



BIOLOGICAL SCIENCES

Pollen morphology of family Solanaceae and its taxonomic significance

SHOMAILA ASHFAQ, MUSHTAQ AHMAD, MUHAMMAD ZAFAR, SHAZIA SULTANA, SARAJ BAHADUR, SIDRA N. AHMED, SABA GUL & MOONA NAZISH

Abstract: The pollen micro-morphology of family Solanaceae from the different phytogeographical region of Pakistan has been assessed. In this study, thirteen species belonging to ten genera of Solanaceae have been studied using light and scanning electron microscopy for both qualitative and quantitative features. Solanaceae is a eurypalynous family and a significant variation was observed in pollen size, shape, polarity and exine sculpturing. Examined plant species includes, *Brugmansia suaveolens*, *Capsicum annum*, *Cestrum parqui*, *Datura innoxia*, *Solanum lycopersicum*, *Nicotiana plumbaginifolia*, *Petunia hybrida*, *Physalis minima*, *Solanum americanum*, *Solanum erianthum*, *Solanum melongena*, *Solanum surattense* and *Withania somnifera*. The prominent pollen type is tricolporate and shed as a monad. High pollen fertility reflects that observed taxa are well-known in the study area. Based on the observed pollen traits a taxonomic key was developed for the accurate and quick identification of species. Principal Component Analysis was performed that shows some morphological features are the main characters in the identification. Cluster Analysis was performed that separate the plant species in a cluster. The findings highlight the importance of Palyno-morphological features in the characterization and identification of Solanaceous taxa. It is concluded that both LM and SEM significantly play a key role in correct identification of taxa studied.

Key words: Solanaceae, pollen morphology, principal component analysis, taxonomic significance.

INTRODUCTION

Solanaceae is a eurypalynous family including 94 genera and about 2950 species, distribution is sub cosmopolitan and more prominent in tropical America. In Pakistan, it was represented by 14 genera and 50 species. Some leading genera of the family Solanaceae are *Solanum*, *Atropa*, *Datura*, *Capsicum*, *Nicotiana*, *Lycium*, *Hyoscyamus*, *Lycopersicon* and *Petunia* (Perveen & Qaiser 2007). Solanaceae is one of the important family having unique floristic, phytochemistry, economic and ethnobotanical importance (Ahmed et al. 2019, Ashfaq et al.

2019b, Adedeji & Akinniyi 2015, Bahadur et al. 2020a, Waheed et al 2020). Palynological studies are essential to explore some aspects related to pollen class and shape, P/E ratio, polar and equatorial measurements, and pollen outline because morphological evidence sometimes to depend upon alone is not adequate to demarcate plant species. Pollen grains morphology is important for the identification of plants found in numerous conditions. Palyno-morphological data provide valuable validation about closely related genera and species (Arora & Modi 2008). Morphology of pollen is highly diverse and have

unique taxonomic importance (Tellería & Daners 2003).

Morphology of pollen grains of the family Solanaceae has been studied by different researchers (Persson et al. 1999, Plowman & Knapp 1998, Gentry 1986). Pollen morphology of the genus *Solanum* was investigated by Du et al. (2018) using light microscopy and scanning electron microscopy and demonstrated sufficient pollinic heterogeneity in their shapes, aperture feature and sexine ornamentation to enable their palynological characterization. Pollen morpho diversity of some genera viz., *Capsicum*, *Datura*, *Petunia* and *Solanum* of the family Solanaceae were investigated by Rajurkar (2017) and observed that pollen grains vary from sub-oblate, oblate-spheroidal to prolate spheroidal and striate-reticulate, striate, perforate, scabrate, psilate and granulate exine sculpturing. Murry et al (1971) reported the pollen grains of Solanaceous genera using light microscopy and statistical analysis.

Pollen morphology of seven genera viz., *Datura*, *Lycium*, *Hyoscyamus*, *Nicotiana*, *Physalis*, *Solanum* and *Withania* of the Solanaceae from Pakistan have been investigated by using light and scanning electron microscope (SEM) and reported that pollen grains were isopolar, radially symmetrical, prolate-spheroidal to oblate-spheroidal and sub-oblate to sub-prolate, tricolporate, tectal surface commonly scabrate to verrucate (Perveen & Qaisar 2007). Anil et al. 2015 investigated pollen morphology of genus *Solanum* and analyzed the palynological and evolutionary relationship shared with both wild and cultivated species.

There is no comprehensive study for the unique palyno-morphological characteristics of family Solanaceae in Pakistan. Therefore, this research project was design and conducted with the aim (1) to evaluate the taxonomic significance of pollen features of Solanaceae

taxa, (2) to enlighten the morphology of pollen grains, pollen types and their relationship within the family using light and scanning electron microscopy and (3) to develop a taxonomic key for the accurate and quick identification of Solanaceae taxa.

MATERIALS AND METHODS

Collection and identification

Thirteen Solanaceous species were collected in different flowering seasons during the period from April 2016 to May 2017. Five samples were collected from each locality randomly (Table I and Figure 1). The collected plant species were dried, preserved, poisoned and mounted on herbarium sheet following the previously published protocol of Sufyan et al. (2018). The International Plant Name Index (IPNI) was used to get the accurate botanical names authorization for taxonomic validation. The voucher specimens were submitted in the Herbarium of Pakistan (ISL), Quaid-i-Azam University Islamabad, Pakistan.

Exploration of taxonomic characters using light microscopy

According to the previously published protocol of Ashfaq et al. (2018) anthers from the flowers where separated and they were dipped into 1-2 drops of acetic acid. Finally, they were crushed and mounted on a slide and with glycerin jelly staining was done. With the help of Leica Dialup 20 light microscope, pollen morphology and several parameters were measured. The quantitative measurements were conducted by using the SPSS 16.0 software for observing minimum, mean, standard error and a maximum of each quantitative character is denoted as 37.50(38 ±0.30)38.75 and given in (Table II).

Table I. Locality and voucher numbers of the taxa studied.

Species name	Voucher No.	Locality	Altitudes (m)
<i>Brugmansia suaveolens</i> (Humb. and Bonpl. ex Willd.) Sweet	QAU-SA-501	Attock	2758
<i>Capsicum annuum</i> L	QAU-SA-702	Jhelum	234
<i>Cestrum parqui</i> (Lam.) L'Hér.	QAU-SA-143	Rawalpindi	508
<i>Datura innoxia</i> Mill	QAU-SA-434	Bhakkar	159
<i>Solanum lycopersicum</i> Mill.	QAU-SA-310	Jhelum	234
<i>Nicotiana plumbaginifolia</i> Viv.	QAU-SA-500	Attock	2758
<i>Petunia hybrida</i> M.C.Ferguson and Ottley	QAU-SA-121	Rawalpindi	508
<i>Physalis minima</i> L.	QAU-SA-117	Jhelum	234
<i>Solanum americanum</i> Mill.	QAU-SA-300	Rawalpindi	508
<i>Solanum erianthum</i> D. Don	QAU-SA-119	Bhakkar	159
<i>Solanum melongena</i> L.	QAU-SA-302	Attock	2758
<i>Solanum surattense</i> Burm. f.	QAU-SA-332	Attock	2758
<i>Withania somnifera</i> (L.) Dunal	QAU-SA-314	Mianwali	210

Scanning electron microscopy: a tool for identification

For scanning electron microscopy (SEM) the anthers were crushed in few drops of acetic acid following the method of Bahadur et al. (2018) and directly transferred to double-sided tape fixed to a stub with a gold-coated sputter and fine pipette to 150 Å. The specimens were scanned and photographed with Scanning Electron Microscope (Model JEOL JSM- 5910) installed in the Central Resource Library (CRL), Department of Physics University of Peshawar, Pakistan.

