

Phytoplankton diversity in the middle Rio Doce lake system of southeastern Brazil

Cristiane Freitas de Azevedo Barros^{1,3}, Aline Morena Menezes dos Santos² and Francisco Antônio Rodrigues Barbosa²

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ABSTRACT

This study presents results of the inventory of algal flora conducted between August 2007 and May 2008 in 18 lakes of the middle Rio Doce lake system, most of which is in the state of Minas Gerais, Brazil. We recorded 481 taxa, increasing the known total phytoplankton diversity of the region (gamma diversity) by 80%. The following classes were represented: Zygnematophyceae (171 taxa), Cyanobacteria (101), Chlorophyceae (71), Bacillariophyceae (42), Euglenophyceae (43), Trebouxiophyceae (24), Dinophyceae (8), Xanthophyceae (8), Chrysophyceae (6), Cryptophyceae (6) and Oedogoniophyceae (1). We identified 221 taxa that were rare (restricted to one or two lakes), and 101 that were considered representative (present in at least nine lakes). *Botryococcus braunii*, *Elakatothrix genevensis*, *Planktolyngbya limnetica*, *Peridinium pusillum*, *Trachelomonas volvocina*, *Cosmarium contractum*, *Staurastrum forficulatum*, *Staurastrum leptocladum*, *Staurastrum rotula*, and *Staurodesmus dejectus* were present in all lakes. Richness varied from 95 taxa (in Lake Gambazinho) to 168 taxa (in Lake Palmeirinha). Jaccard indices were low, and the highest similarities between lakes were 53% (Ferrugem/Ferruginha), 47% (Central/Almecega) and 46% (Águas Claras/Palmeirinha), demonstrating high environmental and biotic dissimilarities between lakes. Geographic distance was not significantly associated with floristic similarity, suggesting that local factors are more important than are regional ones in shaping the phytoplankton composition of lakes.

Key words: algal flora, inventory, tropical lakes

Introduction

Species richness is considered one of the simplest measures to express and quantify biological complexity in a given region (Nabout *et al.* 2007). In aquatic environments, species richness is influenced by several factors, including water temperature, mixing patterns of the water column, light, nutrient availability, and herbivory (Reynolds, 1987). Therefore, knowledge of richness patterns is essential to proposals for monitoring strategies and for biodiversity conservation activities (Downing & Leibold 2002; Declerck *et al.* 2005; Nogueira *et al.* 2008).

Diversity can be assessed on different scales. Local, or alpha, diversity is given by the total number of species in each habitat. Regional, or gamma, diversity is the total number of species observed in a range of habitats (Magurran 2004). The term beta diversity was introduced by Whittaker in the 1960s. At first the term was used in order to describe changes in species composition along gradients of altitude and humidity through differences in rates of gain and loss of species. However, beta diversity is now defined as the

taxonomic difference between samples, whether occurring along an environmental gradient or not (Veech *et al.* 2002).

The middle Rio Doce lake system is a large freshwater system in southeastern Brazil, formed as a result of a mass of sediments (from the original drainages of Rio Doce and its tributaries) that acted as a natural dam, giving rise to a dense network of lakes (Pflug 1969 cited in de Meis & Tundisi 1997). The biological and ecological importance of the system was recently demonstrated by its recognition as an international Ramsar site, making it the 11th site in Brazil to be added to the Ramsar list of wetlands of international importance (Ramsar 2010).

Limnological research in the middle Rio Doce lake system was initiated in the 1970s (Tundisi & Saijo 1997). Since then, various aspects have been investigated (Barbosa & Tundisi 1980; Henry & Barbosa 1989; Rocha *et al.* 1989; Tundisi & Saijo 1997), increasing knowledge of the geological, morphological, physical, chemical, and biological characteristics of the lakes. However, specific studies on phytoplankton were mainly focused on Lake Dom Helvécio and Lake Carioca (Hino *et al.* 1986; Taniguchi *et al.* 2003;

¹ Universidade do Estado de Minas Gerais, Frutal, MG, Brazil

² Universidade Federal de Minas Gerais, Instituto de Ciências Biológicas, Laboratório de Limnologia, Belo Horizonte, MG, Brazil

³ Author for correspondence crisfabarros@gmail.com

Barros *et al.* 2006; Souza *et al.* 2008). However, a recent survey evaluated plankton diversity in a larger number of lakes (Maia-Barbosa *et al.* 2006). Therefore, there is still little information on phytoplankton diversity in this lake system, as shown by Barbosa *et al.* (1994). The present study aims to contribute to the knowledge of phytoplankton diversity of this lake complex by presenting the results of an inventory of the algal flora in 18 lakes.

Material and methods

Study area

The Rio Doce basin is located at in southeastern Brazil, between the state of Minas Gerais (86% of the total area) and the state of Espírito Santo (14% of the total area), encompassing 83,400 km² (Marques & Barbosa, 2002). Two lake systems compose this basin: one, at the middle course, comprising ca. 250 lakes, distinct in their trophic status (Maillard *et al.* 2011), and another, at the lower course, comprising ca. 70 lakes (Cavati & Fernandes 2008). Approximately 50 lakes in the middle course are protected within Rio Doce State Park, a conservation unit created in 1939 and representing the largest contiguous remnant of the Atlantic Forest in Minas Gerais (359.76 km²). Lakes located in the surrounding area are affected mainly by hardwood (*Eucalyptus* spp.) plantations and pastureland, among several municipalities.

For the purposes of this study, 18 lakes were selected: eight located inside the Rio Doce State Park limits and ten in the surrounding areas (Fig. 1; Tab. 1). In selecting the lakes, we took into account their greatest physiographic differences and their accessibility, mainly during the rainy (summer) season. The climate of the region is classified as tropical semi-humid with 4-5 months of dry weather, exhibiting mesothermal characteristics (Nimer, 1989) with temperatures of approximately 25°C. According to Tundisi (1997), the monthly precipitation is highest in December (350 mm) and lowest in July and August (10 mm).

Samplings

Field work was conducted quarterly, in August 2007, November 2007, February 2008, and May 2008. Samples were collected from a fixed point in the limnetic region of each lake. Samplings were authorized by the Minas Gerais State Forestry Institute (permit no. 005/07). Water transparency was estimated *in situ* by Secchi disk measurements (Cole 1983). Samples were collected for total phosphorus quantification (Mackereth *et al.* 1978).

Samples for qualitative analysis of phytoplankton were collected by successive vertical and horizontal throws with a 20-µm mesh plankton net, then fixed with 4% formaldehyde solution. For each qualitative sample (four samples/lake), eight slides were analyzed, for a total of 32 slides per lake. Organisms were identified under light microscopy down to

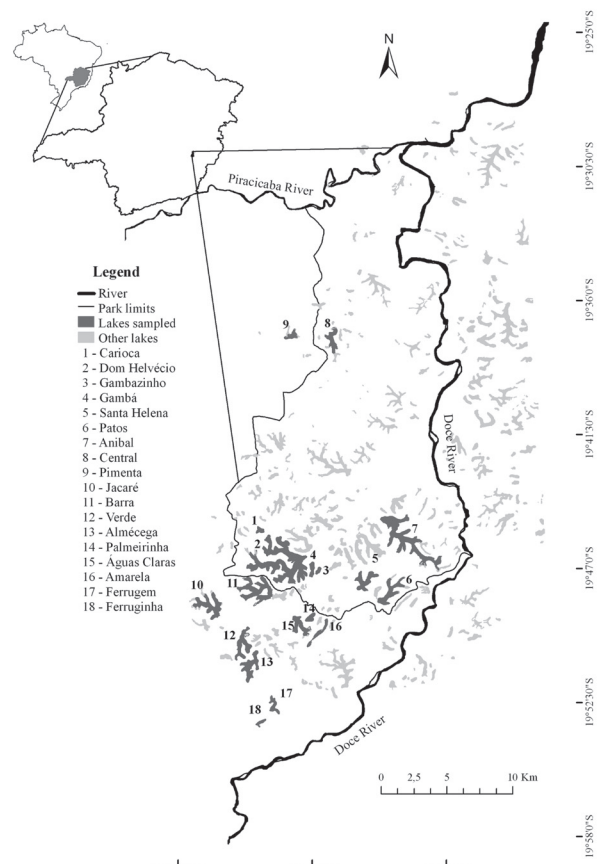


Figure 1. Middle Rio Doce lake system, showing the outline of Rio Doce State Park, sampled lakes (1 to 18), and limnetic fixed points of sampling (+). Digital base (shapefiles) provided by the Minas Gerais State Forestry Institute.

the lowest possible taxonomic level using a specific bibliography: Föster (1969; 1974), Prescott *et al.* (1975; 1977; 1981; 1982), Komárek & Fott (1983), Sant'Anna (1984), Komárek & Anagnostidis (1989; 1999), Menezes *et al.* (1995), and Bicudo & Menezes (2006). Samples for quantitative analyses were collected with van Dorn bottles at three depths (100%, 10%, and 1% of incident light, as defined with Secchi disk measurements) and fixed with Lugol's solution. Quantitative analysis followed the method described by Utermöhl (1958).

