

## MUSCLE INVOLVEMENT IN LEPROSY

### STUDY OF THE ANTERIOR TIBIAL MUSCLE IN 40 PATIENTS

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**ABSTRACT** - The involvement of skeletal striated muscle in leprosy is considered secondary due to peripheral neuropathy, but some studies point it to a primary muscle lesion. In order to investigate the muscle involvement in leprosy, we studied 40 patients (lepromatous 23, tuberculoid 13, borderline 2 and indeterminate 2). The motor nerve conduction of the peroneal nerves had a reduction of the velocity, decreased compound muscle action potential and sometimes absence of potentials. The electromyographic study of the anterior tibial muscle showed signs of recent and chronic denervation in 77.5% of the cases and no myopathic potentials. The anterior tibial muscle biopsy revealed denervation in 45% of the cases, interstitial inflammatory myopathy in 30% and mixed (myopathic and neuropathic) pattern in 12.5%. Acid fast bacillus was detected in 25% of the cases, always in the interstitial tissue. Inflammatory reaction was present in the interstitial space and in patients with the lepromatous type. The histological findings clearly defined the presence of the so-called "Leprous Interstitial Myositis" on the top of denervation signs.

**KEY WORDS:** leprosy, skeletal muscle, muscle biopsy, myositis.

#### **Alterações musculares na lepra: estudo do músculo tibial anterior em 40 pacientes**

**RESUMO** - O envolvimento do músculo estriado na lepra é considerado secundário à lesão dos nervos periféricos, mas alguns estudos relataram acometimento muscular primário. A fim de verificar esta controvérsia estudamos 40 pacientes com lepra, sendo 23 da forma lepromatosa, 13 da tuberculóide, 2 borderline e 2 indeterminada. Realizamos a neurocondução do nervo peroneiro, junto com eletromiografia e biópsia do músculo tibial anterior. Encontramos redução de velocidade de condução, da amplitude e algumas vezes ausência de potenciais no nervo peroneiro. A eletromiografia do tibial anterior mostrou sinais de desinervação recente e crônica em 77,5% dos casos e não foi encontrada evidência de padrão "miopático". A biópsia do músculo tibial anterior revelou desinervação em 45% dos casos, miopatia inflamatória intersticial em 30% e padrão misto (miopático e neuropático) em 12,5%. Bacilos álcool-ácido resistentes foram encontrados em 25% dos casos, sempre localizados no perimísio e endomísio. Na forma lepromatosa, a reação inflamatória estava presente no espaço intersticial. Os dados histológicos claramente definiram a presença de "Miosite Lepromatosa Intersticial" sobreposta às alterações histológicas encontradas em desinervação.

**PALAVRAS-CHAVES:** lepra, músculo esquelético, biópsia muscular, miosite.

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Leprosy is a chronic infection illness caused by the *Mycobacterium leprae* (M.leprae) that preferably attacks peripheral nerves. This disease is endemic all over the world, specifically in Brazil, and it is considered the commonest cause of treatable peripheral neuropathy<sup>1-3</sup>.

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The involvement of the skeletal striated muscle in leprosy has been characterized as a consequence of the peripheral neuropathy with subsequent muscular denervation<sup>2,4,5</sup>. However, some experimental studies with inoculation of the *M. leprae* in animals point to a primary muscle involvement<sup>6-10</sup>. This primary muscle involvement has also been reported<sup>11-15</sup> and an inflammatory reaction was described, being called lepromatous myositis<sup>16-18</sup>, leprosy interstitial myositis<sup>19</sup> or leprosy nodular interstitial myositis<sup>20</sup>. However, these studies do not give the incidence of this abnormality and histological/histochemical details are lacking.

We undertook this study in order to verify the histologic and electromyographic alterations in the muscle of patients with leprosy, as well as to look for their possible relationship.

