt, 1811) based on pelage coloration and divergences of *Cytb* sequences. Later, Ferrari et al. (2014) considered *C.ayresi* as a subspecies of *C. melanocephalus* and resurrected *C. ouakary* as a valid species. Ferrari et al. (2014) also refer the nominal subspecies *C. m. melanocephalus* as Neblina black-faced uacari, which is the designation given by Boubli et al. (2008) to *C. hosomi*. Surprisingly, Ferrari et al. (2014) did not mention *hosomi* along the full main text but it is implicit that *C. m. melanocephalus*, as established there, includes *hosomi*. Ferrari et al. (2014) also argued that pelage differentiation between *ayresi* and *melanocephalus* (including *hosomi*) represents a natural geographic gradient from darker to lighter patterns while the genetic distances between the two clades is far below than interspecific distances observed in other mammalian lineages, including other primate genera. Moreover, Ferrari et al. (2014) call attention to the utilization of a single mitochondrial gene by Boubli et al. (2008), as well as the lack of complementary evidences such as cranial or morphometric analyses. Another important question is the proper definition of *melanocephalus*. The holotype of *Cacajao melanocephalus* (Humboldt, 1812) is a pet with imprecise origin, but referred as "Exact locality unknown, in the region of the Mission of San Francisco Solano on the Canal Cassiquiare, Amazonas, Venezuela" (Boubli et al. 2008). Boubli et al. (2008) designated a neotype for *C. melanocephalus* (MN68616; type locality: Serraria, right bank of Rio Negro, Amazonas, Brazil). Surprisingly, Ferrari et al. (2014), disregarding Boubli et al. (2008) designation, selected another neotype for *C. melanocephalus* (NMNH 406425, type locality: Río Mavaca, 108 km SSE Esmeralda, Amazonas state, Venezuela), claiming that this specimen is closest to the Humboldt's holotype coloration and geographical origin. Nevertheless, this second designation has no validity (ICZN 1999, Article 75.4). The phylogenetic analysis of Boubli et al. (2008) revealed genetic distances (Tamura-Nei+G) between *ayresi* and *melanocephalus* (including sequences of neotype MN68616) ranging from 0.021 to 0.025 while distances between *hosomi* (= *C. m. melanocephalus* sensu Ferrari et al. [2014]) and *ayresi* were conspicuously lower, ranging from 0.003 to 0.006. The *hosomi-ayresi* range overlaps with the intraspecific distances found for both

hosomi (0.000-0.003) and *ayresi* (0.001-0.004) (Boubli et al. 2008). Furthermore, *hosomi* and *ayresi* are sister clades in the *Cytb* phylogenetic tree and the dichotomy between theses clades is overly shallower when compared to *melanocephalus* and *hosomi* + *ayresi* dichotomy (Fig. 2 in Boubli et al. 2008). The divergence between *hosomi* and *ayresi* Cyt*b* sequences is restricted to one C-T transition (Table II in Boubli et al. 2008). In addition, Figueiredo-Ready et al. (2013) found very low *Cytb* divergence between *ayresi* and *melanocephalus* (content not specified), ranging from 0.0004 to 0.007. Thus, to our understanding, (1) there is no genetic divergence that justifies the treatment of *hosomi* and *ayresi* as distinct species, and (2) the darker dorsal coloration of first in relation to the later may represent intraspecific geographic variation. Considering these issues, we recommend to treat *ayresi* Boubli et al. 2008, *hosomi* Boubli et al. 2008 and *melanocephalus* [sensu Boubli et al. (2008) and sensu Ferrari et al. (2014)] as *Cacajao melanocephalus* (Humboldt, 1812) until the light of a more integrative approach, including phylogenetic analyses using mtDNA and nuDNA markers associated to robust morphological analyses (e.g. craniometrics, cranial geometric morphometrics) of a broader sample (including specimens from Venezuela). In relation to *C. ouakary*, Figueiredo-Ready et al. (2013) found a considerable *Cytb* divergence between *ouakary* (including specimens from Solimões river) and *melanocephalus* (including *ayresi*) sequences (0.021-0.038). Thus, considering this evidence, it seems appropriate to treat *ouakary* and *melanocephalus* as distinct species. Further studies are still needed to clarify the taxonomy of *Cacajao melanocephalus* species complex.

- 5. Byrne et al. (2016) performed a phylogenetic analysis of *Callicebus* (sensu Hershkovitz 1988) using a dataset of 20 nuDNA and two mtDNA markers. The authors found deep structuring and proposed nomenclatural changes, erecting genera *Cheracebus* and *Plecturocebus* to allocate divergent clades composed of species formerly treated as *Callicebus*. Byrne et al. (2016) restricted *Callicebus* Thomas, 1903 to *personatus*, *coimbrai*, *barbarabrownae*, *melanochir* and *nigrifons*.
- 6. The genus *Plecturocebus* was erected by Byrne et al. (2016) to allocate a highly divergent clade of species formerly treated as *Callicebus* (see remark 5).
- 7. Byrne et al. (2016) considered *Callicebus dubius* Hershkovitz, 1988 a junior synonym of *Callicebus caligatus* (Wagner, 1842) (allocated in the new genus *Plecturocebus*) in view of the monophyletism of *caligatus* and *dubius* mtDNA sequences and their very low divergence of nuDNA and concatenated nuDNA and mtDNA sequences. The low divergence between *caligatus-dubius* is also corroborated by Hoyos et al. (2016) (*Cytb*) and Carneiro et al. (2016) (*Alu* elements, 16S, COI, and *Cytb*). Meanwhile, Serrano-Villavicencio et al. (2016) analyzed the geographic variation in the pelage color of *Callicebus* [*Plecturocebus* according to Byrne et al. (2016)] occurring between the Madeira and Purus rivers and considered *Callicebus caligatus* as a polytypic species with two subspecies: *Callicebus caligatus caligatus* and

Callicebus caligatus dubius. Therefore, herein *C. dubius* is not considered a full species.

- 8. *Plecturocebus grovesi* (Boubli et al. 2019) (type locality: the community of Novo Horizonte, left bank of the Rio Teles Pires, municipality of Alta Floresta, Mato Grosso state, Brazil) was described from specimens from a small area of Amazon Forest in northern Mato Grosso state.
- 9. *Plecturocebus parecis* Gusmão et al. 2019 (type locality: "the Rondon II hydroelectric dam on the middle Rio Comemoração, a tributary of the left margin of the Rio Ji-Paraná, municipality of Pimenta Bueno, Rondônia, Brazil") was described from specimens collected in western Mato Grosso and eastern Rondonia (Gusmão et al. 2019).
- 10. Byrne et al. (2016), considered *Plecturocebus stephennashi* Roosmalen, Roosmalen & Mittermeier, 2002 a valid species but without including it in their phylogenetic analysis. Serrano-Villavicencio et al. (2016) based on the pelage color considered *stephennashi* (as *Callicebus*) a hybrid form between *C. c. caligatus* and *C. c. dubius*. Moreover, both authors raise the question about the origin of *stephennashi* type series. Herein, we tentatively considered *P. stephennashi* as valid species, but we recommend additional efforts of new sampling and molecular analyses aiming to clarify the status of this taxon.
- 11. The genus *Cheracebus* was erected by Byrne et al. (2016) to allocate a highly divergent clade of species formerly treated as *Callicebus* (see remark 5).
- 12. Serrano-Villavicencio et al. (2019) analyzed the pelage color of *Pithecia irrorata* species group (sensu Marsh 2014) and considered *Pithecia mittermeieri* Marsh, 2014, *P. pissinattii* Marsh, 2014 and *P. rylandsi* Marsh, 2014 junior synonyms of *Pithecia irrorata* Gray, 1843, alleging lack of robust diagnoses and poorly defined geographic distributions.
- 13. *Pithecia vanzolinii* Hershkovitz, 1987 (type locality: "Santa Cruz, Rio Eirú, a east bank (southern) tributary of the Rio Juruá, Amazonas, Brazil.") was originally described as a subspecies of *Pithecia irrorata* Gray, 1843 and erected to full species by Marsh (2014) based on highly distinctive pelage characters. The specific status of *P. vanzolinii* was also corroborated by Serrano-Villavicencio et al. (2019), which analyzed the pelage color of *Pithecia irrorata* species group (sensu Marsh 2014).

Order Lagomorpha

- 1. *Sylvilagus tapetillus* Thomas, 1913 was recognized as a distinct species apart from *Sylvilagus brasiliensis* based on phenotypic, molecular and chromosomal traits (Bonvicino et al. 2015, Ruedas et al. 2017). The current geographic range of *S. tapetillus* is uncertain. Confirmed records are restricted to the Serra do Mar, Rio de Janeiro state (Bonvicino et al. 2015, Ruedas et al. 2017, Silva et al. 2019).

Order Rodentia

- 1. *Brucepattersonius nebulosus* Abreu-Júnior, Vilela, Christoff, Valiati & Percequillo, 2019 (type locality: Brazil, São Paulo, Bananal municipality, Estação Ecológica do Bananal) was described from specimens collected in Atlantic Forest localities in the states of São Paulo, Rio de Janeiro and Minas Gerais (Abreu-Júnior & Percequillo 2019).

- 2. *Calomys mattevii* Gurgel-Filho, Feijó & Langguth, 2015 (type locality: Fazenda Regalito, Rio Santa Maria, 20 km E of Flores de Goiás, Municipality Flores de Goiás, Goiás, Brazil) was considered a junior synonym of *Calomys expulsus* by Gutiérrez & Marinho-Filho (2017). However, phylogenetic analyses show that specimens of *C. mattevii* (karyotype 2n= 66/NF= 68) from the Cerrado and Caatinga areas of Goiás, Minas Gerais, Bahia, Piauí, Ceará and Pernambuco states form a distinct monophyletic clade apart from individuals with the same karyotype as *C. expulsus* (2n = 36/ NF= 66) (Gurgel-Filho et al. 2015, Campos B.A.T.P., unpublished data). Therefore, considering the chromosomal and phylogenetic evidences, we herein considered *C. mattevii* as valid species.
- 3. Pardinãs et al. (2016) erected the genus *Castoria* to allocate the fossil species *Habrothrix angustidens* Winge, 1887 and the living species *Akodon serrensis* Thomas, 1902, which were considered synonyms. The erection of the new genus was needed considering that Cyt*b* and IRBP sequences of *Akodon serrensis* fell out of *Akodon* clade within the Akodontini radiation (D'Elía 2003, D'Elía et al. 2003, Smith & Patton 2007, Ventura et al. 2013, Abreu et al. 2014, Pardiñas et al. 2014)
- 4. *Deltamys araucaria* (type locality [original]: São Francisco de Paula, Rio Grande do Sul State, Brazil) was described from specimens collected in grassland – Araucaria Forest mosaic (Mixed Ombrophilous Forest, Atlantic Forest biome) in Serra Geral highlands of northeastern Rio Grande do Sul state (Quintela et al. 2017). Pardiñas (2018) restricted the type locality of *D. araucaria* to "5 km by road W Centro de Pesquisas e Conservação da Natureza Pró-Mata/ PUCRS, Rio Grande do Sul State, Brazil".
- 5. *Hylaemys seuanezi* (Weksler, Geise & Cerqueira, 1999) (type locality: Fazenda União, Município Casimiro de Abreu, estado do Rio de Janeiro) was considered valid species in the list of Percequillo & Gregorin (2017). Meanwhile, this species has been previously recognized by Percequillo (2015) as a synonym of *Hylaeamys laticeps* (Lund, 1840) (type locality: "Rio das Velhas's Floddal", Lagoa Santa, Minas Gerais) without justifications. The type locality of *H. laticeps* (Lagoa Santa) is inserted in an area of Cerrado nearly to the transitional zone between this biome and the Atlantic Forest in Minas Gerais state and it is inhabited both by representatives of the rodent fauna typical from Cerrado (e.g. *Calomys*, *Cerradomys*, *Necromys*; Brennand et al. 2013) as well as Atlantic Forest forms (*Blarinomys*, *Delomys*, *Thaptomys*; Ávila-Pires 1960). Although the species limits of *Hylaeamys* from Brazilian Atlantic Forest is satisfactorily defined through qualitative morphology, craniometrics and karyology (Brennand et al. 2013, considering *seuanezi* as valid), there is a lack of an integrative analysis of the genus, including the use of molecular markers. In view of this, we considered *seuanezi* as valid in accordance with Brennand et al. (2013), but we recommend the application of molecular techniques aiming to better clarify the taxonomy of *Hylaemys* in Brazil.
- 6. *Juliomys ximenezi* (type locality: Parque Nacional de Aparados da Serra [Aparados da Serra National Park], municipality of Cambará do Sul, Rio Grande do

Sul State, Brazil) was described from specimens collected in Araucaria Forest (Mixed Ombrophilous Forest, Atlantic Forest biome) in Serra Geral highlands of northeastern Rio Grande do Sul state (Christoff et al. 2016). Only specimens from type locality are known.

- 7. Hurtado & Pacheco (2017) recognized *Neacomys amoenus* Thomas, 1903 as a full species, distinguished from *Neacomys spinosus* (Thomas, 1882) by morphological characters and Cyt*b* sequences. The authors also recognized two subspecies, *N. a. amoenus* Thomas, 1903 and *N. a. carceleni* Hershkovitz, 1940. *Neacomys spinosus* (Thomas, 1882) was restricted to mountain cloud forests of Peruvian Amazonia (Hurtado & Pacheco 2017).
- 8. *Neusticomys peruviensis* (Musser & Gardner, 1974) was recorded for Parque Nacional de Pacaás Novos, Rondônia state (Percequillo et al. 2017).
- 9. *Oecomys bicolor* (Tomes, 1860) was considered a species complex based on mitochondrial and nuclear sequences (Suárez-Villota et al. 2017). Four well-supported lineages were recovered within *Oecomys bicolor* species group. However, the sequence closest to the type locality of *O. bicolor* [sequence from Peru; type locality of *Oecomys bicolor* (Tomes, 1860): "Gualaquiza," Rio Gualaquiza, 885 m, Morona-Santiago, Ecuador (Carleton & Musser 2015)] comprised a clade apart from the Brazilian clades. The name *bicolor* was herein maintained for the Brazilian Amazon lineages of *O. bicolor* group until further analyses and formal description of such forms.
- 10. *Oecomys catherinae* Thomas, 1909 [type locality: "Joinville, Santa Catherina [Santa Catarina], S. Brazil" (Carleton & Musser 2015)] was considered a species complex based on phylogenetic analyses (Suárez-Villota et al. 2017), although no topotypical sequences were employed. One lineage occurs in southeastern Atlantic Forest, one in Cerrado and three in Amazon Forest (Suárez-Villota et al. 2017). The name *catherinae* was herein maintained but is highly recommended to perform further analyses including sequences from type locality and other localities in southern Atlantic Forest (Santa Catarina and Paraná states).
- 11. *Oecomys cleberi* Locks, 1981 [type locality: "Fazenda Agua Limpa, da Universidade de Brasília, Distrito Federal, Brasil (Carleton & Musser 2015)] was considered a species complex based on mitochondrial and nuclear sequences (Suárez-Villota et al. 2017), comprising two lineages. One lineage, which includes sequences from the holotype, is distributed in Cerrado of central Brazil. The other occurs in southern Amazon, including Mato Grosso state (Suárez-Villota et al. 2017).
- 12. *Oecomys franciscorum* was originally described from specimens collected in Argentinean provincies of Formosa and Chaco (type locality: Provincia de Formosa, Departamento de Formosa, Estación de Animales Silvestres Guaycolec, 0.4 km NW of the junction between Ruta Nacional 11 and Riacho Pilagá) (Pardiñas et al. 2016). *Cytb* sequences of specimens from Brazilian Pantanal (Mato Grosso do Sul state) and treated as *Oecomys* cf. *franciscorum* by Pardiñas et al. (2016) grouped with sequences from type specimens in a moderated-supported clade (BPP =

0.96). Following, Suárez-Villota et al. (2017) performed Bayesian phylogenetic analyses using those sequences plus additional specimens from Brazilian Pantanal (including a new locality, Parque Nacional do Pantanal, Mato Grosso state) and found similar relationships and very low K2P distances (0.0 – 1.1%), considering the clade (*O. franciscorum* – *O.* cf. *franciscorum*) a single species.

