



CHEMICAL SCIENCES

Current extraction methods and potential use of essential oils for quality and safety assurance of foods

JÉSSICA M.F. DE ALMEIDA-COUTO, JÉSSICA B. RESSUTTE, LÚCIO CARDOZO-FILHO & VLADIMIR F. CABRAL

Abstract: Essential oils (EOs) or vegetable oils have become the focus of several studies because of their interesting bioactive properties. Their application has been successfully explored in active packaging, edible coatings, and as natural flavoring to extend the shelf life of various types of food products. In addition, alternative methods of extraction of EOs (ultrasound-assisted extraction, microwave-assisted extraction, pressurized liquid extraction and supercritical fluid extraction) have been shown to be more attractive than traditional methods since they present better efficiency, shorter extraction times and do not use toxic solvents. This review paper provides a concise and critical view of extraction methods of EOs and their application in food products. The researchers involved in the studies approached in this review were motivated mainly by concern about food quality. Here, we recognize and discuss the major advances and technologies recently used to enable shelf life extension of food products.

Key words: Food control, shelf life, food preservatives, essential oils, extraction techniques.

INTRODUCTION

Growing consumer preference for safe, non-toxic foods with increased shelf-life has stimulated research on natural food preservatives (Wu et al. 2019). In recent years, the use of essential oils (EOs) has become a very useful technology due to their wide range of natural biologically active compounds, which are capable of aiding in the preservation of food systems (Freitas & Cattelan 2018, Khorshidian et al. 2018).

The EOs have been studied mainly due to their antioxidant, antimicrobial, insecticidal, antitumor and antidiabetic properties, and have been mainly applied in active packaging, edible coatings, and as natural flavoring (Brahmi et al. 2016, Khorshidian et al. 2018, Periasamy et al. 2016, Yen et al. 2015).

In the food industry, the desire to offer packaging that can protect food against external factors and ensure the safety of the food has stimulated studies about the development of packaging incorporated with EOs. These packaging systems, known as active packaging, interact with the food and gradually release bioactive compounds capable of minimizing or eliminating the presence of pathogenic microorganisms and even inhibiting lipid oxidation (Ribeiro-Santos et al. 2017, Sirocchi et al. 2017).

The incorporation of bioactive compounds aims to benefit food products by extending their shelf life. This has become the focus of several studies, and the application of nanotechnology has assisted in overcoming technical challenges,

such as solubility and stability of the bioactive compounds. Nanoemulsion contributes efficiently by promoting the application of EOs in real food systems as a means of natural conservation, thus increasing antimicrobial activity and, consequently, food safety. Nanoemulsion has contributed, for example, to significant advances in the development of edible coatings in food products (Abbas et al. 2015, Donsì & Ferrari 2016, Ma et al. 2016).

Furthermore, several materials used in food packaging may be updated to incorporate EOs. The current trend in food packaging is to use polymeric matrixes obtained from renewable and biodegradable resources, such as lipids, proteins and polysaccharides, thus contributing to environmental sustainability (Ribeiro-Santos et al. 2017, Romani et al. 2017).

Another point related to food preservation is the use of additives. There is a considerable interest in use of EOs, because those substances are considered safe food additives by the Food and Drug Administration. Moreover, due to the increase in consumer demand, there is a tendency to research natural additives, since synthetical ones are associated with negative side effects to human health (Cacho et al. 2016).

In this context, the main objective of the present work is to present a literature review in order to disclose the major advances and future trends regarding EO applications in food products. This review focuses on relevant papers published in the last eight years.

Essential oils

EOs are aromatic substances produced from secondary metabolites of plants belonging to the angiosperm family. They can be used for different purposes in several fields, such as pharmaceutical, cosmetic, agricultural and food sectors. Their complexity may vary from 20 to 60 components (Asbahani et al. 2015) and they are

characterized by two or three major components which are considered as such due to their high concentration (20 to 70%) in comparison to other components that define the physicochemical properties of the oil. The components existing in lower concentrations are also important to EO composition, due to the synergistic effect of the combined components (Asbahani et al. 2015, Pavela 2015).

Composition and quality of EOs depend on a plant's characteristics, its stage of development, origin, part of the plant used, and age, time, and condition at which the plant has been harvested. In addition to those factors, EOs are also affected by extraction method, analysis conditions and the type of solvent used, therefore, it is fundamental to choose the most suitable method (Ribeiro-Santos et al. 2017, Asbahani et al. 2015).

Conventional extraction methods

These are classic extraction methods based on the distillation of water by heating, traditionally used to recover EOs from oilseeds and medicinal or aromatic plants. Next, some of the main extraction techniques will be presented.

Steam distillation

Although EOs are produced by different methods, the majority (93%) are produced by steam distillation (Masango 2005). In practice, the process uses water as an extraction agent to vaporize or release volatile compounds from the raw material. The compounds are volatilized by absorbing heat from the steam and are then diffused in the vapor phase. The vapor phase is cooled and condensed before the water is separated from the organic phase based on its immiscibility (Prado et al. 2015).

Steam distillation can be combined with other extraction methods, such as microwave or ultrasound, to increase efficiency. This

combination can provide faster extraction kinetics, lower costs, reduce environmental impact, and provide a product similar to that obtained by conventional hydrodistillation (Palma et al. 2013). Variants of steam distillation are hydrodistillation and hydrodiffusion, presented in the following sections.

Hydrodistillation

This method is the simplest and oldest method used for the extraction of EO. Avicenna (980-1037), was the first to develop extraction using the still, extracting the first pure essential oil from the rose (Asbahani et al. 2015). The hydrodistillation (HD) system for extracting EO is equipped with a Clevenger type device. In this process, plant materials immersed in water are heated in a balloon; the water evaporates and flows towards the condenser until the EO is released (Gavahian & Farahnaky 2018). This technique is efficient in isolating a wide variety of EOs. However, it requires large amounts of energy and its high temperatures can cause changes in the compounds, with possible degradation (Pavlič et al. 2015).

Consumer demands, unpredictable energy costs in the future, and environmental constraints drive the development of clean technologies that prevent the use of chemicals and consume less energy (Zermane et al. 2016). The HD process with ohmic heating is a relatively new and innovative technique that has gained increasing interest in the last decade for promoting time and energy savings (Gavahian & Farahnaky 2018).

Moreover, several improved modules have been developed in recent years, such as microwave compressed hydrodistillation, microwave accelerated rod distillation, microwave vacuum hydrodistillation, and microwave-assisted hydrodistillation (Singh et al. 2019).

In current studies, HD has been used to obtain oils of pink pepper (Dannenberg et al. 2017, Dannenberg et al. 2016), rosemary (Sirocchi et al. 2017), orange leaves (Alparslan et al. 2016), ginger (Noori et al. 2018), oregano (Asensio et al. 2015, Hashemi et al. 2017), mint (Smaoui et al. 2016), and citronella (Gavahian et al. 2018).

Hydrodiffusion

This is a particular type of steam distillation, where the flow of vapors occurs from the top of the generator (Asbahani et al. 2015). This method has now been improved with the addition of microwave technology. The technique using microwave hydrodiffusion and gravity (MHG) is a solventless extraction method, based on the drilling of oil glands and subsequent oil drainage by gravity (Singh et al. 2019).

The use of MHG technology improved the extraction rate of rosemary, and only 20 min was sufficient to achieve a yield comparable to that obtained in 3 h by the conventional HD method. Also, a mixture of molecules with different properties (for example, polarity and volatility) can be extracted in a single step, such as essential oils and phenolic compounds (Ferreira et al. 2020).

In the extraction of EO from cumin seeds, the researchers observed that the chemical composition was approximately similar for the MHG and HD methods, with a drastic reduction in the extraction time from 150 min of HD to 16 min (200 W) of MHG (Benmoussa et al. 2018).

Organic solvent extraction

The plant material is macerated in an organic solvent; the extract is concentrated by removing the solvent under reduced pressure. This technique avoids the chemical changes and artifacts produced by cold extraction compared to hydrodistillation (Asbahani et al. 2015).

For solvent extraction, organic solvents such as n-hexane, alcohol, chloroform, water, and acetone are used, which provide efficient lipid recovery. (Pavlić et al. 2015). Hexane is also an excellent solvent for oil because of the high oil solubility, and because the oil can be easily recovered by distillation. The main drawback of the use of hexane is its high toxicity. As a result, other solvents have been used to substitute hexane in oil recovery including some medium polarity alcohols such as isopropanol and ethanol (Palma et al. 2013).

Soxhlet is one of the oldest extraction procedures. It is the standard extraction process (Zygler et al. 2012), since the solvent is recirculated in the sample until the oil is completely removed (Luque de Castro & Priego-Capote 2010).

The Soxhlet procedure has many disadvantages. For example, the average time for extraction is 1 to 72 hours; the solutes extracted by this method are obtained in high volume and diluted form and, therefore, need

to be concentrated before analysis. Perhaps the biggest disadvantage of this method is the need for expensive, toxic, and high purity organic solvents (Yousefi et al. 2019).

Several modified Soxhlet systems have been designed to overcome the drawbacks of the classical technique. Most of them focus on speeding up the process in an attempt to reduce the solvent consumption and the thermal degradation of the target compounds (Palma et al. 2013). Some alternatives to increase the speed at which the matrix releases components include applying microwaves or ultrasound (Luque de Castro & Priego-Capote 2012).

Alternative extraction methods

Most of the conventional methods need long-term extraction, in addition to the use of high-quality solvents. Alternative processes present some advantages, such as reduction in extraction time and power consumption, moreover, they increase the oil yield and improve quality (Asbahani et al. 2015).