## METHOD

We studied 40 patients with an established diagnosis of leprosy through clinical examination, skin biopsy and bacilloscopy; 34 (85%) were male and 6 (15%) female. Their mean age was 45.6 years and the average disease duration was 3.85 years. Of these patients, 57.5% had the lepromatous form (Virchow form), 32.5% the tuberculoid form, 5% the borderline form (dimorphous) and 5% an indeterminate form (Table 1). None of the patients had any clinical symptoms suggesting polymyositis or any other myopathy; four had mononeuropathy, 24 had multiple mononeuropathy and 10 had polyneuropathy (Table 1). Four patients (10%) were studied before treatment and 36 (90%) under drug therapy (36 with dapsone, 6 with rifampin, 5 with clofazimine, 6 with thalidomide and 5 with prednisone). The mean treatment time was 3.48 years (one month to 18 years).

White and red blood cell counts, erythrocyte sedimentation rate (ESR), serum glucose, creatinine, syphilis Venereal Disease Research Laboratory test (VDRL), creatinekinase (CK), aldolase, lactic dehydrogenase (LDH), aspartate aminotransferase (SGOT) and alanineamino transferase (SGPT) were done in all patients.

We studied the motor conduction velocity of the peroneal nerve with cutaneous electrodes on both sides. We stimulated below the head of the fibula and just above the ankle with recording electrodes placed over the extensor digitorum brevis. We recorded the proximal and distal latencies, the amplitude and duration of the compound motor muscle potential and calculated the conduction velocity after measuring the distance between the stimulation points<sup>22</sup>.

Electromyography (EMG) was performed on the anterior tibial muscle with a coaxial needle electrode. We chose to study the anterior tibial muscle because it is a distal and superficial one, possible with lower temperature. Also the peroneal nerve enervates the anterior tibial muscle, the second most common nerve involved in leprosy. At rest we registered spontaneous activity like insertion activity, fibrillation, fasciculation, positive waves, high frequency and myotonic discharges. At voluntary contraction, we evaluated the duration, amplitude, degree of polyphasic potentials, as well as the recruitment pattern during weak and strong contraction<sup>21,22</sup>. We called it recent denervation when we found only spontaneous potentials with all the other parameters being normal; chronic denervation, when we had increased duration, large amplitude, excess of polyphasic potentials and decreased recruitment; recent and chronic with the combination of both, and borderline when we had only few or one type of abnormality present.

Table 1. Clinical findings.

| Clinical type            | Lepromatous | Tuberculoid | Borderline | Indeterminate |
|--------------------------|-------------|-------------|------------|---------------|
| Number of cases          | 23          | 13          | 2          | 2             |
| Mean age (years)         | 43.2        | 50.5        | 50.5       | 36            |
| Disease duration (years) | 3.9         | 4.3         | 1.5        | 2.5           |
| Mononeuropathy           | 1           | 2           | 0          | 1             |
| Multiple mononeuropathy  | 13          | 7           | 2          | 1             |
| Polyneuropathy           | 7           | 3           | -          | -             |
| Nerve enlargement        | 20          | 9           | 2          | 0             |

All the patients were submitted to an anterior tibial muscle biopsy, always on the opposite side of the electromyography. The muscle fragment was studied with fresh-frozen sections and submitted to the following staining and histochemical reaction: hematoxylin-eosin, modified Gomori trichrome, oil red O, PAS, cresyl violet, sirius red, NADH-tetrazolium reductase, ATPases pH 4.3, 4.6, 9.4, myophosphorylase, non-specific esterase, alkaline phosphatase, acid phosphatase, succinic dehydrogenase and cytochrome c-oxidase<sup>23</sup>.

The final histological diagnoses were reported as normal (no alterations found), interstitial inflammatory myopathy (inflammatory infiltrate in the perimysium, diffuse or in the perivascular space, necrosis, phagocytosis, excess of internal nuclei, basophilic fibers, increased acid phosphatase in the fibers and in the interstitial cells), active denervation (atrophic angulated fibers in the NADH-tetrazolium reductase and non-specific esterase and rare necrotic fibers), chronic denervation (when found target fibers, type grouping, absence of necrosis and phagocytosis, type 1 and 2 atrophy, type 1 and 2 hypertrophy), active and chronic denervation (combination of all elements of chronic and active denervation) and mixed (combination of elements of interstitial inflammatory myopathy, active and chronic denervation). Ziehl-Neelsen coloration was done on all specimens using Fite-Faraco method. Each specimen had several serial slides section and an extensive search for acid fast bacilli (AFB) was conducted<sup>21</sup>.