Statistical analysis

Cluster analysis

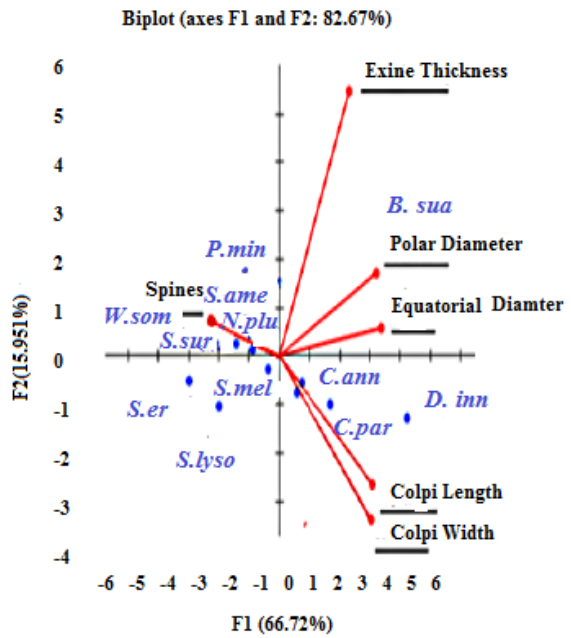
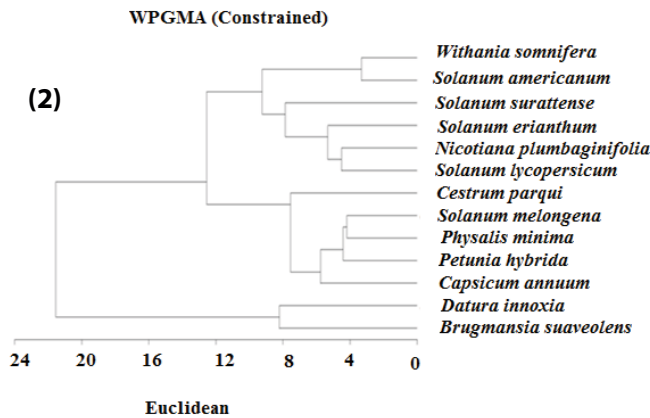
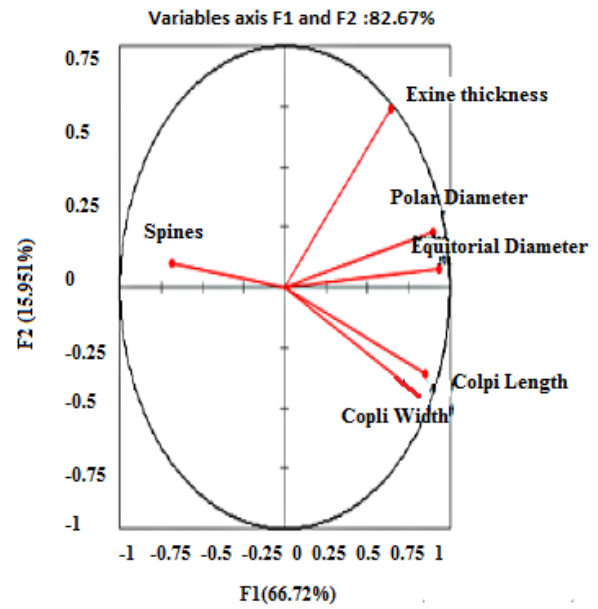
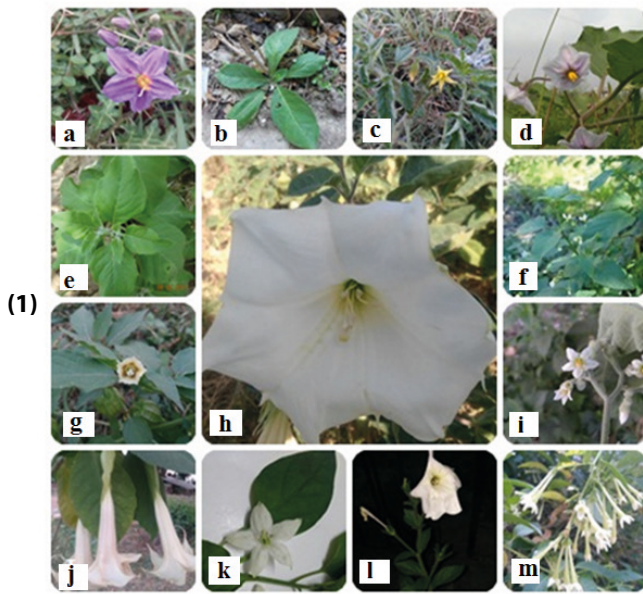
To show the relationship between the species, Cluster analysis was performed based on Euclidean Distances by using the MVSP version 3.22 statistical software.

Principal component analysis

According to (Harman 1976) Principal Component Analysis (PCA) was performed with the help of statistical software XLSTAT. Five metric variables for the PCA were used i.e polar diameter, equatorial diameter, colpi length, colpi width, and exine thickness.

RESULTS

Different palynological characters of thirteen Solanaceae taxa were examined. Pollen was measured both qualitatively and quantitatively and shown in Table II and III. Solanaceous taxa exhibited a wide range of variation in sculpturing and size showing their potential taxonomic value (Figure 2 to 5). The maximum size of the pollen was observed in *Brugmansia suaveolens* (38) μm and *Datura innoxia* (37.25) μm in polar views and minimum in *Solanum erianthum* (14.3) μm . While rest of the species are intermediate in size.



(3)

Figure 1. (1) Studied taxa of family Solanaceae (A) *Solanum surattense* (B) *Nicotiana plumbaginifolia* (C) *Solanum lycopersicum* (D) *Solanum melongena* (E) *Withania somnifera* (F) *Solanum americanum* (G) *Physalis minima* (H) *Datura innoxia* (I) *Solanum erianthum* (J) *Brugmansia suaveolens* (K) *Capsicum annum* (L) *Petunia hybrida* (M) *Cestrum parqui* (2) Cluster analyses showing the relationship among the studied Solanaceous species based on the palynological characters. (3) Principal Component Analysis (PCA) among the studied Solanaceous species based on palynological characters. B.sua: *Brugmansia suaveolens*, C.ann: *Capsicum annum*, C.par: *Cestrum parqui*, D.inn: *Datura innoxia*, L.esc: *Solanum lycopersicum*, N.plu: *Nicotiana plumbaginifolia*, P.alb: *Petunia hybrida*, P.min: *Physalis minima*, S.ame: *Solanum americanum*, S.eri: *Solanum erianthum*, S.mel: *Solanum melongena*, S.sur: *Solanum surattense*, W.som: *Withania somnifera*.

Table II. Quantitative data for the pollen of Solanaceous species.

