

Acta Botanica Brasilica - 32(1): 12-19. January-March 2018. doi: 10.1590/0102-33062017abb0121

Disentangling plant establishment in sandy coastal systems: biotic and abiotic factors that determine *Allagoptera arenaria* (Arecaceae) germination

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Received: April 19, 2017 Accepted: August 15, 2017

ABSTRACT

Germination rate and establishment success of plants in harsh environments depend on the ability of seeds to withstand unfavorable environmental conditions and avoid predators. Brazilian coastal plains, known as *restinga*, are subject to environmental factors that seriously limit plant establishment and survival (e.g. salinity, desiccation, oligotrophy, flooding, high temperature and radiation levels). We tested, both in field and laboratory experiments, conditions for germination and establishment of *Allagoptera arenaria*, a palm tree often found in *restinga* ecosystems of southeastern Brazil, and which have a principal role in plant community dynamics. Our results showed that the absence of mesocarp, high radiation exposure, and temperature were the main drivers of seed germination. In the field, the highest germination rate was linked to nude seeds buried in open areas. High temperatures and/or predation damaged seeds that remained on the soil surface, especially if they were close to the mother plant and alongside dung piles made by dispersers. Under controlled conditions, seeds exhibited optimum germination at 35 °C. Therefore, the germination and establishment of *A. arenaria* depend as much on environmental conditions as on a network of interactions including vertebrates and invertebrates, which allow this species to colonize harsh, open areas in *restinga* ecosystems.

Keywords: Arecaceae, facilitation, resting, secondary dispersion, seedling

Introduction

The structure of plant populations is intimately linked to recruitment, and seed germination and seedling establishment are periods of high risk in the plant life cycle, which must occur at the right time and under appropriate conditions (Silvertown & Charlesworth 2001; Tobe *et al.* 2005). Many post-dispersal factors can have profound

influence on recruitment (Roberts & Heithaus 1986; Schupp 1990; Howe 1993; Shepherd & Chapman 1998) including environmental conditions such as temperature, radiation, and water availability that greatly influence the development of the embryonic axis during germination (Carvalho & Nakagawa 2000; Vandelook *et al.* 2008; Baskin & Baskin 2014), and seed predation (Janzen 1971; Howe & Smallwood 1982; Schupp 1990). Dealing with unfavorable environmental conditions (Hölzel & Otte 2004) and escaping

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predation often involve mutualistic interactions, such as long seed dispersal away from the mother plant (Janzen 1970; Wicklow et al. 1984; Estrada & Coates-Estrada 1991; Comita et al 2014). This is the case in restinga communities, which occur in sandy plains along the Brazilian coast and show great spatial and temporal fluctuations in water availability (Martin et al. 1993). Even during the rainy season, periods of water shortage can occur due to dry spells and the low water retention capacity of sandy soil (Cavalin & de Mattos 2007; Braz & Mattos 2010).

The ability to germinate under environmental constraints gives a species the chance to become established in a community (Bolfrey-Arku et al. 2011). In contrast to species unable to stand high temperatures and low water availability (Fialho & Furtado 1993; Pinheiro & Borghetti 2003; Mantovani & Iglesias 2008), some restinga species establish in open, sandy sites (Zaluar & Scarano 2000; Scarano 2002) and often act as facilitators of other plant species (Scarano 2002; Scarano et al. 2004).

The palm tree, *Allagoptera arenaria* grows in *restinga* areas in southeastern Brazil (Menezes & Araujo 2000), showing high recruitment rates on bare soil, and acting as nucleus of vegetation patches (Zaluar & Scarano 2000). Mature palms modify the environment under their canopy through litter accumulation, increased soil nutrient content, shade, and decreased wind velocity (Menezes & Araujo 1999). Although we have a good understanding of *A. arenaria* life history and dynamics in *restinga* environments, there is a lack of knowledge regarding recruitment conditions in exposed *restinga* soils.

Assuming that seed desiccation and predation are the main post-dispersal causes limiting plant establishment, and that long-distance dispersion from mother plants reduce predation rates (Janzen-Connell effect), we explored mechanisms by which *A. arenaria* thrives in *restinga* habitats. Our hypotheses were that establishment does occur through processes that include mitigation of adverse conditions, especially by burial, and protection against predators through a network of positive interactions involving several species of vertebrates and invertebrates.

Materials and methods

Field site and species

Our field site was at the *Restinga* de Marambaia (23°02'S 43°35'W, sea level), Rio de Janeiro State, where vegetation patches are scattered over a sand matrix with a sparse herbaceous layer (Menezes & Araujo 2005). Climate is tropical rainy (Aw in Köeppen classification) with a mean annual temperature of 23.7 °C, being February the warmest month (mean of 26.8 °C) and August the coldest (20.9 °C). Mean annual precipitation is 1,240 mm, 37 % of which

falls in summer. Winter months are significantly drier, with only about 15 % of the annual precipitation falling in approximately 21 days (Mattos 2005). *Restinga* sites show significant spatial variation in environmental conditions, which are buffered in vegetation patches; surface temperature of bare sand can reach 70 °C on a typical summer day (Dias *et al.* 2005), while maximum soil temperatures beneath vegetation patches remain near 25 °C (Mantovani & Iglesias 2008).