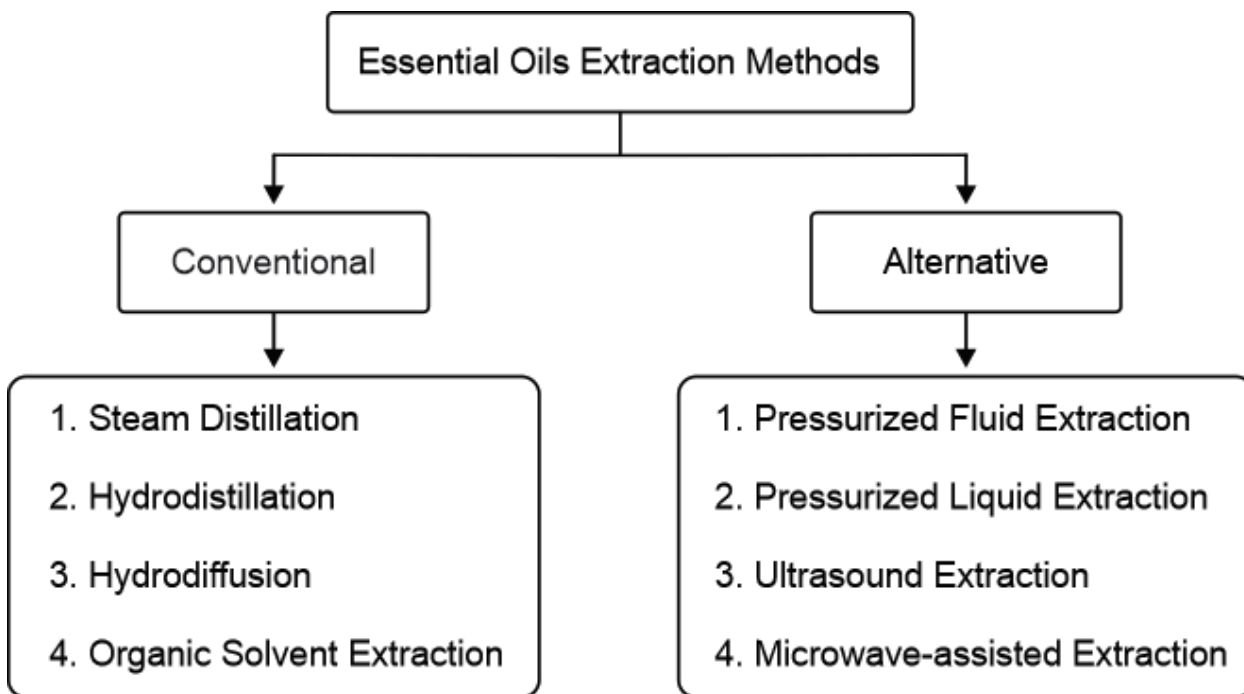


Figure 1. Extraction methods used for obtaining essential oils.

Therefore, several alternative extraction methodologies are reported in literature: ultrasound-assisted extraction (Tekin et al. 2015, Fernandes et al. 2016), microwave-assisted extraction (Franco-Vega et al. 2016, Chen et al. 2017), pressurized liquid extraction (PLE) (Rai et al. 2017), and supercritical fluid extraction (SFE) (Mustafa & Turner 2011, Dawidowicz et al. 2012).

These techniques are considered efficient and economically viable for the extraction of EOs. Green technologies are better options mainly because they are ecologically correct and there are fewer toxic solvents used in the extraction process. In addition, selective extraction, in most cases, occurs through changes in process parameters and can operate at elevated temperatures and pressures, reducing extraction time (Bubalo et al. 2018).

Pressurized fluid extraction

Regarding alternative methods, pressurized fluids under subcritical or supercritical conditions are indicated as a promising technique for the extraction of compounds with high purity (Moncada et al. 2014, Zheng et al. 2017). A supercritical fluid is any substance whose temperature and pressure conditions are above its critical parameters (Sánchez-Camargo et al. 2017). In this state, the substance acquires an intermediate behavior between gases and liquids (Shivonen et al. 1999). Small variations in temperature and pressure cause significant changes in the properties of the supercritical fluid (Panja et al. 2018).

Commonly used in supercritical extraction, carbon dioxide (CO₂) has advantageous properties such as low reactivity, low toxicity, and low cost, and is recognized as safe for use in food products (Cornelio-Santiago et al. 2017, Panja et al. 2018). It is a non-polar solvent and, consequently, has an affinity for other non-polar solvents (Herrero et al. 2006). This disadvantage

is usually overcome by the addition of modifiers or co-solvents capable of altering the solvation power of the supercritical fluid, facilitating the solubility and desorption of the analyte. However, to avoid a reduction in selectivity, a maximum of 10% v/v must be added (Pinto et al. 2018).

Meanwhile, the subcritical state occurs when an extraction solvent is used at a temperature between its boiling point and the critical temperature, at a pressure sufficient to maintain the solvent in a liquid state (Lu et al. 2014). It is considered more advantageous than the supercritical state because of the lower pressures used, resulting in shorter extraction times (Miao et al. 2013). Currently, the subcritical pressurized n-propane has been efficient in extracting fatty acids and active compounds such as phytosterols and tocopherols present in oils (Knez Hrnčič et al. 2018, Lopes et al. 2020, Trentini et al. 2017, Zanqui et al. 2020).

An alternative method involves the addition of propane as a solubility modifier. Propane's solvent power is superior to CO₂, requiring lower solvent/feed ratios and lower operating pressures. Also, propane can be easily removed from the oil after extraction by simple solvent depressurization (Palla et al. 2014) and can offer convenient selectivity and safety properties (Hegel et al. 2013), since one of the main disadvantages of extraction with pressurized propane is its flammability.

Pressurized liquid extraction

Pressurized liquid extraction (PLE) is another alternative technique which shows satisfactory results in relation to oil recovery and extraction time, when compared to conventional methods. In this technique, there is no need for filtration steps, since compounds are dissolved in the solvent and may remain inside the extractor. The PLE process consists of placing the sample

into the extractor and extracting its compounds using a solvent, with temperatures going up to 200°C and pressures ranging from 4 to 20MPa. The solvent is pumped into the extractor containing the sample and remains there for a period that may vary depending on the type of solvent and matrix used. Then, the extracted material is transferred to a sample container and undergoes specific analyses (Nieto et al. 2010, Dawidowicz et al. 2012).

This method has been successfully used in the extraction of green coffee oil (Xu et al. 2019), pomegranate bark oil (Santos et al. 2019), crambe seed oil (Mello et al. 2019) and cypress oil (Dawidowicz et al. 2012).

Despite being an efficient technique, PLE is not able to separate compounds from similar phenolic classes, and the extracts produced contain a wide mixture of components. Solid adsorbents (solid-phase extraction - SPE) can be used to separate specific classes and phenolic compounds. In this context, the PLE method has been improved through online SPE padding for simultaneous extraction and fractionation (Silva et al. 2020).

Ultrasound extraction

In the ultrasound extraction technique, the ultrasound apparatus promotes a higher rate of solvent penetration in the sample, caused by cavitation bubbles formed during the application of sonic waves. In general, the use of ultrasound causes vibrations in the matrix, thus enhancing the contact surface between the matrix and the solvent and resulting in a higher solvent recovery in a short period of time. Additionally, the technique requires low temperatures, which facilitates the recovery of volatile compounds in EOs (Tekin et al. 2015, Samaram et al. 2014).

Current studies have used this method in pomegranate bark (Sharayei et al. 2019), moringa seed (Zhong et al. 2018), ginger (Fernandes et al.

2016), papaya seed (Samaram et al. 2014), garlic (Tekin et al. 2015) and grape marc (Goula et al. 2016).

A combined method of ultrasound-assisted and microwave-assisted extractions was adopted for the extraction of polyphenols in distilled water (Yu et al. 2017). Another combined method included ultrasound, followed by supercritical CO₂ to extract polyphenols from grape marc (Porto et al. 2015).

Microwave-assisted extraction

Among alternative methods, we also highlight microwave-assisted extraction. This is an emerging technique which improves material recovery, and reduces the time and energy needed in the process. It uses microwave radiation as a heating source in the extraction process. Microwaves, through dipole rotation and ionic conduction, cause instantaneous heating inside the sample, thus leading to faster extractions (Franco-Vega et al. 2016). Hibiscus chalice (Cassol et al. 2019), lemon peel (Rodsamran & Sothornvit 2019), grape marc (Garrido et al. 2019) and orange peel (Franco-Vega et al. 2016) are examples of raw material used in the microwave assisted extraction.

Emerging improvements in microwave-assisted extraction include combining it with other technologies to maximize the yield of target food components. The integration of alternative and environmentally friendly solvents (ionic liquids, deep eutectic solvents, multiphase solvents and nonionic surfactants at cloud point temperature) promotes overall extraction efficiency (Ekezie et al. 2017).

Lately, enormous attention has been given to extraction of food constituents using the synergistic application of microwaves to ultrasound irradiation techniques (Yu et al. 2017), negative pressure cavitation (Yao et al. 2015), sub or supercritical extraction (Matusiewicz &

Ślachciński 2014), enzymatic extraction (Rashed et al. 2017) and hydrodiffusion (Singh et al. 2019).

Applications of essential oils in food

When applied to food, EOs can act as flavoring, antioxidant and antimicrobial agents, with special importance given to the last two functions. EOs can be added directly to food or incorporated into material used for packaging (Ribeiro-Santos et al. 2017, Falleh et al. 2020). Table I presents some important uses of EOs in the food industry, such as their use in active packaging, edible coatings and food additives.

Additives

The inclusion of EOs in food as conserving agents is an alternative to the use of synthetic additives. Antigo et al. (2017) produced milk caramel spread (*dulce de leche*) with an addition of clove and cinnamon EOs. Lipid oxidation, microbiological, physical, chemical and sensorial attributes of the product were analyzed. There were no alterations related to composition, texture, color and sensory attributes. Microbiological analyses indicated the EOs used are active antimicrobial components and including them in *dulce de leche* provided general sensory acceptance similar to the traditional product. The sample with cinnamon EO showed less lipid oxidation during storage.