Data analysis

For each lake, species richness was assessed on the basis of the number of taxa identified, considering the qualitative and quantitative data. Species richness for the sampled region (gamma diversity) was estimated with the first-order jackknife estimator (Nabout *et al.* 2007), using Stimata S software (Colwell 2006). Beta diversity was estimated by the difference between gamma diversity (total species recorded for the set of lakes) and average alpha diversity (mean species richness per lake), as suggested by Crist *et al.* (2003):

$$\beta = \gamma - \text{mean } \alpha$$

Table 1. Geographic coordinates, depth, margin development index, area, transparency, and total phosphorous concentration of 18 lakes sampled in the middle Rio Doce lake system, between August 2007 and May 2008.

Location	Lake (abbreviation)	Geographic coordinates	Depth (m)	DL	Area (km ²)	Secchi (m)	Total phosphorous (µg/L)
State Park	Aníbal (AN)	19°06'47.1"S; 42°29'54.5"W	6.0	4.29	2.79	2.44 ± 0.13	30.12 ± 18.74
	Carioca (CA)	19°45'26.0"S; 42°37'06.2"W	10.0	1.28	0.13	1.70 ± 0.32	17.31 ± 6.75
	Central (CE)	19°37'39.0"S; 42°34'12.5"W	5.0	2.03	0.44	0.81 ± 0.18	30.44 ± 10.72
	Dom Helvécio (DH)	19°46'55.7"S; 42°35'28.9"W	28.0	4.93	5.27	2.48 ± 0.55	20.13 ± 8.86
	Gambá (GA)	19°47'15.1"S; 42°35'01.0"W	12.0	1.13	0.22	3.23 ± 0.67	13.27 ± 4.46
	Gambazinho (GN)	19°47'07.7"S; 42°34'45.5"W	10.0	2.9	0.09	1.78 ± 0.22	23.29 ± 8.51
	Patos (PT)	19°48'19.9"S; 42°32'12.7"W	8.0	2.01	1.09	2.47 ± 0.82	27.33 ± 6.69
	Santa Helena (SH)	19°47'48.8"S; 42°33'04.7"W	10.5	2.42	0.86	2.26 ± 0.44	26.77 ± 11.81
	Águas Claras (AC)	19°49'06.9"S; 42°35'42.5"W	9.5	2.24	0.62	3.28 ± 1.35	16.16 ± 7.35
	Almécega (AL)	19°51'25.4"S; 42°37'31.9"W	7.0	2.44	1.3	2.81 ± 0.55	19.21 ± 14.10
Surroundings	Amarela (AM)	19°49'23.1"S; 42°34'28.7"W	2.5	1.82	0.27	0.86 ± 0.11	75.46 ± 64.80
	Barra (BA)	19°48'11.1"S; 42°37'43.6"W	7.0	3.45	1.94	2.45 ± 0.76	47.91 ± 32.13
	Ferrugem (FE)	19°52'39.0"S; 42°36'34.3"W	3.5	1.61	0.42	0.93 ± 0.25	49.96 ± 21.92
	Ferruginha (FN)	19°53'17.5"S; 42°36'59.4"W	4.0	1.4	0.12	1.03 ± 0.05	32.99 ± 12.53
	Jacaré (JA)	19°48'37.8"S; 42°38'57.0"W	8.5	1.28	1.22	1.95 ± 0.17	42.49 ± 18.30
	Palmeirinha (PA)	19°49'41.8"S; 42°36'25.4"W	6.0	2.83	0.23	2.26 ± 0.60	28.36 ± 17.93
	Pimenta (PI)	19°37'27.4"S; 42°35'44.3"W	3.5	1.63	1.24	1.13 ± 0.25	37.78 ± 22.38
	Verde (VE)	19°49'55.2"S; 42°37'54.1"W	19.0	2.29	0.83	2.19 ± 0.58	14.38 ± 4.24

DL – development index.

where β is beta diversity, γ is gamma diversity, and α is alpha diversity.

Differences in relation to water transparency and total phosphorus levels were determined using ANOVA. Spearman's correlation coefficient was used in order to test for relationships between the morphometric features (area, depth, and margin development index) and physico-chemical variables. Hierarchical cluster analysis using Jaccard distance and Ward's method (Ward 1963) were performed in order to assess similarity between lakes in terms of the phytoplankton species composition. These statistical analyses were conducted using Past 1.90 software (Hammer *et al.* 2001).

Results and discussion

Morphometric features, such as depth, margin development index, and area, varied among lakes (Table 1). Secchi depth and total phosphorus concentrations also differed among lakes ($F=996.888$; $p=0.000$ and $F=730.533$; $p=0.000$, respectively) and correlated with depth ($p<0.05$; $r=0.630$ for Secchi disk measurements and $r=0.596$ for total phosphorus concentrations). Lakes that were shallower (<5 m: Pimenta, Central, Amarela, Ferrugem, and Ferruginha) showed lower

Secchi disappearance depths (>1.2 m) and higher levels of total phosphorus (>30 µg/L; especially Lakes Amarela and Ferrugem, in which total phosphorus was >50 µg/L), than did the lakes that were deeper (>7 m: Dom Helvécio, Águas Claras, Almécega, Gambá, and Verde), which showed greater water transparency (down to 2.2 m) and lower values of the trophy indicator (below 21 µg/L of phosphorus).

Richness extrapolation indices, such as the jackknife, although not usually used for phytoplankton (Nabout *et al.* 2007; Nogueira *et al.* 2008), can be important tools to assess the representativeness of the sampling effort. In the present study, a total of 481 taxa were recorded (Fig. 2), corresponding to 77% of the expected richness, estimated using first-order jackknife (jackknife 1 = 624). The relationship between the observed and estimated values for richness indicated that our methods were appropriate for a diversity survey. In addition, the known phytoplankton diversity in the middle Rio Doce lake system was high in comparison with the 267 species reported for seven lakes in the system by Maia-Barbosa *et al.* (2006). Gamma diversity increased by 80% with the expansion of the number of studied environments, reinforcing the observed environmental and biotic heterogeneity.

Eleven classes were identified: Zygnematophyceae (171 taxa), Cyanobacteria (101), Chlorophyceae (71), Bacillario-

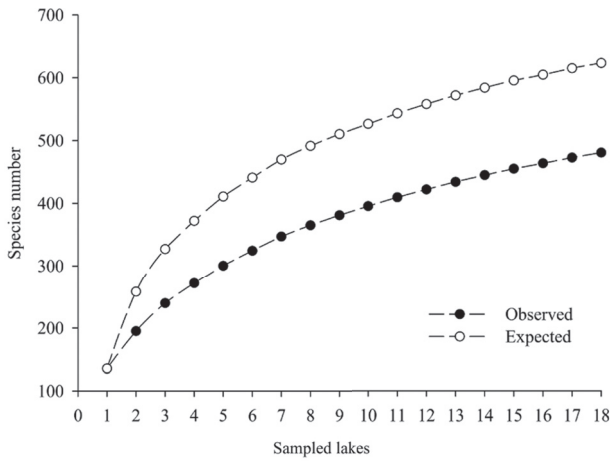


Figure 2. Species accumulation curve of 18 lakes sampled in the middle Rio Doce lake system, between August 2007 and May 2008.

phyceae (42), Euglenophyceae (43), Trebouxiophyceae (24), Dinophyceae (8), Xanthophyceae (8), Chrysophyceae (6), Cryptophyceae (6) and Oedogoniophyceae (1); Appendix 1. The predominance of desmids, especially those of the genera *Cosmarium*, *Staurastrum* and *Staurodesmus*, in the middle Rio Doce lakes was previously reported by Reynolds (1997), who attributed the dominance of this group to oligotrophic conditions and good preservation of the lakes. It has also been suggested that the thermal stratification pattern known as atelomixis (characterized by unusual, irregular circulation periods of short duration) is a key factor for

desmid prevalence (Barbosa & Padisák 2002; Souza *et al.* 2008), because it allows these species, which have relatively high specific density (Padisák *et al.* 2003), to remain within the upper layers of the water column.

Of the 481 taxa identified, 221 were rare, occurring exclusively in one or two lakes, and 101 exhibited high frequency, occurring in at least nine lakes (Tab. 2). In addition, 10 species were common to all lakes: *Botryococcus braunii* Kützing, *Elakatothrix genevensis* (Reverdin) Hindák, *Planktolyngbya limnetica* (Lemmerman) Komárkova-Legnerová and Cronberg, *Peridinium pusillum* (Pénard) Lemmerman, *Trachelomonas volvocina* Ehrenberg, *Cosmarium contractum* Kirchner, *Staurastrum forficulatum* Lundell, *Staurastrum leptocladum* Nordstedt, *Staurastrum rotula* Nordstedt, and *Staurodesmus dejectus* (Brébisson) Teiling.