Allagoptera arenaria is a palm species about 2 m high which forms dense populations in certain restinga sites (Menezes & Araujo 1999) and has a small distribution range between southern Bahia State and the northern coast of São Paulo State (Lorenzi et al. 2004). Infructescences contain five to 130 orange to yellowish one-seeded fruits, 12-20 mm long and 10-13 mm in diameter with a sweet fibrous mesocarp and a stony endocarp. The corolla is persistent, covering the fruit. The micropile is lateral and the three germ pores are located slightly basal or lateral. The endosperm is homogeneous and highly rich in lipids (Henderson et al. 1995; Moraes 1996; Henderson 2006; Grenha et al. 2010). Fruits are mainly dispersed by the crab-eating fox, Cerdocyon thous, and the raccoon, Procyon cancrivorus (Gatti et al. 2006; Andreazzi et al. 2009). As other palm species, A. arenaria fruits are highly predated by Bruchinae beetles (Grenha et al. 2008) and exit holes of Bruchinae species evidencing fruit predation, are easy to recognize at naked eye. The type of germination in *A. arenaria* seeds is the remote tubular. After germination, the seedling develops a plagiotropous form and becomes upright after a short or long period of growth (Moraes 1996).

The A. arenaria seeds used in the experiments were collected at the Marambaia restinga in November 2004. Twenty-five thousand fruits were collected, 4 fruits per individual, to guarantee maximum genetic variability in the sample of the tested seeds. The ripe fruits were collected directly from the inflorescences selecting only the yellow staining ones. After harvesting, the fruits were taken to the laboratory and the epicarp and mesocarp were mechanically removed with a knife. A manual selection was performed to discard wilted and damaged seeds, and then the samples were homogenized. Those that did not reach 1.2 cm in width were discarded, since this was the limit found by Leite (1990) for aborted and unviable seeds.

Field experiments

Germination rate of *A. arenaria* was determined under four conditions: either seeds with or without mesocarp were placed on the soil surface or buried 5 cm deep. This burial depth was selected, as temperature 5 cm below the soil surface does not exceed 30 °C at midday (Scarano 2002). We used 8 replicates of 25 seeds each, totaling 1600 seeds.



Experimental plots were protected by wire mesh to avoid predation and seeds were not surface-sterilized. Buried seeds were extracted every two weeks along a 150-day period to determine germination status; non-germinating seeds were buried again. The germination criterion used was the emergence of the 1 mm-long cotyledonary petiole. At the end of the experiment, seeds that did not germinate were sectioned to look for the presence of Bruchinae larvae.

To test for the Janzen-Connell effect we collected five fresh fruits with mesocarp from under 20 *A. arenaria* individuals and five seeds from 20 dung piles of dispersers animals selected 50 m far from the nearest *A. arenaria* individual. Predation criteria were either the presence of Bruchinae larvae or exit holes.

Lab experiments

Four thousand *A. arenaria* seeds were collected in November 2004 from approximately 800 individuals. Mesocarps were removed manually using a knife, and old seeds and those damaged by insects were discarded; seeds less than 1.2 cm wide were also excluded as non-viable (Leite 1990).

To examine the combined effects of temperature and light on germination, seeds were exposed to five treatments at steady temperature (15, 20, 25, 30 and 35 °C) under constant white light (fluorescent light at 30 $\mu mol\ m^{-2}\ s^{-1}$) and dark conditions, using five replicates of 16 seeds each (800 seeds in total). Before the experiment, seeds were surface-sterilized by soaking them three times for 3 min in 5 % sodium hypochlorite solution and then rinsed three times for 3 min with deionized water. Seeds were subsequently sown onto sterilized sand in germination boxes and periodically watered with distilled water. Seeds were checked for germination on a weekly basis for 150 days.