EOs have been used to reduce the addition of nitrites in meat products. In this context, a study evaluated the effect of adding coriander EO in concentrations of 0.075–0.150 µL/ g on the characteristics of cooked pork sausages, produced with different concentrations of sodium nitrite (0.50 and 100 mg/ kg). The combination of 60 mg/ kg of nitrite with 0.12 µL/ g of EO resulted in a better microbial and oxidative stability and satisfactory values of red color, therefore, it is possible to use the coriander EO to reduce the amount of sodium

nitrite added to cooked pork sausages, while retaining high quality and shelf life (Šojić et al. 2019).

The *Melaleuca alternifolia* EO, also known as tea tree EO, is widely used due to its broad-spectrum antimicrobial activity and powerful anti-inflammatory properties. Thus, the objective of one study was to evaluate the antimicrobial potential of this EO (1.5%) in the inhibition of *Listeria monocytogenes* in ground beef. The samples were inoculated with four different suspensions of *L. monocytogenes* (1.5×10^8 , 4.6×10^4 , 9.2×10^3 , and 1.2×10^2 CFU/ mL) and stored at 4 °C for 14 days. Except for the sample inoculated with the suspension at 1.5×10^8 CFU/ mL, the tests showed that tea tree EO had antimicrobial activity (Silva et al. 2019a).

Shange et al. (2019) evaluated the effect of adding oregano EO (1%) on the shelf life of wildebeest *biceps femoris* muscles, stored anaerobically at 2.6 °C for 9 days. Lipid oxidation was stabilized at <9 mgMDA/ kg for the sample with EO, while the same was not observed for the control. Samples with EO also showed lower total viable counts (TVC), coliform counts, and lactic acid bacteria (LAB) counts. The count limit for TVC and LAB for this product was reached 3 days later than in the control group. In addition, bacterial growth rates for TVC and LAB were >1.4-fold slower for the samples with EO. *Hyptis suaveolens* EO has also been studied due to its antibacterial, antioxidant and antifungal action. Based on this, Mihin et al. (2019) evaluated the addition of this EO and its effect on the shelf life of beef. During *in vitro* tests, the EO showed inhibitory activity against 11 microbial strains and was able to preserve the quality of the meat for 7 days.

Another alternative to extend the shelf life of meat is the use of EOs in marinades, as in the study by Haute et al. (2016) which consisted of immersing meat (pork fillet, pork bacon, chicken

Table I. Examples of essential oils application in food.

Application	Food	Essential oil	Property	Reference
Additive	Milk caramel spread (<i>dulce de leche</i>)	Clove and Cinnamon	Antimicrobial Antioxidant	Antigo et al. 2017
	Cooked pork sausages	Coriander	Antimicrobial Antioxidant	Šojić et al. 2019
	Ground beef	Tea tree	Antimicrobial	Silva et al. 2019
	<i>Wildebeest biceps femoris</i> muscles	Oregano	Antimicrobial	Shange et al. 2019
	Ground beef	<i>Hyptis suaveolens</i>	Antimicrobial Antioxidant	Mihin et al. 2019
	Meat	Cinnamon, Oregano, and Thyme	Antimicrobial	Haute et al. 2016
	Pork meat	Cinnamon	Antimicrobial	Haute et al. 2017
	Minas frescal cheese	Oregano and Rosemary	Antimicrobial	Diniz-Silva et al. 2020
	Pressed ewes' cheese	Thyme, Lemongrass and Basil	Antimicrobial	Licon et al. 2020
	Fruit juice	Japanese Mint and Pepper Mint	Antimicrobial	Guedes et al. 2016
	Pomegranate	Eucalyptus, Galbanum, Thymus, and Clove	Antimicrobial Antioxidant	Jahani et al. 2020
	Rice	Nutmeg	Antimicrobial	Das et al. 2020
	Seeds	Lemongrass	Antimicrobial Antioxidant	Deepika et al. 2020
	Bread	Oregano and Thyme	Antimicrobial	Rosa et al. 2020
	Fish burger	Lemon	Antioxidant	Hasani et al. 2020
Beef patties	Cinnamon	Antimicrobial	Ghaderi-Ghahfarokhi et al. 2017	
Active packaging	Cheese	Pink Pepper	Antimicrobial	Dannenberg et al. 2017
	Peach	Ginger and Angelica	Antioxidant	Jiang et al. 2020
	Corn	Oregano and Cinnamon	Antimicrobial	Mateo et al. 2017
	Cherry tomato	Oregano	Antimicrobial	Kwon et al. 2017
	Bread	Lemongrass	Antimicrobial	Oliveira et al. 2020
	Chicken fillets	<i>Polylophium involucreatum</i>	Antimicrobial	Javaherzadeh et al. 2020
	Shrimp	Clove	Antimicrobial	Ejaz et al. 2017
	<i>Otolithes ruber</i> fish	<i>Zataria multiflora</i> and pepper mint	Antimicrobial Antioxidant	Heydari-Majd et al. 2019
	Beef	Chrysanthemum	Antimicrobial Antioxidant	Lin et al. 2019
	Fresh poultry meat	Rosemary	Antimicrobial Antioxidant	Souza et al. 2019

Table I. Continuation.

Edible coatings	Cheese	<i>Pimpinella saxifraga</i>	Antimicrobial Antioxidant	Ksouda et al. 2019
	Beef slices	Cumin	Antimicrobial	Behbahani et al. 2020
	Pistachio	Thyme	Antimicrobial Antioxidant	Hashemi et al. 2020
	Mango	Cinnamon	Antioxidant	Yin et al. 2019
	Table grape	Thymus	Antimicrobial	Pina-Barrera et al. 2019
	Guava	Cinnamon and Lemon	Antioxidant	Murmu and Mishra 2018
	Case gooseberry	Rue	Antimicrobial Antioxidant	González-Locarno et al. 2020
	Strawberry	Lemongrass	Antioxidant	Silva et al. 2019
	Rainbow trout fillets	<i>Ferulago angulata</i>	Antimicrobial Antioxidant	Shokri et al. 2020
	Chicken breast	Ginger	Antimicrobial Antioxidant	Noori et al. 2018
	Cheese	Oregano	Antimicrobial	Artiga-Artigas et al. 2017

fillet, chicken skin and salmon) in a solution marinated with cinnamon, oregano, and thyme EOs. It was observed that yeast growth was inhibited by immersion in 1% cinnamon EO in all matrixes. Haute et al. (2017) used a marinade with 1% cinnamon EO in pork and salmon and subsequently packed them with modified atmosphere (MAP) or vacuum. Cinnamon EO extended shelf life of pork packed in MAP and vacuum against microbial growth but did not have the same effect on salmon.

Diniz-Silva et al. (2020) evaluated the incorporation of oregano and rosemary EOs in the processing of Minas Frescal cheese stored under refrigeration temperature (7 °C). In the first 15 days, a significant reduction in *Escherichia coli* counts was observed in the analyzed samples. The addition of EOs to cheese also had a positive impact on sensory analysis. Another cheese study evaluated the addition of different EOs in the production of pressed ewes' cheese. The thyme EO was the most effective in completely inhibiting the growth of *Penicillium verrucosum* and in reducing the *Clostridium tyrobutyricum* count, without affecting the

natural flora present in the cheese (Licon et al. 2020).

Traditionally, safety and stability of fruit juices were reached through thermal processing and use of chemical preservatives. Guedes et al. (2016) highlighted the use of EOs as an alternative for the reduction of pathogenic microorganisms in fruit juices. In this study, Japanese mint and peppermint EOs were evaluated. Such EOs induced reductions in counts of *Escherichia coli*, *Listeria monocytogenes* and *Salmonella enteritidis* in cashew, guava, mango, and pineapple juices. Incorporation of these EOs in fruit juices promoted a reduction in pathogenic bacteria without altering their physical-chemical properties; however, it significantly affected the flavour.

In order to reduce the application of synthetic antifungals in pomegranate fruits, Jahani et al. (2020) evaluated the addition of different concentrations of eucalyptus, galbanum, thymus, and clove EOs and their inhibitory effects against *Aspergillus niger*. All analyses were performed on the first and tenth days of storage. In *in vitro* analyses, the growth of *A. niger* was completely inhibited on the first

and tenth days by the application of clove EO in the concentrations of 200, 400, 600 and 800 $\mu\text{L/L}$. Thyme EO was effective in the concentration of 800 $\mu\text{L/L}$ on the tenth day. The fruits treated with thyme EO at a concentration of 800 $\mu\text{L/L}$ showed the least weight loss and the highest firmness in comparison with fruits treated with other EOs. In addition, the highest anthocyanin content was obtained with eucalyptus EO at 800 $\mu\text{L/L}$.

The nanoencapsulation of EOs for use in food as a preservative is a recent and promising research field. EOs stored under ambient conditions have some disadvantages, such as insolubility in water, easy oxidation, instability, volatilization and degradation of bioactive compounds in a short period of time. These factors can reduce their effectiveness when used for practical applications. To overcome these drawbacks and keep the original characteristics of EOs, nanoencapsulation is an efficient method (Das et al. 2020). Emulsification, spray drying, ionic gelation and coacervation are the most adopted nanoencapsulation techniques (Chaudhari et al. 2019).

A large number of nanoencapsulation carrier matrixes (nanoencapsulates) can be used to encapsulate EO and their bioactive compounds, such as starch, chitosan, zein, cyclodextrin and cellulose. These encapsulants must be biodegradable and safe for human health. Depending on the method adopted, the nanoencapsulated EOs can take different forms, such as nanoemulsions, nanoparticles, nanotubes, nanogel, nanosponge, nanofibers and nanoliposomes. Among them, nanoemulsion, nanoparticle and nanogel are the most frequently used systems in the food sector (Chaudhari et al. 2019).