Species name	Polar axis (μm)	Equatorial diameter(μm)	P/E ratio	No of Colpi	Colpi		Exine thickness (μm)
					Length (μm)	Width(μm)	
<i>Brugmansia suaveolens</i>	37.50(38 \pm 0.30) 38.75	40(40.75 \pm 0.50) 42.50	1.14	3	5(5.25 \pm 0.25) 6.25	10(10.25 \pm 0.25) 11.25	1.5-2
<i>Capsicum annum</i>	22.50(23 \pm 0.30) 23.75	30(30.75 \pm 0.50) 32.50	0.83	3	5(5.25 \pm 0.25) 6.25	10(10.25 \pm 0.25) 11.25	0.8-1
<i>Cestrum parqui</i>	25(25.75 \pm 0.5) 27.50	32.50(34.75 \pm 0.61) 36.25	0.83	3	5(5.25 \pm 0.25) 6.25	13.75(14.50 \pm 0.30) 15	0.8-1
<i>Datura innoxia</i>	36.25(37.25 \pm 0.25) 37.50	40(42 \pm 0.63) 43.75	0.09	3	8.75(9.25 \pm 0.30) 10	16.25(17.25 \pm 0.25) 17.50	1-1.1
<i>Lycopersicon esculantum</i>	15(15.55 \pm 0.33) 16.50	17.50(17.75 \pm 0.25) 18.75	1.05	3	2.50(3.45 \pm 0.24) 3.75	7.50(7.75 \pm 0.25) 8.75	0.5-0.6
<i>Nicotiana plumbaginifolia</i>	17.50(17.75 \pm 0.25) 18.75	20(21.25 \pm 0.55) 22.50	0.91	3-4	5(5.1 \pm 0.1) 5.50	6.25(7.15 \pm 0.24) 7.50	0.8-1
<i>Petunia hybrida</i>	27.50(28 \pm 0.30) 28.75	27.50(29.25 \pm 0.75) 31.25	1.2	3	5(6 \pm 0.46) 7.50	7.50(10.75 \pm 1.08) 12.50	0.7-0.8
<i>Physalis minima</i>	27.50(27.75 \pm 0.25) 28.75	27.50(28 \pm 0.30) 28.75	1.08	3	2.50(3 \pm 0.3) 3.75	7.50(7.6 \pm 0.1) 8	1-1.2
<i>Solanum americanum</i>	22.50(23 \pm 0.30) 23.75	22.50(22.75 \pm 0.25) 23.75	1.6	3	2.50(2.90 \pm 0.25) 3.75	6.50(7.15 \pm 0.21) 7.50	1-1.1
<i>Solanum erianthum</i>	13.50(14.3 \pm 0.31) 15	21.25(22.25 \pm 0.25) 22.50	1.64	3	1(1.2 \pm 0.5) 1.25	6.25(6.65 \pm 0.25) 7.50	0.4-0.5
<i>Solanum melongena</i>	25(26 \pm 0.46) 27.50	25(25.75 \pm 0.50) 27.50	1.28	3	5(5.4 \pm 0.27) 6.25	8.75(9.5 \pm 0.30) 10	0.8-0.9
<i>Solanum surattense</i>	16.25(17.25 \pm 0.25) 17.50	27.50(28.10 \pm 0.37) 29.25	0.78	3	2.50(2.75 \pm 0.25) 3.75	7.50(7.75 \pm 0.25) 8.75	0.9-1
<i>Withania somnifera</i>	25(25.45 \pm 0.27) 25.25	20(22 \pm 0.5) 22.50	1.57	3	2.50(2.75 \pm 0.15) 3.25	8.75(9.25 \pm 0.3) 10	0.8-0.9

The polar and equatorial relationship (P/E ratio) was also noted. *Datura innoxia* is characterized by a low P/E ratio 0.09, while that of *Solanum erianthum* is high 1.64. The exine thickness was highest in *Brugmansia suaveolens* (1.5-2) μm , *Datura innoxia*, *Solanum americanum* (1-1.1) μm and *Physalis minima* (1-1.2) μm . Tricolporate pollen was noted as a prominent feature among the Solanaceous flora. Subspheroidal pollen shape was observed in *Brugmansia suaveolens*, *Capsicum annum*, *Cestrum parqui*, *Datura innoxia*, *Lycopersicon esculantum*, *Petunia hybrida*, *Physalis minima*, *Solanum melongena*, and *Solanum surattense*.

Prolate shape pollen was noted in *Solanum americanum*, *Solanum erianthum*, and *Withania somnifera*, while peroblate shape was observed in *Nicotiana plumbaginifolia* (Table III).

Large size colpi length was noted in *Datura innoxia* (9.25) μm and width is (17.25) μm . While small size colpi were observed in *Solanum erianthum* (1.2) μm and width are (6.65) μm respectively. Pollen fertility percentage was also noted for the Solanaceous taxa. Fertility levels show the highest percentage of fertility in *Withania somnifera* 90% and lowest in *Capsicum annum* 69% (Table IV).

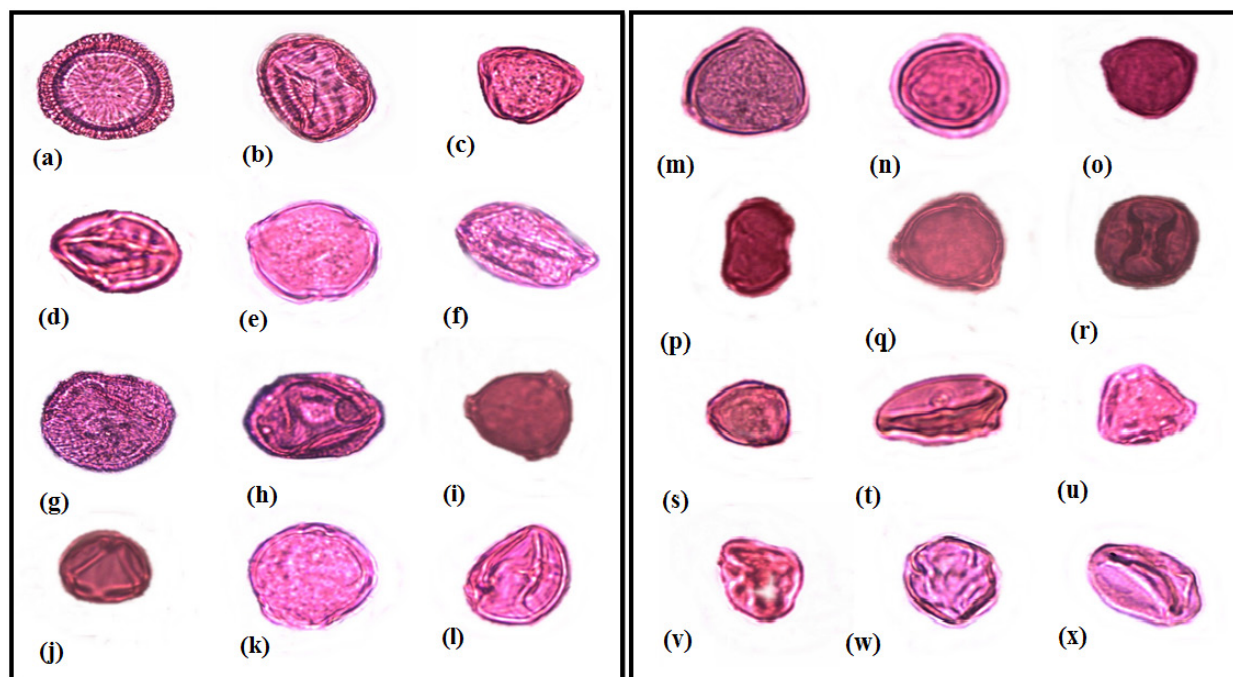


Figure 2. Light microscope photomicrographs of Solanaceae pollen grains. PV = polar view; EV =equatorial view (a) *Brugmansia suaveolens* (PV), (b) *Brugmansia suaveolens* (EV), (c) *Capsicum annuum* (PV) general view (d) *Capsicum annuum* (EV), (e) *Cestrum parqui* (PV), (f) *Cestrum parqui* (EV), (g) *Datura innoxia* (PV), (h) *Datura innoxia* (EV), (i) *Solanum lycopersicum*(PV), (j) *Solanum lycopersicum* (EV), (k) *Physalis minima* (PV), (l) *Physalis minima* (EV), (m) *Solanum americanum* (PV), (n) *Solanum americanum* (EV), (o) *Solanum erianthum* (PV), (p) *Solanum erianthum* (EV), (q) *Solanum melongena* (PV), (r) *Solanum melongena* (EV), (s) *Solanum surattense* (PV), (t) *Solanum surattense* (EV), (u) *Withania somnifera* (PV), (v) *Withania somnifera* (EV), (w) *Nicotiana Plumbaginifolia* (PV), (x) *Petunia× alba* (EV).