Data analyses

We used t-test and Analysis of Variance (ANOVA), followed by Tukey test to compare mean number of germinated and infested *A. arenaria* seeds when data conformed to parametric premises either before or after transformation. If data did not conform to parametric premises, we compared mean number of germinated and infested *A. arenaria* seeds with the randomization test suggested by Pillar & Orlóci (1996). Post-hoc tests to evaluate significance of differences between means compared with the randomization procedure were done using contrasts between levels of factors (Pillar & Orlóci 1996). Randomization and post-hoc (contrasts) tests were conducted with the MULTIV software package (Pillar 1997). Data are presented as mean ± sd throughout the manuscript. Parametric analyses were performed using the basic interface of the software R v3.0.1 (R Development Core Team 2013)

Results

Seed germination in the field

Seed germination of Allagoptera arenaria in the field depended on the presence of mesocarp (Q = 165.8; P = 0.001, Fig. 1) being the germination rate greater when mesocarp was absent than when it was present (3.7 \pm 5.5 vs. 0.5 \pm 1.0, respectively). Position was also a determinant for A. arenaria germination (Q = 276.4; P = 0.001), being higher when seeds were buried than when they were on the soil surface $(4.2 \pm 5.2 \text{ vs. 0}, \text{ Fig. 1})$. Finally, light condition also influenced germination (Q = 118.3; P = 0.001), and seeds fully exposed to sunlight germinated more than those in shade $(3.4 \pm 5.6 \text{ vs. } 0.7 \pm 1.2)$. Interaction between factors was significant (Q = 83.3; P = 0.006) and showed A. arenaria germination rate was affected by the interaction between the three factors (presence of mesocarp, light condition and seed position [buried or not]). Optimal conditions for *A*. arenaria germination would then be buried seeds without mesocarp in areas fully exposed to sunlight (Fig. 1).

Seed predation by bruchinae beetles

The frequency of seeds with Bruchinae larvae depended on the presence of mesocarp (Q = 19.1; P = 0.01) since the number of seeds with beetles was greater when mesocarp was present than when it was absent $(2.4 \pm 2.9 \text{ vs } 1.3 \pm 1.8)$. When seeds were laying on the soil surface, the presence of Bruchinae larvae were higher (Q = 206.64; P = 0.001) than when seeds were buried (3.7 \pm 2.4 vs. 0.1 \pm 0.3), and under shade (Q = 15.01; P = 0.007) than in fully exposed sunlight (2.4 \pm 2.7 vs. 1.4 \pm 2.2). There were significant interactions between seed position (sowing conditions) and light condition (Q = 9.8; P = 0.03) and between sowing conditions and presence of mesocarp (Q = 13.1; P = 0.01). Allagoptera arenaria seeds were more exposed to Bruchinae larvae when laying on the soil surface under shade and with mesocarp (Fig. 2A-B). There were no interactions between seed light condition and presence/absence of mesocarp (Q = 1.3; P = 0.44 - Fig. 2C) nor between all three factors (Q = 0.14; P = 0.87 - Fig. 2D). Additionally, we found that A. arenaria seeds laying on the soil surface next to the mother plant were more predated by beetles than seeds laying in dung piles 50 m apart from a mother plant (t = 13.125, sd= 37.998, p-value < 0.0001).

Laboratory seed germination

We found strong effects of mesocarp, light and temperature on *A. arenaria* germination rate in the lab. Seeds without mesocarp had greater germination rate than seeds with mesocarp (t = -4.7; gl = 8; p-value = 0.001, Fig. 3A);

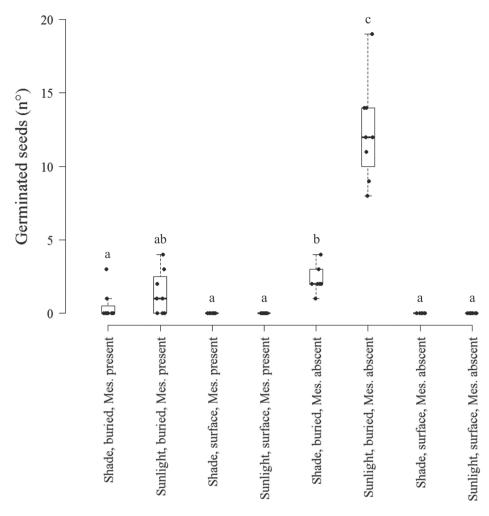


Figure 1. Germination rate of *Allagoptera arenaria* according to habitat, presence of mesocarp and environmental factors. Bars with the same letters are not significantly different (P > 0.05).

seeds germinated more in light than in the dark (Q = 4.8; P = 0.005, Fig. 3B), and germination rate increased linearly with temperature (Q = 74.4; P = 0.001, Fig. 3C). The interaction between light and temperature was not significant (Q = 2.2; P = 0.398).

Discussion

As expected, seeds of *Allagoptera arenaria* strongly depended for germination and establishment on environmental conditions and interactions with dispersers. In both, field and laboratory experiments, the absence of mesocarp, exposure to light, and temperature were critical for seed germination. Field experiments showed that *A. arenaria* seeds placed directly onto the soil surface had poor germination rates, both in the open and under the canopy. In addition, the presence of mesocarp likewise limited germination. Temperature oscillations may trigger the

germination process by breaking the testa in some seeds and in palms could interfere with the process of clearing the germ pore, weakening the pore plate or the operculum (Henderson 2006; Baskin & Baskin 2014). Thus, and even though there is greater water availability beneath tree canopies (Pugnaire *et al.* 1996; Zaluar & Scarano 2000) the buffering of temperatures did not favor seed germination. This pattern is typical of pioneer species (*sensu* Swaine & Whitmore 1988) such as *A. arenaria*, which prefer open areas even though seeds would be more exposed to predators (Grenha *et al.* 2008).