Phytoplankton richness ranged from 95 taxa (Lake Gambazinho) to 168 taxa (Lake Palmeirinha). In general, the most representative groups for each lake were the same observed for the data set: Zygnematoephyceae > Chlorophyceae > Cyanobacteria; exceptions occurred for Lake Amarela, where the number of Euglenophyceae species (25) exceeded that of Cyanobacteria species (15), and for Lake Gambazinho, which had more Bacillariophyceae species (22) than Cyanobacteria species (12) (Fig. 3). The predominance of Euglenophyceae in Lake Amarela, previously reported by Reynolds (1997), is associated with its late successional stage, shallowness, and broad macrophyte coverage of the surface area, mainly with *Nymphaea* sp., *Utricularia* sp., and *Eleocharis* sp.

Table 2. List of the most common species in the 18 lakes sampled in the middle Rio Doce lake system between August 2007 and May 2008

50-79%			80-100%	
Bacillariophyceae	Chrysophyceae	Oedogoniophyceae	Bacillariophyceae	Euglenophyceae
<i>Cyclotella</i> sp.	<i>Dinobryon bavaricum</i>	<i>Oedogonium</i> NI	<i>Synedra acus</i>	<i>Trachelomonas volvocina</i>
<i>Encyonema</i> sp. 1	<i>Dinobryon sertularia</i>	Xanthophyceae	Chlorophyceae	Zygnemaphyceae
<i>Eunotia lineolata</i>	Cryptophyceae	<i>Isthmochloron lobulatum</i>	<i>Botryococcus braunii</i>	<i>Cosmarium asphaerosporum</i>
<i>Gomphonema gracile</i>	<i>Cryptomonas</i> sp2	<i>Tetraplekton</i> sp1	<i>Botryococcus terribilis</i>	<i>Cosmarium contractum</i>
<i>Stenopterobia curvula</i>	<i>Cryptomonas</i> sp4	Zygnemaphyceae	<i>Chlorella</i> sp.	<i>Cosmarium moniliforme</i>
<i>Urosolenia</i> cf. <i>longiseta</i>	<i>Cryptomonas</i> sp5	<i>Cosmarium bioculatum</i>	<i>Closteriopsis</i> sp. 1	<i>Cosmarium pseudoconnatum</i>
Chlorophyceae	Cyanobacteria	<i>Cosmarium depressum</i>	<i>Elakatothrix genevensis</i>	<i>Staurastrum forficulatum</i>
<i>Ankistrodesmus fusiformis</i>	<i>Anabaena</i> cf. <i>solitaria</i>	<i>Cosmarium monomazum</i>	<i>Eutetramorus planctonicus</i>	<i>Staurastrum ionatum</i>
<i>Ankistrodesmus</i> sp.2	<i>Aphanocapsa delicatissima</i>	<i>Cosmarium quadrum</i>	<i>Monoraphidium contortum</i>	<i>Staurastrum laeve</i>
<i>Botryococcus protuberans</i>	<i>Aphanocapsa</i> sp.	<i>Micrasterias truncata</i>	<i>Monoraphidium</i> sp.	<i>Staurastrum leptocladum</i>
<i>Coelastrum pulchrum</i>	<i>Coelomorum</i> sp.	<i>Spondylosium panduriforme</i>	<i>Nephrocytium agardhianum</i>	<i>Staurastrum rotula</i>
<i>Coelastrum reticulatum</i>	<i>Coelosphaerium</i> sp.	<i>Staurastrum cerastes</i>	<i>Oocystis</i> sp. 4	<i>Staurastrum smithii</i>
<i>Coelastrum sphaericum</i>	<i>Cylindrospermopsis raciborskii</i>	<i>Staurastrum</i> cf. <i>muticum</i>	<i>Tetraedron caudatum</i>	<i>Staurastrum trifidum</i>
<i>Crucigenia tetrapedia</i>	<i>Limnothrix redekei</i>	<i>Staurastrum chaetoceras</i>	Cryptophyceae	<i>Staurastrum</i> sp5
<i>Crucigeniella retangularis</i>	<i>Lyngbya</i> sp.	<i>Staurastrum manfeldtii</i>	<i>Cryptomonas brasiliensis</i>	<i>Staurodesmus crassus</i>

Continues

Table 2. Continuation.

50-79%			80-100%	
<i>Desmodesmus armatus</i>	<i>Merismopedia tenuissima</i>	<i>Staurastrum setigerum</i>	Cyanobacteria	<i>Staurodesmus dejectus</i>
var. <i>bicaudatus</i>	<i>Microcystis aeruginosa</i>	<i>Staurastrum tetracerum</i>	<i>Aphanocapsa elachista</i>	<i>Staurodesmus jaculiferus</i>
<i>Dictyosphaerium pulchellum</i>	<i>Oscillatoria</i> sp1	<i>Staurastrum</i> sp6	<i>Chroococcus minutus</i>	<i>Staurodesmus</i> sp3
<i>Keratococcus</i> sp.	<i>Planktolyngbya contorta</i>	<i>Staurodesmus convergens</i>	<i>Planktolyngbya limnetica</i>	<i>Teilingia granulata</i>
<i>Kirchneriella lunaris</i>	<i>Pseudanabaena limnetica</i>	<i>Staurodesmus cuspidatus</i>	<i>Pseudanabaena galeata</i>	
<i>Oocystis lacustris</i>	Dinophyceae	<i>Staurodesmus subulatus</i>	Dinophyceae	
<i>Oocystis</i> sp. 3	<i>Peridinium cf. africanum</i>		<i>Gymnodinium</i> sp.	
<i>Scenedesmus bijugus</i>	Euglenophyceae		<i>Peridinium pusillum</i>	
<i>Scenedesmus quadricauda</i>	<i>Phacus longicauda</i>		<i>Peridinium baliense</i>	
<i>Tetraedron minimum</i>	<i>Phacus raciborskii</i>			

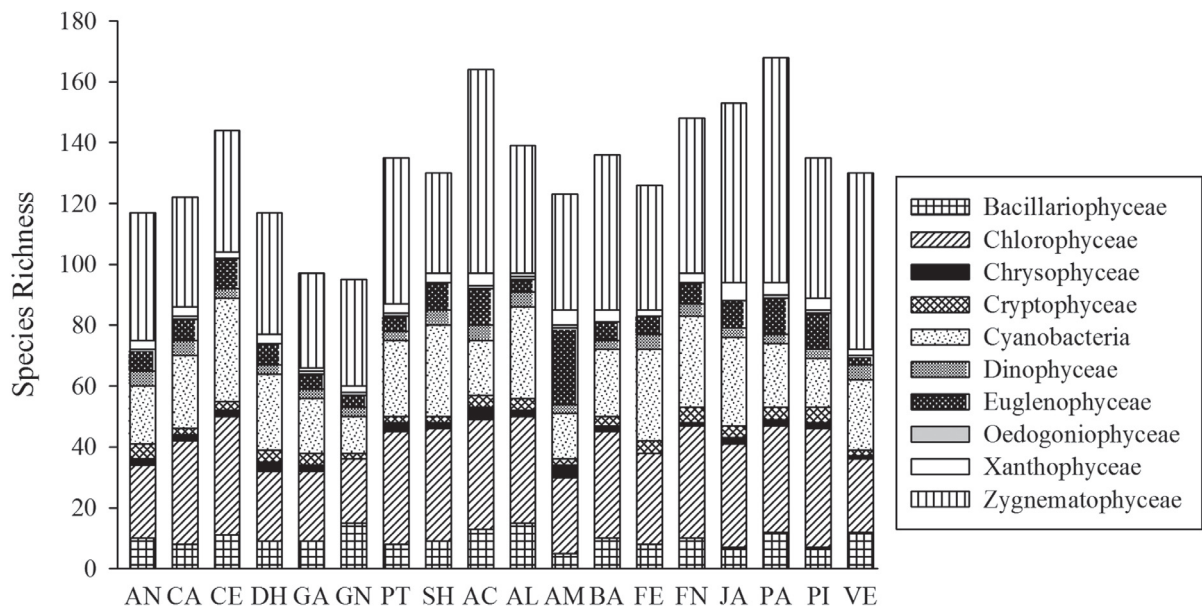


Figure 3. Species richness and contribution of each taxonomic class of 18 lakes sampled in the middle Rio Doce lake system, between August 2007 and May 2008: Águas Claras (AC), Alméciga (AL), Amarela (AM), Anibal (AN), Barra (BA), Carioca (CA), Central (CE), Dom Helvécio (DH), Ferrugem (FE), Ferruginha (FN), Gambá (GA), Gambazinho (GN), Jacaré (JA), Palmeirinha (PA), Patos (PT), Pimenta (PI), Santa Helena (SH), and Verde (VE).

Lake Amarela showed the highest number of exclusive taxa (26), followed by Lakes Gambazinho (15), Jacaré (13), and Palmeirinha (13). The lowest numbers of exclusive species were observed in Lakes Ferrugem (1), Carioca (2), and Ferruginha (2).