- 13. *Oecomys mamorae* (Thomas, 1906) [type locality: type locality "Mosetenes, Upper Mamoré, Yungas, [Cochabamba,] Bolivia." (Carleton & Musser 2015)] was considered a species complex based on mitochondrial and nuclear sequences, encompassing three lineages (Suárez-Villota et al. 2017). Sequences from Brazil (Pantanal biome, Mato Grosso state), however, were recovered apart from Bolivian lineage, comprising an exclusive clade. The name *mamorae* is herein maintained until further analyses.
- 14. *Oecomys paricola* (Thomas, 1904) [type locality: "Igarapé-Assu, near Pará, [Para, Brazil]. Alt. 50 m" (Carleton & Musser 2015)], was considered a species complex based on mitochondrial and nuclear sequences, comprising three lineages (Suárez-Villota et al. 2017). The three lineages are distributed in Brazilian territory, being two in Amazon forest of Para and Mato Grosso states and the other in Cerrado (Suárez-Villota et al. 2017).
- 15. *Oecomys roberti* (Thomas, 1904) [type locality: "Santa Ana de Chapada, a village situated at an altitude of about 800 m, on the Serra do Chapada, some thirty miles N.E. of Cuyabá [Cuiabá]," Mato Grosso, Brazil (Carleton & Musser 2015)] was revealed as a species complex based on mitochondrial and nuclear sequences, comprising three lineages all occurring in Brazil. One lineage is restricted to Western Amazon, one is widespread from central Amazon to western Cerrado and Pantanal, and the last is restricted to northern Cerrado (Suárez-Villota et al. 2017).
- 16. *Oxymycterus itapeby* (type locality: Brazil, São Paulo, Itapevi, Transurb district, "Condomínio Vila Verde") was described from specimens collected in Atlantic Forest-Cerrado transitional areas of São Paulo and Paraná states (Peçanha et al. 2019).
- 17. *Rhagomys longilingua* Luna & Patterson, 2003 was recorded to Brazilian Amazon in Rondônia state (Hydroelectric Dam Jirau) (Percequillo et al. 2017).
- 18. The type locality of *Ctenomys brasiliensis* (Blainville, 1826) is indicated as "St Paul, prov. Las Minas" and it was long thought to refer to the state of Minas Gerais, southeastern Brazil, a region apart from the distribution of *Ctenomys* genus. In view of this, Fernandes et al. (2012) analyzed the holotype of *C. brasiliensis* through qualitative characters and skull geometric morphometrics. Their results indicated that *C. brasiliensis* is closely related to *C. pearsoni* and *C. torquatus*, respectively distributed in Uruguay and Rio Grande do Sul state and Uruguay. Thus, it is very likely that "Minas" indicated in *C. brasiliensis* holotype label could be related to the municipality of Minas, the capital of Lavalleja department, Uruguay, a region within the distribution of *C. pearsoni*. Considering this evidences, we did not consider *C. brasiliensis* as occurring in Brazil.

- 19. *Agouti silvagarciae* Van Roosmalen et Van Hoof, 2015 was described from two specimens, one collected by hunters at the left bank of Aripuanã river, and the other vouchered in Museu Paraense Emílio Goeldi, without provenance. Acording to Van Roosmalen (2015), *A. silvagarciae* differs from *C. paca* by overall large size. The author also states that "One complete mitochondrial Dloop and two nuclear SINE PRE-1 DNA sequences of Silva Garcia's giant paca were carried out and compared with Genbank sequences of the sympatric common paca (*A. paca*). The results (15.5% difference between species) clearly support the distinction into valid species." However, the author makes no distinction about the distance found for each marker, and no reference to the methods used for phylogenetic inference, nodal supports, and the *C. paca* sequences used in the analysis was added. In view of the lack of such crucial information, we herein did not consider *A. silvagarciae* as valid.
- 20. *Dasyprocta aurea* Cope, 1889, *Dasyprocta catrinae* Thomas, 1917 and *Dasyprocta nigriclunis* Osgood, 1916 were considered by Iack-Ximenes (2019) as valid species. *Dasyprocta aurea* and *D. catrinae* were considered synonyms of *Dasyprocta azarae* Lichtenstein, 1823 while *D. nigriclunis* is referred as a synonym of *D. prymnolopha* Wagler, 1831 by Patton & Emmons (2015a).
- 21. *Phyllomys centralis* (type locality: Fazenda Água Limpa (15°57'4.42'S, 47°57'48.85'W), APA Gama Cabeça de Veado, Distrito Federal, Brazil, elevation of 1,100 m) was described from specimens collected in Cerrado of Distrito Federal state and Atlantic Forest-Cerrado transitional area in Minas Gerais state (Machado et al. 2018).
- 22. *Trinomys panema* (Moojen, 1948) was considered a synonym of *Trynomys gratiosus* (Moojen, 1948) by Patton & Emmons (2015b) and herein is not considered as full species.
- 23. *Coendou baturitensis* Feijó & Langguth, 2013 (type locality: Community Sitio Barreiros, municipality of Aratuba, Baturite Range, Ceará, Brazil) was considered a junior synonym of *Coendou prehenselis* by Voss (2015) and Gutiérrez & Marinho-Filho (2017). Recent morphology and Cyt*b* phylogenetic analyses revealed that individuals of *C. baturitensis* cluster apart from *C. prehenselis* and shows several diagnostic phenotypic traits (Menezes F.H., unpublished data). Therefore, we herein considered *C. baturitensis* a valid species.
- 24. *Coendou ichillus* Voss & da Silva, 2001 was reported in Brazil based on one individual collected in the margins of Rio Japurá, Limoeiro, Amazonas state (Menezes et al. 2020).
- 25. *Hadrosciurus ignitus* (Gray, 1867) was previously considered as a subspecies of *Notosciurus pucheranii* (Fitzinger, 1867). Abreu-Júnior et al. (2020), based on mitogenomic phylogeny, considered it as a full species and part of the genus *Hadrosciurus* Allen, 1915. In Brazil, *H. ignitus* is recorded in the states of Acre and Amazonas.

Order Chiroptera

- 1. *Eumops chimaera* (type locality: Parque Estadual do Rio Doce [PERD], municipality of Marliéria, state of Minas Gerais, Brazil) was described from specimens collected in the Atlantic Forest of Minas Gerais

state and the Bosque Chiquitano of Bolivia (Gregorin et al. 2016). *Eumops chimaera* diverges from other *Eumops* species by Cyt*b* sequences and external and craniodental morphology (Gregorin et al. 2016).

- 2. *Pteronotus* (*Phyllodia*) *alitonus* Pavan, Bobrowiec & Percequillo, 2018 was described from specimens from Suriname, French Guiana, and the Brazilian Amazon, previously referred as *Pteronotus* sp. 1 (Pavan & Marroig 2016), *Pteronotus* sp. 3 (Clare et al. 2013), *Pteronotus rubiginosus* and *P. parnellii* (see Pavan et al. 2018). *Pteronotus* (*Phyllodia*) *alitonus* differs from other *Pteronotus* species by morphological traits, echolocation calls and COI sequences (Pavan et al. 2018). *Pteronotus parnellii* Gray, 1843 (type locality: Jamaica) was excluded from the Brazilian fauna because none of the Brazilian specimens analyzed shared haplotypes with the Jamaican lineage (see Thoisy et al. 2014, López-Wilchis et al. 2016).
- 3. *Pteronotus* Gray, 1838 was organized in three subgenera (*Pteronotus*, *Phyllodia*, *Chilonycteris*) by Smith (1972) based on a robust craniometric dataset. However, molecular analyses performed by Pavan & Marroig (2016) using mitochondrial and nuclear markers (COI, Cyt*b*, *Dby*, RAG2, STAT5A) revealed *Chilonycteris* as an artificial group and suggested the erection of a new subgenus for *Pteronotus personatus* (Wagner, 1843). Since there is still no available name, we kept a binomial classification for *P. personatus*.
- 4. *Chiroderma salvini* Dobson, 1878 was recorded for Brazil by Rocha et al. (2016) based on two specimens from Rondônia and Mato Grosso states. However, analysing a large sample of *Chiroderma,* Brandão et al. (2019) argued that both specimens reported by Rocha et al. (2016) are actually *C. villosum*. Therefore, we did not include *C. salvini* as part of the Brazilian fauna.
- 5. Garbino et al. (in press) revised the genus *Chiroderma* using morphological and molecular datasets and recognized *Chiroterma vizottoi* Taddei & Lim, 2010 as a subspecies of *Chiroderma doriae* Thomas, 1891. Therefore, *C. vizottoi* is not included in the present list.
- 6. The genus *Gardnerycteris* Hurtado & Pacheco, 2014 was erected to allocate two species previously included in genus *Mimon*, which was revealed as polyphyletic from a phylogenetic analysis of 91 morphological characters (Hurtado & Pacheco 2014). Later, the monophyly of *Gardnerycteris* and its contents were sustained by molecular data (Cyt*b*, COI, RAG2) (Hurtado & D'Elía 2018). *Gardnerycteris* currently comprises three valid species (*G. koepckeae* [type species], *G. crenulatum* and *G. keenani*) and *G. crenulatum* is the single species occurring in Brazil (Hurtado & D'Elía 2018).
- 7. *Hsunycteris pattoni* (Woodman & Timm, 2006), originally described as *Lonchophylla pattoni* (type locality: Reserva Cusco Amazónico, north bank of the Río Madre de Dios; 14 km east of Puerto Maldonado; Tambopata Province; Madre de Dios Department; southeastern Peru), was assigned to Brazilian Amazon in the states of Amazonas and Pará (Velazco et al. 2017).
- 8. *Lonchophylla inexpectata* Moratelli & Dias, 2015 (type locality: Barra, Bahia, Brazil) was described from specimens previously identified as *Lonchophylla*

mordax Thomas, 1903. *Lonchophylla inexpectata* is distinguishable from other *Lonchophylla* species by the fur color, cranial size and dental morphological (Moratelli & Dias 2015).

- 9. *Micronycteris brosseti* Simmons & Voss, 1998 (type locality: Paracou, French Guiana) was not considered in the present list. The referred material of *M. brosseti* from Brazil includes only one specimen from "Rio Juquiá, Barra, São Paulo", deposited in the Field Museum of Natural History, Chicago (FMNH 92997) (Simmons & Voss 1998). Garbino (2016) disagrees with this identification, taking into account a personal communication from Ricardo Moratelli, who alleged that FMNH 92997 could not be conclusively identified as *M. brosseti*. Garbino (2016) also called attention to the over 3,000 km gap between the São Paulo record and the type locality of *M. brosseti*. Considering these issues, we opted for the exclusion of *M. brosseti* until further clarification.
- 10. *Sturnira giannae* Velazco & Patterson, 2019 (type locality: Paracou (5°17'N, 53°55'W, 210 m), near Sinnamary, Cayenne, French Guiana) was described from specimens collected in Guianas, Venezuela, Ecuador, Peru, Bolívia and the Brazilian states of Amazonas and Pará (Velazco & Patterson 2019).
- 11. *Tonatia maresi* Williams, Willig & Reid, 1995 (type locality: Blanchisseuse, Trinidad and Tobago) was originally described as a subspecies of *Tonatia saurophila* Koopman & Williams, 1951. Basantes et al. (2020) elevated *maresi* and *T. saurophila bakeri* Williams, Willig & Reid, 1995 to species level based on morphological and genetic (Cytb and nuclear exon RAG2) differences. *Tonatia* currently comprises three valid species (*T. saurophila* [considered extinct and restricted to Jamaica], *T. bakeri*, and *T. maresi*), and only *T. maresi* is recorded in Brazil (Basantes et al. 2020).
- 12. *Histiotus diaphanopterus* Feijó, Rocha & Althoff, 2015 (type locality: Boqueirão do Onça, village of São Pedro do Lago, municipality of Sento Sé, state of Bahia, Brazil) was described from specimens collected in Caatinga and Cerrado of Brazil (Bahia, Ceará, Paraíba and Maranhão states) and Bosque Chiquitano of Bolivia.

Order Carnivora

- 1. The name *Conepatus semistriatus* (Boddaert, 1785) has been historically applied for the form occurring in Cerrado and Caatinga of Brazil (e.g. Cavalcanti et al. 2013, 2014, Dias 2017, Tomas et al. 2017). However, phylogenetic analyses using mtDNA markers (*Cyt*b, COI, CR) recovered sequences from a specimen from Mexico (type locality of *C. semistriatus*) in a clade apart from sequences of specimens from South America, including individuals from Brazilian Cerrado-Caatinga (states of Goiás, Minas Gerais and Piauí) (Schiaffini et al. 2013). This aspect was observed by Feijó & Langguth (2013), who stated that *C. amazonicus* (Lichtenstein, 1838) is the name applicable to *Conepatus* from Cerrado and Caatinga in Brazil. Herein we follow this new classification.
- 2. Kitchener et al. (2017) considered the jaguarundi as belonging to genus *Herpailurus* Severtzov, 1858, only stating "As used here, this is a monotypic genus, but it may be included within *Puma*". Herein, *Herpailurus yagouaroundi* is used instead of *Puma yagouaroundi*.

- 3. Nascimento et al. (2020), using an integrative taxonomic approach, recognized five species of pampas cats (*Leopardus braccatus*, *L. colocola*, *L. garleppi*, *L. munoai*, and *L. pajeros*). These five species show distinct skull and skin traits, inhabit distinct climatic niches and are supported by phylogeny and molecular species delimitation. In Brazil, two species of pampas cats are present. *Leopardus braccatus* (Cope, 1889) (type locality: Chapada dos Guimarães, Mato Grosso, Brazil) is distributed in central Brazil (from Cerrado of south-western Piauí to Pantanal of Mato Grosso do Sul), Paraguay, and open areas of Bolivia and northern Argentina.
- 4. *Leopardus emiliae* (Thomas, 1914) (type locality: "Ipu, Ceará, N.E. Brazil. Alt. 300 m") has long been considered a junior synonym of *Leopardus tigrinus* (Wozencraft 2005). Nascimento & Feijó (2017) resurrected *L. emiliae* as a full species, distinguishable from *L. tigrinus* (Schreber, 1775) and *L. guttulus* (Hensel, 1872) by pelage color and cranial differences.
- 5. *Leopardus munoai* (Ximénez, 1961) (type locality: 'Arroyo Perdido, Departamento de Soriano', Uruguay) is considered as a valid species by Nascimento et al. (2020) (see remark 3). It occurs in the pampas of Rio Grande do Sul Brazilian state, Uruguay and northeastern Argentina (Nascimento et al. 2020).