Allagoptera anrenaria, is often a critical component of the restinga community as it contributes to the survival of other species acting as facilitator (Menezes & Araujo 2000; Zaluar & Scarano 2000; Scarano 2002; Scarano et al. 2004). Facilitators are positively associated to other species and are instrumental in increasing vegetation complexity over time (Armas & Pugnaire 2005). This palm tree shows high recruitment rates on bare soil, becoming dominant and occupying a central position in vegetation patches that



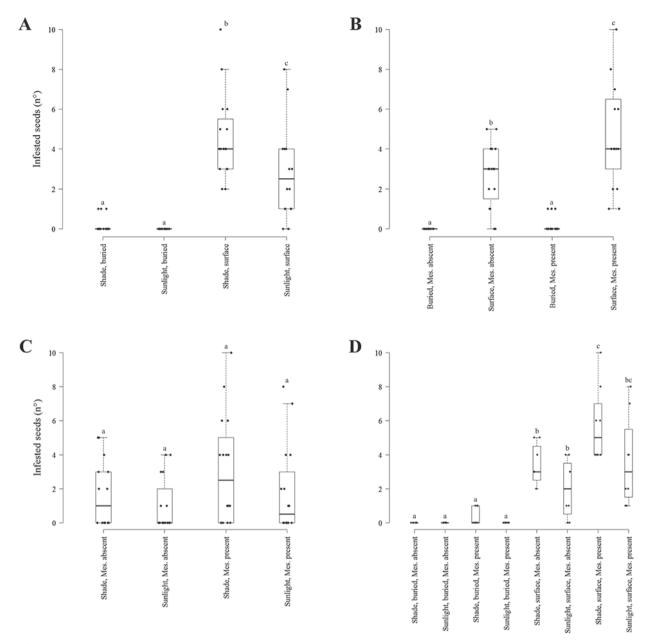


Figure 2. Mean incidence of *Allagoptera arenaria* seed predation by Bruchinae beetles under different conditions. First and second order interactions between environment, mesocarp and sowing condition are shown: **A.** between microhabitat and environmental condition, **B.** between sowing and mesocarp presence; **C.** between environment and mesocarp; **D.** between environment, mesocarp and sowing microhabitat. Bars with the same letters are not significantly different (P> 0.05).

thrive around them, showing a positive spatial association with seedlings of other species (Zaluar & Scarano 2000). Mature individuals act as facilitator species by modifying the environment under their canopy through accumulation of litter, increased soil nutrient content, lowering soil surface and air temperature as well as decreasing wind velocity (Menezes & Araujo 1999).

Our results suggest that a complex sequence of interactions is required for germination success of this *restinga* species. For instance, the high germination rate of naked *A. arenaria* seeds buried in open areas is consistent

with dispersal by the Scarabaeidae Ateuchus squalidus (Leite 1990). These Scarabaeidae use mammalian dung and fruits as food and/or as oviposition sites (Cambefort & Walter 1991; Hanski 1991; Halffter & Matthews 1966). In A. arenaria seeds this beetle is essential for germination. Beetles remove fruits and bury them ca. 5 cm deep, where they feed on the pulp (Leite 1990). Just slightly below the soil surface, temperatures do not exceed 30 °C, close to the optimal germination range for this species. By contrast, seeds remaining on the soil surface in the open would rapidly be damaged by desiccation and

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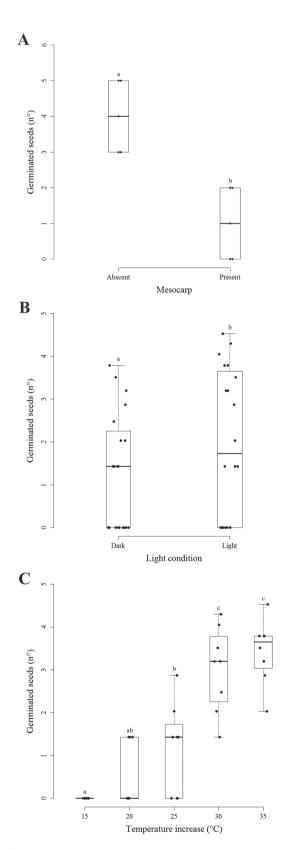


Figure 3. Mean number of seeds germinated in lab conditions **A.** with and without mesocarp, **B.** in conditions of light and dark; and **C.** in five temperature conditions. Black dots represent the dispersion of raw data for each treatment of temperature. Bars with the same letters are not significantly different (P > 0.05).

high temperatures, often reaching 70 °C (Scarano 2002). Therefore, reproductive success of *A. arenaria* strongly depends on the feeding behaviour of *A. squalidus*, an interaction that could be termed as mutualistic. While mutualistic relationships are often difficult to recognize, they are very important in terms of plant population and community structure (Price 1997).