The Solanaceous taxa studied were separated by using cluster analysis WPGMA. Cluster analysis represents the similarity of species based on anatomical characters (Figure 1).

The dendrogram distributed the species into three major clusters. C1 (cluster 1) includes species *Datura innoxia* and *Brugmansia suaveolens* which entirely distinct from other species. C2 (cluster II) contained *Cestrum parqui*, *Solanum melongena*, *Physalis minima*, *Petunia hybrida*, *Capsicum annuum* whereas the third cluster retains *Withania somnifera*, *Solanum americanum*, *Solanum surattense*, *Solanum erianthum*, *Nicotiana plumbaginifolia*, *Solanum lycopersicum*. The highest similarities were observed among *Solanum melongena*,

Physalis minima in C2 (cluster 2), *Nicotiana plumbaginifolia* and *Solanum lycopersicum* in C3 (cluster 3).

Principal Component Analysis (PCA) reveals the maximum contribution to the total variability at each axis (Figure 1). The Eigen values characterize the total number of variable which is often used to ascertain that how many factors to hold. In the current study, three PC's limited more than one eigen value and 93.10% of total variation (Table V). Moreover, PC1 demonstrated 66.721 % of variability which had high positive loading component of the polar axis, equatorial diameter, colpi length, colpi width and exine thickness whereas negative loading of species. Second PC (PC-II) accounted 15.951% of the variation and strongly positive associated with species, polar axis, and equatorial diameter

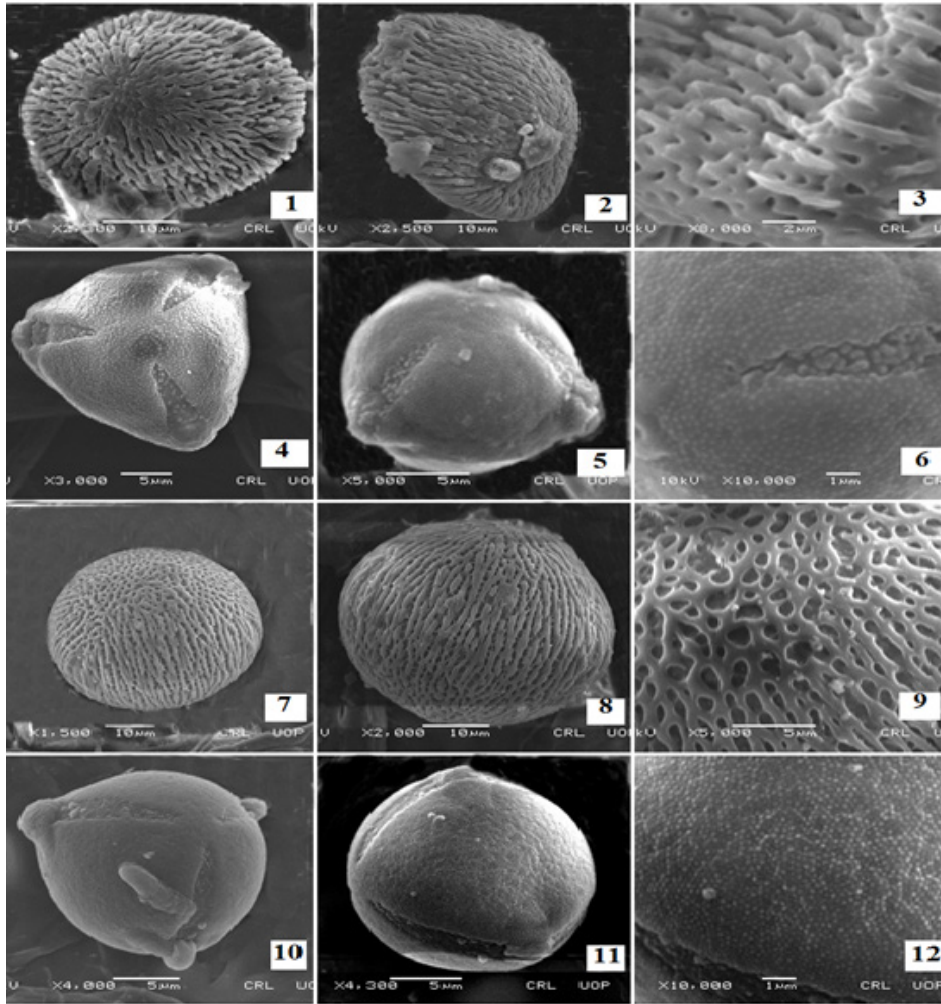


Figure 3. S Scanning electron microscope photomicrographs (SEM) of Solanaceous pollen grains. The scale bar represents 5 and 10µm for whole pollen grains and 1µm for detailed exine ornamentation. PV = polar view; EV =equatorial view; ES= exine sculpture. (1) *Brugmansia suaveolens* (PV), (2)*Brugmansia suaveolens* (EV), (3) *Brugmansia suaveolens* (ES). (4) *Capsicum annuum* (PV), (5) *Capsicum annuum* (EV), (6) *Capsicum annuum* (ES). (7) *Datura innoxia* (PV), (8)*Datura innoxia* (EV), (9) *Datura innoxia* (ES). (10)*Solanum lycopersicum*(PV), (11) *Solanum lycopersicum*(EV), (12) *Solanum lycopersicum*(ES).

and exine thickness whereas the third PC (PCIII) contained 10.433% variability in which species, polar axis, equatorial diameter, colpi length, and width were important.

Taxonomic key based on pollen micro-morphological features

- 1. Striate-reticulate exine
- 2. Pollen small, circular, oval to elliptic shaped, P/E ratio is 1.14..... *Brugmansia suaveolens*
- 1'. Rugulate and gemmate exine
- 2'. Pollen small, triangular shaped, P/E ratio is 1.57..... *Withania somnifera*
- 3. Micro-echinate and perforate exine

- 4. Pollen medium size, elliptic shaped *Capsicum annuum*
- 3'. Perforate, verrucate, and psilate exine
- 4'. Pollen medium size, circular shaped..... *Cestrum parqui*
- 5. Exine striate and reticulate, reticulate arrange in series to form long striation
- 6. P/E ratio is 0.09.....*Datura innoxia*
- 5'. Rugulate and perforate exine
- 6'. P/E ratio is 0.91.....*Nicotiana plumbaginifolia*
- 7. Exine striate and reticulate
- 8. Pollen trilobate shaped, P/E ratio is 1.2 *Petunia hybrida*
- 7'. Scabrate and micro-echinate exine

Table III. Qualitative characters for pollens of Solanaceous species from arid zone of Pakistan.