Order Perissodactyla

- 1.*Tapirus kabomani* was described from specimens collected in Rondônia state, southern Brazilian Amazonia (Cozzuol et al. 2013). The authors concluded that the new tapir differs from *T. terrestris* by external and cranial external morphology and molecular characters (Cyt*b*, COI, COII). Meanwhile, Voss et al. (2014b) discussed the validity of *T. kabomani* claiming the very low molecular divergence from *T. terrestris* (1.3%) and the inconsistence between molecular and morphological characters, among other aspects. Herein we tentatively considered *T. kabomani* as a valid species, but in agreement with Voss et al. (2014b), we believe that further molecular analysis including nuclear markers could provide a better view on *T. kabomani* taxonomic status.

Order Artiodactyla

- 1. The name Cetartiodactyla Montgelard, Catzeflis & Douzery, 1997 was proposed as a "solution" for the paraphyletic condition of Artiodactyla in relation to Cetacea Brisson, 1762. The International Code of Zoological Nomenclature (ICZN) does not regulate order names, and considering that Cetacea is a clade within Artiodactyla (Montgelard et al. 1997), there is no restriction for the usage of Order Artiodactyla as a conservative name. Therefore, herein we adopted Order Artiodactyla.
- 2. *Mazama tienhoveni* van Roosmalen & van Hooft, 2015 was described from two skins, one skull and mandible, and one spike in possession of hunters from Tucunaré village, lower Aripuanã River, Central Amazon. In our point of view, the new species presents some inconsistencies, including: 1) The holotype is poorly defined. According to the author the "holotypus" MR204 consists of "complete head with partly damaged mandible, adult female, on May 12, 2006 killed for food by a local hunter along the

left bank of the Rio Aripuanã near the settlement of Tucunaré, skull, spike and skin". The author mentions "two skins" in "Examined material" section but also mentions that "No paratypes have been collected thus far" in "Variability" section. In "Remarks" section, Van Roosmalen (2015) also included: "The males of *M. tienhoveni* n. sp. do not have the distinct crest of hairs on the forehead as *M. nemorivaga* has, neither do the males of *Mazama americana*". These information raised the following questions: Does this material, assigned as "holotypus MR204" comprise more than one individual? This doubt arose taking into account the references of: a) two skins; b) a spike as part of the material designated as an "adult female". 2) Van Roosmalen (2015) performed a dated linearized minimum-evolution tree using a very small fragment consisted of 233 bp of *Cytb*. We believe that the small fragment used for phylogenetic inferences is little informative. The resultant phylogenetic tree, in turn, showed a basal polytomy, with unsolved relationships (*Mazama* genus was recovered as polyphyletic) and undetermined or weak supports of clades. In summary, the molecular dataset used by Roosmalen is insufficient for a phylogenetic inference within *Mazama* and the support of a specific-level divergent form. 3) Van Roosmalen (2015) made comparisons between *M. tienhoveni* and the other Amazonic representatives of *Mazama* genus, *M. americana* and *M. nemorivaga*. Those comparisons were based on pelage coloration, morphometrics (spike size, 16 cranial measurements, tail length, ear length, hindfoot length, head-body length, cranial) and body weight. However, in our understanding, the material comprised by two skins, a skull and mandible and one spike is insufficient for a solid comparison of high variable characters such as body and skull measurements and pelage color. Furthermore, head-body length and body mass were not directly obtained but were assumed to be intermediate between *M. americana* and *M. nemorivaga*, based on hunter's information. Craniometric dataset of other species were also very limited, comprising 11 specimens of *M. nemorivaga* and five specimens of *M. americana*, without any distinction on age classes or sex. Moreover, only means of *M. americana* and *M. nemorivaga* cranial measurements were presented while ranges were not disclosed. Considering these aspects, we herein did not consider *Mazama tienhoveni* a valid species due to the lack of genetic and morphological support.

- 3. *Tursiops gephyreus* Lahille, 1908 was revalidated for *Tursiops* forms occurring in estuaries of Brazilian states of Paraná, Santa Catarina and Rio Grande do Sul state, Uruguay and Argentina), based on morphometrics and qualitative morphological characters (Wickert et al. 2016). According to these authors, *Tursiops truncatus* (Montagu, 1821) also occurs in Brazilian coast, along all coastline, but more associated with open waters. Meanwhile, posterior molecular analyses (*CR*, nuclear microsatellites) revealed few support for the designation of two species of bottlenose dolphins occurring in Brazilian coast (Oliveira et al. 2019). First, "*truncatus* haplogroup" and "*gephyreus* haplogroup" are separated by a single mutational step (see Fig. 2 in

Oliveira et al. 2019). Second, there was a lack of complete structuring between the genotypes of the morphotypes attributed to *gyphereus* and *truncatus* (Oliveira et al. 2019). In view of this, Oliveira et al. (2019) recommended further integrative analyses on a broader geographical sample for a formal decision on the taxonomic status of *T. gephyreus*. In agreement, we considered only *T. truncatus* as valid.

- 4. *Inia araguaiaensis* Hrbek, Farias, Dutra & da Silva, 2014 (type locality: near the entrance of Lake Jurumirim, Araguaia River, state of Goiás) was described from three specimen collected in Araguaia River, state of Goiás. *Inia araguaiaensis* differs from other *Inia* species by morphological and molecular (Cyt*b*, *COI*, nuDNA microsatellite markers) characters. The status of *I. araguaiaensis*, however, was argued by The Society for Marine Mammalogy (2019), alleging sampling gaps which would leave doubts about the genetic difference found by Hrbek et al. (2014) represents a specific level divergence or an effect of isolation by distance in a specific clade. The Society for Marine Mammalogy (2019) also argues about the morphological analysis based on only two specimens of *I. araguaiaensis* and nine specimens of *I. geoffrensis*, which could not account possible effects of sexual dimorphism between the species. Herein we recognized *I. araguaiaensis* in view of the clear marked genetic structure (see Hrbek et al. 2014), but we also recognize that morphological analyzes on a broader sample could give more support to the specific status of *I. araguaiaensis*.

Introduced (non-native) species

To date, eight species of mammals are known to have been introduced to Brazil:

Small rodents (*Mus musculus* Linnaeus, 1758, *Rattus rattus* (Linnaeus, 1758) and *Rattus norvegicus* (Berkenhout, 1769)): the three murid rodents were unintentionally introduced in South America in the sixteenth century, during the European colonization of the continent (Pimentel 2011). The three species, originally from Eurasia, spread into Brazil and now occur in all biomes and states of the country, in both conserved and disturbed areas, including protected areas (Rosa et al. 2017a).

Lepus europaeus Pallas, 1778: in the late nineteenth century, the European hare *L. europaeus*, native in Eurasia, arrived in Argentina and Chile to serve as hunting. It spreaded to Southern Brazil in the 1950s by natural dispersal across the border with Uruguay and through deliberate introductions (Grigera & Rapoport 1983, Costa & Fernandes 2010). Nowadays, *L. europaeus* occupies mainly pastures, agricultural areas and forest edges of Pampa, Cerrado and Atlantic Forest biomes in the Brazilian states of Rio Grande do Sul, Santa Catarina, Paraná, São Paulo, Minas Gerais, Goiás and Bahia.

Sus scrofa Linnaeus, 1758: the wild boar *S. scrofa* is native in Eurasia and Africa and was first introduced to South America in the sixteenth century for meat consumption by explorers and settlers. By the late twentieth century, *S. scrofa* had reached Brazil from Uruguay and in the beginning of the twenty-first century several independent introductions were made throughout Brazil for meat production and hunting (Oliveira C.H.S., unpublished data). There are currently both voluntary introductions for hunting purposes and involuntary introductions of animals escaping from illegal domestic breeding (Oliveira C.H.S., unpublished data). Nowadays, *S. scrofa* occupies both conserved

and disturbed areas throughout Brazil, including protected areas, of all biomes in the states of Rio Grande do Sul, Santa Catarina, Paraná, São Paulo, Minas Gerais, Espírito Santo, Rio de Janeiro, Goiás, Tocantins, Mato Grosso, Mato Grosso do Sul, Bahia, Ceará, Pará, Maranhão and Rondônia.

Axis axis (Erxleben, 1777): original from Asia, the chital *A. axis* was first recorded in 2010 at Espinilho State Park, Rio Grande do Sul state, in Southern Brazil, probably coming from Argentina where the species was introduced for hunting (Sponchiado et al. 2011, Rosa et al. 2017a). Since then, *A. axis* have been reported in several municipalities of Rio Grande do Sul (Rosa et al. 2017a), occupying grasslands of Pampa biome where feral populations may have established.

Bubalus bubalis (Linnaeus, 1758): original from Asia, the water buffalo *B. bubalis* was intentionally introduced to Brazil in the nineteenth century for food and labor animal (Rosa et al. 2017a). Nowadays *B. bubalis* has scattered populations in both conserved and disturbed areas, including protected areas, in the Pampa, Atlantic Forest, Pantanal, Cerrado and Amazon biomes in the states of Rio Grande do Sul, Santa Catarina, Paraná, São Paulo, Rio de Janeiro, Goiás, Minas Gerais, Mato Grosso, Mato Grosso do Sul, Pará, Amapá, Rondônia, Bahia, Tocantins, Maranhão and Piauí (Rosa et al. 2017a).

Rusa unicolor (Kerr, 1792): the sambar *R. unicolor* is original from Asia and was intentionally introduced during the twentieth century into non fenced areas of Atlantic Forest, likely for sport hunting (Pimentel 2011). Nowadays three isolated populations of *R. unicolor* are known, one in Paraná state and two in São Paulo state (Pimentel 2011, Rosa et al. 2017a).

Native displaced species

Seven species have been translocated from their natural habitats in Brazil to other locations inside the country:

Callithrix sp. (*Callithrix jacchus* (Linnaeus, 1758), *Callithrix geoffroyi* (Humboldt, 1812), *Callithrix penicillata* (É. Geoffroy St.-Hilaire, 1812) and their hybrids): Three species of marmosets from *Callithrix* genus have been introduced through Brazil, including islands and protected areas, as a result of pet escapes and misguided releases of confiscated pet animals (Pimentel 2011). *Callithrix jacchus* naturally occurs in the scrub forests of the Atlantic Forest in Northeast Brazil (Bezerra et al. 2018). Invasive populations of the *C. jacchus* have been reported in several Atlantic Forest regions in states of Bahia, Minas Gerais, Rio de Janeiro, São Paulo, Espírito Santo, Paraná and Santa Catarina (Rosa et al. 2017a). *Callithrix penicillata* has a wide native distribution in Brazil, occurring in gallery forests, dry forests, and forest patches in the Cerrado, Caatinga and Atlantic Forest (Bicca-Marques et al. 2018). Invasions of *C. penicillata* have been reported in Atlantic Forest habitats of states of Rio Grande do Sul, Santa Catarina, Paraná, Minas Gerais, São Paulo, Rio de Janeiro, Bahia and Paraíba. *Callithrix geoffroyi* originally occurs in the Atlantic Forest of Minas Gerais, Bahia and Espírito Santo (Rylands & Mendes 2018) and is known to have been introduced in southern Brazilian Atlantic Forest, into Florianópolis Island in the state of Santa Catarina. The three species hybridize with each other and with *Callithrix flaviceps, Callithrix kuhlli* and *Callithrix aurita.* Hybrids were registered in the states of Santa Catarina, Paraná, Espírito Santo, Rio de Janeiro, São Paulo, Minas Gerais, Pernambuco and Bahia (Rosa et al. 2017a, Aximoff et al. 2020).

Saimiri sciureus (Linnaeus, 1758): the common squirrel monkey *S. sciureus* is a small Amazonian primate and was intentionally

introduced to the Atlantic Forest. The first record of *S. sciureus* outside its natural range was in 1987 at the Saltinho Biological Reserve, Pernambuco, during a pet release operation of individuals apprehended from illegal fauna trade (Camarotti et al. 2015). Currently, established populations of *S. sciureus* occur in forest fragments of Atlantic Forest in the states of Pernambuco, Rio de Janeiro and Alagoas. The species has potential to expand its distribution into non-native environments due illegal pet release (Rosa et al. 2017b).

Leontopithecus chrysomelas (Kuhl, 1820): the golden-head lion tamarin *L. chrysomelas* is an endemic and threatened species of the Atlantic Forest of Bahia state (Coimbra-Filho & Mittermeier 1973, Kierulff et al. 2008). In the 1990s, individuals of *L. chrysomelas* were accidentally introduced by a private collector into an urban Atlantic Forest remnant in the state of Rio de Janeiro, where established an invasive population (Kierulff et al. 2012, Molina et al. 2017). Recently, some individuals of *L. chrysomelas* were also seen in forest fragments of Atlantic Forest in Camaragibe municipality, Pernambuco state (Rosa et al. In Press).

Myocastor coypus (Molina, 1782): the nutria *M. coypus* is a large rodent native from open areas of southern South America, including the south of Brazil (Ojeda et al. 2016). In Brazil *M. coypus* was introduced by breeders for purposes of use of their skin (Bueno 2013). The species established non-native populations in states of São Paulo, Rio de Janeiro and Minas Gerais (Rosa et al. In Press).

Kerodon rupestris (Wied Neuwied, 1820): the rock cavy *K. rupestris* is a rodent native to the Brazilian Caatinga biome. In 1967, the species was introduced to Fernando de Noronha Archipelago by military personnel as hunting game, and an established population occurs until today (Pimentel 2011, Rosa et al. 2017a). No other non-native populations of *K. rupestris* are known to occur in Brazil.