Our results show that *A. arenaria* seeds in dung piles away (50 m) were less predated than those near the parent tree, evidencing the Janzen-Connell effect. Species of the Pachymerini beetle tribe (Bruchinae) feed almost exclusively on palm seeds (Johnson *et al.*1995), being fruit exposure the main cause of bruchinae predation (Silvius & Fragoso 2002). We recorded that mesocarp presence increased seed predation rate. There is a misconception that palm fruits only become predated by Bruchiane beetles after their consumption by frugivores (Wright 1983). However, in our case intact mesocarps significantly contributed to greater Bruchinae predation rates.

Predation by invertebrates is density-dependent, so that the greater accumulation of seeds the greater number of beetles (Wilson & Janzen 1972; Shepherd & Chapman 1998; Galetti et al. 2001). However, Bruchinae beetles generally have low dispersal capacity and move short distances between plants, so that individuals are often restricted to the understory of a palm tree for their whole life (Janzen 1971). Frugivores, by contrast, take fruits away from the mother plant, preventing the accumulation of fruits and thus reducing Bruchinae predation. Therefore, dung piles deposited by dispersers somehow are a way of seed protection against Bruchinae while helping dispersal of seeds to significant distances from the mother plant (Fragoso 1997; Quiroga-Castro & Roldán 2001; Fragoso et al. 2003) avoiding competition for resources (Janzen 1970; Pimentel & Tabarelli 2004; Rios & Pacheco 2006). Bruchinae predation can reach up to 100 % of exposed seeds in some palm species (Wenny 2000; Russo & Augspurger 2004) in contrast to low predation rates in dung piles (Quiroga-Castro & Roldán 2001; Rios & Pacheco 2006). For instance, only 2 % of *Attalea maripa* seeds in dung piles were predated by Bruchinae beetles as compared to 77 % of seeds lying under parent palm trees (Fragoso 1997); and only 6 % of Attalea phalerata seeds in mammalian dung piles were predated, against 61 % of seeds under parent trees (Quiroga-Castro & Roldán 2001).

Long-distance dispersal of *A. arenaria* seeds in *restinga* environments depends on two vertebrates, *Cerdocyon thous* and *Procyon cancrivorus*, in which more than 80 % of scats contain palm seeds (Gatti *et al.* 2006; Andreazzi *et al.* 2009). They are also responsible for the clumped distribution of *A. arenaria*. Dung beetles use both fruits and mammalian dung as fodder and oviposition sites (Cambefort & Walter 1991; Hanski 1991; Halffter & Matthews 1966), acting as secondary dispersers and positively affecting seed survival



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by preventing predation (Wicklow et al. 1984; Estrada & Coates-Estrada 1991; 1996). The minimum temperature for A. arenaria germination was 20 °C (2.7 %), with the greatest germination rate at 35 °C (40 %), unusually high compared to other tropical species (Mello & Barbedo 2007; Cardoso & Pereira 2009; Pires et al. 2009; Pimenta et al. 2010). Optimum temperature for germination corresponds to temperatures usually experienced at the time of seedling emerge (Baskin & Baskin 2014; Bell et al. 1993) which in A. arenaria is the summer period, when water availability is greatest. Temperature control of germination could determine the geographical range of many plant species (Thompson 1973; Probert 1992) and for A. arenaria may represent a limiting factor as temperatures decrease further south.

Conclusion

Our results show that *A. arenaria* colonizes open areas in *restinga* systems mostly through mutualistic interactions that help escape both, unfavorable conditions and predation, thus confirming our hypotheses. On the one hand, vertebrates contribute to long-distance dispersal and the protection of seeds within dung piles, crucial against Bruchinae beetles, the main predators of *A. arenaria* seeds. On the other hand, secondary dispersal by the beetle *Ateuchus squalidus* contributes to germination success. Then, establishment of *A. arenaria* in open areas of *restinga* is the outcome of a complex interaction network between dispersers and predators, which place seeds in the best possible microhabitat for establishment. This interactions network is essential to maintain *restinga* community dynamics.

Acknowledgements

We are grateful to Fátima Piña Rodrigues for advice, assistance and technical help; Fabio Rubio Scarano for comments on an earlier draft of this manuscript. The Centro de Avaliações do Exército (Brazilian Army) for research permission and access to the study area. This work was supported by the Fundação Coordenação de Aperfeiçoamento de Pessoal de Nível Superior under Grant CAPES - Proc. 2543/13-3, and Fundação o Boticário de Proteção à Natureza under Grant Proc. 0512-20012.