A study using nanoencapsulation technology evaluated the antifungal activity of nutmeg EO applied to the chitosan nano-matrix in order to

control post-harvest losses of rice grains. The EO was tested against 15 food-borne fungi. In comparison with free EO, nanoencapsulated EO showed greater efficacy against the evaluated fungi, and at lower doses, was able to inhibit the aflatoxin B1 biosynthesis by *Aspergillus flavus* strain LHP R14. In situ efficacy of nanoencapsulated and unencapsulated EO on stored rice seeds showed effective protection against lipid peroxidation, fungal infestation and aflatoxin B1 contamination (Das et al. 2020).

Deepika et al. (2020) tested the potential of lemongrass EO encapsulated into chitosan nanoparticles against *Aspergillus flavus* and 15 other fungi, in order to control the deterioration of stored food. After nanoencapsulation, the EO showed better effectiveness in inhibiting the growth of fungi and production of aflatoxin B1 by *A. flavus*. Furthermore, the nanoencapsulated lemongrass EO exhibited remarkable antioxidant activity and did not have adverse effects on seed germination.

In the research developed by Rosa et al. (2020), oregano and thyme EOs were encapsulated using zein nanocapsules. Nanoencapsulated EOs have been shown to be more effective against gram-positive bacteria compared to gram-negative bacteria. In addition, nano-encapsulated EOs were also effective in preserving bread, protecting against the proliferation of molds and yeasts.

Lemon EO is an antimicrobial and antioxidant compound, used mainly as a food additive. Thus, a study aimed to evaluate the antioxidant effect of lemon EO, (0.5 and 1%) nanoencapsulated in chitosan/modified starch, in fish burgers stored for 18 days. The nanoencapsulated EO in the concentration chitosan: modified starch (1.5: 8.5%) improved the quality characteristics of the fish burgers. This improvement was due to the reduction in the values of peroxides (PV),

thiobarbituric acid value (TBA) and total volatile nitrogen base (TVB-N) (Hasani et al. 2020).

Ghaderi-Ghahfarokhi et al. (2017) evaluated the addition of cinnamon EO incorporated into chitosan nanoparticles in beef patties. Both free and nanoencapsulated cinnamon EO were effective in reducing the microbial population of samples compared to the control (without addition of EO) over an 8-day storage period at 4°C. At the end of the storage period, the best formulations in thiobarbituric acid reactive substances (TBARS) test were the samples with 0.05% of ascorbic acid and 0.1% of encapsulated EO. In addition, it was observed that the color of the samples containing nanoencapsulated EO changed slightly, while for the sample with free EO, there were significant changes in color. In the sensory analysis, the beef patties with free cinnamon EO showed lower consumer acceptability in the color and odor parameters.

Active packaging

Through interaction with products, active packaging increases the shelf life of food, improving or maintaining its properties. Due to the antimicrobial and antioxidant properties of EOs, the development of active packaging for food with EO incorporation has become the focus of many research studies (Ribeiro-Santos et al. 2017).

One of these studies developed and evaluated active films made of cellulose acetate incorporated with pink pepper EO. Active films were evaluated based on their action in sliced mozzarella cheese against *Staphylococcus aureus*, *Listeria monocytogenes*, *Escherichia coli* and *Salmonella Typhimurium*. Concentrations of 2, 4 and 6% of the added EO were active against *L. monocytogenes* and *S. aureus*. The tests showed that affinity between nonpolar EO molecules and the lipid components of cheese

allow migration of antimicrobial properties to food (Dannenberg et al. 2017).

Ginger and angelica EOs are known for their antimicrobial and antioxidant properties. Thus, a study aimed to develop films based on polylactic acid (PLA) and EO of ginger and angelica for the preservation of peaches. The film with the addition of angelica EO showed the highest antioxidant activity and had the best effect on the preservation of peach samples. Due to the delay in the oxidation process, the shelf life of this fruit was extended to more than 15 days (Jiang et al. 2020).

Mateo et al. (2017) developed a packaging made of ethylene-vinyl alcohol copolymer incorporated with oregano and cinnamon EOs to control the usual fungi associated with aflatoxin contamination in maize grains. The bioactive film that contained an effective dose of cinnamaldehyde was the most efficient in controlling *Aspergillus flavus* and *Aspergillus parasiticus*. Antimicrobial activity was also tested in cherry tomatoes with active packaging containing microencapsulated oregano EO. Results showed that tomatoes' quality and their physical properties were preserved. The packaging with 2% oregano EO was the most efficient, reducing 91.64% of the microbial load (Kwon et al. 2017).

Oliveira et al. (2020) developed cashew gum and gelatin films, incorporated with ferulic acid and lemongrass EO, for application as bread packaging. The packaging with EO provided six days of storage for bread compared to three days for commercial packaging. The increase in the shelf life of the bread samples suggests antimicrobial action of the lemongrass EO packaging.

EO nanoencapsulation techniques have also been widely used for application in active packaging. A study investigated the effect of applying a polylactic acid film

(PLA) incorporated with nanochitosan and *Polylophium involucreatum* EO in chicken fillets stored for 10 days at refrigerated temperature. In the packaged samples, the total microbial population was reduced by approximately 1–3 log CFU/ g. In addition, the films extended the shelf life of the chicken fillet by more than 10 days, without producing adverse sensorial properties (Javaherzadeh et al. 2020).

Ejaz et al. (2018) used clove oil to develop active films for peeled shrimp. The authors produced a not very flexible film with high mechanical resistance, combining nanocomposites, zinc oxide and clove EO. In this study, films with 50% of clove EO showed the maximum antibacterial activity against *Listeria monocytogenes* and *Salmonella typhimurium*.

Heydari-Majd et al. (2019) produced films based on PLA containing 1.5% zinc oxide nanoparticles and different concentrations of *Zataria multiflora* and peppermint EOs. The films were applied to *Otolithes ruber* fish, stored at 4 °C for 16 days. Compared to the control sample, the shelf life of the packaged fish fillet samples increased from 8 to 16 days. The lowest values of TBARS and TVB-N were obtained for samples packed with films containing 1.5% *Zataria multiflora* EO.

EO extracted from the chrysanthemum plant is used mainly as an organic pesticide and as an insect repellent. This EO has also exhibited antimicrobial properties. Therefore, Lin et al. (2019) incorporated chrysanthemum EO into chitosan nanofibers for application as packaging in beef. After 7 days of storage, the nanofibers with EO were effective against *Listeria monocytogenes* bacteria, with inhibition rates of 99.91%, 99.97% and 99.95% at temperatures of 4 °C, 12 °C and 25 °C, respectively. Due to the release of antioxidant components present in the EO by nanofibers, the TBARS value in beef

was 0.135 MDA/ kg lower compared to the control sample, after 12 days at 4 °C.

Souza et al. (2019) developed bionanocomposites based on chitosan and montmorillonite, with incorporation of rosemary EO in different concentrations (0.5, 1 and 2%) to use as primary packaging for fresh poultry meat. The meat samples were packaged and stored for 15 days at 5 °C. In comparison to the control, the samples packaged showed a reduction of 1.2–2.1 log UFC/ g in the total count of microorganisms. EO films were also able to delay the lipid peroxidation and discoloration of the fresh poultry meat.

Edible coatings

The coating technique consists of applying a thin layer of a biodegradable and edible material on the food surface in order to prolong the shelf life of a wide variety of products. Some of the functions of the edible coating are to prevent undesirable chemical reactions, to serve as a barrier against moisture loss and to prevent deterioration by microorganisms. A wide variety of polymeric matrixes can be used, such as chitosan, sodium alginate and gelatin, which can be incorporated into EOs to increase the effectiveness of these coatings (Ju et al. 2019, Pina-Barrera et al. 2019).

One of the edible coating studies investigated the effect of adding a coating based on sodium alginate and EO of *Pimpinella saxifrage*, at a concentration of 1–3%, to cheese samples. The EO enrichment of sodium alginate coating, particularly at 3%, improved the preservation of the analyzed samples. The preservation of pH and color were observed, as well as reduction of weight loss and enhanced oxidative and bacterial stability (Ksouda et al. 2019).

Cumin EO is known for its anti-inflammatory and antimicrobial properties. Thus, one study aimed to develop a coating based on Shahri

Balangu and cumin EO to improve the shelf life of beef slices, stored for 9 days under refrigerated temperature. The counts of psychrotrophic bacteria, coliforms, *Escherichia coli*, *Staphylococcus aureus*, molds and yeasts were significantly reduced. Moreover, there was a reduction in lipid oxidation, and the coated samples showed no adverse effects on the sensory characteristics (Behbahani et al. 2020).

Hashemi et al. (2020) evaluated the effects of coatings made with different concentrations of alginate and thyme EO on the postharvest characteristics of fresh pistachios stored for 39 days at 3 °C and 80% relative humidity. The addition of coatings on the fruits contributed to the maintenance of higher antioxidant activity and phenolic content in comparison with the control. In addition, the coated samples reduced mold and yeast growth. The values of free fatty acids and peroxides were also significantly lower in pistachios with the addition of the coating enriched with thyme EO.

Application of edible coatings incorporated with EO in fruits has become a promising field. Yin et al. (2019) evaluated the addition of a coating on fresh mangoes stored for 14 days, at 25 °C and 50% relative humidity. The samples were packaged in multilayer coatings made from chitosan, cinnamon EO microcapsules and alginate solutions. Compared to the control, the coated fruits could effectively inhibit the decrease of vitamin C content, slow down weight loss and delay the appearance of respiration peaks. Additionally, the mangoes coated with five layers still maintained their commercial value during the evaluated period, although the same was not observed for the control samples.