DISCUSSION

Brazilian mammalogy is passing through an accelerated period of investigative taxonomy. The first compilation of Brazilian mammals (Fonseca et al. 1996) listed 524 species. Ten years later, Reis et al. (2006) indicated the occurrence of 652 species to the country. This number was increased to 694 species after five years (Reis et al. 2011). The following compilation, the annotated checklist of Paglia et al. (2012), published only one year later, listed a total of 701 species. After five years, the last list of mammals occurring in Brazil (Percequillo & Gregorin 2017) presented a total of 721 species. During the short period of two years between the publication of Percequillo & Gregorin (2017) list and the annotated checklist herein presented, 15 new taxa were described (Miranda et al. 2018, Pavan et al. 2017, Quintela et al. 2017, Machado et al. 2018, Pavan et al. 2018, Voss et al. 2018, Boubli et al. 2019, Pavan 2019, Peçanha et al. 2019, Velazco & Patterson 2019, Costa-Araújo et al. 2019, Abreu-Júnior & Percequillo 2019), most of them supported by genetic analyses. This, added to synonimizations (e.g. Byrne et al. 2016, Serrano-Villavicencio et al. 2019) and to the new records for the country (e.g. Bezerra et al. 2019, Percequillo et al. 2017, Menezes et al. 2020), resulted in 751 species herein listed. Notwithstanding, our checklist represents the entire mammalian diversity so far known for the country, the available evidence pointed out a still underestimated diversity. Based on recent taxonomic revisions, some of the taxa currently listed clearly represent species complex that invites further studies (e.g. *Oecomys*; Suárez-Villota et al. 2017). In addition, several lineages

divergent at specific level identified through molecular phylogenetic analyses still await formal description. These lineages comprise new forms of rodent genera *Deltamys* (Quintela et al. 2017), *Holochilus* (D'Elía et al. 2015), and *Phyllomys* (Machado et al. 2018), a new form of long-nosed armadillo (*Dasypus*) from Guiana Shield region (Feijó et al. 2019), among others. Thus, the new era of integrative taxonomic studies combining morphology and molecular tools is unveiling an overlooked diversity and driving a new period in Brazilian mammalogy. Hence, a considerable increment in the Brazilian mammalian diversity is expected. The increase in the number of researchers dedicated to systematics and taxonomy of mammals, together with greater field survey efforts in new regions, are also important factors that have contributed to advancement on the knowledge of mammalian diversity in Brazil.

The mammalian richness in Brazil is considerably higher than those reported in surrounding countries, including Argentina (409 spp.; Teta et al. 2018), Bolivia (406 spp; Aguirre et al. 2019), Paraguay (184 spp.; de la Sancha et al. 2017, 2019), Peru (508 spp; Pacheco et al. 2009) and Uruguay (73 spp; Queirolo 2016). This higher diversity may be a reflex of the larger area and greater diversity of environments in Brazil in relation to neighboring countries.

The negative effects of introduced (non-native or native) mammalian species are far known (see Rosa et al. 2017a). Reis et al. (2006, 2011) also concerned on listing the non-native species established in natural environments in Brazil and in that time this species were restricted to the murid rodents *M. musculus*, *R. novergicus* and *R. rattus*, the European hare *L. europaeus*, the wild boar *S. scrofa* and the water buffalo *B. bubalis*. Since them, the cervids *Axis axis* and *Rusa unicolor* were detected in the wild respectively in Atlantic Forest-Cerrado transitional area and Pampa environments, raising to eight the number of non-native mammals introduced to the country. Another very relevant issue is the occurrence of native displaced species, comprising five species of primates and two species of hystricomorph rodents. The potential impacts of both native and non-native species, as well as the recommendations for management and policy, are discussed in Rosa et al. (2017a).

Around 10% (80 species) of the Brazilian mammalian fauna is threatened in a global scale. However, the Brazilian Red Book of Threatened Species of Fauna (ICMBio 2018) listed a total of 110 taxa (15% of the total species), which implies that many species considered regionaly threatened (e.g. *Thylamys macrurus*, *T. velutinus*, *Ozotoceros bezoarticus*, *Leopardus geoffroyi*, *L. guttulus*, *Puma concolor*) are not considered globally threatened. This situation is especially notable for order Carnivora considering that only two from the 12 species included in the Brazilian list (ICMBio 2018) were considered as threatened by IUCN (2019). On the other hand, most of the primate species considered regionally threatened were also evaluated as globally threatened. Together with these two orders, rodents also contribute substantially for both Brazilian and global lists. Most of the primate and rodent threatened species are endemic to Brazil. In general, the Brazilian mastofauna has an impressive endemism rate, comprising more than a quarter of the species accounted for the country. Around 22% of the endemic species (50 species) are threatened and a similar percentage has Data Deficient for conservation according to IUCN (2019). Therefore, endemic species comprise the greatest part of the threatened mammalian fauna in Brazil (62%). Many of these species are distributed mainly in non-protected areas as well as the non-endemic threatened taxa. The loss, fragmentation and

decharacterization of the natural environments represent the greatest threats to the mammalian fauna in Brazil (Costa et al. 2005, ICMBio 2018), and outside the protected areas, populations are extremely susceptible to these deleterious factors. Thus, the creation of new conservation units and the expansion of the established protected areas are crucial for mammalian conservation in Brazil.

The near future of mammalogy in Brazil is a quite delicate matter. On the one hand, we have an enormous potential for the discovery of new species and a wide field of research for many systematic, evolutionary, ecology, phylogeography, population genetics open questions. On the other hand, we have an unfavorable scenario for the biological conservation in the country. The current Brazilian Forest Code (established in 2012) is very ineffective for the maintenance of the areas covered by native vegetation, and is especially damaging to Atlantic Forest and Cerrado biomes, where only 20% of the natural areas is required as preserved within private properties. Atlantic Forest and Cerrado are recognized biodiversity hotspots (Myers et al. 2000) and have a crucial role in mammalian conservation in Brazil (Costa et al. 2005). Moreover, the new Brazilian government is absolutely antagonistic to the scientific development and biological conservation. The Ministry of Environment has drastically weakened on its crucial role of regularization of rural properties and control of the protected areas. Additionally, there has been a dramatic budget reduction in scientific research in recent years. In summary, the current panorama of uncertainty and instability of Brazilian science and weaken environmental rules are not consistent with the status of the country as the world's highest biodiversity, which includes an impressive mammalian fauna.

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Quintela FM idealized the study, performed the literature survey and wrote the manuscript. Rosa CA wrote the 'introduced species' section of the study. Feijó A performed the literature survey and wrote the manuscript.

APPENDIX

List of mammal species occurring in Brazil and their conservation status based on the Brazilian (ICMBio 2018) and international lists of threatened species (IUCN 2019). Acronyms: CR = critically endangered, DD = Deficient Data, EN = endangered, EX = presumable extinct, LC = least concern, NT = near threatened, PE = pending (re)evaluation, VU = vulnerable. Numbers between parenthesis, after authorship and date of description, indicate remarks included in the main text (Results section), separated by order. Asteriscs (*) indicate the species endemic to the country.

Taxon	Conservation Status	
Class Mammalia Linnaeus, 1758	**IUCN**	**ICMBio**
Infraclass Marsupialia		
Order Didelphimorphia Gill, 1872		
Family Didelphidae Gray, 1821		
***Caluromys* Allen, 1900**		
Caluromys philander (Linnaeus, 1758)	LC	
Caluromys lanatus (Olfers, 1818)	LC	
***Caluromysiops* Sanborn, 1951**		
Caluromysiops irrupta Sanborn, 1951	LC	CR
***Chironectes* Illiger, 1811**		

Continuation.

Chironectes minimus (Zimmermann, 1780)	LC	
***Cryptonanus* Voss, Lunde & Jansa, 2005**		
Cryptonanus agricolai (Moojen, 1943) *	DD	
Cryptonanus chacoensis (Tate, 1931)	LC	
Cryptonanus guahybae (Tate, 1931) *	DD	
***Didelphis* Linnaeus, 1758**		
Didelphis albiventris Lund, 1840	LC	
Didelphis aurita Wied-Neuwied, 1826	LC	
Didelphis imperfecta Mondolfi & Pérez-Hernández, 1984	LC	
Didelphis marsupialis Linnaeus, 1758	LC	
***Glironia* Thomas, 1912**		
Glironia venusta Thomas, 1912	LC	
***Gracilinanus* Gardner & Creighton, 1989**		
Gracilinanus agilis (Burmeister, 1854)	LC	
Gracilinanus emiliae (Thomas, 1909)	DD	
Gracilinanus microtarsus (Wagner, 1842)	LC	
Gracilinanus peruanus (Tate, 1931) [(1)]	PE	
***Hyladelphys* Voss, Lunde & Simmons, 2001**		
Hyladelphys kalinowskii (Hershkovitz, 1992)	LC	
***Lutreolina* Thomas, 1910**		
Lutreolina crassicaudata (Desmarest, 1804)	LC	
***Marmosa* Gray, 1821**		
Marmosa (Marmosa) murina (Linnaeus, 1758)	LC	
Marmosa (Micoureus) constantiae (Thomas, 1904)	LC	
Marmosa (Micoureus) demerarae (Thomas, 1905)	LC	
Marmosa (Micoureus) paraguayana Tate, 1931	LC	
Marmosa (Micoureus) phaea Thomas, 1899 [(2)]	VU	
Marmosa (Micoureus) rapposa Thomas, 1899 [(3)]	PE	
Marmosa (Micoureus) regina (Thomas, 1898)	LC	
Marmosa (Micoureus) rutteri Thomas, 1924 [(4)]	PE	
Marmosa (Stegomarmosa) lepida (Thomas, 1888) [(5)]	LC	
***Marmosops* Matschie, 1916 [(6,7)]**		
Marmosops (Marmosops) incanus (Lund, 1840) *	LC	
Marmosops (Marmosops) neblina Gardner, 1989	LC	
Marmosops (Marmosops) noctivagus (Tschudi, 1845)	LC	
Marmosops (Marmosops) ocellatus (Tate, 1931)	LC	
Marmosops (Marmosops) paulensis (Tate, 1931) *	LC	VU
Marmosops (Sciophanes) bishopi (Pine, 1981)	LC	

Continuation.

Marmosops (Sciophanes) parvidens (Tate, 1931)	LC	
Marmosops (Sciophanes) pinheiroi (Pine, 1981)	LC	
***Metachirus* Burmeister, 1854**		
Metachirus myosuros (Temminck, 1824) [8]	PE	
Metachirus nudicaudatus (Desmarest, 1817)	PE	
***Monodelphis* Burnett, 1830** [9,10]		
Monodelphis (Microdelphys) americana (Müller, 1776) *	LC	
Monodelphis (Microdelphys) scalops (Thomas, 1888)	LC	
Monodelphis (Microdelphys) iheringi (Thomas, 1888) *	DD	
Monodelphis (Monodelphis) arlindoi Pavan, Rossi & Schneider, 2012	PE	
Monodelphis (Monodelphis) brevicaudata (Erxleben, 1777)	LC	
Monodelphis (Monodelphiops) dimidiata (Wagner, 1847)	LC	
Monodelphis (Monodelphis) domestica (Wagner, 1842)	LC	
Monodelphis (Monodelphis) glirina (Wagner, 1842)	LC	
Monodelphis (Monodelphis) touan (Shaw, 1800)	PE	
Monodelphis (Monodelphis) vossi Pavan, 2019 [11] *	PE	
Monodelphis (Monodelphiops) unistriata (Wagner, 1842)	CR	
Monodelphis (Mygalodelphys) handleyi Solari, 2007 [12]	DD	
Monodelphis (Mygalodelphys) kunsi Pine, 1975	LC	
Monodelphis (Mygalodelphys) pinocchio Pavan, 2015 *	PE	
Monodelphis (Mygalodelphys) saci Pavan, Mendes-Oliveira & Voss, 2017 [13] *	PE	
Monodelphis (Pyrodelphys) emiliae (Thomas, 1912)	LC	
***Philander* Brisson, 1762**		
Philander andersoni (Osgood, 1913)	LC	
Philander canus (Osgood, 1913) [14]	PE	
Philander quica (Temminck, 1824) [15]	PE	
Philander mcilhennyi Gardner & Patton, 1972	LC	
Philander opossum (Linnaeus, 1758)	LC	
Philander pebas Voss, Díaz-Nieto & Jansa, 2018 [16]	PE	
***Thylamys* Gray, 1843**		
Thylamys karimii (Petter, 1968) *	VU	
Thylamys macrurus (Olfers, 1818)	NT	EN
Thylamys velutinus (Wagner, 1842) *	NT	VU
Infraclass Placentalia		
Superordem Afrotheria		
Order Sirenia Illiger, 1811		
Family Trichechidae Gill, 1872		
***Trichechus* Linnaeus, 1758** [1]		

Continuation.

Trichechus inunguis (Natterer, 1883)	VU	VU
Trichechus manatus Linnaeus, 1758	VU	EN
Superorder Xenarthra		
Order Cingulata Illiger, 1821		
Family Dasypodidae Gray, 1821		
***Dasypus* Linnaeus, 1758** [(1)]		
Dasypus (Dasypus) novemcinctus Linnaeus, 1758	LC	
Dasypus (Hyperoambon) beniensis Lönnberg, 1942 [(2)]	PE	
Dasypus (Hyperoambon) kappleri Krauss, 1862 [(3)]	PE	
Dasypus (Hyperoambon) pastasae (Thomas, 1901) [(4)]	PE	
Dasypus (Muletia) septemcinctus Linnaeus, 1758 [(5)]	LC	
Family Chlamyphoridae Bonaparte, 1850 [(6)]		
***Cabassous* McMurtrie, 1831** [(7)]		
Cabassous tatouay (Desmarest, 1804)	LC	
Cabassous squamicaudis (Lund, 1845) [(8)]	PE	
Cabassous unicinctus (Linnaeus, 1758)	PE	
***Euphractus* Wagler, 1830**		
Euphractus sexcinctus (Linnaeus, 1758)	LC	
***Priodontes* Cuvier, 1825**		
Priodontes maximus (Kerr, 1792)	VU	VU
***Tolypeutes* Illiger, 1811**		
Tolypeutes matacus (Desmarest, 1804)	NT	
Tolypeutes tricinctus (Linnaeus, 1758) *	VU	EN
Order Pilosa Flower, 1883		
Family Bradypodidae Gray, 1821		
***Bradypus* Linnaeus, 1758**		
Bradypus torquatus Illiger, 1811 *	VU	VU
Bradypus tridactylus Linnaeus, 1758	LC	
Bradypus variegatus Schinz, 1825	LC	
Family Megalonychidae Gervais, 1855		
***Choloepus* Illiger, 1811**		
Choloepus didactylus (Linnaeus, 1758)	LC	
Choloepus hoffmanni Peters, 1858	LC	
Family Cyclopedidae Pocock, 1924		
***Cyclopes* Gray, 1821** [(1)]		
Cyclopes didactylus (Linnaeus, 1758) [(2)]	PE	
Cyclopes ida Thomas, 1900 [(3)]	PE	
Cyclopes rufus Miranda, Casali, Perini, Machado & Santos, 2017 [(4)] *	PE	

Continuation.