References

- Andreazzi CS, Pires AS, Fernandes FAS. 2009. Mamíferos e palmeiras neotropicais: interações em paisagens fragmentadas. Oecologia Brasiliensis 13: 554-574.
- Armas C, Pugnaire FI. 2005. Plant interactions govern population dynamics in a semiarid plant community. Journal of Ecology 93: 978-989.
- Baskin CC, Baskin JM. 2014. Seeds: ecology, biogeography, and evolution of dormancy and germination. San Diego, San Diego Academic Press.
 Bell DT, Plummer JA, Taylor SK. 1993. Seed germination ecology in southwesternWestern Australia. Botanical Review 59: 286-305.

- Bolfrey-Arku GEK, Chauhan BS, Johnson DE. 2011. Seed germination ecology of itchgrass (*Rottboellia cochinchinensis*). Weed Science 59: 182-187.
- Braz MIG, Mattos EA. 2010. Seed dispersal phenology and germination characteristics of a drought-prone vegetation in south-eastern Brazil. Biotropica 42: 327-335.
- Cambefort Y, Walter P. 1991. Dung beetles in tropical forests in Africa. In: Hanski I. Cambefort Y. (ed.) Dung beetle ecology. Princeton, Princeton University Press. p. 198-210.
- Cardoso VJM, Pereira FJM. 2009. Dependência térmica da germinação de sementes de *Drymaria cordata* (L.) Willd. ex Roem. & Schult. (Cariophyllaceae). Acta Botanica Brasilica 23: 305-312.
- Carvalho NM, Nakagawa J. 2000. Sementes: ciência, tecnologia e produção. 4th. edn. Jaboticabal, FUNEP.
- Cavalin PO, Mattos EA. 2007. Spatio-temporal variation of photosynthetic pigments in CAM tree *Clusia hilariana* Schlechtendal associated with dry spells during rainy season in south-eastern Brazil. Trees 21: 671-675.
- Comita LS, Queenborough SA, Murphy SJ, et al. 2014. Testing predictions of the Janzen-Connell hypothesis: a meta-analysis of experimental evidence for distance- and density-dependent seed and seedling survival. Journal of Ecology 102: 845-856.
- Dias ATC, Zaluar HT, Ganade G, Scarano FR. 2005. Canopy composition influencing plant patch dynamics in Brazilian sandy coastal plain. Journal of Tropical Ecology 21: 343-347.
- Estrada A, Coates-Estrada R. 1991. Howler monkeys (*Alouatta palliata*), dung beetles (Scarabaeidae) and seed dispersal: ecological interactions in the tropical rain forest of Los Tuxtlas, Mexico. Journal of Tropical Ecology 7: 459-474.
- Estrada A, Coates-Estrada R. 1996. Tropical rain forest fragmentation and wild populations of primates at Los Tuxtlas, Mexico. International Journal of Primatology 17: 759-784.
- Fialho RF, Furtado ALS. 1993. Germination of *Erythroxylum ovalifolium* (Erythroxylaceae) seeds within the terrestrial bromeliad *Neoregelia cruenta*. Biotropica 25: 359-362.
- Fragoso JMV. 1997. Tapir-generated seed shadows: Scale- dependent patchiness in the Amazon rain forest. Journal of Ecology 85: 519-529.
- Fragoso JMV, Silvius KM, Correa JA. 2003. Long-distance seed dispersal by tapirs increases seed survival and aggregates tropical trees. Ecology 84: 1998-2006.
- Galetti M, Keuroghlian A, Hanada L, Morato MI. 2001. Frugivory and seed dispersal by the lowland tapir (*Tapirus terrestris*) in southeast Brazil. Biotropica 33: 723-726.
- Gatti A, Bianchi R, Rosa CRX, Mendes SL. 2006. Diet of two sympatric carnivores, *Cerdocyon thous* and *Procyon cancrivorus*, in a *restinga* area of Espirito Santo State, Brazil. Journal of Tropical Ecology 22: 227-230.
- Grenha V, Macedo MV, Monteiro RF. 2008. Seed predation on *Allagoptera arenaria* (Gomes) O'Kuntze (Arecaceae) by *Pachymerus nucleorum* Fabricius (Coleoptera, Chrysomelidae, Bruchinae). Revista Basileira de Entomologia 52: 50-56.
- Grenha V, Macedo MV, Pires AS, Monteiro RF. 2010. The role of Cerradomys subflavus (Rodentia, Cricetidae) as seed predator and disperser of the palm Allagoptera arenaria. Mastozoologia Neotropical 17: 61-68.
- Halffter G, Matthews EG. 1966. The natural history of dung beetles of the subfamily Scarabaeinae (Coleoptera, Scarabaeidae). Folia Entomologica Mexicana 38: 29-107
- Hanski I. 1991. The dung insect community. In: Hanski I, Cambefort Y. (eds.) Dung beetle ecology. New Jersey, Princeton University Press. p. 5-21.
- Henderson A, Galeano G, Bernal R. 1995. Field guide to the palms of the Americas. New Jersey, Princeton University Press.
- Henderson F. 2006. Morphology and anatomy of palm seedlings. Botanical Review 72: 273-329.
- Hölzel N, Otte A. 2004. Assessing soil seed bank persistence in flood-meadows: the search for reliable traits. Journal of Vegetation Science 15: 93-100.
- Howe HF. 1993. Aspects of variation in a neotropical seed dispersal system. Vegetatio 107/108: 149-162.