## RESULTS

Laboratory investigation found an increase of ESR in 17 cases, hypochromic and microcytic anemia in 12, eosinophilia and intestinal parasites in 8, positive VDRL with negative FTA-ABS in 7. Two cases had a slight elevation of the SGOT and SGPT, but they had also liver involvement (Hepatic granuloma on liver biopsy). All the cases had normal CK, aldolase and LDH.

The nervous conduction of the right peroneal showed reduction of the velocity in 28.2% of the cases, normal conduction in 48.7% and absence of conduction in 23%. The distal latency was increased in 36.6% and there was a reduction in the amplitude in 60% of the cases. The left peroneal nerve had reduction of the velocity in 17% of the cases, normal conduction in 64% and absence of conduction in 17.6%. The distal latency was increased in 32.1% and there was a reduction in the amplitude of motor response in 60% of the cases (Table 2).

The electromyographic examinations showed denervation signs in 31 (77.5%) cases, borderline signs of denervation in 3 (7.5%) and were normal in 6 cases (15.0%) (Table 3).

The most frequent abnormalities found in the muscle biopsy of all the cases were: variation of muscle fiber diameter, type 1 fiber predominance, atrophic rounded fibers, type 2 fiber atrophy, type 1 fiber atrophy, type grouping, atrophic dark angulated fibers in the non-specific esterase and NADH-tetrazolium reductase. There were some differences in the incidence of abnormalities regarding the type of leprosy, but no statistical difference was found between the tuberculoid and lepromatous types (Chi-Square test) (Table 4). The inflammatory infiltrate was more prominent in the perimysium and less in the endomysium, with rare fibers with necrosis and phagocytosis (Figs 1,2,3,4). The inflammatory reaction of the perimysium was also present into the nerve branches, which could be responsible for the denervation found in the muscle (Fig 5). The AFB were found exclusively in the lepromatous type and they were located in the interstitial space, inside macrophages, vessels walls and intramuscular nerves (Fig 6) (Table 4).

The muscle biopsy abnormalities permitted the following histological diagnosis: denervation in 45% of the cases, inflammatory myopathy in 30%, (17.5% with the presence of AFB), mixed abnormalities with myopathic and neuropathic findings in 12.5% of the cases, and non specific abnormalities in 15% of the cases. The AFB in the muscle was detected in 25% of the cases (Tables 4 and 5).

Comparing the electromyographic abnormalities and the histological diagnosis, we found absence of a myopathic pattern in the EMG. All the cases with interstitial inflammatory reaction had a denervation pattern in the EMG. Also, 14 cases had denervation in the histology and denervation type of EMG. Of the four cases with non-specific findings, only one had a normal biopsy (Table 6).

Table 2. Nervous conduction velocity: peroneal nerve.