Species name	Pollen Size	Pollen Shapes	amb	Pollen type	Pollen description
<i>Brugmansia suaveolens</i>	Medium	Subspheroidal	Circular and oval to elliptic shaped	Tricolporate	Striate-reticulate exine. Each striae are long, broad and arranged in series to form a network like structure. having psilate and perforate lumina. It was sparsely scabrate at the apocolpium and lobed at the mesocolpium
<i>Capsicum annum</i>	Medium	Subspheroidal	Circular, triangular, elliptic	Tricolporate	Pollen shed as monad, exine micro-echinate, scabrate and perforate. Pollen outline lobate in polar view. Aperture are sunken having scabrate and gemmate surface. whereas surface of the oriented lobe at the mesocolpium was psilate to sparsely scabrate.
<i>Cestrum parqui</i>	Medium	Subspheroidal	Circular in polar view	Tricolporate	Pollen shed as monad, exine perforate, verrucate, and psilate.
<i>Datura innoxia</i>	Medium	Subspheroidal	Circular	Tricolporate	Pollen shed as monad, exine striate-reticulate, each striae are narrow and arrange in series to form a network like structure. However it was reticulate at the polar area. Surface of the colpus was scabrate to sparsely gemmate
<i>Lycopersicon esculantum</i>	Small	Subspheroidal	Circular, lobate, elliptic shaped.	Tricolporate	Exine scabrate, and sparsely gemmate. colpus was long having pointed tip. Surface of the colpus was scabrate to verrucate while oriented lobe have psilate surface at the mesocolpium
<i>Nicotiana plumbaginifolia</i>	Small	Peroblate	Circular to lobate	Tri and tetra-colporate	Pollen unit monad. Exine rugulate, perforate. Surface of the colpus verrucate, and surface of the lobe at the mesocolpium was psilate to scabrate.
<i>Petunia hybrida</i>	Medium	Subspheroidal	Elliptic shaped in equatorial view, circular and trilobate in polar view.	Tricolporate	Pollen shed as monad and lobate outline. Exine striate, reticulate. Each striae are long and narrow. It was rugulate along the colpus area and psilate, rugulate at the apocolpium. Surface of the colpus was verrucate, gemmate and scabrate. While surface of the lobe at the mesocolpium was psilate to rugulate.

Table III. (continuation)

Species name	Pollen Size	Pollen Shapes	amb	Pollen type	Pollen description
<i>Physalis minima</i>	Medium	Subspheroidal	Circular shape in polar view and elliptic shape in equatorial.	Tricolporate	Pollen unit monad, exine microechinate, scabrate. Surface of the colpus psilate and sparsely scabrate while tip of the colpus obtuse.
<i>Solanum americanum</i>	Small	Prolate	Circular shape in polar view and elliptic shape in equatorial view	Tricolporate	Pollen outline lobate and shed as monad. Exine scabrate. Colpus was long and wide having pointed tip. surface of the colpus densely gemmate to verrucate while surface of the lobe was psilate to gemmate.
<i>Solanum erianthum</i>	Small	Prolate	Circular	Tricolporate	Pollen unit monad, exine micro-echinate, scabrate and verrucate. Colpus was long and narrow having pointed tip. While surface of the colpus was gemmate, verrucate and scabrate.
<i>Solanum melongena</i>	Small	Subspheroidal	Circular, rectangular and elliptic shaped	Tricolporate	Exine thin, scabrate and perforate. Colpus was long and wide having obtuse end. while well oriented lobed at mesocolpium having densely verrucate and gemmate surface.
<i>Solanum surattense</i>	Medium	Subspheroidal	Circular shaped in polar view elliptic shaped in equatorial view	Tricolporate	Pollen shed as monad and exine scabrate to psilate. Colpus was long, narrow, sunken and lobed at mesocolpium.
<i>Withania somnifera</i>	Small	Prolate	Triangular in polar view	Tricolporate	Exine rugulate and gemmate. Colpus was long and very wide with well develop lobe at mesocolpium.

8'. Pollen circular to elliptic shaped, P/E ratio is 1.08..... *Physalis minima*

9. Micro-echinate and verrucate exine

10. Pollen circular shaped, P/E ratio is 1.64..... *Solanum erianthum*

9'. Scabrate exine

10'. Pollen elliptic shaped, P/E ratio is 1.6 *Solanum americanum*

11. Lobe depressed having scabrate to psilate surface

12. Pollen elliptic shaped, P/E ratio is 0.78 *Solanum surattense*

11'. Lobe well oriented having densely verrucate and gemmate surface

12'. Pollen rectangular shaped, P/E ratio is 1.28 *Solanum melongena*

13. Exine gemmate..... *Solanum lycopersicum*

DISCUSSION

Palyno-morphological characters show heterogeneous features. Tricolporate pollen type is a prominent feature in the family Solanaceae,

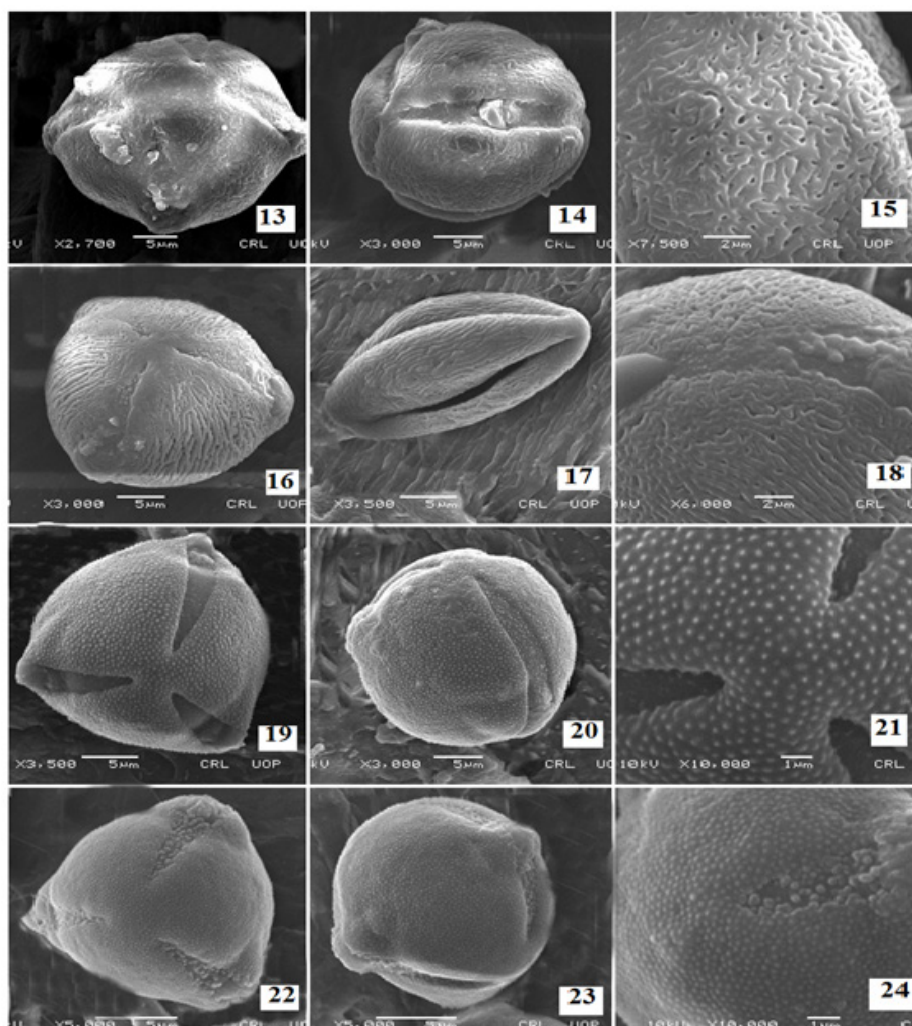


Figure 4. Scanning electron microscope photomicrographs of Solanaceous pollen grains. The scale bar represents 5 and 10 μ m for whole pollen grains and 1 μ m for detailed exine ornamentation. PV = polar view; EV = equatorial view; ES = exine sculpture. (13) *Nicotiana Plumbaginifolia* (PV), (14) *Nicotiana Plumbaginifolia* (EV), (15) *Nicotiana Plumbaginifolia* (ES). (16) *Petunia alba* (PV), (17) *Petunia alba* (EV), (18) *Petunia alba* (ES). (19) *Physalis minima* (PV), (20) *Physalis minima* (EV), (21) *Physalis minima* (ES). (22); *Solanum americanum* (PV), (23) *Solanum americanum* (EV), (24) *Solanum americanum* (ES).

however, pollen grains are typically isopolar, radially symmetrical, prolate- spheroidal to sub prolate. Generally, tricolporate pollen grains having a coarsely granulated colpal membrane. Different type of tectum was found i.e reticulate, regulate, verrucate and striate. In Solanaceous taxa, scabrate tectum with tricolporate pollen was observed. However, conspicuous variation was noted in pollen shape, class, type, aperture and tectal membrane. Smallest pollen size of pollen grains supports to the fact that pollens are wind pollinated rather than flowers pollinated (Barbola et al. 2006).