In another study, Pina-Barrera et al. (2019) developed a multisystemic coating based on pullulan and polymeric nanocapsules containing thymus EO, in order to increase the shelf life of table grapes. The shelf life study showed that the

coated grapes maintained their characteristics of firmness, total acidity, color, and total soluble solids for a longer time compared to the control. Furthermore, the coating inhibited the growth of undesirable microorganisms and reduced oxidative stress induced during the postharvest period. Murmu & Mishra (2018) demonstrated improved antioxidant activity in guava coated with arabic gum, sodium caseinate, cinnamon and lemon EO-based coating.

González-Locarno et al. (2020) evaluated the effect of coatings made from chitosan and rue EO in different concentrations for application on cape gooseberries stored at 18 °C for 12 days. The fruits coated with 0.5% EO suffered lower weight loss compared to the uncoated samples. The application of coatings with 1.0 and 1.5% delayed the growth of aerobic mesophilic bacteria, molds and yeasts. The coating also preserved the antioxidant properties of the fruits after 12 days. Silva et al. (2019b) obtained similar results in his research on edible coatings made of pectin, cellulose nanocrystals, glycerol and lemongrass EO, for application on strawberries under refrigeration temperature, during 8 days of storage. Application of the coatings reduced the weight loss and the anthocyanin content of the fruits.

The use of nanoemulsions offers clear advantages, such as better antimicrobial activity, reduced interactions with other components of the food matrix and greater stability to EO compounds (Prakash 2018). Therefore, Shokri et al. (2020) evaluated the efficiency of nanoemulsions in improving the characteristics of a coating based on chitosan and *Ferulago angulata* EO, and the coating's potential to extend the shelf life of Rainbow trout fillets stored at 4 °C for 16 days. Nanoemulsions with 3% EO showed the best inhibitory effect on the growth of bacteria in the fish fillet samples. In addition, nanoemulsions improved the

effectiveness of the coating in retarding the increase of lipid peroxidation and TVB-N of the analyzed samples.

The work of Noori et al. (2018) reported the use of nanoemulsion with ginger EO in a sodium caseinate edible coating applied to chicken breast refrigerated for 12 days. Coating with 6% of EO ginger nanoemulsion resulted in significant decrease of total aerobic bacteria. Although antioxidant activity was not significant, samples coated with nanoemulsion showed less difference in color and cooking loss, and proved effective in prolonging the shelf life of the product.

Artiga-Artigas et al. (2017) applied nanoemulsion-based coatings containing oregano EO incorporated with mandarin fibers and sodium alginate in low-fat cheese in order to extend its shelf life. The authors observed that the cheese's native microbiota was controlled and growth of *Staphylococcus aureus* was decreased from 6.0 to 4.6 log CFU/g after 15 days.

Legal aspects of the use of essential oils in food

EOs are classified as flavorings by the European Commission (EC) (EC 2008). Since 2012, the European Union has adopted a list of flavorings approved for use, which is updated frequently. EOs are also classified and registered as flavorings by the United States Food and Drug Administration (FDA) and considered Generally Recognized as Safe (GRAS) (FDA 2020).

The EOs classified as GRAS comprise a series of EOs most commonly used, such as oregano, coriander, ginger, thyme, clove, basil, cinnamon, nutmeg, and menthol, among others. Despite being classified as GRAS, EOs have a recommended intake limit, since some of their components can cause allergies. The Codex Alimentarium, the Council of Europe (CoE), Food

Chemical Codex (FCC), Manufacturers Association (FEMA), and the International Organization of Flavor Industries (IOFI) have adopted specific protocols to check the toxicity of EOs and their components, as well as established security restrictions.

Some of the EOs, such as lavender, eucalyptus and laurel, have been linked to allergic reactions. Consequently, each EO added to a food matrix must be validated by its safe intake limit for humans, considering the classification of EOs and their limits already pre-established by health organizations, such as the FDA (Falleh et al. 2020).

Future perspectives

The application of EOs in food matrixes has emerged from an increasing trend to replace synthetic preservatives. EOs used as natural additives offer a clear advantage (Falleh et al. 2020). Several studies have been successful in incorporating EOs in food matrixes, either in their free form or added to materials to produce active packaging and edible coating. Ecological technologies capable of increasing the bioactive potential of EOs have also been studied and used, such as the alternative extraction methods and micro and nanoencapsulation techniques previously described in this literature review.

In addition to EOs that already have pre-established limits for use in food, regulated by food safety organizations, it is expected that in the near future, the standardization of new types of EOs will occur, so that they may be safely applied to foods in doses capable of producing desirable results. Moreover, the synergism between different EOs for food applications presents a field of research with promising future perspectives. Future research combining alternative EO extraction methods with emerging technologies such as nanoencapsulation is also expected.

CONCLUSION

The study of the properties and extraction methods of EOs, as well as their application in food, has proved to be a subject of extreme relevance, since safe and high-quality food products have become a requirement of consumers in recent years. Based on this literature review, it was possible to identify the main advances made and technologies used in the development of active packaging, edible coatings and food additives. In addition, the ability of EOs to control microbiological action and extend shelf life of products, thereby providing safe products, has become clear. Thus, future studies should be conducted to further explore the interactions between EOs and food.

Acknowledgments

We thank Coordenação de Aperfeiçoamento de Pessoal de Nível Superior - CAPES for the financial support.

REFERENCES

- ABBAS S, KARANGWA E, BASHRI M, HAYAT K, HONG C, SHARIF HR & ZHANG X. 2015. Fabrication of polymeric nanocapsules from curcumin-loaded nanoemulsion templates by self-assembly. *Ultrason Sonochem* 23: 81-92.
- ALPARSLAN Y, YAPICI HH, METIN C, BAYGAR T, GÜNLÜ A & BAYGAR T. 2016. Quality assessment of shrimps preserved with orange leaf essential oil incorporated gelatin. *LWT. Food Sci Technol* 72: 457-466.
- ANTIGO J, CESTARI L, SCAPIM M, SANTOS SS, MORITZ CF & MADRONA GS. 2017. Clove and cinnamon essential oils in dulce de leche. *Nutr Food Sci* 47: 101-107.
- ARTIGA-ARTIGAS M, ACEVEDO-FANI A & MARTÍN-BELOSO O. 2017. Improving the shelf life of low-based edible coatings containing oregano essential oil and mandarin fiber. *Food Control* 76: 1-12.
- ASBAHANI AE, MILADI K, BADRI W, SALA M, ADDI EHA, CASABIANCA H, MOUSADIK AE, HARTMANN D, JILALE A, RENAUD FNR & ELAISSARI A. 2015. Essential oils: From extraction to encapsulation. *Int J Pharm* 483: 220-243.
- ASENSIO CM, GROSSO NR & JULIANI HR. 2015. Quality preservation of organic cottage cheese using oregano essential oils. *Food Sci Technol* 60: 664-671.
- BEHBAHANI BA, NOSHAD M, & JOOYANDEH H. 2020. Improving oxidative and microbial stability of beef using Shahri Balangu seed mucilage loaded with Cumin essential oil as a bioactive edible coating. *Biocatal Agric Biotechnol* 24: 101563.
- BENMOUSSA H, ELFALLEH WHES, ROMDHANE M, BENHAMOU A & CHAWECH R. 2018. Microwave hydrodiffusion and gravity for rapid extraction of essential oil from Tunisian cumin (*Cuminum cyminum* L.) seeds: Optimization by response surface methodology. *Ind Crops Prod* 124: 633-642.
- BRAHMI F, ABDNOUR A, BRUNO M, SILVA P, ALESSANDRA P, DANILO F, DRIFA Y, FAHMI EM, KHODIR M & MOHAMED C. 2016. Chemical composition and in vitro antimicrobial, insecticidal, and antioxidant activities of the essential oils of *Mentha pulegium* L. and *Mentha rotundifolia* (L) Huds growing in Algeria. *Ind Crop Prod* 88: 96-105.
- BUBALO MC, VIDOVIĆ S, REDOVNIKOVIC IR & JOKIĆ S. 2018. New perspective in extraction of plant biologically active compounds by green solvents. *Food Bioprod Process* 109: 52-73.
- CACHO JI, CAMPILLO N, VIÑA P & HERNÁNDEZ-CÓRDOBA M. 2016. Determination of synthetic phenolic antioxidants in edible oils using microvial insert large volume injection gas-chromatography. *Food Chem* 200: 249-254.
- CASSOL L, RODRIGUES E & ZAPATA NOREÑA CP. 2019. Extracting phenolic compounds from *Hibiscus sabdariffa* L. calyx using microwave assisted extraction. *Ind Crop Prod* 133: 168-177.
- CHAUDHARI AK, DWIVEDY AK, SINGH VK, DAS S, SINGH A & DUBEY NK. 2019. Essential oils and their bioactive compounds as green preservatives against fungal and mycotoxin contamination of food commodities with special reference to their nanoencapsulation. *Environ Sci Pollut Res* 26: 25414-25431.
- CHEN F, JIA J, ZHANG Q, GU H & LEI YANG L. 2017. A modified approach for isolation of essential oil from fruit of *Amorphafruticosa* Linn using microwave-assisted hydrodistillation concatenated liquid-liquid extraction. *J Chromatogr A* 1524: 254-265.
- CORNELIO-SANTIAGO HP, GONÇALVES CB, OLIVEIRA NA & OLIVEIRA AL. 2017. Supercritical CO₂ extraction of oil from green coffee beans: Solubility, triacylglycerol composition, thermophysical properties and thermodynamic modelling. *J Supercrit Fluid* 128: 386-394.
- DANNENBERG GS, FUNCK GD, CRUXEN CES, MARQUES JL, SILVA WP & FIORENTINI AM. 2017. Essential oil from pink pepper as an antimicrobial component in cellulose acetate film: Potential for application as active packaging for sliced cheese. *LWT - Food Sci Technol* 81: 314-318.