| Case Number | Right          |              |              | Left           |              |              |
|-------------|----------------|--------------|--------------|----------------|--------------|--------------|
|             | Velocity (m/s) | Latency (ms) | Voltage (mV) | Velocity (m/s) | Latency (ms) | Voltage (mV) |
| 1           | 39.4           | 5.6          | 3.0          | 49.7           | 5.4          | 1.2          |
| 2           | A              | A            | A            | 34.6           | 3.3          | 2.0          |
| 3           | 44.8           | 3.5          | 9.0          | 45.4           | 4.0          | 8.0          |
| 4           | 35.7           | 12.1         | 4.0          | 32.0           | 12.4         | 4.0          |
| 5           | A              | A            | A            | -              | -            | -            |
| 6           | A              | A            | A            | A              | A            | A            |
| 7           | A              | A            | A            | A              | A            | A            |
| 8           | 41.9           | 4.0          | 10.0         | 51.7           | 4.7          | 8.0          |
| 9           | A              | A            | A            | A              | A            | A            |
| 10          | A              | A            | A            | -              | -            | -            |
| 11          | 49.0           | 7.3          | 1.5          | 32.5           | 6.3          | 1.0          |
| 12          | 46.6           | 4.0          | 4.0          | 45.6           | 4.5          | 4.0          |
| 13          | 45.9           | 3.9          | 2.0          | 45.6           | 4.5          | 2.0          |
| 14          | 68.5           | 3.2          | 4.0          | 54.5           | 3.6          | 3.0          |
| 15          | 52.0           | 3.4          | 0.4          | 51.8           | 3.6          | 5.0          |
| 16          | 43.4           | 5.1          | 5.0          | 42.6           | 3.9          | 4.0          |
| 17          | A              | A            | A            | 38.1           | 6.9          | 0.3          |
| 18          | A              | A            | A            | 33.8           | 6.1          | 0.4          |
| 19          | 48.2           | 4.4          | 8.0          | 50.0           | 4.7          | 10.0         |
| 20          | 50.0           | 11.0         | 0.6          | 60.0           | 11.5         | 0.4          |
| 21          | 36.0           | 5.3          | 2.0          | 46.0           | 5.1          | 2.0          |
| 22          | 36.5           | 5.3          | 2.0          | 46.1           | 5.1          | 2.0          |
| 23          | 45.2           | 4.6          | 1.0          | 45.5           | 3.8          | 3.0          |
| 24          | 31.2           | 14.2         | 2.0          | -              | -            | -            |
| 25          | 46.8           | 5.1          | 3.0          | -              | -            | -            |
| 26          | 43.0           | 4.6          | 1.5          | 53.5           | 4.3          | 1.5          |
| 27          | A              | A            | A            | A              | A            | A            |
| 28          | 48.5           | 3.4          | 10.0         | 50.0           | 3.3          | 4.0          |
| 29          | -              | -            | -            | -              | -            | -            |
| 30          | 34.0           | 6.2          | 0.7          | -              | -            | -            |
| 31          | 39.4           | 4.0          | 2.0          | A              | A            | A            |
| 32          | 60.4           | 3.9          | 3.0          | 54.9           | 3.5          | 4.0          |
| 33          | 45.6           | 3.9          | 2.0          | 51.7           | 3.7          | 2.0          |
| 34          | 37.6           | 4.4          | 6.0          | 45.8           | 3.8          | 4.0          |
| 35          | 59.0           | 4.9          | 0.8          | 50.9           | 4.6          | 1.5          |
| 36          | 53.3           | 3.3          | 10.0         | 57.1           | 3.7          | 8.0          |
| 37          | 26.3           | 16.8         | 0.15         | A              | A            | A            |
| 38          | 52.6           | 4.3          | 10.0         | 58.0           | 7.4          | 0.8          |
| 39          | 32.2           | 6.7          | 0.1          | 40.2           | 5.8          | 0.3          |
| 40          | 37.2           | 5.7          | 4.0          | 35.2           | 5.2          | 1.0          |

A, Absence of potential; (-), not realized. Normal values: Velocity > 40.0 m/s; Latency < 5.1 ms; Voltage > 4000  $\mu$ V.

Table 3. Electromyographic abnormalities (%).

| Type of abnormality present<br>Number of cases | Total<br>40 | lepromatous<br>23 | Tuberculoid<br>13 |
|--|-------------|-------------------|-------------------|
| Insertion activity                             |             |                   |                   |
| Increased                                      | 5.0         | -                 | 15.4              |
| Decreased                                      | 2.5         | 4.3               | -                 |
| Fibrillation potentials                        | 20.0        | 21.7              | 23.1              |
| Positive sharp waves                           | 25.0        | 21.7              | 30.8              |
| Fasciculation potentials                       | 5.0         | -                 | 7.7               |
| Complex repetitive discharges                  | 7.5         | 4.3               | 15.4              |
| Duration of motor unit potentials              |             |                   |                   |
| Increased                                      | 47.5        | 47.8              | 38.5              |
| Decreased                                      | 2.5         | -                 | 7.7               |
| Amplitude of motor unit potentials             |             |                   |                   |
| Increased                                      | 5.0         | 4.3               | -                 |
| Decreased                                      | 2.5         | -                 | 7.7               |
| Short polyphasic motor units                   | 10.0        | 13.0              | 7.7               |
| Long polyphasic motor units                    | 90.0        | 95.7              | 84.6              |
| Recruitment pattern                            |             |                   |                   |
| Increased                                      | 2.5         | -                 | 7.7               |
| Decreased                                      | 80.0        | 91.3              | 61.5              |