Cyclopes thomasi Miranda, Casali, Perini, Machado & Santos, 2017 [5]	PE	
Cyclopes xinguensis Miranda, Casali, Perini, Machado & Santos, 2017 [6] *	PE	
Family Myrmecophagidae Gray, 1825		
***Myrmecophaga* Linnaeus, 1758**		
Myrmecophaga tridactyla Linnaeus, 1758	VU	VU
***Tamandua* Gray, 1825**		
Tamandua tetradactyla (Linnaeus, 1758)	LC	
Superorder Euarchontoglires		
Order Primates Linnaeus, 1758		
Family Aotidae Poche, 1908		
***Aotus* Illiger, 1811**		
Aotus azarae (Humboldt, 1811)	LC	
Aotus infulatus (Kuhl, 1820) *	NT	
Aotus nancymaae Hershkovitz, 1983	VU	
Aotus nigriceps Dollman, 1909	LC	
Aotus trivirgatus (Humboldt, 1811)	LC	
Aotus vociferans (Spix, 1823)	LC	
Family Atelidae Gray, 1825		
***Alouatta* Lacépède, 1799**		
Alouatta belzebul (Linnaeus, 1766) *	VU	VU
Alouatta caraya (Humboldt, 1812)	LC	
Alouatta discolor (Spix, 1823) *	VU	VU
Alouatta guariba (Humboldt, 1812)	LC	
Alouatta juara Elliot, 1910	LC	
Alouatta macconnelli Elliot, 1910	LC	
Alouatta nigerrima Lönnberg, 1941*	LC	
Alouatta puruensis Lönnberg, 1941	LC	
Alouatta seniculus (Linnaeus, 1766)	LC	
Alouatta ululata Elliot, 1912 *	EN	EN
***Ateles* É. Geoffroy St.-Hilaire, 1806**		
Ateles belzebuth É. Geoffroy Saint-Hilaire, 1806	EN	VU
Ateles chamek (Humboldt, 1812)	EN	VU
Ateles marginatus (É. Geoffroy Saint-Hilaire, 1809) *	EN	EN
Ateles paniscus (Linnaeus, 1758)	VU	
***Brachyteles* Spix, 1823**		
Brachyteles arachnoides (É. Geoffroy Saint-Hilaire, 1806) *	CR	EN
Brachyteles hypoxanthus (Kuhl, 1820) *	CR	CR
***Lagothrix* É Geoffroy Saint-Hilaire, 1812**		

Continuation.

Lagothrix cana (É Geoffroy Saint-Hilaire, 1812)	EN	
Lagothrix lagothrica (Humboldt, 1812)	VU	VU
Lagothrix poeppigii Schinz, 1844	VU	VU
Family Callitrichidae Thomas, 1903		
***Callimico* Miranda-Ribeiro, 1912**		
Callimico goeldii (Thomas, 1904)	VU	
***Callithrix* Erxleben, 1777**		
Callithrix aurita (É. Geoffroy Saint-Hilaire, 1812) *	VU	EN
Callithrix flaviceps (Thomas, 1903) *	EN	EN
Callithrix geoffroyi (Humboldt, 1812) *	LC	
Callithrix jacchus (Linnaeus, 1758) *	LC	
Callithrix kuhlii Coimbra-Filho, 1985 *	NT	
Callithrix penicillata (É. Geoffroy Saint-Hilaire, 1812) *	LC	
***Cebuella* Gray, 1866**		
Cebuella niveiventris Lönnberg, 1940 [(1)]	LC	
Cebuella pygmaea (Spix, 1823)	LC	
***Leontopithecus* Lesson, 1840**		
Leontopithecus caissara Lorini & Persson, 1990 *	CR	EN
Leontopithecus chrysomelas (Kuhl, 1820) *	EN	EN
Leontopithecus chrysopygus (Mikan, 1823) *	EN	EN
Leontopithecus rosalia (Linnaeus, 1766) *	EN	EN
***Mico* Lesson, 1840**		
Mico acariensis (Roosmalen, Roosmalen, Mittermeier & Rylands, 2000) *	LC	
Mico argentatus (Linnaeus, 1771) *	LC	
Mico chrysoleucus (Wagner, 1842) *	LC	
Mico emiliae (Thomas, 1920) *	LC	
Mico humeralifer (É. Geoffroy Saint-Hilaire, 1812) *	DD	
Mico humilis (Roosmalen, Roosmalen, Mittermeier & Fonseca, 1998) *	LC	
Mico intermedius (Hershkovitz, 1977) *	LC	
Mico leucippe (Thomas, 1922) *	VU	
Mico marcai (Alperin, 1993) *	DD	
Mico mauesi (Mittermeier, Schwartz & Ayres, 1992) *	LC	
Mico melanurus (É. Geoffroy Saint-Hilaire, 1812)	LC	
Mico munduruku Costa-Araújo, Farias & Hrbek, 2019 [(2)] *	PE	
Mico nigriceps (Ferrari & Lopes, 1992) *	DD	
Mico rondoni Ferrari, Sena, Schneider & Silva Júnior, 2010 *	VU	VU
Mico saterei (Silva Junior & Noronha, 1998) *	LC	
***Saguinus* Hoffmannsegg, 1807**		

Continuation.

Saguinus inustus (Schwartz, 1951)	LC	
Saguinus (Leontocebus) cruzlimai Hershkovitz, 1966 *	LC	
Saguinus (Leontocebus) fuscicollis (Spix, 1823)	LC	
Saguinus (Leontocebus) fuscus (Lesson, 1840)	LC	
Saguinus (Leontocebus) nigricollis (Spix, 1823)	LC	
Saguinus (Leontocebus) weddelli (Deville, 1849)	LC	
Saguinus (Saguinus) bicolor (Spix, 1823) *	EN	CR
Saguinus (Saguinus) martinsi (Thomas, 1912) *	LC	
Saguinus (Saguinus) midas (Linnaeus, 1758)	LC	
Saguinus (Saguinus) niger (É Geoffroy Saint-Hilaire, 1803) *	VU	VU
Saguinus (Saguinus) ursula Hoffmannsegg, 1807 *	PE	
Saguinus (Tamarinus) imperator (Goeldi, 1907)	LC	
Saguinus (Tamarinus) labiatus (É Geoffroy Saint-Hilaire, 1812)	LC	
Saguinus (Tamarinus) mystax (Spix, 1823)	LC	
Family Cebidae Bonaparte, 1831		
***Cebus* Erxleben, 1777** [3]		
Cebus albifrons (Humboldt, 1812)	LC	
Cebus kaapori Queiroz, 1992 *	CR	CR
Cebus olivaceus Schomburgk, 1848	PE	
Cebus unicolor Spix, 1823	PE	
***Saimiri* Vogt, 1831**		
Saimiri boliviensis (I. Geoffroy & Blainville, 1834)	LC	
Saimiri cassiquiarensis (Lesson, 1840)	LC	
Saimiri collinsi Osgood, 1916 *	PE	
Saimiri macrodon Elliot, 1907	LC	
Saimiri sciureus (Linnaeus, 1758)	LC	
Saimiri ustus (I. Geoffroy Saint-Hilaire, 1843)	NT	
Saimiri vanzolinii Ayres, 1985 *	VU	VU
***Sapajus* Kerr, 1792** [3]		
Sapajus apella (Linnaeus, 1758)	LC	
Sapajus cay (Illiger, 1815)	LC	VU
Sapajus flavius (Schreber, 1774) *	CR	EN
Sapajus libidinosus (Spix, 1823) *	LC	
Sapajus macrocephalus (Spix, 1823)	LC	
Sapajus nigritus (Goldfuss, 1809)	NT	
Sapajus robustus (Kuhl, 1820) *	EN	EN
Sapajus xanthosternos (Wied-Neuwied, 1826) *	CR	EN
Family Pitheciidae Mivart, 1865		

Continuation.

***Cacajao* Lesson, 1840** [4]		
Cacajao calvus (I. Geoffroy Saint-Hilaire, 1847)	VU	
Cacajao melanocephalus (Humboldt, 1812)	LC	
***Callicebus* Thomas, 1903** [5]		
Callicebus barbarabrownae Hershkovitz, 1990 *	CR	CR
Callicebus coimbrai Kobayashi & Langguth, 1999 *	EN	EN
Callicebus melanochir Wied-Neuwied, 1820 *	VU	VU
Callicebus nigrifrons (Spix, 1823) *	NT	
Callicebus personatus (É. Geoffroy Saint-Hilaire, 1812) *	VU	VU
***Plecturocebus* Byrne, Rylands, Carneiro, Lynch Alfaro, Bertuol, da Silva, Messias, Groves, Mittermeier, Farias, Hrbek, Schneider, Sampaio & Boubli, 2016** [6]		
Plecturocebus baptista Lönnberg, 1939 *	LC	
Plecturocebus bernhardi Roosmalen, Roosmalen & Mittermeier, 2002 *	LC	
Plecturocebus brunneus (Wagner, 1842)	LC	
Plecturocebus caligatus (Wagner, 1842) [7] *	LC	
Plecturocebus cinerascens (Spix, 1823) *	LC	
Plecturocebus cupreus (Spix, 1823)	LC	
Plecturocebus donacophilus (d'Orbigny, 1836)	LC	
Plecturocebus grovesi Boubli, Byrne, da Silva, Silva-Júnior, Araújo, Bertuol, Gonçalves, Melo, Rylands, Mittermeier, Silva, Dash, Canale, Alencar, Rossi, Carneiro, Sampaio, Farias, Schneider, Hrbek, 2019 [8] *	PE	
Plecturocebus hoffmannsi Thomas, 1908 *	LC	
Plecturocebus moloch (Hoffmannsegg, 1807) *	LC	
Plecturocebus pallescens Thomas, 1907	LC	
Plecturocebus parecis Gusmão, Messias, Carneiro, Schneider, Alencar, Calouro, Dalponte, Mattos, Ferrari, Buss, Azevedo, Júnior, Nash, Rylands & Barnett, 2019 [9] *	PE	
Plecturocebus stephennashi Roosmalen, Roosmalen & Mittermeier, 2002 [10] *	DD	
Plecturocebus vieirai (Gualda-Barros, Nascimento & Amaral, 2012) *	DD	
***Cheracebus* Byrne, Rylands, Carneiro, Lynch Alfaro, Bertuol, da Silva, Messias, Groves, Mittermeier, Farias, Hrbek, Schneider, Sampaio & Boubli, 2016** [11]		
Cheracebus lucifer Thomas, 1914	LC	
Cheracebus lugens (Humboldt, 1811)	LC	
Cheracebus purinus Thomas, 1927 *	LC	
Cheracebus regulus Thomas, 1927 *	LC	
Cheracebus torquatus (Hoffmannsegg, 1807) *	LC	
***Chiropotes* Lesson, 1840**		
Chiropotes albinasus (I. Geoffroy & Deville, 1848) *	EN	
Chiropotes chiropotes (Humboldt, 1811)	LC	

Continuation.

Chiropotes sagulatus (Traill, 1821)	PE	
Chiropotes satanas (Hoffmannsegg, 1807) *	CR	CR
Chiropotes utahicki Hershkovitz, 1985 *	EN	VU
***Pithecia* Desmarest, 1804**		
Pithecia albicans Gray, 1860 *	LC	
Pithecia cazuzai Marsh, 2014 *	DD	
Pithecia chrysocephala (I. Geoffroy Saint-Hilaire, 1850)	LC	
Pithecia hirsuta (Spix, 1823)	DD	
Pithecia inusta (Spix, 1823)	LC	
Pithecia irrorata Gray, 1843 [(12)]	PE	
Pithecia monachus (É. Geoffroy Saint-Hilaire, 1812)	LC	
Pithecia pithecia (Linnaeus, 1766)	LC	
Pithecia vanzolinii Hershkovitz, 1987 [(13)] *	DD	
Order Lagomorpha Brandt, 1855		
Family Leporidae Fischer, 1817		
***Sylvilagus* Gray, 1867**		
Sylvilagus brasiliensis (Linnaeus, 1758)	PE	
Sylvilagus tapetillus Thomas, 1913 [(1)] *	PE	
Order Rodentia Bowdich, 1821		
Family Caviidae Fischer, 1817		
***Cavia* Pallas, 1766**		
Cavia aperea Erxleben, 1777	LC	
Cavia fulgida Wagler, 1831 *	LC	
Cavia intermedia Cherem, Olimpio & Ximenez, 1999 *	LC	CR
Cavia magna Ximenez, 1980	LC	
***Galea* Meyen, 1833**		
Galea flavidens (Brandt, 1835) *	LC	
Galea spixii (Wagler, 1831)	LC	
***Hydrochoerus* Brisson, 1762**		
Hydrochoerus hydrochaeris (Linnaeus, 1766)	LC	
***Kerodon* Cuvier, 1823**		
Kerodon acrobata Moojen, Locks & Langguth, 1997 *	DD	VU
Kerodon rupestris (Wied-Neuwied, 1820) *	LC	VU
Family Cricetidae Fischer, 1817		
***Abrawayaomys* Cunha & Cruz, 1979**		
Abrawayaomys ruschii Cunha & Cruz, 1979 *	LC	
***Akodon* Meyen, 1833**		

Continuation.

Akodon azarae (Fischer, 1829)	LC	
Akodon cursor (Winge, 1887) *	LC	
Akodon lindberghi Hershkovitz, 1990 *	DD	
Akodon montensis Thomas, 1913	LC	
Akodon mystax Hershkovitz, 1998 *	DD	VU
Akodon paranaensis Christoff, Fagundes, Sbalqueiro, Mattevi & Yonenaga-Yassuda, 2000	LC	
Akodon reigi González, Langguth & Oliveira, 1998	LC	
Akodon sanctipaulensis Hershkovitz, 1990 *	DD	
Akodon toba Thomas, 1921	LC	
***Bibimys* Massoia, 1979**		
Bibimys labiosus (Winge, 1887)	LC	
***Blarinomys* Thomas, 1896**		
Blarinomys breviceps (Winge, 1887)	LC	
***Brucepattersonius* Hershkovitz, 1998**		
Brucepattersonius griserufescens Hershkovitz, 1998 *	DD	
Brucepattersonius igniventris Hershkovitz, 1998 *	DD	
Brucepattersonius iheringi (Thomas, 1896)	LC	
Brucepattersonius nebulosus Abreu-Júnior, Vilela, Christoff, Valiati & Percequillo, 2019 [(1)] *	PE	
Brucepattersonius soricinus Hershkovitz, 1998 *	DD	
***Calassomys* Pardiñas, Lessa, Teta, Salazar-Bravo & Camara, 2014**		
Calassomys apicalis Pardiñas, Lessa, Teta, Salazar-Bravo & Camara, 2014 *	PE	
***Calomys* Waterhouse, 1837**		
Calomys callidus (Thomas, 1916)	LC	
Calomys callosus (Rengger, 1830)	LC	
Calomys cerqueirai Bonvicino, Oliveira & Gentile, 2010 *	PE	
Calomys expulsus (Lund, 1840) *	LC	
Calomys laucha (Fischer, 1814)	LC	
Calomys mattevii Gurgel-Filho, Feijó & Langguth 2015 [(2)] *	PE	
Calomys tener (Winge, 1887)	LC	
Calomys tocantinsi Bonvicino, Lima & Almeida, 2003 *	LC	
***Castoria* Pardiñas, Geise, Ventura & Lessa, 2016 [(3)]**		
Castoria angustidens (Winge, 1887) [(3)]	PE	
***Cerradomys* Weksler, Percequillo & Voss, 2006**		
Cerradomys akroai Bovincino, Casado & Weksler, 2014 *	PE	
Cerradomys goytaca Tavares, Pessôa & Gonçalves, 2011 *	PE	EN
Cerradomys langguthi Percequillo, Hingst-Zaher & Bonvicino, 2008 *	PE	
Cerradomys maracajuensis (Langguth & Bonvicino, 2002)	LC	

Continuation.