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- Howe HF, Smallwood J. 1982. Ecology of seed dispersal. Annual Review of Ecology and Systematics. 13: 201-228.
- Janzen DH. 1970. Herbivores and the number of tree species in tropical forests. American Naturalist 104: 501-528.
- Janzen DH. 1971. Seed predation by animals. Annual Review of Ecology, Evolution and Systematics 2: 465-492.
- Johnson CD, Zona S, Nilsson JA. 1995. Bruchid beetles and palm seeds: recorded relationships. Principes 39: 25-35.
- Leite CO. 1990. Biologia de reprodução de *Allagoptera arenaria* (Gomes) O. Kuntze (*Diplothemium maritimum* MART.) Palmae, Rio de Janeiro. MSc. Thesis, Universidade Federal do Rio de Janeiro, Rio de Janeiro.
- Lorenzi H, Sousa HM, Costa JTM, Cerqueira LSC, Ferreira E. 2004. Palmeiras Brasileiras: nativas e exóticas cultivadas. Nova Odessa, Instituto Plantarum.
- Mantovani A, Iglesias RR. 2008. Factors limiting seed germination of terrestrial bromeliads in the sandy coastal plains (*restinga*) of Maricá, Rio de Janeiro, Brazil. Rodriguésia 59: 135-150.
- Martin L, Suguio K, Flexor JM. 1993. As flutuações de nível do mar durante o quaternário superior e a evolução geológica de "deltas" brasileiras. Boletim do Instituto de Geologia-USP 15 Publicação Especial:1186
- Mattos CLV. 2005. Caracterização Climática da Restinga da Marambaia. In Menezes LFT, Peixoto AL, Araujo DSD. (eds.) História Natural da Marambaia. Seropédica, Editora da Universidade Federal Rural do Rio de Janeiro. p. 55-66.
- Mello JIO, Barbedo CJ. 2007. Temperatura, luz e substrato para a germinação de sementes de pau-brasil *Caesalpinia echinata* Lam., Leguminosae-Caesalpiniodeae. Revista Árvore 31: 645-655.
- Menezes LFT, Araujo DSD. 1999. Estrutura de duas formações vegetais do cordão externo da restinga de Marambaia-RJ. Acta Botanica Brasilica 13: 223-235.
- Menezes LFT, Araujo DSD. 2000. Variação da biomassa aérea de *Allagoptera* arenaria (Gomes) O. Kuntze numa comunidade arbustiva de Palmae na Restinga da Marambaia, RJ. Brazilia Journal of Biology 60: 47-157.
- Menezes LFT, Araujo DSD. 2005. Formações vegetais da restinga de Marambaia, Rio de Janeiro. In: Menezes LFT, Peixoto AL, Araujo DSD. (eds.) Historia Natural da Marambaia. Seropédica, EDUR. p. 67-12.
- Moraes M. 1996. Allagoptera (Palmae) Flora Neotropica 73:1-34. Pillar VP.1997. Multivariate exploratory analysis and randomization
- Pillar VP.1997. Multivariate exploratory analysis and randomization testing using Multiv. Colnoses 12: 145-148.
- Pillar VP, Orlóci L. 1996. On randomization testing in vegetation science: multifactor comparisons of relevé groups. Journal of Vegetation Science 7: 585-592.
- Pimenta RS. Luz PB, Pivetta KL, Castro A, Pizetta PUC. 2010. Efeito da maturação e temperatura na germinação de sementes de *Phoenix canariensis* Hort. ex Chabaud Arecaceae. Revista Árvore 34: 31-38.
- Pimentel DS, Tabarelli M. 2004. Seed dispersal of the palm Attalea oleifera in a remnant of the Brazilian Atlantic Forest. Biotropica 36: 74-84
- Pinheiro F, Borghetti F. 2003. Light and temperature requirements for germination of seeds of *Aechmea nudicaulis* (L.) Griesebachand and *Streptocalyx floribundus* (Martius ex Schultes F.) Mez (Bromeliaceae). Acta Botanica Brasilica 17: 27-35.
- Pires LA, Cardoso VJM, Joly VA, Rodrigues RR. 2009. Germinação de Ternstroemia brasiliensis Cambess. (Pentaphylacaceae) de Floresta de Restinga. Acta Botanica Brasilica 23: 57-66. 2009.
- Price PW. 1997. Insect Ecology. 3rd. edn. NewYork, John Willey & Sons. Probert RJ. 1992. The role of temperature in germination ecophysiology. In: Fenner M. (ed.) Seeds: The Ecology of regeneration in plant communities. Wallingford, CAB International. p. 285-325.
- Pugnaire FI, Haase P, Puigdefábregas J. 1996. Facilitation between higher plant species in a semiarid environment. Ecology 77: 1420-1426.