Significant variation was observed in micromorphology of the pollen. Pollen

morphology of Solanaceae is obscure. Sometimes different tribes or subtribes have a similar type of pollen or vice versa genera referred to same tribe or subtribe may have a different type of pollen (Erdtman 1952). For instance, two species of the genus *Withania* i.e., *W. coagulans* and *W. somnifera*, have somewhat dissimilar pollen grains and fall under different types. *Withania* (*W. coagulans* and *W. somnifera*) similarity affinities with the other genera than with each other. Pollen of *W. coagulans* is very similar to grains of genus *Solanum*, while *W. somnifera*, pollen is closely related to *Lycium* grains. Eurypalynous nature of subfamily Solanoideae

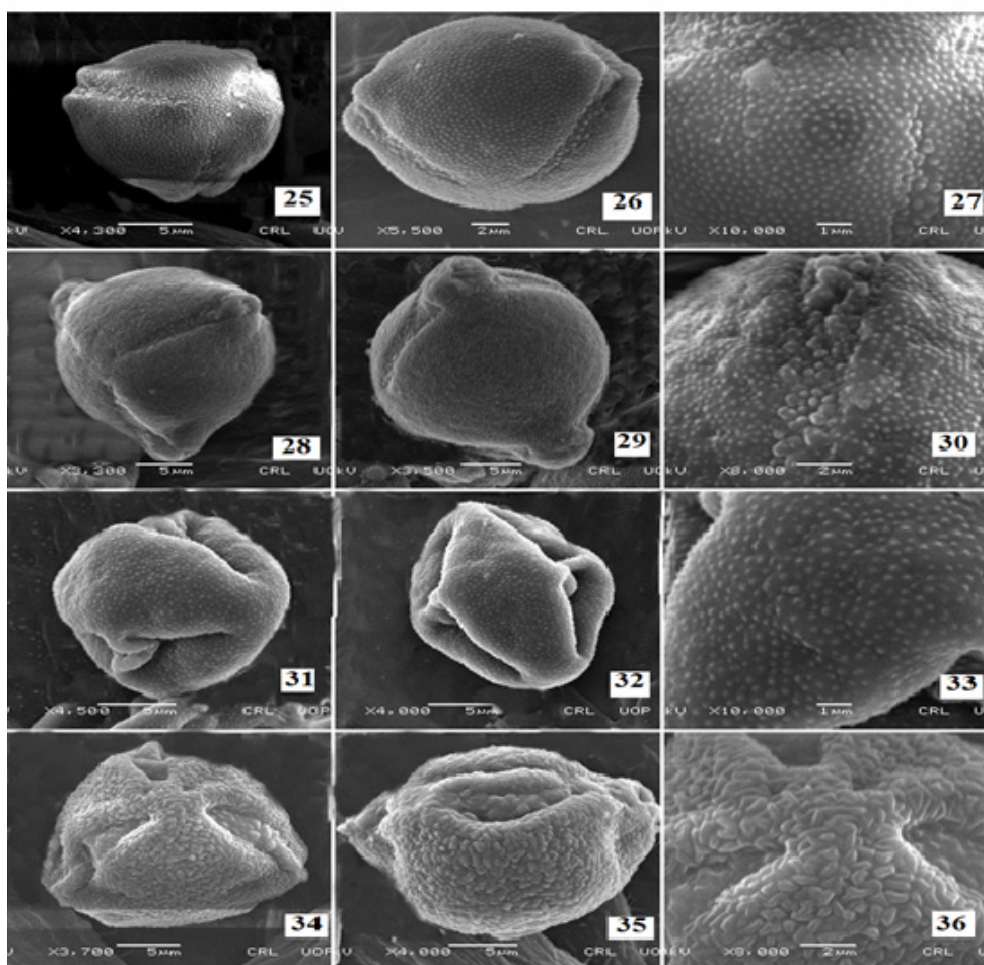


Figure 5. Scanning electron microscope photomicrographs of Solanaceous pollen grains. The scale bar represents 5 and 10 μ m for whole pollen grains and 1 μ m for detailed exine ornamentation. PV = polar view; EV = equatorial view; ES = exine sculpture. (25) *Solanum erianthum* (PV), (26) *Solanum erianthum* (EV), (27) *Solanum erianthum* (ES). (28) *Solanum melongena* (PV), (29) *Solanum melongena* (EV), (30) *Solanum melongena* (ES). (31) *Solanum surattense* (PV), (32) *Solanum surattense* (EV), (33) *Solanum surattense* (ES). (34) *Withania somnifera* (PV), (35) *Withania somnifera* (EV), (36) *Withania somnifera* (ES).

suggests its polyphyletic origin that supports (D'Arcy 1978, 1991).

According to Hunziker (1979), Solanaceae is divided into 2 sub families, Solanoideae and Cestoideae. However, the more recent study of subfamily Solanoideae by (Olmstead et al. 1999, Knapp 2002) indicates that the Solanoideae is monophyletic and derived from the Cestoideae. Only a single genus *Nicotiana* of subfamily Cestoideae having different mesocolpium and apocolpium pattern but not as diverse as of *Datura*. Subpsilate apocolpium and fossulate to the rugulose mesocolpium region are found in *Nicotiana*. Reported highest fertility percentage shows that the species is well adopted in the

area. Perveen & Qaiser (2007) investigated seven genera of family Solanaceae including *Datura Capsicum*, *Solanum* and *Petunia* species. The investigated pollen was medium size oblate spheroidal, radially symmetrical, prolate- spheroidal. Which are quite like our present findings. Vijayakumari & Vilasini (2005) expose a trizonocolporate and granulate exine ornamentation in *Solanum melongena*. In present findings, *Solanum melongena* shows thin exine, scabrate and perforate. Colpus was long and wide having an obtuse end, while well oriented lobed at mesocolpium having densely verrucate and gemmate surface.

Table IV. Pollen fertility percentages for Solanaceous species from arid zone of Pakistan.

Species name	Fertile Pollen	Sterile Pollen	Fertility (%)
<i>Brugmansia suaveolens</i>	70	15	82
<i>Capsicum annuum</i>	90	40	69
<i>Cestrum parqui</i>	77	10	88
<i>Datura innoxia</i>	99	12	89
<i>Solanum lycopersicum</i>	88	22	80
<i>Nicotiana plumbaginifolia</i>	55	9	86
<i>Petunia hybrida</i>	70	13	84
<i>Physalis minima</i>	72	10	88
<i>Solanum americanum</i>	87	20	81
<i>Solanum erianthum</i>	80	17	82
<i>Solanum melongena</i>	54	11	83
<i>Solanum surattense</i>	90	21	81
<i>Withania somnifera</i>	86	9	90

Table V. Correlation coefficients for metric variables of principal component analysis (PCA). Ordination in for Solanaceous species.