The fact that the incidence of exclusive species was highest in Lakes Amarela and Gambazinho indicates the great importance of the shoreline regions, given that, in the case of Lake Gambazinho, the main representative species were diatoms of the order Pennales (6 taxa) and large desmids (5 taxa), the latter also prevailing in Lake Amarela (12 taxa), together with

Euglenophyceae (7 taxa). Those two lakes are quite distinct from the other lakes of the region and from those presented here. In Lake Amarela, high amounts of organic matter reflect its advanced state of eutrophication, as evidenced by the total phosphorous and transparency values. Lake Gambazinho has a polymictic pattern of thermal stratification, in contrast to the more common warm-monomictic pattern of thermal stratification observed in the majority of the middle Rio Doce lakes. Such characteristics seem to have been responsible for the higher numbers of typically periphytic species in the samples collected from these lakes.

We obtained low Jaccard indices. The highest similarities were 53% for Lake Ferrugem versus Lake Ferruginha, followed by 47% for Lake Central versus Lake Almécga, and 46% for Lake Águas Claras versus Lake Palmeirinha. Lake Amarela showed the lowest similarity with the other lakes, ranging from 14% to 25%.

On the basis of phytoplankton species composition, we identified five clusters of lakes (Fig. 4): cluster 1-Lakes Aníbal, Carioca, Pimenta, Dom Helvécio, and Santa Helena; cluster 2-Lakes Central, Almécga, Ferrugem, and Ferruginha; cluster 3-Lakes Águas Claras, Palmeirinha, Patos, Barra, Jacaré, and Verde; cluster 4-Lakes Gambá and Gambazinho, and cluster 5-Lake Amarela. Geographic distance between lakes did not correlate significantly with interlake similarity in phytoplankton composition ($r = 0.03$; $p > 0.05$).

With the exception of a few pairs of lakes that are geographically proximal and were all included in the same cluster-Gambá and Gambazinho (500 m apart); Ferrugem and Ferruginha (1200 m apart); and Águas Claras and Palmeirinha (1500 m apart)-it should be noted that the clusters were composed of lakes that were distant from one another, some located within the Rio Doce State Park and others located in the surrounding areas. In addition, the difference between the gamma diversity and the mean alpha diversity, considered here as an estimate of beta diversity, was high: 347 species. This suggests that the phytoplankton species composition of such lakes is more dependent on local factors than on regional factors.

Considering that diversity and rarity are important criteria for assessing the conservation value of a given region (Coesel 2001), the possibility of protecting the middle Rio Doce lake system as a whole should be considered. That will require specific strategies for the environments surrounding the state park, with the primary objective of maintaining phytoplankton diversity at the regional level.

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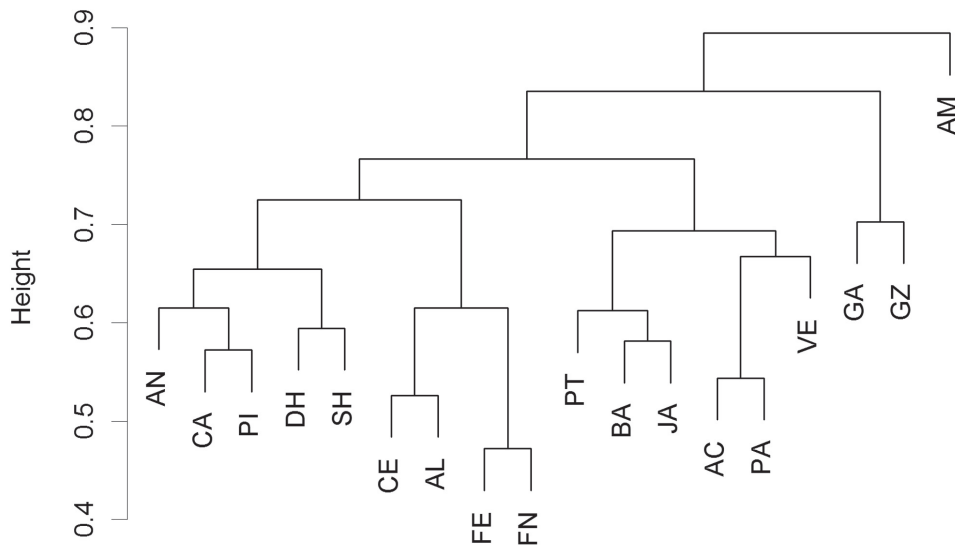


Figure 4. Cluster obtained from data of 18 lakes sampled in the middle Rio Doce lake system, between August 2007 and May 2008, considering presence and absence of species. Cluster 1-Aníbal (AN), Carioca (CA), Pimenta (PI), Dom Helvécio (DH), and Santa Helena (SH); Cluster 2-Central (CE), Almécga (AL), Ferrugem (FE), and Ferruginha (FN); Cluster 3-Águas Claras (AC), Palmeirinha (PA), Patos (PT), Barra (BA), Jacaré (JA), and Verde (VE); Cluster 4-Gambá (GA) and Gambazinho (GN); Cluster 5-Amarela (AM).

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Appendix 1. List of species recorded between August 2007 and May 2008 in the 18 lakes sampled in the middle Rio Doce lake system: **Águas Claras (AC)**, **Almécega (AL)**, **Amarela (AM)**, **Aníbal (AN)**, **Barra (BA)**, **Carioca (CA)**, **Central (CE)**, Dom Helvécio (DH), Ferrugem (FE), Ferruginha (FN), Gambá (GA), Gambazinho (GN), Jacaré (JA), Palmeirinha (PA), Patos (PT), Pimenta (PI), Santa Helena (SH), and Verde (VE).

Taxa	Occurrence
Bacillariophyceae	
<i>Aulacoseira ambigua</i> (Grunow) Simonsen	GN; AL
<i>Aulacoseira granulata</i> (Ehrenberg) Simonsen	SH; AC
<i>Aulacoseira</i> sp. 1	CE; PT; AL; FE; FN; VE
<i>Aulacoseira</i> sp. 2	AL; FN
<i>Caloneis</i> cf.	GN
<i>Cyclotella</i> sp.	All except GA, PT; FE, FN
<i>Encyonema</i> sp. 1	AN; CA; CE; PT; SH; AC; FN; PA; VE
<i>Encyonema</i> sp. 2	DH; SH; FE; JA
<i>Encyonema</i> sp. 3	GN
<i>Encyonema</i> sp. 4	JA
<i>Eunotia lineolata</i> Hustedt	All except CA, GA, GN, PT
<i>Eunotia zygodon</i> Ehrenberg	GA; PT
<i>Eunotia</i> sp. 1	AN; CA; GA; SH; AC; AL; PA
<i>Eunotia</i> sp. 2	DH; AC
<i>Eunotia</i> sp. 3	GN; AL; PA
<i>Eunotia</i> sp. 4	PT; AC; VE
<i>Eunotia</i> sp. 5	GN
<i>Frustulia crassinervia</i> (Brébisson) Costa	GN; PA; VE
<i>Frustulia krammeri</i> Lange-Bertalot e Metzeltin	CE; GA; GN
<i>Frustulia</i> sp. 1	FN
<i>Gomphonema gracile</i> Ehrenberg	All except CA; GA; GN
<i>Gomphonema subtile</i> Ehrenberg	PI
<i>Hantzschia amphioxys</i> (Ehrenberg) Grunow	GA
<i>Melosira varians</i> Agardh	AL; VE
<i>Navicula</i> sp.	CE; DH; PT; AL; BA; FN; PA; VE
<i>Neidium</i> sp.	GA; GN; BA
<i>Nitzschia</i> sp.	AN
<i>Pinnularia viridis</i> (Nitzsch) Ehrenberg	AN; CA; CE; AC; BA
<i>Pinnularia</i> sp. 1	AL
<i>Pinnularia</i> sp. 2	GN
<i>Pinnularia</i> sp. 3	GN
<i>Pinnularia</i> sp. 4	AC
<i>Pinnularia</i> sp. 5	PA
<i>Stauroneis phoenicenteron</i> (Nitzsch) Ehrenberg	GN; AM
<i>Stenopterobia curvula</i> (Wm. Smith) Krammer	All except CE; SH; AL; AM; JA
<i>Surirella linearis</i> W. Smith	CA; GN; AL; VE
<i>Synedra acus</i> Kützing	All except GN; VE
<i>Urosolenia</i> cf. <i>eriensis</i> (H.L.Smith) Round & R.M. Crawford	GA; BA
<i>Urosolenia</i> cf. <i>longiseta</i> (Zacharias) Edlung & Stoermer	AN; CA; CE; DH; GA; SH; AC; AL; FE; FN; PA; PI; VE
Pennales NI 1	GN;
Pennales NI 2	CE; AL; FE; VE
Pennales NI 3	BA; JA

Continues

Appendix 1. Continuation.