- DANNENBERGGS, FUNCKGD, MATTEI FJ, SILVA WP & FIORENTINI ÂM. 2016. Antimicrobial and antioxidant activity of essential oil from pink pepper tree (*Schinus terebinthifolius* Raddi) in vitro and in cheese experimentally contaminated with *Listeria monocytogenes*. *Innov Food Sci Emerg Technol* 36: 120-127.
- DAS S, SINGH VK, DWIVEDY AK, CHAUDHARI AK, UPADHYAY N, SINGH A, DEEPIKA & DUBEY NK. 2020. Fabrication, characterization and practical efficacy of *Myristica fragrans* essential oil nanoemulsion delivery system against postharvest biodeterioration. *Ecotoxicol Environ Saf* 189: 110000.
- DAWIDOWICZ AL, CZAPCZYNSKA NB & WIANOWSKA D. 2012. The loss of essential oil components induced by the purge time in the pressurized liquid extraction (PLE) procedure of *Cupressus sempervirens*. *Talanta* 94: 140-145.
- DEEPIKA, SINGH A, CHAUDHARI AK, DAS S & DUBEY NK. 2020. Nanoencapsulated *Monarda citriodora* Cerv. ex Lag. essential oil as potential antifungal and antiaflatoxigenic agent against deterioration of stored functional foods. *J Food Sci Technol*.
- DINIZ-SILVA HT, BRANDÃO LR, GALVÃO MS, MADRUGA MS, MACIEL JF, SOUZA EL & MAGNANI M. 2020. Survival of *Lactobacillus acidophilus* LA-5 and *Escherichia coli* O157:H7 in Minas Frescal cheese made with oregano and rosemary essential oils. *Food Microbiol* 86: 103348.
- DONSÌ, F & FERRARI G. 2016. Essential oil nanoemulsions as antimicrobial agents in food. *J Biotechnol* 233: 106-120.
- EKEZIE FGC, SUN DW & CHENG JH. 2017. Acceleration of microwave-assisted extraction processes of food components by integrating technologies and applying emerging solvents: A review of latest developments. *Trends Food Sci Technol* 67: 160-172.
- EUROPEAN COMMISSION (EC). 2008. Regulation (EC) No 1334/2008. Official Journal of the European Union. Retrieved from: <https://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=OJ:L:2008:354:FULL&from=EN>. Access: June, 2020.
- FALLEH H, JEMAA MB, SAADA M & KSOURI R. 2020. Essential oils: A promising eco-friendly food preservative. *Food Chem* 330: 127268.
- FERNANDES RVB, BORGES SV, SILVA EK, SILVA YF, SOUZA HJB, CARMO EL, OLIVEIRA CR, YOSHIDA MI & BOTREL DA. 2016. Study of ultrasound assisted emulsions on microencapsulation of ginger essential oil by spray drying. *Ind Crop Prod* 94: 413-423.
- FERREIRA DF, LUCAS BN, VOSS M, SANTOS D, MELLO PA, WAGNER R, CRAVOTTO G & BARIN JS. 2020. Solvent-free simultaneous extraction of volatile and non-volatile antioxidants from rosemary (*Rosmarinus officinalis* L.) by microwave hydrodiffusion and gravity. *Ind Crops Prod* 145: 112094.
- FOOD AND DRUG ADMINISTRATION (FDA). 2020. Code of federal regulations (CFR). Retrieved from: <https://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/CFRSearch.cfm?fr=182.20>. Access: June, 2020.
- FRANCO-VEJA A, RAMÍREZ-CORONA N, PALOU E & LOPEZ-MALO, A. 2016. Estimation of mass transfer coefficients of the extraction process of essential oil from orange peel using microwave assisted extraction. *J Food Eng* 170: 136-143.
- FREITAS IR & CATTELAN MG. 2018. Antimicrobial and Antioxidant Properties of Essential Oils in Food Systems—An Overview. In: *Microbial Contamination and Food Degradation*, Amsterdam: Elsevier, p. 443-470.
- GARRIDO T, GIZDAVIC-NIKOLAIDIS M, LECETA I, URDANPILLETA M, GUERRERO P, DE LA CABA K & KILMARTIN PA. 2019. Optimizing the extraction process of natural antioxidants from chardonnay grape marc using microwave-assisted extraction. *J Waste Manag* 88: 110-117.
- GAVAHIAN M & FARAHNAKY A. 2018. Ohmic-assisted hydrodistillation technology: A review. *Trends Food Sci Technol* 72: 153-161.
- GAVAHIAN M, LEE YT & CHU YH. 2018. Ohmic-assisted hydrodistillation of citronella oil from Taiwanese citronella grass: Impacts on the essential oil and extraction medium. *Innovative Food Sci Emerging Technol* 48: 33-41.
- GHADERI-GHAHFAROKHI M, BARZEGAR M, SAHARI MA, AHMADI GAVLIGHI H & GARDINI F. 2017. Chitosan-cinnamon essential oil nano-formulation: Application as a novel additive for controlled release and shelf life extension of beef patties. *Int J Biol Macromol* 102: 19-28.
- GONZÁLEZ-LOCARNO M, PAUTT YM, ALBIS A, LÓPEZ EF & TOVAR CDG. 2020. Assessment of chitosan-rue (*Ruta graveolens* L.) essential oil-based coatings on refrigerated cape gooseberry (*Physalis peruviana* L.) quality. *Appl Sci* 10: 2684.
- GOULA AM, THYMIATIS K & KADERIDES K. 2016. Valorization of grape pomace: Drying behavior and ultrasound extraction of phenolics. *Food Bioprod Process* 100: 132-144.
- GUEDES JPS, MEDEIROS JAC, SILVA RRS, SOUSA JMB, CONCEIÇÃO ML & SOUZA EL. 2016. The efficacy of *Mentha arvensis* L. and *M. piperita* L. essential oils in reducing pathogenic bacteria and maintaining quality characteristics in cashew, guava, mango, and pineapple juices. *Int J Food Microbiol* 238: 183-192.

- HASANI S, GHORBANI M & HASANI M. 2020. Nano-encapsulation of lemon essential oil approach to reducing the oxidation process in fish burger during refrigerated storage. *J Food Biosci Technol* 10: 35-46.
- HASHEMI M, DASTJERDI AM, SHAKERARDEKANI A, & MIRDEHGHAN S H. 2020. Effect of alginate coating enriched with Shirazi thyme essential oil on quality of the fresh pistachio (*Pistacia vera* L.). *J Food Sci Technol* 58: 34-43.
- HASHEMI SMB, NIKMARAM N, ESTEGHLAL S, MOUSAVI KHANEGHAH A, NIAKOUSARI M, BARBA FJ, ROOHINEJAD S & KOUBAA M. 2017. Efficiency of Ohmic assisted hydrodistillation for the extraction of essential oil from oregano (*Origanum vulgare* subsp. *viride*) spices. *Innovative Food Sci Emerging Technol* 41: 172-178.
- HAUTE SV, RAES K, DEVLIEGHERE F & SAMPERS I. 2017. Combined use of cinnamon essential oil and MAP/vacuum packaging to increase the microbial and sensorial shelf life of lean pork and salmon. *Food Packag Shelf Life* 12: 51-58.
- HAUTE SV, RAES K, MEEREN PVD & SAMPERS I. 2016. The effect of cinnamon, oregano and thyme essential oils in marinade on the microbial shelf life of fish and meat products. *Food Control* 68: 30-39.
- HEGEL P, MABE G, BRIGNOLE EA & PEREDA S. 2013. Phase equilibrium engineering of jojoba oil extraction with mixed-CO₂ + propane solvent. *J Supercrit Fluids* 79: 114-122.
- HERRERO M, CIFUENTES A & IBAÑEZ E. 2006. Sub- and supercritical fluid extraction of functional ingredients from different natural sources: Plants, food-by-products, algae and microalgae - A review. *Food Chem* 98: 136-148.
- HEYDARI-MAJD M, GHANBARZADEH B, SHAHIDI-NOGHABI M, NAJAFI MA & HOSSEINI M. 2019. A new active nanocomposite film based on PLA/ZnO nanoparticle/essential oils for the preservation of refrigerated *Otolithes ruber* fillets. *Food Packag Shelf Life* 19: 94-103.
- JAHANI M, PIRA M & AMINIFARD MH. 2020. Antifungal effects of essential oils against *Aspergillus niger* in vitro and in vivo on pomegranate (*Punica granatum*) fruits. *Sci Hortic (Amsterdam)* 264: 109188.
- JAVAHERZADEH R, TABATABAEE BAFROEE AS & KANJARI A. 2020. Preservation effect of *Polylophium involucreatum* essential oil incorporated poly lactic acid/ nanochitosan composite film on shelf life and sensory properties of chicken fillets at refrigeration temperature. *LWT - Food Sci Technol* 118: 108783.
- JIANG J, GONG L, DONG Q, KANG Y, OSAKO K & LI L. 2020. Characterization of PLA-P3,4HB active film incorporated with essential oil: Application in peach preservation. *Food Chem* 313: 126134.
- JU J, XIE Y, GUO Y, CHENG Y, QIAN H & YAO W. 2019. Application of edible coating with essential oil in food preservation. *Crit Rev Food Sci Nutr* 59: 2467-2480.
- KHORSHIDIAN N, YOUSEFI M, HANNIRI E & MORTAZAVIAN AM. 2018. Potential application of essential oils as antimicrobial preservatives in cheese. *Innov Food Sci Emerg Technol* 45: 62-72.
- KNEZ HRNČIČ M, CÖR D & KNEZ Ž. 2018. Subcritical extraction of oil from black and white chia seeds with n-propane and comparison with conventional techniques. *J Supercrit Fluids* 140: 182-187.
- KSOUDA G, SELLIMI S, MERLIER F, FALCIMAIGNE-CORDIN A, THOMASSET B, NASRI M & HAJJI M. 2019. Composition, antibacterial and antioxidant activities of *Pimpinella saxifraga* essential oil and application to cheese preservation as coating additive. *Food Chem* 288: 47-56.
- KWON S, CHANG Y & HAN J. 2017. Oregano essential oil-based natural antimicrobial packaging film to inactivate *Salmonella enterica* and yeasts/molds in the atmosphere surrounding cherry tomatoes. *Food Microbiol* 65: 114-121.
- LICON CC, MORO A, LIBRÁN CM, MOLINA AM, ZALACAIN A, BERRUGA MI & CARMONA M. 2020. Volatile transference and antimicrobial activity of cheeses made with Ewes' milk fortified with essential oils. *Foods* 9: 35.
- LIN L, MAO X, SUN Y, RAJIVGANDHI G & CUI H. 2019. Antibacterial properties of nanofibers containing chrysanthemum essential oil and their application as beef packaging. *Int J Food Microbiol* 292: 21-30.
- LOPES NDEL, ALMEIDA-COUTO JMFDE, SILVA CDA, BISINOTTO PEREIRA M, COLOMBO PIMENTEL T, BARÃO CE & CARDOZO-FILHO L. 2020. Evaluation of the effects of pressurized solvents and extraction process parameters on seed oil extraction in *Pachira aquatica*. *J Supercrit Fluids* 161: 104823.
- LU J, FENG X, HAN Y & XUE C. 2014. Optimization of subcritical fluid extraction of carotenoids and chlorophyll a from *Laminaria japonica* Aresch by response surface methodology. *J Sci Food Agric* 94: 139-145.
- LUQUE DE CASTRO MD & PRIEGO-CAPOTE F. 2010. Soxhlet extraction: Past and present panacea. *J Chromatogr A* 1217: 2383-2389.
- LUQUE DE CASTRO MD & PRIEGO-CAPOTE F. 2012. Soxhlet extraction versus accelerated solvent extraction. In: *Comprehensive Sampling and Sample Preparation*. Amsterdam: Elsevier, p. 83-102.