	PC1	PC2	PC3	PC4	PC5	PC6
Species	-0.690	0.102	0.712	0.031	-0.081	-0.011
Polar diameter	0.895	0.232	0.266	0.038	0.269	-0.030
Equitorial diameter	0.938	0.079	0.069	-0.275	-0.126	-0.134
Colpi Length	0.852	-0.360	0.055	0.359	-0.095	-0.059
Colpi Width	0.840	-0.456	0.199	-0.147	-0.024	0.159
Exine thicknes	0.645	0.741	-0.039	0.097	-0.121	0.094
Eigenvalue	4.003	0.957	0.626	0.238	0.119	0.057
Variability (%)	66.721	15.951	10.433	3.966	1.986	0.943
Cumulative %	66.721	82.671	93.104	97.071	99.057	100.000

Cluster analysis presented the similarities of species based on their palynological characters. Attained cluster also supported the taxonomic key. Cluster analysis helps to identify the distinctness and similarities among different species which can be useful to determine the phylogeny. Cluster analysis showed the similarity of species within the family based on their palynological characters. In our

results, the similarities were observed between *Withania somnifera*, *Solanum americanum*, *Nicotiana plumbaginifolia*, *Solanum lycopersicum*, *Solanum melongena* and *Physalis minima* while distinctness was observed in *Datura innoxia* and *Brugmansia suaveolens*. Similarly, Shah et al. (2018) characterized Dryopteridaceae Family into five clusters based on foliar morphology and stomatal traits.

According to Sharma & Paliwal (2007), Principal Component Analysis (PCA) reflects the maximum contribution to the total variability at each axis. The Eigen values represent the total number of variable which is often used to ascertain that how many factors to retain. PCA was based on both the qualitative and quantitative data analysis, which shows a highly dominant variation of 66% to 82%. Both PCA and Cluster analysis used as a statistical method for the separation of species and genera (Shah et al. 2018). In the present study, we used the PCA and Cluster analysis of Pollen Morphology of Solanaceous species. A similar analysis was done on leaf anatomy of many plants group (Ozcan 2017, De Luna et al. 2017).

The observed pollen micromorphological features are taxonomically significant examined under SEM to identify Solanaceae taxa at the micro level. Similarly, Amina et al. 2020, Arshad et al. 2019, Ashfaq et al. 2018, 2019a, Bahadur et al. 2018a, 2018b, 2020b, Bano et al. 2018, Gul et al. 2019a, b, Kosenko 1999, Naz et al. 2019, Nazish et al. 2019, Qureshi et al. 2019, Rashid et al. 2018, Sufyan et al. 2018 and Ullah et al. 2018 investigated pollen of various taxa, using light and scanning electron microscopy techniques and reported that the pollen traits were of important characters for the taxonomic identifications of species and genera in different plant families. Furthermore, phytochemical, antimicrobial, and molecular studies are recommended to strengthen the knowledge about the family Solanaceae in detail (Azam et al. 2020, Hussain et al. 2019, Naeem et al. 2020, Rubab et al. 2020, Sajad et al. 2020, Sarah et al. 2019).

CONCLUSION

Pollen micromorphology of thirteen Solanaceous species provides significant taxonomic information about the qualitative and quantitative characteristics. A significant various was observed in pollen morphological features. Palyno-morphological characteristics considered to be a very supportive tool at taxonomic level. Based on the diagnostic characteristics of the pollen a taxonomic key was developed that is considered supportive in the delimitation of Solanaceous taxa. The observed pollen features through LM and SEM is a good source of taxonomic information that can help the species and genera delimitation, identification and strengthen their systematic position. The main purposes of the study are to expand the pollen morphological study thus, this study contributes to taxonomic aspects and subsequent knowledge of the family Solanaceae from Pakistan. So far, micromorphological description of other Pakistani Solanaceae taxa is recommended to use these characters in a broader taxonomic and evolutionary context. It is further suggested that chemotaxonomically and DNA characteristics of Solanaceous taxa will be more useful to develop monograph to solve the taxonomic problem for the correct identification of the species.

Acknowledgments

The authors are very grateful to the Central Resource Library (CRL) Department of Physics University of Peshawar for providing the facility of scanning electron microscopy. No funding sources.

REFERENCES

ADEDEJI O & AKINNIYI TA. 2015. Pollen morphology of some species in the family Solanaceae. *J Adv Lab Res Biol* 6(4): 125-129.

- AHMED SN, AHMAD M, ZAFAR M, RASHID S, YASEEN G, ASHFAQ S & SULTANA S. 2019. Taxonomical and phytochemical characterization of two highly traded medicinal species of genus berberis. *Proceedings of the Pakistan Academy of Sciences. B Life Environ Sci* 56(3): 121-125.
- AMINA ET AL. 2020. Microscopic investigation of pollen morphology of Brassicaceae from central Punjab-Pakistan. *Microsc Res Tech* 83(4): 446-454.
- ANIL KUMAR VS, MAYA NAIR C & MURUGAN K. 2015. Pollen morphology of selected taxa of the genus *Solanum* from Southern Western Ghats, Kerala, India. *Rheedea* 25(2): 128-145.
- ARORA A & MODI A. 2008. An acetolysis technique for pollen slide preparation. *Indian J Microbiol* 21(2): 90-91.
- ARSHAD S ET AL. 2018. Role of trees in climate change and their authentication through electron microscopy. *Microsc Res Tech* 82(2): 92-100.
- ASHFAQ S, AHMAD M, ZAFAR M, SULTANA S, BAHADUR S & ABBAS N. 2019b. Medicinal plant biodiversity used among the rural communities of arid regions of northern Punjab, Pakistan. *Indian J Tradit Knowl* 18(2): 226-241.
- ASHFAQ S ET AL. 2019a. Foliar micromorphology of Convolvulaceae species with special emphasis on trichome diversity from the arid zone of Pakistan. *Flora* 255: 110-124.
- ASHFAQ S, ZAFAR M, AHMAD M, SULTANA S, BAHADUR S, KHAN A & SHAH A. 2018. Microscopic investigations of palynological features of convolvulaceous species from arid zone of Pakistan. *Microsc Res Tech* 81(2): 228-239.
- AZAM ET AL. 2020. Isolating Soil-Born fungi and determining their phytotoxicity against weeds in millet. *Pol J Environ Stud* 29(3): 2055-2062.
- BAHADUR S ET AL. 2018a. Palyno-anatomical studies of monocot taxa and its taxonomic implications using light and scanning electron microscopy. *Microsc Res Tech* 82(4): 372-393.
- BAHADUR S ET AL. 2020a. Traditional usage of medicinal plants among the local communities of Peshawar valley, Pakistan. *Acta Entomol Sin* 40(1): 1-29.
- BAHADUR S ET AL. 2020b. Taxonomic study of one generic and two new species record to the flora of Pakistan using multiple microscopic techniques. *Microsc Res Tech* 83(4): 345-353.
- BAHADUR S, AHMAD M, MIR S, ZAFAR M, SULTANA S, ASHFAQ S & ARFAN M. 2018b. Identification of monocot flora using pollen features through scanning electron microscopy. *Microsc Res Tech* 81(6): 599-613.
- BANO A ET AL. 2018. Microscopic investigations of some selected species of Papilionaceae through SEM and LM from Skardu valley, northern Pakistan. *Microsc Res Tech* 82(4): 452-458.
- BARBOLA IF, LAROCA S, ALMEIDA MCD & NASCIMENTO ED. 2006. Floral biology of *Stachytarpheta maximiliani* Scham.(Verbenaceae) and its floral visitors. *Rev Bras Entomol* 50(4): 498-504.
- D'ARCY WG. 1978. A preliminary synopsis of Salpiglossis and other Cestreae (Solanaceae). *Ann Mo Bot Gard* 65(2): 698-724.
- D'ARCY WG. 1991. The Solanaceae since 1976, with a review of its biogeography. In: Hawkes JG, Lester RN, Nee M & Estrada N (Eds), *Solanaceae III Taxonomy, chemistry, evolution* Roy Bot Gard: Kew and Linnean Soc: London: 75-137.
- DE LUNA BN, DE FÁTIMA FREITAS M, BAAS P, DE TONI KLG & BARROS C. 2017. Leaf anatomy of five neotropical genera of Primulaceae. *Int J Plant Sci* 178(5): 362-377.
- GENTRY J. 1986. Solanaceae: Biology and Systematics. In: D'Arcy WG (Ed), *Pollen studies in the Cestreae (Solanaceae)*. New York: Columbia University Press, 138-158.
- GUL S ET AL. 2019a. Foliar epidermal anatomy of Lamiaceae with special emphasis on their trichomes diversity using scanning electron microscopy. *Microsc Res Tech* 82(3): 206-223.
- GULETAL. 2019b. Taxonomic significance of foliar epidermal morphology in Lamiaceae from Pakistan. *Microsc Res Tech* 82(9): 1507-1528.
- HARMAN HH 1976. *Modern factor analysis*. University of Chicago Press. Linnean Society symposium series, 1979 p.
- HUSSAIN F, SHAH SZ, SHUAIB M, BAHADUR S & MUHAMMAD I. 2019. Optimization conditions for native microalgal strains grown on high ammonia-containing wastewater and their biomass utilization. *Limnol Review* 19(4): 191-198.
- KNAPP S. 2002. Tobacco to tomatoes: a phylogenetic perspective on fruit diversity in the Solanaceae. *J Exp Bot* 53(377): 2001-2022.
- KOSENKO VN. 1999. Contributions to the pollen morphology and taxonomy of the Liliaceae. *Grana* 38(1): 20-30.
- MURRY LE & ESHBAUGH WH 1971. A palynological study of the Solaninae (Solanaceae). *Grana* 11(2): 65-78.
- NAEEM I ET AL. 2020. Feasible regeneration and agro bacterium-mediated transformation of *Brassica juncea*