Taxa	Occurrence
Chlorophyceae	
<i>Ankistrodesmus densus</i> Korsikov	CE; SH; VE
<i>Ankistrodesmus falcatus</i> (Corda) Ralfs	FN
<i>Ankistrodesmus fusiformis</i> Corda	CA; CE; DH; GA; PT; SH; AC; AL; BA; FE; JA; PI
<i>Ankistrodesmus gracilis</i> (Reinsch) Korsikov	AC
<i>Ankistrodesmus spiralis</i> (Turner) Lemmermann	PT; FN; PA; VE
<i>Ankistrodesmus</i> sp. 1	SH
<i>Ankistrodesmus</i> sp. 2	CA; DH; GA; PT; AC; AM; BA; FE; PA; VE
<i>Ankyra judayi</i> (G.M. Smith) Fott	CE; PT; SH; AL; AM; PI; VE
Chlamydomonadaceae NI2	AC; PA
<i>Coelastrum cambricum</i> Archer	CE; GA; AL; FE; FN; PA; PI
<i>Coelastrum microporum</i> Nägeli	CE; PT; AL; JA
<i>Coelastrum pseudomicroporum</i> Korsikov	JA
<i>Coelastrum pulchrum</i> Schmidle	CE; DH; GN; SH; AC; JA; PA; PI; VE
<i>Coelastrum reticulatum</i> (P.A.Dangeard) Senn	AN; CA; CE; GN; PT; AL; FE; FN; PI; VE
<i>Coelastrum sphaericum</i> Nägeli	AN; CA; CE; AC; AL; FE; JA; PA; VE
<i>Desmodesmus armatus</i> var. <i>bicaudatus</i> (Guglielmetti) Hegewald	CA; CE; SH; AC; BA; FN; JA; PA; PI
<i>Desmodesmus denticulatus</i> (Lagerheim) S.S.An, T.Friedl & E.Hegewald	AN; AC; PA; PI
<i>Dimorphococcus</i> cf. <i>lunatus</i> A. Braun	AC; AM
<i>Elakatothrix</i> cf. <i>biplex</i> (Nyg.) Hindák	GN
<i>Elakatothrix genevensis</i> (Reverdin) Hindák	All lakes
<i>Eudorina elegans</i> Ehrenberg	DH; PT; BA; PA
<i>Eutetramorus planctonicus</i> (Korsikov) Bourrelly	All except AN; DH; AM
<i>Glaucocystis</i> cf. <i>nostochinearum</i> Itzigsohn	SH
<i>Gloeomonas</i> sp.	AN; CE; SH; VE
<i>Golenkinia radiata</i> Chodat	CA; AC; BA; JA; PA
<i>Gregiochloris</i> cf.	PI
<i>Kirchneriella lunaris</i> (Kirchner) K. Möbius	All except AN; PT; SH; JA
<i>Kirchneriella</i> cf. <i>obesa</i> (W.West) Schmidle	CA; CE; AL; FN
<i>Kirchneriella rosolata</i> Hindák	PT; SH
<i>Korschikoviella</i> sp.	AM
<i>Monoraphidium contortum</i> (Thuret) Komárková-Legnerová	All except GA; AL; AM
<i>Monoraphidium</i> sp.	All except PI; VE
<i>Nephrocytium agardhianum</i> Nägeli	All except SH; PI; VE
<i>Nephrocytium</i> cf. <i>lunatum</i> W. West	AM
<i>Pectodictyon</i> aff.	DH
<i>Pediastrum duplex</i> Meyen	CA; AC; PI
<i>Pediastrum</i> sp. 1	AM
<i>Planktonema</i> cf.	PT; SH
<i>Quadrigula closterioides</i> (Bohlin) Printz	CE; DH; PT; AL; FN; PA
<i>Radiococcus</i> cf.	PT; SH; VE
<i>Scenedesmus acuminatus</i> (Lagerheim) Chodat	CA; CE; SH; AC; BA; FE; FN; PI
<i>Scenedesmus acuminatus</i> f. <i>maximus</i> Uherkovich	JA; PI
<i>Scenedesmus arcuatus</i> Lemmermann	AN; CE; AC; BA; PI

Continues

Appendix 1. Continuation.

Taxa	Occurrence
<i>Scenedesmus bijugus</i> (Turpin) Kutzing	All except DH; GA; FE; PI
<i>Scenedesmus bijugus</i> var. <i>disciformis</i> (Chodat) Leite	PT; AC; BA; FN; JA; PA; PI
<i>Scenedesmus acunae</i> Comas	GN; AC; PA
<i>Scenedesmus quadricauda</i> (Turpin) Brébisson	CA; GN; SH; AC; AL; FE; FN; PA; PI; VE
<i>Scenedesmus regularis</i> Swir	PI
<i>Scenedesmus spinosus</i> Chodat	CE; AL
<i>Selenastrum</i> sp. 1	GA; FN
<i>Sphaerocystis</i> sp. 1	CE; GA; PT; BA; FE
<i>Sphaerocystis</i> sp. 2	AL
<i>Stauridium privum</i> (Printz) Hegewu	PT; AL; BA; FN; PA; PI
<i>Stauridium tetras</i> (Ehrenberg) Ralfs	FN; PI
<i>Tetraedron caudatum</i> (Corda) Hansgirg	All except AM
<i>Tetraedron incus</i> (Teiling) G.M. Smithi	GA; PT; BA
<i>Tetraedron minimum</i> (A. Braun)	AN; CA; CE; DH; GA; PT; AL; BA; FE; FN; JA; PA
<i>Tetrallantos lagerheimii</i> Teiling	DH; SH; AC; FE; FN; JA; PI
<i>Tetranephris brasiliense</i> Leite & C. Bicudo	CA; SH
<i>Tetranephris</i> cf.	CA; DH
<i>Ulothrix</i> sp. 1	PI
<i>Ulothrix</i> sp. 2	AM
<i>Ulothrix</i> sp. 3	AM
Volvocales NI	SH; PI
Chaetophorales NI	GN
Chlorococcales NI1	GA; FN
Chlorococcales NI2	GN
Chlorococcales NI3	JA
Chlorococcales NI4	SH
Chlorococcales NI5	SH
Chrysophyceae	
<i>Dinobryon bavaricum</i> Imhof	CA; CE; DH; GA; PT; AC; AL; AM; JA; PA; PI
<i>Dinobryon sertularia</i> Ehrenberg	All except GN; PT; FE; JA
<i>Dinobryon</i> sp.	AC
<i>Mallomonas</i> cf.	PT; SH; AC; AM; BA; JA
<i>Pteromonas</i> cf.	DH; PT; AM
<i>Synura</i> cf.	AN
Cryptophyceae	
<i>Cryptomonas brasiliensis</i> Castro, C. Bicudo & D. Bicudo	All except PT; AM; VE
<i>Cryptomonas marssonii</i> Skuja	AN; GA; PT; AC; AL; AM; BA; FN; JA; PA; PI; VE
<i>Cryptomonas</i> sp. 1	AN; DH; AC; AL; PI
<i>Cryptomonas</i> sp. 2	CE; GA; AC; AM; FE; FN; JA; PA; VE
<i>Cryptomonas</i> sp. 3	All except SH; AC; AM; VE
<i>Rhodomonas</i> sp.	AN; DH; SH; FE; FN; PI
Cyanobacteria	
<i>Anabaena planctonica</i> Brunthaler	PT; AL; PA
<i>Anabaena</i> cf. <i>solitaria</i> Klebahn	AN; DH; GA; PT; SH; AM; FE; JA; PA; PI

Continues

Appendix 1. Continuation.

Taxa	Occurrence
<i>Anabaena</i> sp. 1	AM
<i>Anabaena</i> sp. 2	DH; SH; AC; FE; JA; PI; VE
<i>Anabaena</i> sp. 3	CA; AL; BA; JA
<i>Aphanizomenon</i> sp.	AN
<i>Aphanocapsa delicatissima</i> W. West & G. S. West	CA; CE; GA; GN; PT; SH; AL; BA; FE; FN; JA; VE
<i>Aphanocapsa elachista</i> W. West & G. S. West	All except AM
<i>Aphanocapsa holsatica</i> (Lemmermann) Cronberg & Komárek	GA; GN; PT; JA
Cyanobacteria	
<i>Aphanocapsa planctonica</i> (G.G.Smith) Komárek et Anagnostidis	CE; AL
<i>Aphanocapsa</i> sp.	AN; CA; CE; SH; AL; BA; FE; FN; PI; VE
<i>Aphanothece</i> cf. <i>stagnina</i> (Sprengel) A Braun	AN; DH; SH; AL; FE; VE
<i>Aphanothece</i> sp. 1	CE; GA; GN; FN
<i>Aphanothece</i> sp. 2	FE
<i>Aphanothece</i> sp. 3	CA; DH; SH
<i>Aphanothece</i> sp. 4	JA; PA
<i>Asterocapsa</i> sp.	CA; SH; JA
<i>Calothrix</i> sp.	CA; DH; PT; AC; AL; AM; PA
<i>Chroococcus minutus</i> (Kütz) Nägeli	All except GN; PI
<i>Chroococcus</i> sp. 1	PT; AM; JA
<i>Coelomorum</i> sp.	CE; DH; SH; AC; AL; FE; FN; JA; VE
<i>Coelosphaerium</i> sp. 1	CA; CE; DH; GN; SH; AL; BA; FE; FN; JA
<i>Coelosphaerium</i> sp. 2	SH
<i>Cyanodictyon</i> cf. <i>iac</i> Cronberg & Komárek	AL; FN; VE
<i>Cyanodictyon imperfectum</i> Cronberg & Weibull	CE; AL; FE; FN
<i>Cylindrospermopsis raciborskii</i> (Woloszynska) Seenayya & Subba Raju	DH; PT; SH; AL; BA; FE; FN; PI; VE
<i>Epigloeosphaera</i> sp. 1	CE; DH; PT; SH; AL; FE; FN; PA
<i>Epigloeosphaera</i> sp. 2	CE; FE; FN
<i>Eucapsis</i> sp.	CE
<i>Geitlerinema</i> cf. <i>amphibium</i> (Agardh ex Gomont) Anagnostidis	AN; DH; SH
<i>Geitlerinema splendidum</i> (Greville ex Gomont) Anagnostidis	AN; PT; AM
<i>Geitlerinema unigranulatum</i> (Singh) Komárek & Azevedo	CE; AC; JA
<i>Johannesbaptistia</i> sp.	AM
<i>Leptolyngbya</i> sp.	AC; AM
<i>Limnothrix redekei</i> (Van Goor) Meffert	All except AL; AM; JA; PA
<i>Lyngbya</i> sp.	AN; CA; CE; DH; PT; AC; AM; BA; JA; VE
<i>Merismopedia glauca</i> (Ehrenberg) Kützing	AN; CA; PT; BA; FE; FN
<i>Merismopedia tenuissima</i> Lemmermann	All except AN; GN; AM; VE
<i>Microcrocis</i> cf.	JA
<i>Microcystis aeruginosa</i> (Kützing) Kützing	CA; CE; PT; SH; AL; BA; FE; FN; JA; PA; VE
<i>Microcystis novacekii</i> (Komárek) Compère	SH; FE; JA
<i>Microcystis protocystis</i> Crow	CA; CE; SH; AL; BA; FE; JA; VE
<i>Microcystis wesenbergii</i> Komárek	AN; SH; AL; FN; JA
<i>Nostoc</i> sp. 1	VE
<i>Nostoc</i> sp. 2	SH