- MA Q, ZHANG Y, CRITZER F, DAVISON PM, ZIVANOVIC S & ZHONG Q. 2016. Physical, mechanical, and microbial properties of chitosan films with microemulsions of cinnamon bark oil and soybean oil. *Food Hydrocoll* 52: 533-542.
- MASANGO P. 2005. Cleaner production of essential oils by steam distillation. *J Cleaner Prod* 13: 833-839.
- MATEO EV, GÓMEZ JV, DOMÍNGUEZ I, GIMENO-ADELANTADO JV, MATEO CASTRO R, GAVARA R & JIMÉNEZ M. 2017. Impact of bioactive packaging systems based on EVOH films and essential oils in the control of aflatoxigenic fungi and aflatoxin production in maize. *Int J Food Microbiol* 254: 36-46.
- MATUSIEWICZ H & ŚLACHCIŃSKI M. 2014. Development of a one-step microwave assisted subcritical water extraction for simultaneous determination of inorganic elements (Ba, Ca, Cu, Fe, Mg, Mn, Na, Pb, Sr, Zn) in reference materials by microwave induced plasma spectrometry. *Microchem J* 115: 6-10.
- MELLO BTF, IWASSA IJ, CUCO RP, DOS SANTOS GARCIA VA & SILVA C. 2019. Methyl acetate as solvent in pressurized liquid extraction of crambe seed oil. *J Supercrit Fluid* 145: 66-73.
- MIAO J, CHE K, XI R, HE L, CHEN X, GUAN X, ZHUANG X, WEN X & CAO Y. 2013. Characterization and benzo[a]pyrene content analysis of camellia seed oil extracted by a novel subcritical fluid extraction. *J Am Oil Chem Soc* 90: 1503-1508.
- MIHIN HB, SOMDA MK, KABORE D, SANON S, AKAKPO AY, SEMDE Z, TRAORE A S & OUATTARA AS. 2019. Biopreservation of meat using the essential oil from *Hyptis suaveolens* Poit. (Lamiaceae) in Burkina Faso. *African J Biotechnol* 18: 808-818.
- MONCADA J, TAMAYO JA & CARDONA CA. 2014. Techno-economic and environmental assessment of essential oil extraction from Citronella (*Cymbopogon winteriana*) and Lemongrass (*Cymbopogon citrus*): A Colombian case to evaluate different extraction technologies. *Ind Crop Prod* 54: 175-184.
- MURMU SB & MISHRA HN. 2018. The effect of edible coating based on Arabic gum, sodium caseinate and essential oil of cinnamon and lemon grass on guava. *Food Chem* 245: 820-828.
- MUSTAFA A & TURNER C. 2011. Pressurized liquid extraction as a green approach in food and herbal plants extraction: a review. *Anal Chim Acta* 703: 8-18.
- NIETO A, BORRULL F, POCURULL E & MARCÉ RM. 2010. Pressurized liquid extraction: A useful technique to extract pharmaceuticals and personal-care products from sewage sludge. *Trac-Trend Anal Chem* 29: 752-764.
- NOORI S, ZEYNALI F & ALMASI H. 2018. Antimicrobial and antioxidant efficiency of nanoemulsion based edible coating containing ginger (*Zingiberofficinale*) essential oil and its effect on safety and quality attributes of chicken breast fillets. *Food Control* 84: 312-320.
- OLIVEIRA MA, GONZAGA M LC, BASTOS MSR, MAGALHÃES HCR, BENEVIDES S D, FURTADO RF, ZAMBELLI RA & GARRUTI DS. 2020. Packaging with cashew gum/gelatin/essential oil for bread: Release potential of the citral. *Food Packag Shelf Life* 23: 100431.
- PALLA C, HEGEL P, PEREDA S & BOTTINI S. 2014. Extraction of jojoba oil with liquid CO₂ + propane solvent mixtures. *J Supercrit Fluids* 91:37-45.
- PALMA M, BARBERO GF, PIÑEIRO Z, LIAZID A, BARROSO CG, ROSTAGNO MA, PRADO JM & MEIRELES, M A A. Extraction of natural products: Principles and fundamental aspects. In: *Natural Product Extraction: Principles and Applications*. Oxford: Woodhead Publishing, 450 p.
- PANJA P. 2018. Green extraction methods of food polyphenols from vegetable materials. *Curr Opin Food Sci* 23: 173-182.
- PAVELA R. 2015. Essential oils for the development of eco-friendly mosquito larvicides: A review. *Ind Crop Prod* 76: 174-187.
- PAVLIĆ B, VIDOVIĆ S, VLADIĆ J, RADOSAVLJEVIĆ R & ZEKOVIĆ Z. 2015. Isolation of coriander (*Coriandrum sativum* L.) essential oil by green extractions versus traditional techniques. *J Supercrit Fluid* 99: 23-28.
- PERIASAMY VS, ATHINARAYANAN J & ALSHATWI AA. 2016. Anticancer activity of an ultrasonic nanoemulsion formulation of *Nigella sativa* L. essential oil on human breast cancer cells. *Ultrason Sonochem* 31: 449-455.
- PINA-BARRERA, A M, ALVAREZ-ROMAN, R, BAEZ-GONZALEZ, J G, AMAYA GUERRA, C A, RIVAS-MORALES, C, GALLARDO-RIVERA CT, & GALINDO- RODRIGUEZ SA. 2019. Application of a multisystem coating based on polymeric nanocapsules containing essential oil of *Thymus Vulgaris* L. to increase the shelf life of table grapes (*Vitis Vinifera* L.). *IEEE Trans Nanobioscience* 18: 549-557.
- PINTO GMF, PINTO JF & JARDIM ICSF. 2018. Extração com fluido supercrítico. *Rev Chemkeys* 4: 1-13.
- PORTO CDA, NATOLINO A & DECORTI D. 2015. The combined extraction of polyphenols from grape marc: Ultrasound assisted extraction followed by supercritical CO₂ extraction of ultrasound-raffinate. *LWT - Food Sci Technol* 61: 98-104.
- PRADO JM, VARDANEGA R, DEBIEN ICN, MEIRELES MADEA, GERSCHENSON LN, SOWBHAGYA HB & CHEMAT S. 2015.