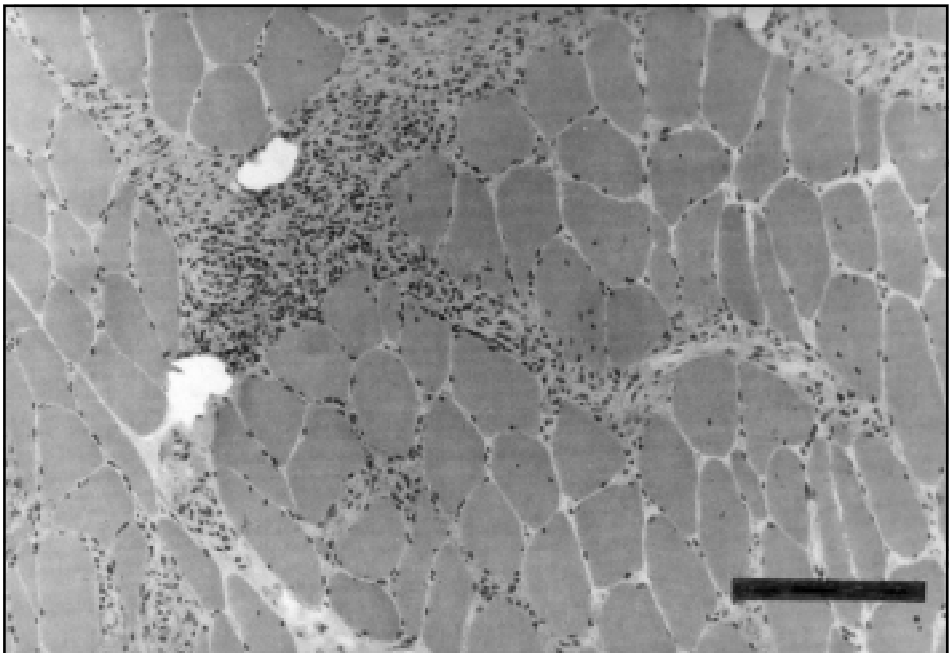


Fig 1. Inflammatory infiltrate in the perimysium spreading through the vasculo-nervous terminals to the endomysium. Haematoxylin & Eosin. Bar 200  $\mu$ .

Table 4. Anterior tibial muscle biopsy abnormalities (%).