Continues

Appendix 1. Continuation.

Taxa	Occurrence
<i>Nostoc</i> sp. 3	BA
<i>Oscillatoria</i> sp. 1	CA;DH; GA; PT; SH;AC; AM; BA; FN;JA;PA;PI
<i>Oscillatoria</i> sp. 2	AN; CE; AM; FE; FN; JA; PA; VE
<i>Planktolyngbya contorta</i> (Lemmermann) Anagnostidis & Komárek	CA;CE;DH;GA;GN; SH; AL; BA; FE; FN; PI; VE
<i>Planktolyngbya microspira</i> Komárek et Cronberg	GN; BA
<i>Planktolyngbya</i> sp.	VE
<i>Planktolyngbya limnetica</i> (Lemmermann) J.Komárková-Legnerová & G.Cronberg	All lakes
<i>Planktothrix</i> sp.	CA; DH; AC; AM
<i>Porphyrosiohon</i> cf.	DH
<i>Pseudanabaena catenata</i> Lauterborn	AN; SH; BA; VE
<i>Pseudanabaena galeata</i> Böcher	All except AN; AM; PA
<i>Pseudanabaena limnetica</i> (Lemmermann) Komárek	AN; CA; CE; GA; AL; BA; FE; FN; PA; PI; VE
<i>Pseudanabaena mucicola</i> (Nauman & Hubber-Pestalozzi) Bourrelly	DH; GN; FE
<i>Pseudanabaena</i> sp. 1	AN; PT; PI
<i>Pseudanabaena</i> sp. 2	SH; AC; JA; PA
<i>Rabdoderma</i> cf.	FN
<i>Radiocystis</i> cf.	CE; GA
<i>Radiocystis fernandoi</i> Komárek & Komárková-Legnerová	SH; AL; FE; FN
<i>Raphidiopsis</i> cf.	CE
<i>Snowella</i> cf.	JA
<i>Sphaerocavum brasiliense</i> Azevedo & Sant'Anna	DH
<i>Spirulina</i> sp. 1	PT; PI
<i>Spirulina</i> sp. 2	PT; AM
<i>Synechococcus</i> cf.	DH; AC; JA; PA
<i>Synechocystis</i> cf.	AL; BA; FE
Chroococcales NI 1	CE; AL
Chroococcales NI 2	CE; AL
Chroococcales NI 3	PT; FN
Chroococcales NI 4	CE; FE
Chroococcales NI 5	CE
Chroococcales NI 6	FN
Chroococcales NI 7	GA
Chroococcales NI 8	SH
Chroococcales NI 9	PI
Chroococcales NI 10	CA
Chroococcales NI 11	CA; AM
Chroococcales NI 12	GA
Chroococcales NI 13	GA
Chroococcales NI 14	CE; DH; AL; FN
Chroococcales NI 15	AL; FE; FN
Chroococcales NI 16	CE; AL
Nostocales NI 1	CA; GN; AC; JA; VE
Nostocales NI 2	VE

Continues

Appendix 1. Continuation.

Taxa	Occurrence
Nostocales NI 3	PA
Nostocales NI 4	PA
Nostocales NI 5	PI
Nostocales NI 6	PA
Oscillatoriales NI 1	CE; GA
Oscillatoriales NI 2	AN; CE; FN; PA
Oscillatoriales NI 3	AN
Pseudanabaenaceae NI 1	AC
Pseudanabaenaceae NI 2	CE; PA
Pseudanabaenaceae NI 3	PT
Pseudanabaenaceae NI 4	PA
Stigonematales NI 1	PT
Stigonematales NI 2	JA
Dinophyceae	
<i>Gymnodinium</i> sp.	All except AM; BA
<i>Peridinium baliense</i> Lindemann	All except PT; AM
<i>Peridinium cf. africanum</i> Lemmermann	AN; CA; PT; SH; AC; AL; AM; FE; FN; SE
<i>Peridinium cf. volzii</i> Lemmermann	AL; AM; BA; FE
<i>Peridinium pusillum</i> (Pénard) Lemmermann	All lakes
<i>Peridinium</i> sp. 1	VE
<i>Peridinium</i> sp. 2	AN; CA; AC
<i>Peridinium</i> sp. 3	SH
Euglenophyceae	
<i>Euglena acus</i> Ehrenberg	CE; AC; AM; PA; PI
<i>Euglena ehrenbergii</i> Klebs	PT; AC; AL; AM
<i>Euglena oxyuris</i> Schmarda	AN; CA; CE; AC; AM; PA
<i>Lepocinclis fusiformis</i> (H.J.Carter) Lemmermann	AM; PI
<i>Lepocinclis cf. ovum</i> (Ehrenberg) Lemmermann	SH
<i>Lepocinclis</i> sp. 1	CA; CE; DH; AM; PA; VE
<i>Lepocinclis</i> sp. 2	GN
<i>Lepocinclis</i> sp. 3	AM
<i>Lepocinclis</i> sp. 4	JA
<i>Monomorpha cf. pyrum</i> (Ehrenberg) Mereschkowski	DH; SH; AC; PA
<i>Phacus hamatus</i> Pochmann	AN; PT; AM; BA; FN; JA; PI
<i>Phacus longicauda</i> (Ehrenberg) Dujardin	AN; JA; GN; SH; AC; AM; BA; FN; JA; PI
<i>Phacus onyx</i> Pochmann	CA; CE; AM
<i>Phacus raciborskii</i> Drezepolski	CA; CE; JA; GA; PT; AM; JA; PA; PI; VE
<i>Phacus suecicus</i> Lemmermann	AL; AM; BA
<i>Phacus</i> sp. 1	AM
<i>Phacus</i> sp. 2	JA; SH; AC; AM
<i>Phacus</i> sp. 3	GN; FN
<i>Rhabdomonas</i> sp.	GA
<i>Strombomonas cf. encifera</i>	PI
<i>Strombomonas cf. gibberosa</i>	AM

Continues

Appendix 1. Continuation.