- Conventional extraction. In: Food Waste Recovery. San Diego: Academic Press, p. 127-148.
- RAI A, BHARGAVA R & MOHANTY B. 2017. Simulation of supercritical fluid extraction of essential oil from natural products. *J Appl Res Med Aromat Plants* 5: 1-9.
- RIBEIRO-SANTOS R, ANDRADE M, MELO NR & SANCHES-SILVA A. 2017. Use of essential oils in active food packaging: Recent advances and future trends. *Trends Food Sci Technol* 61: 132-140.
- RODSAMRAN P & SOTHORNVIT R. 2019. Extraction of phenolic compounds from lime peel waste using ultrasonic-assisted and microwave-assisted extractions. *Food Biosci* 28: 66-73.
- ROMANI VP, PRENTICE-HERNÁNDEZ C & MARTINS VG. 2017. Active and sustainable materials from rice starch, fish protein and oregano essential oil for food packaging. *Ind Crop Prod* 97: 268-274.
- ROSA CG, MELO APZ, SGANZERLA WG, MACHADO MH, NUNES MR, MACIEL MV OB, BERTOLDI FC & BARRETO PLM. 2020. Application in situ of zein nanocapsules loaded with *Origanum vulgare* Linneus and *Thymus vulgaris* as a preservative in bread. *Food Hydrocoll* 99: 105339.
- SAMARAM S, MIRHOSSEINI H, TAN CP & GHAZALI HM. 2014. Ultrasound assisted extraction and solvent extraction of papaya seed oil: Crystallization and thermal behavior, saturation degree, colour and oxidative stability. *Ind Crop Prod* 52: 702-708.
- SÁNCHEZ-CAMARGO AP, IBÁÑEZ E, CIFUENTES A & HERRERO M. 2017. Bioactives obtained from plants, seaweeds, microalgae and food by-products using pressurized liquid extraction and supercritical fluid extraction. *Compr Anal Chem* 76: 27-51.
- SANTOS MP, SOUZA MC, SUMERE BR, SILVA LC, CUNHA DT, BEZERRA RMN & ROSTAGNO MA. 2019. Extraction of bioactive compounds from pomegranate peel (*Punica granatum* L.) with pressurized liquids assisted by ultrasound combined with an expansion gas. *Ultrason Sonochem* 54: 11-17.
- SHANGE N, MAKASI T, GOUWS P & HOFFMAN LC. 2019. Preservation of previously frozen black wildebeest meat (*Connochaetes gnou*) using oregano (*Oreganum vulgare*) essential oil. *Meat Sci* 148: 88-95.
- SHARAYE P, AZARPAZHOOH E, ZOMORODI S & RAMASWAMY HS. 2019. Ultrasound assisted extraction of bioactive compounds from pomegranate (*Punica granatum* L.) peel. *LWT - Food Sci Technol* 101: 342-350.
- SHIVONEN M, JARVENPAA E, HIETANIEMI V & HUOPALAHTI R. 1999. Advances in supercritical carbon dioxide technologies. *Trends Food Sci Tech* 10: 217-222.
- SHOKRI S, PARASTOUEI K, TAGHDIR M & ABBASZADEH S. 2020. Application an edible active coating based on chitosan-Ferulago angulata essential oil nanoemulsion to shelf life extension of Rainbow trout fillets stored at 4 °C. *Int J Biol Macromol* 153: 846-854.
- SILVA CS, FIGUEIREDO HM, STAMFORD TLM & SILVA LHM. 2019a. Inhibition of *Listeria monocytogenes* by *Melaleuca alternifolia* (tea tree) essential oil in ground beef. *Int J Food Microbiol* 293: 79-86.
- SILVA ISV, PRADO NS, MELO PG, ARANTES DC, ANDRADE MZ, OTAGURO H & PASQUINI D. 2019b. Edible coatings based on apple pectin, cellulose nanocrystals, and essential oil of lemongrass: Improving the quality and shelf life of strawberries (*fragaria ananassa*). *J Renew Mater* 7: 73-87.
- SILVA LCDA, SOUZA MC, SUMERE BR, SILVA LGS, CUNHA DTDA, BARBERO GF, BEZERRA RMN & ROSTAGNO MA. 2020. Simultaneous extraction and separation of bioactive compounds from apple pomace using pressurized liquids coupled on-line with solid-phase extraction. *Food Chem* 318: 126450.
- SINGH CHOUHAN KB, TANDEY R, SEN KK, MEHTA R & MANDAL V. 2019. Critical analysis of microwave hydrodiffusion and gravity as a green tool for extraction of essential oils: Time to replace traditional distillation. *Trends Food Sci Technol* 92: 12-21.
- SIROCCHI V, DEVLIEGHERE F, PEELMAN N, SAGRATINI G, MAGGI F, VITTORI S & RAGAERT P. 2017. Effect of *Rosmarinus officinalis* L. essential oil combined with different packaging conditions to extend the shelf life of refrigerated beef meat. *Food Chem* 221: 1069-1076.
- SMAOUI S, HSOUNA AB, LAHMAR A, ENNOURI K, TIBAA-CHAKCHOUK A, SELLEM I, NAJAH S, BOUAZIZ M & MELLOULI, L. 2016. Bio-preservative effect of the essential oil of the endemic *Mentha piperita* used alone and in combination with BacTN635 in stored minced beef meat. *Meat Sci* 117: 196-204.
- ŠOJIĆ B ET AL. 2019. Coriander essential oil as natural food additive improves quality and safety of cooked pork sausages with different nitrite levels. *Meat Sci* 157: 107879.
- SOUZA VGL, PIRES JRA, VIEIRA ÉT, COELHO IM, DUARTE MP & FERNANDO A L. 2019. Activity of chitosan-montmorillonite bionanocomposites incorporated with rosemary essential oil: From in vitro assays to application in fresh poultry meat. *Food Hydrocoll* 89: 241-252.
- TEKIN K, AKALIN MK & SEKER MG. 2015. Ultrasound bath-assisted extraction of essential oils from clove using central composite design. *Ind Crop Prod* 77: 954-960.

TRENTINI CP, SANTOS KA, SILVA EA, GARCIA VAS, CARDOZO-FILHO L & SILVA C. 2017. Oil extraction from macauba pulp using compressed propane. *J Supercrit Fluid* 126: 72-78.

WU Y, LUO Y, ZHOU B, LEI M, WANG Q & ZHANG B. 2019. Porous 174-178metal organic framework (MOF) carrier for incorporation of volatile antimicrobial essential oil. *Food Control* 98: 174-178.

XU JL, KIM TJ, KIM JK & CHOI Y. 2019. Simultaneous roasting and extraction of green coffee beans by pressurized liquid extraction. *Food Chem* 281: 261-268.

YAO X-H, ZHANG D-Y, LUO M, JIN S, ZU Y-G, EFFERTH T & FU Y-J. 2015. Negative pressure cavitation-microwave assisted preparation of extract of *Pyrola incarnata* Fisch. rich in hyperin, 2-O-galloylhyperin and chimaphilin and evaluation of its antioxidant activity. *Food Chem* 169: 270-276.

YEN HF, HSIEH CT, HSIEH TJ, CHANG FR & WANG CK. 2015. In vitro anti diabetic effect and chemical component analysis of 29 essential oils products. *J Food Drug Anal* 23: 124-129.

YIN C, HUANG C, WANG J, LIU Y, LU P & HUANG L. 2019. Effect of chitosan- and alginate-based coatings enriched with cinnamon essential oil microcapsules to improve the postharvest quality of mangoes. *Materials* 12: 2039.

YOUSEFI M, RAHIMI-NASRABADI M, POURMORTAZAVI SM, WYSOKOWSKI M, JESIONOWSKI T, EHRlich H & MIRSADEGHI S. 2019. Supercritical fluid extraction of essential oils. *TrAC – Trend Anal Chem* 118: 182-193.

YU Q, LI C, DUAN Z, LIU B, DUAN W & SHANG F. 2017. Ultrasonic microwave assisted extraction of polyphenols, flavonoids, triterpenoids, and Vitamin C from *Clinacanthus nutans*. *Czech J Food Sci* 35: 89-94.

ZANQUI AB, SILVA CMDA, RESSUTTE JB, MORAIS DRDE, SANTOS JM, EBERLIN MN, CARDOZO-FILHO L, SILVA EADA, GOMES STM & MATSUSHITA, M. 2020. Extraction and assessment of oil and bioactive compounds from cashew nut (*Anacardium occidentale*) using pressurized n-propane and ethanol as cosolvent. *J Supercrit Fluids* 157: 104686.

ZERMANE A, LARKECHE O, MENIAI A-H, CRAMON C & BADENS E. 2016. Optimization of Algerian rosemary essential oil extraction yield by supercritical CO₂ using response surface methodology. *C R Chim* 19: 538-543.

ZHENG L, SHI LK, ZHAO CW, JIN QZ & WANG XG. 2017. Fatty acid, phytochemical, oxidative stability and in vitro antioxidant property of sea buckthorn (*Hippophaë rhamnoides* L.) oils extracted by supercritical and subcritical technologies. *LWT - Food Sci Technol* 86: 507-513.

ZHONG J, WANG Y, YANG R, LIU X, YANG Q & QIN X. 2018. The application of ultrasound and microwave to increase oil

extraction from *Moringa oleifera* seeds. *Ind. Crops Prod* 120: 1-10.

ZYGLER A, SŁOMIŃSKA M & NAMIEŚNIK J. 2012. Soxhlet Extraction and New Developments Such as Soxtec. In: *Comprehensive Sampling and Sample Preparation*. Canada: Academic Press, 3200 p.

How to cite

DE ALMEIDA-COUTO JMF, RESSUTTE JB, CARDOZO-FILHO L & CABRAL VF. 2022. Current extraction methods and potential use of essential oils for quality and safety assurance of foods. *An Acad Bras Cienc* 93: e20191270. DOI 10.1590/0001-376520220191270.

Manuscript received on October 14, 2019; accepted for publication on December 28, 2020

JÉSSICA M.F. DE ALMEIDA-COUTO¹

<https://orcid.org/0000-0001-7032-8421>

JÉSSICA B. RESSUTTE²

<https://orcid.org/0000-0002-4057-0695>

LÚCIO CARDOZO-FILHO¹

<https://orcid.org/0000-0002-1764-9979>

VLADIMIR F. CABRAL³

<https://orcid.org/0000-0002-5835-9249>

¹Universidade Estadual de Maringá/UEM, Departamento de Engenharia Química, Av. Colombo nº 5.790, 87020-900 Maringá, PR, Brazil

²Universidade Estadual de Londrina/UEL, Departamento de Ciência e Tecnologia de Alimentos/UEL, Rodovia Celso Garcia Cid, 86057970 Londrina, PR, Brazil

³Universidade Estadual de Maringá/UEM, Departamento de Engenharia de Alimentos, Av. Colombo nº 5.790, 87020-900 Maringá, PR, Brazil

Correspondence to: **Jéssica B. Ressutte**

E-mail: jessicaressutte@gmail.com

Author contributions

Manuscript writing: Jéssica M. F. de Almeida-Couto and Jéssica B. Ressutte; Manuscript review: Lúcio Cardozo-Filho and Vladimir F. Cabral.