- Pugnaire FI, Armas C, Maestre FT. 2011. Positive plant interactions in the Iberian Southeast: Mechanisms, environmental gradients, and ecosystem function. Journal of Arid Environments 75: 1310-1320.
- Quiroga-Castro VD, Roldán IA. 2001. The fate of Attalea phalerata (Palmae) seeds dispersed to a tapir latrine. Biotropica 33: 472-477.
- R Development Core Team. 2013. R: A language and environment for statistical computing, R Foundation for Statistical Computing, Vienna, Austria. http://www.R-project.org.
- Rios RS, Pacheco LF. 2006. The effect of dung and dispersal on post dispersal seed predation of *Attalea phalerata* (Arecaceae) by bruchid beetles. Biotropica 38: 778-781.
- Roberts JT, Heithaus ER. 1986. Ants rearrange the vertebrate-generated seed shadow of a neotropical fig tree. Ecology 67: 1046-1051.
- Russo SE, Augspurger CK. 2004. Aggregated seed dispersal by spider monkeys limits recruitment to clumped patterns in *Virola calophylla*. Ecology Letters 7: 1058-1067.
- Scarano FR. 2002. Structure, function and floristic relationships of plant communities in stressful habitats marginal to the Brazilian Atlantic rain forest. Annals of Botany 90: 517-524.
- Scarano FR, Cirne P, Nascimento MT, et al. 2004. Ecologia vegetal: integrando ecossistema, comunidades, populações e organismos. In: Rocha CFD, Esteves FA, Scarano FR. (eds.) Pesquisas de longa duração na Restinga de Jurubatiba: ecologia, história natural e conservação. São Carlos, Editora Rima. p. 77-97.
- Schupp EW. 1990. Annual variation in seed fall, post dispersal predation, and recruitment of a Neo-tropical tree. Ecology 71: 504-515.
- Shepherd VE, Chapman CA. 1998. Dung beetles as secondary seed dispersers: impact on seed predation and germination. Journal of Tropical Ecology 14: 199-215.
- Silvertown J, Charlesworth D. 2001. Introduction to plant population biology, 4th. edn. Oxford, Blackwell.
- Silvius KM, Fragoso JMV. 2002. Pulp handling by vertebrate seed dispersers increases palm seed predation by bruchid beetles in the northern Amazon. Journal of Ecology 90: 1024-1032.
- Swaine MD, Whitmore TC. 1988. On the definition of ecological species groups in tropical rain. Plant Ecology 75: 81-86.
- Thompson PA. 1973. Seed germination in relation to ecological and geographical distribution. In: Heywood VH. (ed.) Taxonomy and ecology. London, Academic Press. p. 93-119.
- Tobe K, Zhang L, Omasa K. 2005. Seed germination and seedling emergence of three annual growing on Desert Sand Dunes in China. Annals of Botany 95: 649-659.
- Vandelook F, Moer V, Assche JA. 2008. Environmental signals for seed germination reflect habitat adaptations in four temperate Caryophyllaceae. Functional Ecology 22: 470-478.
- Wenny DG. 2000. Seed Dispersal, Seed Predation, and Seedling Recruitment of a Neotropical Montane Tree. Ecological Monographs 70: 331-351.
- Wicklow DT, Kumvar R, Lloyd JE. 1984. Germination of blue grain seeds buried by dung beetles (Coleoptera: Scarabacidae). Environmental Entomology 13: 878-881.
- Wilson DE, Janzen DH. 1972. Predation on *Scheelea* palm seeds by bruchid beetles: seed density and distance from the parent palm. Ecology 53: 954-959
- Wright SJ. 1983. The dispersion of eggs by a bruchid beetle among *Scheelea* palm seeds and the effect of distance to the parent palm. Ecology 64: 1116-1021.
- Zaluar HLT, Scarano FR. 2000. Facilitação em restingas de moitas: um século de busca por espécies focais. In: Esteves FA, Lacerda LD. (eds.) Ecologia de restingas e lagoas costeiras. Rio de Janeiro, Nupem-UFRJ. p 3-23.