Cerradomys marinhus (Bonvicino, 2003) *	LC	
Cerradomys scotti (Langguth & Bonvicino, 2002)	LC	
Cerradomys subflavus (Wagner, 1842)	LC	
Cerradomys vivoi Percequillo, Hingst-Zaher & Bonvicino, 2008*	PE	
***Delomys* Thomas, 1917**		
Delomys altimontanus Gonçalves & Oliveira, 2014 *	PE	
Delomys dorsalis (Hensel, 1873)	LC	
Delomys sublineatus (Thomas, 1903) *	LC	
***Deltamys* Thomas, 1917**		
Deltamys araucaria Quintela, Bertuol, González, Cordeiro-Estrela, Freitas & Gonçalves, 2017 [(4)] *	PE	
Deltamys kempi Thomas, 1917	LC	
***Drymoreomys* Percequillo, Weksler & Costa, 2011**		
Drymoreomys albimaculatus Percequillo, Weksler & Costa, 2011 *	NT	
***Euryoryzomys* Weksler, Percequillo & Voss, 2006**		
Euryoryzomys emmonsae (Musser, Carleton, Brothers & Gardner, 1998) *	DD	
Euryoryzomys lamia (Thomas, 1901) *	VU	EN
Euryoryzomys macconnelli (Thomas, 1910)	LC	
Euryoryzomys nitidus (Thomas, 1884)	LC	
Euryoryzomys russatus (Wagner, 1848)	LC	
***Gyldenstolpia* Pardiñas, D'Elía & Teta, 2009**		
Gyldenstolpia fronto (Winge, 1887)	CR	
Gyldenstolpia planaltensis (Avila-Pires, 1972) *	PE	EN
***Holochilus* Brandt, 1835**		
Holochilus brasiliensis (Desmarest, 1819) *	LC	
Holochilus chacarius Thomas, 1906	LC	
Holochilus sciureus Wagner, 1842	LC	
Holochilus vulpinus (Brants, 1827)	PE	
***Hylaeamys* Weksler, Percequillo & Voss, 2006**		
Hylaeamys laticeps (Lund, 1840) *	NT	
Hylaeamys megacephalus (Fischer, 1814)	LC	
Hylaeamys oniscus (Thomas, 1904) *	NT	
Hylaeamys perenensis (Allen, 1901)	LC	
Hylaeamys seuanezi (Weksler, Geise & Cerqueira, 1999) [(5)] *	PE	
Hylaeamys yunganus (Thomas, 1902)	LC	
***Juliomys* González, 2000**		
Juliomys ossitenuis Costa, Pavan, Leite & Fagundes, 2007 *	PE	
Juliomys pictipes (Osgood, 1933)	LC	

Continuation.

Juliomys rimofrons Oliveira & Bonvicino, 2002 *	NT	
Juliomys ximenezi Christoff, Vieira, Oliveira, Gonçalves, Valiati & Tomasi, 2016 [6] *	PE	
***Juscelinomys* Moojen, 1965**		
Juscelinomys candango Moojen, 1965 *	EX	EX
***Kunsia* Hershkovitz, 1966**		
Kunsia tomentosus (Lichtenstein, 1830)	LC	
***Lundomys* Voss & Carleton, 1993**		
Lundomys molitor (Winge, 1887)	LC	
***Microakodontomys* Hershkovitz, 1993**		
Microakodontomys transitorius Hershkovitz, 1993 *	EN	EN
***Neacomys* Thomas, 1900**		
Neacomys amoenus Thomas, 1903 [7]	PE	
Neacomys dubosti Voss, Lunde & Simmons, 2001	LC	
Neacomys guianae Thomas, 1905	LC	
Neacomys minutus Patton, da Silva & Malcolm, 2000 *	LC	
Neacomys musseri Patton, da Silva & Malcolm, 2000	LC	
Neacomys paracou Voss, Lunde & Simmons, 2001	LC	
***Necromys* Ameghino, 1889**		
Necromys lasiurus (Lund, 1841)	LC	
Necromys lenguarum (Thomas, 1898)	LC	
Necromys urichi (Allen & Chapman, 1897)	LC	
***Nectomys* Peters, 1861**		
Nectomys apicalis Peters, 1861	LC	
Nectomys rattus (Pelzeln, 1883)	LC	
Nectomys squamipes (Brants, 1827)	LC	
***Neusticomys* Anthony, 1921**		
Neusticomys ferreirai Percequillo, Carmignotto & Silva, 2005 *	DD	
Neusticomys oyapocki (Dubost & Petter, 1978)	DD	
Neusticomys peruviensis (Musser & Gardner, 1974) [8]	LC	
***Noronhomys* Carleton & Olson, 1999**		
Noronhomys vespuccii Carleton & Olson, 1999 *	EX	
***Oecomys* Thomas, 1906**		
Oecomys auyantepui Tate, 1939	LC	
Oecomys bicolor (Tomes, 1860) [9]	LC	
Oecomys catherinae Thomas, 1909 [10] *	LC	
Oecomys cleberi Locks, 1981 [11]	DD	
Oecomys concolor (Wagner, 1845)	LC	
Oecomys franciscorum Pardiñas, Teta, Salazar-Bravo, Myers & Galliari, 2016 [12]	PE	

Continuation.

Oecomys mamorae (Thomas, 1906) [13]	LC	
Oecomys paricola (Thomas, 1904) [14] *	DD	
Oecomys rex Thomas, 1910	LC	
Oecomys roberti (Thomas, 1904) [15]	LC	
Oecomys rutilus Anthony, 1921	LC	
Oecomys superans Thomas, 1911	LC	
Oecomys trinitatis (Allen & Chapman, 1893)	LC	
***Oligoryzomys* Bangs, 1900**		
Oligoryzomys chacoensis (Myers & Carleton, 1981)	LC	
Oligoryzomys flavescens (Waterhouse, 1837)	LC	
Oligoryzomys mattogrossae (Allen, 1916)	PE	
Oligoryzomys messorius (Thomas, 1901)	PE	
Oligoryzomys microtis (Allen, 1916)	LC	
Oligoryzomys moojeni Weksler & Bonvicino, 2005 *	DD	
Oligoryzomys nigripes (Olfers, 1818)	LC	
Oligoryzomys rupestris Weksler & Bonvicino, 2005 *	DD	EN
Oligoryzomys stramineus Bonvicino & Weksler, 1998 *	LC	
Oligoryzomys utiaritensis (Allen, 1916)	PE	
***Oxymycterus* Waterhouse, 1837**		
Oxymycterus amazonicus Hershkovitz, 1994 *	LC	
Oxymycterus caparaoe Hershkovitz, 1998 *	PE	
Oxymycterus dasytrichus (Schinz, 1821) *	LC	
Oxymycterus delator Thomas, 1903	LC	
Oxymycterus inca Thomas, 1900	LC	
Oxymycterus itapeby Peçanha, Quintela, Ribas, Althoff, Maestri, Gonçalves & Freitas, 2019 [16] *	PE	
Oxymycterus nasutus (Waterhouse, 1837)	LC	
Oxymycterus quaestor Thomas, 1903	LC	
Oxymycterus rufus (G. Fischer, 1814)	LC	
***Phaenomys* Thomas, 1917**		
Phaenomys ferrugineus (Thomas, 1894) *	EN	
***Podoxymys* Anthony, 1929**		
Podoxymys roraimae Anthony, 1929	VU	
***Pseudoryzomys* Hershkovitz, 1962**		
Pseudoryzomys simplex (Winge, 1887)	LC	
***Reithrodon* Waterhouse, 1837**		
Reithrodon typicus Waterhouse, 1837	LC	
***Rhagomys* Thomas, 1917**		

Continuation.

Rhagomys longilingua Luna & Patterson, 2003 [17]	LC	
Rhagomys rufescens (Thomas, 1886) *	VU	
***Rhipidomys* Tschudi, 1845**		
Rhipidomys cariri Tribe, 2005 *	DD	VU
Rhipidomys emiliae (Allen, 1916) *	LC	
Rhipidomys gardneri Patton, da Silva & Malcolm, 2000	LC	
Rhipidomys ipukensis Rocha, Costa & Costa, 2011 *	DD	
Rhipidomys itoan Costa, Geise, Pereira & Costa, 2011 *	LC	
Rhipidomys leucodactylus (Tschudi, 1845)	LC	
Rhipidomys macconnelli de Winton, 1900	LC	
Rhipidomys macrurus (Gervais, 1855) *	LC	
Rhipidomys mastacalis (Lund, 1840) *	LC	
Rhipidomys nitela Thomas, 1901	LC	
Rhipidomys tribei Costa, Geise, Pereira & Costa, 2011 *	DD	EN
Rhipidomys wetzeli Gardner, 1990	LC	
***Scapteromys* Waterhouse, 1837**		
Scapteromys aquaticus Thomas, 1920	LC	
Scapteromys meridionalis Quintela, Gonçalves, Althoff, Sbalqueiro, Oliveira & Freitas, 2014 *	PE	
Scapteromys tumidus (Waterhouse, 1837)	LC	
***Scolomys* Anthony, 1924**		
Scolomys ucayalensis Pacheco, 1991	LC	
***Sigmodon* Say & Ord, 1825**		
Sigmodon alstoni (Thomas, 1881)	LC	
***Sooretamys* Weksler, Percequillo & Voss, 2006**		
Sooretamys angouya (Fischer, 1814)	LC	
***Thalpomys* Thomas, 1916**		
Thalpomys cerradensis Hershkovitz, 1990 *	LC	VU
Thalpomys lasiotis Thomas, 1916 *	LC	EN
***Thaptomys* Thomas, 1916**		
Thaptomys nigrita (Lichtenstein, 1829)	LC	
***Wiedomys* Hershkovitz, 1959**		
Wiedomys cerradensis Gonçalves, Almeida & Bonvicino, 2005 *	DD	
Wiedomys pyrrhorhinos (Wied-Neuwied, 1821) *	LC	
***Wilfredomys* Avila-Pires, 1960**		
Wilfredomys oenax (Thomas, 1928)	EN	EN
***Zygodontomys* Allen, 1897**		
Zygodontomys brevicauda (Allen & Chapman, 1893)	LC	
Family Ctenomyidae Lesson, 1842		

Continuation.

***Ctenomys* Blainville, 1826** [18]		
Ctenomys bicolor Miranda-Ribeiro, 1914 *	PE	EN
Ctenomys bolivensis Waterhouse, 1848	PE	
Ctenomys flamarioni Travi, 1981 *	EN	EN
Ctenomys ibicuiensis Freitas, Fernandes, Fornel & Roratto, 2012 *	PE	
Ctenomys lami Freitas, 2001 *	VU	EN
Ctenomys minutus Nehring, 1887 *	DD	VU
Ctenomys nattereri Wagner, 1848 *	PE	
Ctenomys rondoni Miranda-Ribeiro, 1914 *	PE	
Ctenomys torquatus Lichtenstein, 1830	LC	
Family Cuniculidae Miller & Gidley, 1918		
***Cuniculus* Brisson, 1762** [19]		
Cuniculus paca (Linnaeus, 1766)	LC	
Family Dasyproctidae Bonaparte, 1838		
***Dasyprocta* Illiger, 1811** [20]		
Dasyprocta azarae Lichtenstein, 1823	DD	
Dasyprocta croconota Wagler, 1831 *	DD	
Dasyprocta fuliginosa Wagler, 1832	LC	
Dasyprocta iacki Feijó & Langguth, 2013 *	DD	
Dasyprocta leporina (Linnaeus, 1758)	LC	
Dasyprocta prymnolopha Wagler, 1831 *	LC	
Dasyprocta variegata Tschudi, 1845	DD	
***Myoprocta* Thomas, 1803**		
Myoprocta acouchy (Erxleben, 1777)	LC	
Myoprocta pratti Pocock, 1913	LC	
Family Dinomyidae Alston, 1876		
***Dinomys* Peters, 1873**		
Dinomys branickii Peters, 1873	LC	
Family Echimyidae Gray, 1825		
***Callistomys* Emmons & Vucetich, 1998**		
Callistomys pictus (Pictet, 1841) *	EN	EN
***Carterodon* Waterhouse, 1848**		
Carterodon sulcidens (Lund, 1838) *	DD	
***Clyomys* Thomas, 1916**		
Clyomys laticeps (Thomas, 1909)	LC	
***Dactylomys* I. Geoffroy Saint-Hilaire, 1838**		
Dactylomys boliviensis Anthony, 1920	LC	
Dactylomys dactylinus (Desmarest, 1817)	LC	
***Echimys* Cuvier, 1809**		

Continuation.

Echimys chrysurus (Zimmermann, 1780)	LC	
Echimys vieirai Iack-Ximenes, Vivo & Percequillo, 2005 *	DD	
***Euryzygomatomys* Goeldi, 1901**		
Euryzygomatomys spinosus (Fischer, 1814)	LC	
***Isothrix* Wagner, 1845**		
Isothrix bistriata Wagner, 1845	LC	
Isothrix negrensis Thomas, 1920 *	LC	
Isothrix pagurus Wagner, 1845 *	LC	
***Kannabateomys* Jentink, 1891**		
Kannabateomys amblyonyx (Wagner, 1845)	LC	
***Lonchothrix* Thomas, 1920**		
Lonchothrix emiliae Thomas, 1920 *	LC	
***Makalata* Husson, 1978**		
Makalata didelphoides (Desmarest, 1817)	LC	
Makalata macrura (Wagner, 1842)	LC	
Makalata obscura (Wagner, 1840) *	DD	
***Mesomys* Wagner, 1845**		
Mesomys hispidus (Desmarest, 1817)	LC	
Mesomys occultus Patton, da Silva & Malcolm, 2000 *	LC	
Mesomys stimulax Thomas, 1911 *	LC	
***Myocastor* Kerr, 1792**		
Myocastor coypus (Molina, 1782)	LC	
***Phyllomys* Lund, 1839**		
Phyllomys blainvilii (Jourdan, 1837) *	LC	
Phyllomys brasiliensis Lund, 1840 *	EN	EN
Phyllomys centralis Machado, Loss, Paz, Vieira, Rodrigues & Marinho-Filho, 2018 [(21)] *	PE	
Phyllomys dasythrix Hensel, 1872 *	LC	
Phyllomys kerri (Moojen, 1950) *	DD	
Phyllomys lamarum (Thomas, 1916) *	DD	
Phyllomys lundi Leite, 2003 *	EN	EN
Phyllomys mantiqueirensis Leite, 2003 *	CR	
Phyllomys medius (Thomas, 1909) *	LC	
Phyllomys nigrispinus (Wagner, 1842) *	LC	
Phyllomys pattoni Emmons, Leite, Kock & Costa, 2002 *	LC	
Phyllomys sulinus Leite, Christoff & Fagundes, 2008 *	DD	
Phyllomys thomasi (Ihering, 1871) *	EN	EN
Phyllomys unicolor (Wagner, 1842) *	CR	CR
***Proechimys* Allen, 1899**		

Continuation.