Taxa	Occurrence
<i>Trachelomonas mirabilis</i> var. <i>spinosa</i> Svirenko	AM; PA; PI
<i>Trachelomonas armata</i> (Ehrenberg) Stein	AN; CA; PT; AC; AM; FN; JA; PI
<i>Trachelomonas</i> cf. <i>magdaleniana</i> Deflandre	AM
<i>Trachelomonas</i> cf. <i>oblonga</i> Lemmermann	SH
<i>Trachelomonas</i> cf. <i>spinosa</i> Stokes	JA
<i>Trachelomonas hispida</i> var. <i>coronata</i> Lemmermann	CE; AC
<i>Trachelomonas hispida</i> var. <i>duplex</i> Deflandre	SH; AM; BA; JA; PA; JA
<i>Trachelomonas</i> cf. <i>zingeri</i> Roll	CE; JA; PA
<i>Trachelomonas lacustris</i> Drezepolski	AM; JA; FN; JA; PA; JA
<i>Trachelomonas megalacantha</i> Da Cunha	AM
<i>Trachelomonas volvocina</i> Ehrenberg	All lakes
<i>Trachelomonas</i> sp. 1	SH; AL
<i>Trachelomonas</i> sp. 2	CA; CE; AC; AM; JA; FN
<i>Trachelomonas</i> sp. 3	AN; SH
<i>Trachelomonas</i> sp. 4	AC; JA; PA
<i>Trachelomonas</i> sp. 5	AM
<i>Trachelomonas</i> sp. 6	AM
<i>Trachelomonas</i> sp. 7	DH
Euglenales NI 1	AN; AC; AM; BA; JA; PA; JA
Euglenales NI 2	GA
Euglenales NI 3	GA
Euglenales NI 4	CE
Oedogoniophyceae	
<i>Oedogonium</i> sp.	CA; GA; GN; PT; AC; AL; AM; PA; JA
Trebouxiophyceae	
<i>Actinastrum aciculare</i> Playfair	AN; CA; GA; FE; FN; JA; PA; PI
<i>Actinastrum hantzschii</i> Lagerheim	GA
<i>Botryococcus braunii</i> Kützing	All lakes
<i>Botryococcus protuberans</i> W.West & G.S.West	All except DH; GA; SH; FE
<i>Botryococcus terribilis</i> J. Komárek & P. Marvan	All except GN; BA; VE
<i>Chlorella</i> sp.	All except PA
<i>Closteriopsis</i> sp. 1	All except VE
<i>Closteriopsis</i> sp. 2	CA; GN; SH; AC; JA; PI; VE
<i>Crucigenia</i> cf. <i>fenestrata</i> (Schmidle) Shimidle	AL; BA; JA; VE
<i>Crucigenia</i> cf. <i>quadrata</i> Morren	CE
<i>Crucigenia tetrapedia</i> (Kirchner) W.West & G.S.West	All except DH; GA; GN; FE
<i>Crucigeniella crucifera</i> (Wolle) Komárek	PI
<i>Crucigeniella retangularis</i> (Nägeli) Komárek	AN; CA; CE; PT; AL; AM; BA; FE; JA; PA; PI
<i>Dictyosphaerium erenbergianum</i> Nägeli	GN; AL; FE; JA
<i>Dictyosphaerium pulchellum</i> Wood	All except DH; GA; GN; BA; VE
<i>Keratococcus</i> sp.	AN; CA; CE; PT; SH; AL; BA; FN; PI
<i>Oocystis</i> cf. <i>nephrocytioides</i> Fott & Cado	PT; SH

Continues

Appendix 1. Continuation.

Taxa	Occurrence
<i>Oocystis</i> cf. <i>solitaria</i> Wittrock	AN; GA; PT; FE
<i>Oocystis lacustris</i> Chodat	All except AN; CA; GA; GN
<i>Oocystis</i> sp. 1	CE; GA; AL; BA; PA;
<i>Oocystis</i> sp. 2	All except GA; GN; PI; VE
<i>Oocystis</i> sp. 3	All except AN; GN; PA
<i>Oocystis</i> sp. 4	AM; BA
<i>Oocystis</i> sp. 5	DH
Xanthophyceae	
<i>Centritractus</i> sp.	CA; PT; SH; BA; JA; JA
<i>Isthmochloron lobulatum</i> (Nägeli) Skuja	AN; DH; GN; PT; AL; AM; BA; JA; JA; VE
<i>Isthmochlorum gracile</i> (Reinsch) Skuja	CA; CE; SH; PA; JA
<i>Pseudostaurastrum</i> sp.	AC; AM; JA
<i>Tetraplektron</i> cf. <i>bourelyi</i> Ettl	AN; DH; GA; AC; AM; BA; PI
<i>Tetraplektron torsum</i> (Skuja) Dedus.	AC; AM; FN; JA; PA
<i>Tetraplektron</i> sp. 1	CA; CE; DH; GN; PT; SH; AC; FN; JA; PA; PI; VE
<i>Tetraplektron</i> sp. 2	AN; AM; BA; FE; FN; JA; PA
Zygnematophyceae	
<i>Actinotaenium</i> sp.	AM; JA
<i>Bambusina Brébissonii</i> Kützing	AC; AM
<i>Bourrellyodesmus jolyanus</i> C. Bicudo & Azevedo	PT; SH; AC; JA; PA; VE
<i>Closterium</i> cf. <i>setaceum</i> Ehrenberg	PT; AC; AM; PA; PI; VE
<i>Closterium closterioides</i> (Ralfs) Louis & Peeters	GN; AC
<i>Closterium diana</i> Ehrenberg	CE
<i>Closterium gracile</i> Brébisson ex Ralfs	CA; AC; AM; BA; FN; PA; PI
<i>Closterium kuetzingii</i> Brébisson	AN; CE; DH
<i>Closterium moniliferum</i> (Bory) Ehrenberg ex Ralfs	GN
<i>Closterium</i> cf. <i>turgidum</i> Ehrenberg	AC
<i>Closterium</i> sp. 1	CE; AL
<i>Closterium</i> sp. 2	GN; AM
<i>Closterium</i> sp. 3	GN; AM
<i>Closterium</i> sp. 4	PA
<i>Cosmarium asphaerosporum</i> Wittrock	All except AM
<i>Cosmarium bioculatum</i> Brébisson in Ralfs	AN; CA; GA; SH; AC; BA; FE; FN; JA; PA; PI
<i>Cosmarium conspersum</i> Ralfs	CE; PT; FE; PI
<i>Cosmarium contractum</i> Kirchner	All lakes
<i>Cosmarium depressum</i> (Nägeli) Lundell	
<i>Cosmarium lagoense</i> Nordstedt	AL; BA
<i>Cosmarium moniliforme</i> West & West	All except AM; PI
<i>Cosmarium monomazum</i> P.Lundell	AN; CE; DH; PT; AC; BA; FN; JA; PA; VE
<i>Cosmarium ornatum</i> Ralfs	JA; PA
<i>Cosmarium portianum</i> Archer	GN; AC; JA; PA
<i>Cosmarium pseudoconnatum</i> Nordstedt	All except GA; AM
<i>Cosmarium</i> cf. <i>pseudopyramidatum</i> Lundell	GN

Continues

Appendix 1. Continuation.

Taxa	Occurrence
<i>Cosmarium pyramidatum</i> Brébisson in Ralfs	DH; GA; AC; BA; FE; FN; PA; VE
<i>Cosmarium quadrum</i> P.Lundell	CE; DH; GA; GN; AC; AL; BA; FN; JA; VE
<i>Cosmarium trilobulatum</i> Reinsch	PA
<i>Cosmarium</i> cf. <i>zonatum</i> P.Lundell	AC
<i>Cosmarium</i> sp. 1	CE; AL; AM
<i>Cosmarium</i> sp. 2	AN; AL; PA
<i>Cosmarium</i> sp. 3	BA; FN; PA
<i>Cosmarium</i> sp. 4	BA
<i>Cosmarium</i> sp. 5	AC; FE; FN; JA; PA; VE
<i>Cosmarium</i> sp. 6	BA; VE
<i>Cosmarium</i> sp. 7	VE
<i>Cosmarium</i> sp. 8	CA; GN; PA
<i>Cosmarium</i> sp. 9	AM
<i>Cosmarium</i> sp. 10	AM
<i>Cosmarium</i> sp. 11	AC; AM; PA
<i>Cosmarium</i> sp. 12	AM
<i>Cosmarium</i> sp. 13	AM
<i>Cosmarium</i> sp. 14	AM
<i>Cosmarium</i> sp. 15	PA
<i>Cosmarium</i> sp. 16	PA
<i>Cosmarium</i> sp. 17	JA
<i>Desmidium aptogonum</i> Brébisson	SH; BA
<i>Desmidium grevillei</i> (Kützing ex Ralfs) De Bary	CA
<i>Desmidium swartzii</i> Agardh	SH
<i>Euastrum</i> cf. <i>abruprum</i> Nordstedt	GN; PT; FN
<i>Euastrum didelta</i> (Turpin) Ralfs	GN
<i>Euastrum elegans</i> (Brébisson) Kützing	AM
<i>Euastrum pulchellum</i> Brébisson	PT; BA
<i>Gonatozygon Brébissonii</i> De Bary	AM; BA; PI
<i>Gonatozygon</i> cf. <i>pillosum</i> Wolle	GA; PT
<i>Gonatozygon monotaenium</i> De Bary in West & G.S.West	GN; BA
<i>Haplotaenium minutum</i> (Ralfs) Delponte	DH; AC; PI; VE
<i>Hyaloteca</i> sp.	PT; AC; AL; JA; PA; VE
<i>Micrasterias abrupta</i> West & G.S.West	AC; PA
<i>Micrasterias arcuata</i> Bailey	AC
<i>Micrasterias borgei</i> H. Krieger	GN
<i>Micrasterias crux-melitensis</i> (Ehrenberg) Ralfs	FN; VE
<i>Micrasterias oscitans</i> var. <i>mucronata</i> (R.V.Dixon) J.N.F.Wille	AC; PA
<i>Micrasterias pinnatifida</i> (Kützing) Ralfs	AC; AL; JA; PA
<i>Micrasterias tropica</i> Nordstedt	AM
<i>Micrasterias truncata</i> (Corda) Bréb. ex Ralfs	AN; DH; AC; AL; AM; JA; JA; PA; PI; VE
<i>Mougeotia</i> sp. 1	AN; GN; PT; AC; AM
<i>Mougeotia</i> sp. 2	AM

Continues

Appendix 1. Continuation.