with *Euonymus alatus* diacylglycerol acetyltransferase (EaDacT) gene. Saudi J Biol Sci 27(5): 1324-1332.

NAZ ET AL. 2019. Palynological investigation of lactiferous flora (Apocynaceae) of district Rawalpindi, Pakistan, using light and scanning electron microscopy. Microsc Res Tech 82(9): 1410-1418.

NAZISH M ET AL. 2019. Palyno-morphological investigations of halophytic taxa of Amaranthaceae through SEM from Salt range of Northern Punjab, Pakistan. Microsc Res Tech 82(3): 304-316.

OLMSTEAD RG, SWEERE JA, SPANGLER RE, BOHS L & PALMER JD. 1999. Phylogeny and provisional classification of the Solanaceae based on chloroplast DNA. Sol IV 1(1): 1-137.

OZCAN M. 2017. Cypselia micromorphology and anatomy in *Cirsium sect. Epitrichys* (Asteraceae, Carduoideae) and its taxonomic implications. Nord J Bot 35(6): 653-668.

PERSSON V, KNAPP S & BLACKMORE S. 1999. Pollen morphology and the phylogenetic analysis of *Datura* and *Brugmansia*. In: Nee M, Symon DE, Lester RN & Jessop JP (Eds), Solanaceae IV Advances in biology and utilization Royal Botanic Gardens, Kew: Kew, 171-187.

PERVEEN A & QAISER M. 2007. Pollen morphology of family Solanaceae from Pakistan. Pak J Bot 39(7): 2243-2256.

PLOWMAN T & KNAPP S. 1998. Revision of the South American species of *Brunfelsia* (Solanaceae). J. R. Press. Field Museum of Natural History, Lake Shore, USA

QURESHI MN, TALHA N, AHMAD M, ZAFAR M & ASHFAQ S. 2019. Morpho-palynological investigations of natural resources: A case study of Surghar mountain district Mianwali Punjab, Pakistan. Microsc Res Tech 82(7): 1047-1056.

RASHD S ET AL. 2018. Microscopic investigations and pharmacognostic techniques used for the standardization of herbal drug *Nigella sativa* L. Microsc Res Tech 81(12): 1443-1450.

RUBAB ET AL. 2020. Neuropharmacological potential of various morphological parts of *Camellia sinensis* L. Saudi J Biol Sci 27(1): 567-573.

SAJAD MA ET AL. 2020. Evaluation of chromium phytoremediation potential of some plant species of Dir Lower, Khyber Pakhtunkhwa, Pakistan. Acta Ecol Sin 40(2): 158-165.

SARAH ET AL. 2019. Symbiotic response of three tropical maize varieties to Eco-friendly Arbuscular mycorrhizal fungal inoculation in Marginal soil. Bio Cell 43(5-1): 245-252.

SHAH SN ET AL. 2018. A light and scanning electron microscopic diagnosis of leaf epidermal morphology and its systematic implications in Dryopteridaceae: Investigating 12 Pakistani taxa. Micron 111(1): 36-49.

SHARMA A & PALIWAL KK. 2007. Fast principal component analysis using fixed-point algorithm. Pattern Recognit Lett 28(10): 1151-1155.

SUFYAN M, BADSHAH I, AHMAD M, ZAFAR M, BAHADUR S & RASHID N. 2018. Identification of medicinally used flora using pollen features imaged in the scanning electron microscopy in the lower Margalla Hills Islamabad Pakistan. Microsc Microanal 24(3): 292-299.

TELLERÍA MC & DANERS G. 2003. Pollen types in Southern new world Convolvulaceae and their taxonomic significance. Plant Syst Evol 243(1-2): 99-118.

ULLAH F ET AL. 2018. Pollen morphology of subfamily Caryophylloideae (Caryophyllaceae) and its taxonomic significance. Microsc Res Tech 81(7): 704-715.

WAHEED K, MUHAMMAD SK, SHOMAILA A, MUHAMMAD Z, IZHAR U & ULLAH S. 2020. Antimicrobial activity and phytochemical screening of *Euphorbia helioscopia*. Planta Dani 38(1): 1-11.

How to cite

ASHFAQ S, AHMAD M, ZAFAR M, SULTANA S, BAHADUR S, AHMED SN, GUL S & NAZISH M. 2020. Pollen morphology of family Solanaceae and its taxonomic significance. An Acad Bras Cienc 92: e20181221. DOI 10.1590/0001-3765202020181221.

Manuscript received on December 17, 2018;
accepted for publication on February 11, 2019

SHOMAILA ASHFAQ¹

<https://orcid.org/0000-0002-2892-2467>

MUSHTAQ AHMAD¹

<https://orcid.org/0000-0003-2971-2848>

MUHAMMAD ZAFAR¹

<http://orcid.org/0000-0003-2002-3907>

SHAZIA SULTANA¹

<https://orcid.org/0000-0002-3076-6096>

SARAJ BAHADUR¹

<https://orcid.org/0000-0002-5496-7861>

SIDRA N. AHMED²

<https://orcid.org/0000-0003-4515-6298>

SABA GUL¹

<https://orcid.org/0000-0002-3378-9509>

MOONA NAZISH¹

<https://orcid.org/0000-0001-5486-5470>

¹Quaid-i-Azam University, Department of Plants Sciences, Faculty of Plant Sciences, University Road, 45320 Islamabad, Pakistan

²The Women university Multan, Department of botany, Faculty of Botany, 66000 Multan, Pakistan

Correspondence to: **Shomaila Ashfaq**

E-mail: shomaila_ashfaq87@yahoo.com

Author contributions

All authors contributed to the research work. Shomaila ashfaq, Saraj bahadur, performed experimental work. Sidra Nisar Ahmed, Saba Gul, Moona Nazish did sample collection and paper write up. Mushtaq Ahmad, Muhammad Zafar, Shazia Sultana supervised the work.