Proechimys brevicauda (Günther, 1876)	LC	
Proechimys cuvieri Petter, 1978	LC	
Proechimys echinothrix da Silva, 1998 *	LC	
Proechimys gardneri da Silva, 1998 *	DD	
Proechimys goeldii Thomas, 1905 *	LC	
Proechimys guyannensis (É. Geoffroy Saint-Hilaire, 1803)	LC	
Proechimys hoplomyoides (Tate, 1939)	DD	
Proechimys kulinae da Silva, 1998	DD	
Proechimys longicaudatus (Rengger, 1830)	LC	
Proechimys pattoni da Silva, 1998	LC	
Proechimys quadruplicatus Hershkovitz, 1948	LC	
Proechimys roberti Thomas, 1901 *	LC	
Proechimys simonsi Thomas, 1900	LC	
Proechimys steerei Goldman, 1911	LC	
***Thrichomys* Trouessart, 1880**		
Thrichomys apereoides (Lund, 1839) *	LC	
Thrichomys inermis (Pictet, 1843) *	LC	
Thrichomys laurentius Thomas, 1904 *	DD	
Thrichomys pachyurus (Wagner, 1845)	LC	
***Toromys* Iack-Ximenes, Vivo & Percequillo, 2005**		
Toromys grandis (Wagner, 1845) *	LC	
***Trinomys* Thomas, 1921[(22)]**		
Trinomys albispinus (I. Geoffroy Saint-Hilaire, 1838) *	LC	
Trinomys dimidiatus (Günther, 1877) *	LC	
Trinomys elegans (Lund, 1839) *	PE	
Trinomys eliasi (Pessôa & dos Reis, 1993) *	NT	VU
Trinomys gratiosus (Moojen, 1948) *	LC	
Trinomys iheringi (Thomas, 1911) *	LC	
Trinomys minor (dos Reis & Pessôa, 1995) *	PE	
Trinomys mirapitanga Lara, Patton & Hingst-Zaher, 2002 *	DD	EN
Trinomys moojeni (Pessôa, Oliveira & dos Reis, 1992) *	EN	EN
Trinomys paratus (Moojen, 1948)	DD	
Trinomys setosus (Desmarest, 1817) *	LC	
Trinomys yonenagae (Rocha, 1995) *	EN	EN
Family Erethizontidae Bonaparte, 1845		
***Chaetomys* Gray, 1843**		
Chaetomys subspinosus (Olfers, 1818) *	VU	VU
***Coendou* Lacépède, 1799**		

Continuation.

Coendou baturitensis Feijó & Langguth, 2013 [23] *	DD	
Coendou bicolor (Tschudi, 1844)	LC	
Coendou ichillus Voss & da Silva, 2001 [24]	DD	
Coendou insidiosus (Olfers, 1818) *	LC	
Coendou melanurus (Wagner, 1842)	LC	
Coendou nycthemera (Olfers, 1818) *	DD	
Coendou prehensilis (Linnaeus, 1758)	LC	
Coendou roosmalenorum Voss & da Silva, 2001 *	DD	
Coendou speratus Mendes Pontes, Gadelha, Melo de Sá, Loss, Caldara Junior, Costa & Leite, 2013 *	EN	EN
Coendou spinosus (Cuvier, 1823)	LC	
Family Sciuridae Fischer, 1817		
***Guerlinguetus* Gray, 1821**		
Guerlinguetus aestuans (Linnaeus, 1766)	LC	
Guerlinguetus brasiliensis (Gmelin, 1788) *	PE	
***Hadrosciurus* Allen, 1915**		
Hadrosciurus ignitus (Gray, 1867) [25]	LC	
Hadrosciurus igniventris (Wagner, 1842)	LC	
Hadrosciurus pyrrhinus (Thomas, 1898)	DD	
Hadrosciurus spadiceus (Olfers, 1818)	LC	
***Microsciurus* Allen, 1895**		
Microsciurus flaviventer (Gray, 1867)	LC	
***Sciurillus* Thomas, 1914**		
Sciurillus pusillus (É. Geoffroy Saint-Hilaire, 1803)	LC	
Superorder Laurasiatheria		
Order Chiroptera Blumenbach, 1779		
Family Emballonuridae Gervais, 1856		
***Centronycteris* Gray, 1838**		
Centronycteris maximiliani (Fischer, 1829)	LC	
***Cormura* Peters, 1867**		
Cormura brevirostris (Wagner, 1843)	LC	
***Cyttarops* Thomas, 1913**		
Cyttarops alecto Thomas, 1913	LC	
***Diclidurus* Wied-Neuwied, 1820**		
Diclidurus albus Wied-Neuwied, 1820	LC	
Diclidurus ingens Hernández-Camacho, 1955	DD	
Diclidurus isabella (Thomas, 1920)	LC	
Diclidurus scutatus Peters, 1869	LC	

Continuation.

***Peropteryx* Peters, 1867**		
Peropteryx kappleri Peters, 1867	LC	
Peropteryx leucoptera Peters, 1867	LC	
Peropteryx macrotis (Wagner, 1843)	LC	
Peropteryx pallidoptera Lim, Engstrom, Reid, Simmons, Voss & Fleck, 2010	DD	
Peropteryx trinitatis Miller, 1899	DD	
***Rhynchonycteris* Peters, 1867**		
Rhynchonycteris naso (Wied-Neuwied, 1820)	LC	
***Saccopteryx* Illiger, 1811**		
Saccopteryx bilineata (Temminck, 1838)	LC	
Saccopteryx canescens Thomas, 1901	LC	
Saccopteryx gymnura Thomas, 1901	DD	
Saccopteryx leptura (Schreber, 1774)	LC	
Family Furipteridae Gray, 1866		
***Furipterus* Bonaparte, 1837**		
Furipterus horrens (Cuvier, 1828)	LC	VU
Family Molossidae Gervais, 1856		
***Cynomops* Thomas, 1920**		
Cynomops abrasus (Temminck, 1827)	DD	
Cynomops greenhalli Goodwin, 1958	LC	
Cynomops milleri (Osgood, 1914)	LC	
Cynomops paranus (Thomas, 1901)	DD	
Cynomops planirostris (Peters, 1865)	LC	
***Eumops* Miller, 1906**		
Eumops auripendulus (Shaw, 1800)	LC	
Eumops bonariensis (Peters, 1874)	LC	
Eumops chimaera Gregorin, Moras, Acosta, Vasconcellos, Poma, Santos & Paca, 2016 [(1)]	PE	
Eumops delticus Thomas, 1923	DD	
Eumops glaucinus (Wagner, 1843)	LC	
Eumops hansae Sanborn, 1932	LC	
Eumops maurus (Thomas, 1901)	DD	
Eumops patagonicus Thomas, 1924	LC	
Eumops perotis (Schinz, 1821)	LC	
Eumops trumbulli (Thomas, 1901)	LC	
***Molossops* Peters, 1865**		
Molossops neglectus Williams & Genoways, 1980	DD	
Molossops temminckii (Burmeister, 1854)	LC	

Continuation.

***Molossus* É. Geoffroy Saint-Hilaire, 1805**		
Molossus aztecus Saussure, 1860	LC	
Molossus coibensis Allen, 1904	LC	
Molossus currentium Thomas, 1901	LC	
Molossus molossus Pallas, 1766	LC	
Molossus pretiosus Miller, 1902	LC	
Molossus rufus É. Geoffroy Saint-Hilaire, 1805	LC	
***Neoplatymops* Peterson, 1965**		
Neoplatymops mattogrossensis (Vieira, 1942)	PE	
***Nyctinomops* Miller, 1902**		
Nyctinomops aurispinosus (Peale, 1848)	LC	
Nyctinomops laticaudatus (É. Geoffroy Saint-Hilaire, 1805)	LC	
Nyctinomops macrotis (Gray, 1840)	LC	
***Promops* Gervais, 1856**		
Promops centralis Thomas, 1915	LC	
Promops nasutus (Spix, 1823)	LC	
***Tadarida* Rafinesque, 1814**		
Tadarida brasiliensis (I. Geoffroy Saint-Hilaire, 1824)	LC	
Family Mormoopidae Saussure, 1860		
***Pteronotus* Gray, 1838**		
Pteronotus (Pteronotus) gymnonotus (Wagner, 1843)	LC	
Pteronotus (Phyllodia) alitonus Pavan, Bobrowiec & Percequillo, 2018 [(2)]	PE	
Pteronotus (Phyllodia) rubiginosus (Wagner, 1843)	LC	
Pteronotus personatus (Wagner, 1843) [(3)]	LC	
Family Natalidae Gray, 1866		
***Natalus* Gray, 1838**		
Natalus macrourus (Gervais, 1856)	NT	VU
Family Noctilionidae Gray, 1821		
***Noctilio* Linnaeus, 1766**		
Noctilio albiventris Desmarest, 1818	LC	
Noctilio leporinus (Linnaeus, 1758)	LC	
Family Phyllostomidae Gray, 1825		
***Ametrida* Gray, 1847**		
Ametrida centurio Gray, 1847	LC	
***Anoura* Gray, 1838**		
Anoura caudifer (É. Geoffroy Saint-Hilaire, 1818)	LC	
Anoura geoffroyi Gray, 1838	LC	
***Artibeus* Leach, 1821**		

Continuation.

Artibeus (Artibeus) fimbriatus Gray, 1838	LC	
Artibeus (Artibeus) lituratus (Olfers, 1818)	LC	
Artibeus (Artibeus) obscurus (Schinz, 1821)	LC	
Artibeus (Artibeus) planirostris (Spix, 1823)	LC	
Artibeus (Dermanura) anderseni Osgood, 1916	LC	
Artibeus (Dermanura) bogotensis Andersen, 1906	LC	
Artibeus (Dermanura) cinereus (Gervais, 1856)	LC	
Artibeus (Dermanura) gnomus Handley, 1987	LC	
Artibeus (Koopmania) concolor Peters, 1865	LC	
***Carollia* Gray, 1838**		
Carollia benkeithi Solari & Baker, 2006	LC	
Carollia brevicauda (Schinz, 1821)	LC	
Carollia perspicillata (Linnaeus, 1758)	LC	
***Chiroderma* Peters, 1860** [4,5]		
Chiroderma doriae Thomas, 1891	LC	
Chiroderma trinitatum Goodwin, 1958	LC	
Chiroderma villosum Peters, 1860	LC	
***Choeroniscus* Thomas, 1928**		
Choeroniscus minor (Peters, 1868)	LC	
***Chrotopterus* Peters, 1865**		
Chrotopterus auritus (Peters, 1856)	LC	
***Desmodus* Wied-Neuwied, 1826**		
Desmodus rotundus (É. Geoffroy Saint-Hilaire, 1810)	LC	
***Diaemus* Miller, 1906**		
Diaemus youngi (Jentink, 1893)	LC	
***Diphylla* Spix, 1823**		
Diphylla ecaudata Spix, 1823	LC	
***Dryadonycteris* Nogueira, Lima, Peracchi & Simmons, 2012**		
Dryadonycteris capixaba Nogueira, Lima, Peracchi & Simmons, 2012	DD	
***Gardnerycteris* Hurtado & Pacheco, 2014**		
Gardnerycteris crenulatum (É. Geoffroy Saint-Hilaire, 1803) [6]	LC	
***Glossophaga* É. Geoffroy Saint-Hilaire, 1818**		
Glossophaga commissarisi Gardner, 1962	LC	
Glossophaga longirostris Miller, 1898	LC	
Glossophaga soricina (Pallas, 1766)	LC	
***Glyphonycteris* Thomas, 1896**		
Glyphonycteris behnii (Peters, 1865)	DD	VU
Glyphonycteris daviesi (Hill, 1964)	LC	
Glyphonycteris sylvestris Thomas, 1896	LC	

Continuation.

***Hsunycteris* Parlos, Timm, Swier, Zeballos & Baker, 2014**		
Hsunycteris pattoni (Woodman & Timm, 2006) [7]	DD	
Hsunycteris thomasi (Allen, 1904)	LC	
***Lampronycteris* Sanborn, 1949**		
Lampronycteris brachyotis (Dobson, 1879)	LC	
***Lichonycteris* Thomas, 1895**		
Lichonycteris degener Miller, 1931	LC	
***Lionycteris* Thomas, 1913**		
Lionycteris spurrelli Thomas, 1913	LC	
***Lonchophylla* Thomas, 1903**		
Lonchophylla bokermanni Sazima, Vizotto & Taddei, 1978 *	EN	
Lonchophylla dekeyseri Taddei, Vizotto & Sazima, 1983 *	EN	EN
Lonchophylla mordax Thomas, 1903	NT	
Lonchophylla inexpectata Moratelli and Dias (2015) [8] *	PE	
Lonchophylla peracchii Dias, Esbérard & Moratelli, 2013 *	LC	
***Lonchorhina* Tomes, 1863**		
Lonchorhina aurita Tomes, 1863	LC	VU
Lonchorhina inusitata Handley & Ochoa, 1997	DD	
***Lophostoma* d'Orbigny, 1836**		
Lophostoma brasiliense Peters, 1866	LC	
Lophostoma carrikeri (Allen, 1910)	LC	
Lophostoma schulzi (Genoways & Williams, 1980)	LC	
Lophostoma silvicola d'Orbigny, 1836	LC	
***Macrophyllum* Gray, 1838**		
Macrophyllum macrophyllum (Schinz, 1821)	LC	
***Mesophylla* Thomas, 1901**		
Mesophylla macconnelli Thomas, 1901	LC	
***Micronycteris* Gray, 1866 [9]**		
Micronycteris (Xenoctenes) hirsuta (Peters, 1869)	LC	
Micronycteris (Micronycteris) megalotis (Gray, 1842)	LC	
Micronycteris (Micronycteris) microtis Miller, 1898	LC	
Micronycteris (Schizonycteris) homezorum Pirlot, 1967	PE	
Micronycteris (Schizonycteris) minuta (Gervais, 1856)	LC	
Micronycteris (Schizonycteris) sanborni Simmons, 1996 *	LC	
Micronycteris (Schizonycteris) schmidtorum Sanborn, 1935	LC	
***Mimon* Gray, 1847**		
Mimon bennettii (Gray, 1838)	LC	
***Neonycteris* Sanborn, 1949**		

Continuation.