Taxa	Occurrence
<i>Netrium</i> sp.	PA
<i>Octacanthium mucronulatum</i> (Nordstedt) P.Compère	CA; PA; PI; VE
<i>Octacanthium</i> sp.	JA
<i>Onychonema laeve</i> Nordstedt	PT
<i>Pleurotaenium</i> cf. <i>baculoides</i> (Skuja) Krieger	AC
<i>Pleurotaenium</i> sp.	AM; PA; VE
<i>Sirogonium</i> cf.	AM
<i>Sphaerosozma</i> sp.	CE; DH; SH; FE; FN; JA; PA; PI
<i>Spirogyra</i> sp. 1	AN
<i>Spirogyra</i> sp. 2	VE
<i>Spirogyra</i> sp. 3	AM
<i>Spondylosium panduriforme</i> (Heimerl) Teiling	AN; CE; DH; PT; SH; AL; AM; BA; JA; PA; VE
<i>Spondylosium planum</i> (Wolle) West & G.S.West	JA
<i>Staurastrum avicula</i> Brébisson	All except PT; SH; BA
<i>Staurastrum brasiliense</i> Nordstedt	PT
<i>Staurastrum cerastes</i> Lundell	AN; DH; PT; AL; BA; FN; PA; PI; VE
<i>Staurastrum chaetoceras</i> (Schröder) G.M.Smith	AN; CA; CE; DH; GA; GN; PT; SH; FN; PA; PI
<i>Staurastrum curvimarginatum</i> Scott et Grönblad	PA
<i>Staurastrum depressiceps</i> Scott et Grönblad	CE; BA; FE; JA; PA
<i>Staurastrum forficulatum</i> Lundell	All lakes
<i>Staurastrum gemelliparum</i> Nordstedt	CA; FN
<i>Staurastrum grillatorium</i> Nordstedt	GA; PT; AC; AL; BA; JA; PA; VE
<i>Staurastrum ionatum</i> Wolle	All except AM; FE
<i>Staurastrum</i> cf. <i>hagmannii</i> Grönblad	PT; BA; PA
<i>Staurastrum</i> cf. <i>hirsutum</i> Borge	DH; SH
<i>Staurastrum laeve</i> Ralfs	All except GN; AM
<i>Staurastrum leptacanthum</i> Nordstedt	CA; PT; AC; BA; FE; JA; VE
<i>Staurastrum leptocladum</i> Nordstedt	All lakes
<i>Staurastrum manfeldtii</i> Delponte	PT; SH; AC; AL; FN; JA; PA; PI; VE
<i>Staurastrum minnesotense</i> Wolle	AN; PT; FE; FN; JA
<i>Staurastrum</i> cf. <i>muticum</i> (Brébisson) Ralfs	AN; CE; GA; GN; PT; AC; FE; FN; PA; PI
<i>Staurastrum nudibrachiatum</i> Borge	PT; AL; BA; JA; VE
<i>Staurastrum orbiculare</i> (Ehrenberg) Ralfs	CA; AM; PI
<i>Staurastrum quadrangulare</i> Brébisson	AC; BA; PI
<i>Staurastrum rotula</i> Nordstedt	All lakes
<i>Staurastrum sebalidii</i> Reinsch	SH; PI
<i>Staurastrum setigerum</i> Gleve	All except CA; CE; AM; PA; PI
<i>Staurastrum sexangulare</i> Bulnheim (Rabenhorst)	PT
<i>Staurastrum smithii</i> (G.M. Smith) Teiling	All except GN; AM; VE
<i>Staurastrum taylorii</i> Grönblad	CA; DH; AC; AM; JA; PA; VE
<i>Staurastrum teliferum</i> Ralfs	AN; CE; DH; AC
<i>Staurastrum teliferum</i> Ralfs var. 2	PT; SH; AC; FE; FN; PA; VE

Continues

Appendix 1. Continuation.

Taxa	Occurrence
<i>Staurastrum tentaculiferum</i> Borge	AC; PA; VE
<i>Staurastrum tetracerum</i> (Kützing) Ralfs ex Ralfs	AN;CA; DH; SH; AC; BA;FE;FN; JA; PA; PI;VE
<i>Staurastrum trifidum</i> Nordstedt	All except SH; AM
<i>Staurastrum wolleanum</i> Butler cited in Wolle	AN; PT; AL; PA; VE
<i>Staurastrum</i> sp. 1	All except DH; AM
<i>Staurastrum</i> sp. 2	AN; CE; DH; GA; AC; AL; FE; JA; VE
<i>Staurastrum</i> sp. 3	CE; AC; AL; AM; VE
<i>Staurastrum</i> sp. 4	GN; AL; FE; FN; VE
<i>Staurastrum</i> sp. 5	CE; PT; BA; JA; PA
<i>Staurastrum</i> sp. 6	FN
<i>Staurastrum</i> sp. 7	CA; PT; JA; PA; PI
<i>Staurastrum</i> sp. 8	DH; SH; BA; PA; PI
<i>Staurastrum</i> sp. 9	BA
<i>Staurastrum</i> sp. 10	DH; SH; AC; BA; JA; PA; PI; VE
<i>Staurastrum</i> sp. 11	GA; PT; SH; AC; AM; BA; JA; PA
<i>Staurastrum</i> sp. 12	SH; JA
<i>Staurastrum</i> sp. 13	PI
<i>Staurastrum</i> sp. 14	VE
<i>Staurastrum</i> sp. 15	AM
<i>Staurastrum</i> sp. 16	AN; CE; FE; FN; JA
<i>Staurastrum</i> sp. 17	PA
<i>Staurastrum</i> sp. 18	AC
<i>Staurastrum</i> sp. 19	AC
<i>Staurastrum</i> sp. 20	AC; JA
<i>Staurastrum</i> sp. 21	AC
<i>Staurastrum</i> sp. 22	GA; FE; FN
<i>Stauroidesmus convergens</i> (Ehrenberg Ex Ralfs) Teiling	All except GN; PT; AM; JA; VE
<i>Stauroidesmus convergens</i> (Ehrenberg Ex Ralfs) Teiling var. 2	AC; BA; FN; JA; PI
<i>Stauroidesmus crassus</i> (West & West) Florin	All except GN; AM
<i>Stauroidesmus cuspidatus</i> (Brébisson) Teiling	All except GA; AC; AL; AM; VE
<i>Stauroidesmus dejectus</i> (Brébisson) Teiling	All lakes
<i>Stauroidesmus</i> cf. <i>extensus</i> (Anderson) Teiling	AC; PA
<i>Stauroidesmus incus</i> (Brébisson) Teiling	CA; CE; FE; PI
<i>Stauroidesmus jaculiferus</i> (W. West) Teiling	All except SH; AM
<i>Stauroidesmus lobatus</i> (Borgesen) Bourrelly	AC; AM; BA; FN; JA; PI; VE
<i>Stauroidesmus o'mearii</i> (Archer) Teiling	CA; CE; DH; GA; FN; VE
<i>Stauroidesmus pachyrhynchus</i> (Nordst.) Teiling	AL; BA; FN
<i>Stauroidesmus phimus</i> (Turner) Thomasson	CE; PI; VE
<i>Stauroidesmus subulatus</i> (Kützing) Croasdale	AN;CA;DH; GN;AL;AM; BA; FE; FN; JA; PA;PI
<i>Stauroidesmus</i> sp. 1	DH; AC; AL; FE; FN; JA; PA; VE
<i>Stauroidesmus</i> sp. 2	GA; AC; FN; PA; VE
<i>Stauroidesmus</i> sp. 3	AN; CE; PT; AL
<i>Stauroidesmus</i> sp. 4	FE; PA

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Appendix 1. Continuation.

Taxa	Occurrence
<i>Staurodesmus</i> sp. 5	AN; AL
<i>Staurodesmus</i> sp. 6	JA
<i>Staurodesmus</i> sp. 7	AN; VE
<i>Staurodesmus</i> sp. 8	FN; JA; PA
<i>Staurodesmus</i> sp. 9	PT; PI
<i>Staurodesmus</i> sp. 10	CA; VE
<i>Staurodesmus</i> sp. 11	VE
<i>Staurodesmus</i> sp. 12	AM
<i>Staurodesmus</i> sp. 13	AM; PI
<i>Staurodesmus</i> sp. 14	JA
<i>Staurodesmus</i> sp. 15	AC
<i>Teilingia granulata</i> (J.Roy & Bisset) Bourrelly	All except AM
<i>Teilingia</i> sp. 1	GA
<i>Teilingia</i> sp. 2	GN; AL; VE
<i>Triploceras gracile</i> Bailey	JA; PA
<i>Xanthidium regulare</i> Nordstedt	PA
<i>Zygnema</i> aff.	GN