Neonycteris pusilla (Sanborn, 1949)	DD	
***Phylloderma* Peters, 1865**		
Phylloderma stenops Peters, 1865	LC	
***Phyllostomus* Lacépède, 1799**		
Phyllostomus discolor (Wagner, 1843)	LC	
Phyllostomus elongatus (É. Geoffroy Saint-Hilaire, 1810)	LC	
Phyllostomus hastatus (Pallas, 1767)	LC	
Phyllostomus latifolius (Thomas, 1901)	LC	
***Platyrrhinus* Saussure, 1860**		
Platyrrhinus angustirostris Velazco, Gardner & Patterson, 2010	LC	
Platyrrhinus aurarius (Handley & Ferris, 1972)	LC	
Platyrrhinus brachycephalus (Rouk & Carter, 1972)	LC	
Platyrrhinus fusciventris Velazco, Gardner & Patterson, 2010	LC	
Platyrrhinus incarum (Thomas, 1912)	LC	
Platyrrhinus infuscus (Peters, 1880)	LC	
Platyrrhinus lineatus (É. Geoffroy Saint-Hilaire, 1810)	LC	
Platyrrhinus recifinus (Thomas, 1901) *	LC	
***Pygoderma* Peters, 1843**		
Pygoderma bilabiatum (Wagner, 1843)	LC	
***Rhinophylla* Peters, 1865**		
Rhinophylla fischerae Carter, 1966	LC	
Rhinophylla pumilio Peters, 1865	LC	
***Scleronycteris* Thomas, 1912**		
Scleronycteris ega Thomas, 1912	DD	
***Sphaeronycteris* Peters, 1882**		
Sphaeronycteris toxophyllum Peters, 1882	LC	
***Sturnira* Gray, 1842**		
Sturnira (Sturnira) lilium (É. Geoffroy Saint-Hilaire, 1810)	LC	
Sturnira giannae Velazco & Patterson, 2019 [(10)]	PE	
Sturnira (Sturnira) magna De la Torre, 1966	LC	
Sturnira (Sturnira) tildae De la Torre, 1959	LC	
***Tonatia* Gray, 1827**		
Tonatia bidens (Spix, 1823)	DD	
Tonatia maresi Williams, Willig & Reid, 1995 [(11)]	PE	
***Trachops* Gray, 1847**		
Trachops cirrhosus (Spix, 1823)	LC	
***Trinycteris* Sanborn, 1949**		
Trinycteris nicefori (Sanborn, 1949)	LC	
***Uroderma* Peters, 1865**		

Continuation.

Uroderma bilobatum Peters, 1866	LC	
Uroderma magnirostrum Davis, 1968	LC	
***Vampyressa* Thomas, 1900**		
Vampyressa pusilla (Wagner, 1843)	DD	
Vampyressa thyone (Thomas, 1909)	LC	
***Vampyriscus* Thomas, 1900**		
Vampyriscus bidens (Dobson, 1878)	LC	
Vampyriscus brocki (Peterson, 1968)	LC	
***Vampyrodes* Thomas, 1900**		
Vampyrodes caraccioli (Thomas, 1889)	LC	
***Vampyrum* Rafinesque, 1815**		
Vampyrum spectrum (Linnaeus, 1758)	NT	
***Xeronycteris* Gregorin & Ditchfield, 2005**		
Xeronycteris vieirai Gregorin & Ditchfield, 2005 *	DD	VU
Family Thyropteridae Miller, 1907		
***Thyroptera* Spix, 1823**		
Thyroptera devivoi Gregorin, Gonçalves, Lim & Engstrom, 2006	DD	
Thyroptera discifera (Lichtenstein & Peters, 1855)	LC	
Thyroptera lavali Pine, 1993	DD	
Thyroptera tricolor Spix, 1823	LC	
Thyroptera wynneae Velazco, Gregorin, Voss & Simmons, 2014	DD	
Family Vespertilionidae Gray, 1821		
***Eptesicus* Rafinesque, 1820**		
Eptesicus andinus Allen, 1914	LC	
Eptesicus brasiliensis (Desmarest, 1819)	LC	
Eptesicus chiriquinus Thomas, 1920	LC	
Eptesicus diminutus Osgood, 1915	LC	
Eptesicus furinalis (d'Orbigny & Gervais, 1847)	LC	
Eptesicus taddeii Miranda, Bernardi & Passos, 2006 *	DD	VU
***Histiotus* Gervais, 1856**		
Histiotus alienus Thomas, 1916	DD	
Histiotus diaphanopterus Feijó, Rocha & Althoff, 2015 [(12)]	PE	
Histiotus laephotis Thomas, 1916	PE	
Histiotus montanus (Philippi & Landbeck, 1861)	LC	
Histiotus velatus (I. Geoffroy Saint-Hilaire, 1824)	DD	
***Lasiurus* Gray, 1831**		
Lasiurus (Dasypterus) ega (Gervais, 1856)	LC	
Lasiurus (Lasiurus) blossevillii (Lesson, 1826)	LC	
Lasiurus (Lasiurus) castaneus Handley, 1960	DD	

Continuation.

Lasiurus (Lasiurus) cinereus (Palisot de Beauvois, 1796)	LC	
Lasiurus (Lasiurus) ebenus Fazzolari-Corrêa, 1994 *	DD	
Lasiurus (Lasiurus) egregius (Peters, 1870)	DD	
Lasiurus (Lasiurus) salinae Thomas, 1902	DD	
***Myotis* Kaup, 1829**		
Myotis albescens (É. Geoffroy Saint-Hilaire, 1806)	LC	
Myotis dinellii Thomas, 1902	LC	
Myotis izecksohni Moratelli, Peracchi, Dias & Oliveira, 2011	DD	
Myotis lavali Moratelli, Peracchi, Dias & Oliveira, 2011	LC	
Myotis levis (I. Geoffroy Saint-Hilaire, 1824)	LC	
Myotis nigricans (Schinz, 1821)	LC	
Myotis riparius Handley, 1960	LC	
Myotis ruber (É. Geoffroy Saint-Hilaire, 1806)	NT	
Myotis simus Thomas, 1901	DD	
***Rhogeessa* Allen, 1866**		
Rhogeessa hussoni Genoways & Baker, 1996	DD	
Rhogeessa io Thomas, 1903	LC	
Order Carnivora Bowdich, 1821		
Family Canidae Fischer von Waldheim, 1817		
***Atelocynus* Cabrera, 1940**		
Atelocynus microtis (Sclater, 1883)	NT	VU
***Cerdocyon* Smith, 1839**		
Cerdocyon thous (Linnaeus, 1766)	LC	
***Chrysocyon* Smith, 1839**		
Chrysocyon brachyurus (Illiger, 1815)	NT	VU
***Lycalopex* Burmeister, 1854**		
Lycalopex gymnocercus (Fischer, 1814)	LC	
Lycalopex vetulus (Lund, 1842) *	LC	VU
***Speothos* Lund, 1839**		
Speothos venaticus (Lund, 1842)	NT	VU
Family Mephitidae Bonaparte, 1845		
***Conepatus* Gray, 1837**		
Conepatus chinga (Molina, 1782)	LC	
Conepatus amazonicus (Lichtenstein, 1838) [(1)] *	PE	
Family Mustelidae Fischer von Waldheim, 1817		
***Eira* Smith, 1842**		
Eira barbara (Linnaeus, 1758)	LC	
***Galictis* Bell, 1826**		

Continuation.

Galictis cuja (Molina, 1782)	LC	
Galictis vittata (Schreber, 1776)	LC	
***Lontra* Gray, 1843**		
Lontra longicaudis (Olfers, 1818)	NT	
***Mustela* Linnaeus, 1758**		
Mustela africana Desmarest, 1818	LC	
***Pteronura* Gray, 1837**		
Pteronura brasiliensis (Gmelin, 1788)	EN	VU
Family Otariidae Gray, 1825		
***Arctocephalus* (G. Saint-Hilaire and F. Cuvier, 1826)**		
Arctocephalus australis (Zimmermann, 1783)	LC	
Arctocephalus gazella (Peters, 1875)	LC	
Arctocephalus tropicalis (Gray, 1872)	LC	
***Otaria* Péron, 1816**		
Otaria flavescens (Shaw, 1800)	LC	
Family Phocidae Gray, 1821		
***Hydrurga* Gistel, 1848**		
Hydrurga leptonyx (Blainville, 1820)	LC	
***Lobodon* Gray, 1844**		
Lobodon carcinophaga (Hombron & Jacquinot, 1842)	LC	
***Mirounga* Gray, 1827**		
Mirounga leonina (Linnaeus, 1758)	LC	
Family Procyonidae Gray, 1825		
***Bassaricyon* Allen, 1876**		
Bassaricyon alleni Thomas, 1880	LC	
***Nasua* Storr, 1780**		
Nasua nasua (Linnaeus, 1766)	LC	
***Potos* É. Geoffroy Saint-Hilaire & Cuvier, 1795**		
Potos flavus (Schreber, 1774)	LC	
***Procyon* Storr, 1780**		
Procyon cancrivorus (Cuvier, 1798)	LC	
Family Felidae Fischer von Waldheim, 1817		
***Herpailurus* Severtzov, 1858 [(2)]**		
Herpailurus yagouaroundi (É. Geoffroy Saint-Hilaire, 1803)	LC	VU
***Leopardus* Gray, 1842**		
Leopardus braccatus (Cope, 1889) [(3)]	PE	
Leopardus emiliae (Thomas, 1914) [(4)] *	PE	
Leopardus geoffroyi (d'Orbigny & Gervais, 1844)	LC	VU
Leopardus guttulus (Hensel, 1872)	LC	VU

Continuation.

Leopardus munoai (Ximénez, 1961) [5]	PE	
Leopardus pardalis (Linnaeus, 1758)	LC	
Leopardus tigrinus (Schreber, 1775)	VU	EN
Leopardus wiedii (Schinz, 1821)	NT	VU
***Panthera* Oken, 1816**		
Panthera onca (Linnaeus, 1758)	NT	VU
***Puma* Jardine, 1834**		
Puma concolor (Linnaeus, 1771)	LC	VU
Order Perissodactyla Owen, 1848		
Family Tapiriidae Gray, 1821		
***Tapirus* Brisson, 1762**		
Tapirus kabomani Cozzuol, Clozato, Holanda, Rodrigues, Nienow, de Thoisy, Redondo & Santos, 2013 [1]	PE	
Tapirus terrestris (Linnaeus, 1758)	VU	VU
Order Artiodactyla Owen, 1848 [1]		
Family Cervidae Goldfuss, 1820		
***Blastocerus* Illiger, 1815**		
Blastocerus dichotomus (Illiger, 1815)	VU	VU
***Mazama* Rafinesque, 1817 [2]**		
Mazama americana (Erxleben, 1777)	DD	
Mazama bororo Duarte, 1996 *	VU	VU
Mazama gouazoubira (Fischer, 1814)	LC	
Mazama nana (Hensel, 1872)	VU	VU
Mazama nemorivaga (Cuvier, 1817)	LC	
***Odocoileus* Rafinesque, 1832**		
Odocoileus virginianus (Zimmermann, 1780)	LC	
***Ozotoceros* Ameghino, 1891**		
Ozotoceros bezoarticus (Linnaeus, 1758)	NT	VU
Family Tayassuidae Palmer, 1897		
***Pecari* Reichenbach, 1835**		
Pecari tajacu (Linnaeus, 1758)	LC	
***Tayassu* Fischer, 1814**		
Tayassu pecari (Link, 1795)	VU	VU
Family Balaenidae Gray, 1821		
***Eubalaena* Gray, 1864**		
Eubalaena australis (Desmoulins, 1822)	LC	EN
Family Balaenopteridae Gray, 1864		
***Balaenoptera* Lacépède, 1804**		
Balaenoptera acutorostrata Lacépède, 1804	LC	

Continuation.

Balaenoptera bonaerensis Burmeister, 1867	NT	
Balaenoptera borealis Lesson, 1828	EN	EN
Balaenoptera edeni Anderson, 1879	LC	
Balaenoptera musculus (Linnaeus, 1758)	EN	CR
Balaenoptera physalus (Linnaeus, 1758)	VU	EN
***Megaptera* Gray, 1846**		
Megaptera novaeangliae (Borowski, 1781)	LC	
Family Delphinidae Gray, 1821		
***Cephalorhynchus* Gray, 1846**		
Cephalorhynchus commersonii (Lacépède, 1804)	LC	
***Delphinus* Linnaeus, 1758**		
Delphinus delphis Linnaeus, 1758	LC	
***Feresa* Gray, 1870**		
Feresa attenuata Gray, 1874	LC	
***Globicephala* Lesson, 1828**		
Globicephala macrorhynchus Gray, 1846	LC	
Globicephala melas (Traill, 1809)	LC	
***Grampus* Gray, 1828**		
Grampus griseus (Cuvier, 1812)	LC	
***Lagenodelphis* Fraser, 1956**		
Lagenodelphis hosei Fraser, 1956	LC	
***Lagenorhynchus* Gray, 1846**		
Lagenorhynchus australis (Peale, 1848)	LC	
***Lissodelphis* Gloger, 1841**		
Lissodelphis peronii (Lacépède, 1804)	LC	
***Orcinus* Fitzinger, 1860**		
Orcinus orca (Linnaeus, 1758)	DD	
***Peponocephala* Nishiwaki & Norris, 1966**		
Peponocephala electra (Owen, 1846)	LC	
***Pseudorca* Reinhardt, 1862**		
Pseudorca crassidens (Owen, 1846)	NT	
***Sotalia* Gray, 1866**		
Sotalia fluviatilis (Gervais & Deville, 1853)	DD	
Sotalia guianensis (Van Bénéden, 1864)	NT	VU
***Stenella* Gray, 1866**		
Stenella attenuata (Gray, 1846)	LC	
Stenella clymene (Gray, 1850)	LC	
Stenella coeruleoalba (Meyen, 1833)	LC	
Stenella frontalis (Cuvier, 1829)	LC	

Continuation.

Stenella longirostris (Gray, 1828)	LC	
***Steno* Gray, 1846**		
Steno bredanensis (Cuvier, 1828)	LC	
***Tursiops* Gervais, 1855** [3]		
Tursiops truncatus (Montagu, 1821)	LC	
Family Iniidae Gray, 1846		
***Inia* d'Orbigny, 1834**		
Inia araguaiaensis Hrbek, Farias, Dutra & da Silva, 2014 [4] *	PE	
Inia boliviensis d'Orbigny, 1834	PE	
Inia geoffrensis (Blainville, 1817)	EN	EN
Family Kogiidae Gill, 1871		
***Kogia* Gray, 1846**		
Kogia breviceps (Blainville, 1838)	DD	
Kogia sima (Owen, 1866)	DD	
Family Phocoenidae Gray, 1825		
***Phocoena* Cuvier, 1816**		
Phocoena dioptrica Lahille, 1912	LC	
Phocoena spinipinnis Burmeister, 1865	NT	
Family Physeteridae Gray, 1821		
***Physeter* Linnaeus, 1758**		
Physeter macrocephalus Linnaeus, 1758	VU	VU
Family Pontoporiidae Gray, 1870		
***Pontoporia* Gray, 1846**		
Pontoporia blainvillei (Gervais & d'Orbigny, 1844)	VU	CR
Family Ziphiidae Gray, 1865		
***Berardius* Duvernoy, 1851**		
Berardius arnuxii Duvernoy, 1851	DD	
***Hyperoodon* Lacépède, 1804**		
Hyperoodon planifrons Flower, 1882	LC	
***Mesoplodon* Gervais, 1850**		
Mesoplodon densirostris (Blainville, 1817)	DD	
Mesoplodon europaeus (Gervais, 1855)	DD	
Mesoplodon grayi von Haast, 1876	DD	
Mesoplodon hectori (Gray, 1871)	DD	
Mesoplodon layardii (Gray, 1865)	DD	
Mesoplodon mirus True, 1913	DD	
***Ziphius* Cuvier, 1823**		
Ziphius cavirostris Cuvier, 1823	LC	