|                                  | Histological/histochemical alteration   | Total | Lepromatous | Tuberculoid |
|----------------------------------|---|-------|-------------|-------------|
|                                  | Number of cases                         | 40    | 23          | 13          |
| HE and modified Gomiri trichrome | Connective tissue proliferation         | 15.0  | 13.0        | 15.4        |
|                                  | Adipose tissue infiltration             | 17.5  | 17.3        | 15.4        |
|                                  | Necrosis                                | 15.0  | 8.7         | 23.1        |
|                                  | Phagocytosis                            | 12.5  | 8.7         | 15.4        |
|                                  | Diffuse inflammatory infiltrate         | 32.5  | 43.5        | 23.1        |
|                                  | Perivascular inflammatory infiltrate    | 32.5  | 43.5        | 15.4        |
|                                  | Internal nuclei                         | 25.0  | 8.7         | 46.2        |
|                                  | Basophilic fibers                       | 5.0   | 4.3         | 0           |
|                                  | Hypercontractile fibers                 | 5.0   | 4.3         | 7.7         |
|                                  | Inflammatory infiltrate excessive       | 27.5  | 39.1        | 15.4        |
|                                  | Variation of muscular fibers diameter   | 95.0  | 91.4        | 100.0       |
|                                  | Large agglomerations of atrophic fibers | 7.5   | 0           | 15.4        |
|                                  | Small agglomerations of atrophic fibers | 15.0  | 4.3         | 30.8        |
|                                  | Atrophic rounded fibers dispersed       | 60.0  | 65.2        | 53.9        |
|                                  | Hypertrophic fibers dispersed           | 30.0  | 21.7        | 53.9        |
|                                  | Nuclear clumps                          | 22.5  | 13.0        | 30.8        |
|                                  | Granular fibers (ragged reds)           | 2.5   | 0           | 7.7         |
|                                  | Vacuoles                                | 10.0  | 4.3         | 15.4        |
|                                  | Cytoplasmatic inclusion body            | 2.5   | 4.3         | 0           |
|                                  | Splitting fibers                        | 2.5   | 0           | 7.7         |
| NADH-tetrazolium reductase       | Atrophic angulated fibers               | 42.5  | 43.5        | 30.8        |
|                                  | Target fibers                           | 5.0   | 0           | 7.7         |
|                                  | Moth-eaten fibers                       | 5.0   | 0           | 7.7         |
|                                  | Ring fibers                             | 10.0  | 4.3         | 23.1        |
|                                  | Focal increase NBT granules             | 47.5  | 52.2        | 46.2        |
|                                  | Type grouping                           | 45.0  | 47.8        | 30.8        |
| ATPase pH 9.4,4.3,4.6            | Type 1 fiber atrophy                    | 50.0  | 60.9        | 38.5        |
|                                  | Type 2 fiber atrophy                    | 57.5  | 60.9        | 53.8        |
|                                  | Type 1 fiber hypertrophy                | 25.0  | 13.0        | 46.2        |
|                                  | Type 2 fiber hypertrophy                | 27.5  | 21.7        | 38.5        |
|                                  | Type 2 fiber predominance               | 2.5   | 4.3         | 0           |
|                                  | Type 2 deficiency fibers                | 12.5  | 8.7         | 23.1        |
| Non-specific esterase            | Motor plate - Increased size            | 5.0   | 4.3         | 7.7         |
|                                  | Atrophic angulated fibers               | 42.5  | 34.8        | 46.2        |
|                                  | Interstitial activity in mononuclears   | 37.5  | 52.2        | 15.4        |
| Acid phosphatase                 | Focal increased into fibers             | 37.5  | 21.7        | 46.2        |
|                                  | Positive fibers                         | 7.5   | 4.3         | 15.4        |
|                                  | Interstitial isolated mononuclear cells | 35.0  | 43.4        | 23.1        |
|                                  | Phagocytes near or in necrotic fibers   | 10.0  | 8.7         | 7.7         |
|                                  | Nerves (Mononuclear/myelin sheath)      | 17.5  | 26.1        | 0           |
| Alkaline phosphatase             | Positive fibers                         | 7.5   | 0           | 15.4        |
|                                  | Interstitial increase                   | 37.5  | 47.8        | 15.4        |
|                                  | Nerves (Mononuclear/myelin sheath)      | 5.1   | 8.7         | 0           |
| Oil Red O                        | Type 1 fiber increased                  | 5.0   | 0           | 7.7         |
| Succinic dehydrogenase           | Sub-sarcolemmal accumulation            | 50.0  | 52.2        | 38.5        |
| Cytochrome c-oxidase             | Sub-sarcolemmal accumulation            | 32.5  | 39.1        | 15.4        |
| Sirus red                        | Interstitial tissue increase            | 10.0  | 0           | 23.1        |
| Fite-Faraco for acid             | Endomysium and perimysium               | 17.5  | 30.4        | 0           |
| Fast bacilli                     | Inside macrophage                       | 20.0  | 34.8        | 0           |
|                                  | Around blood vessel                     | 7.5   | 13.0        | 0           |
|                                  | Inside intra-muscular nerves            | 10.0  | 17.4        | 0           |

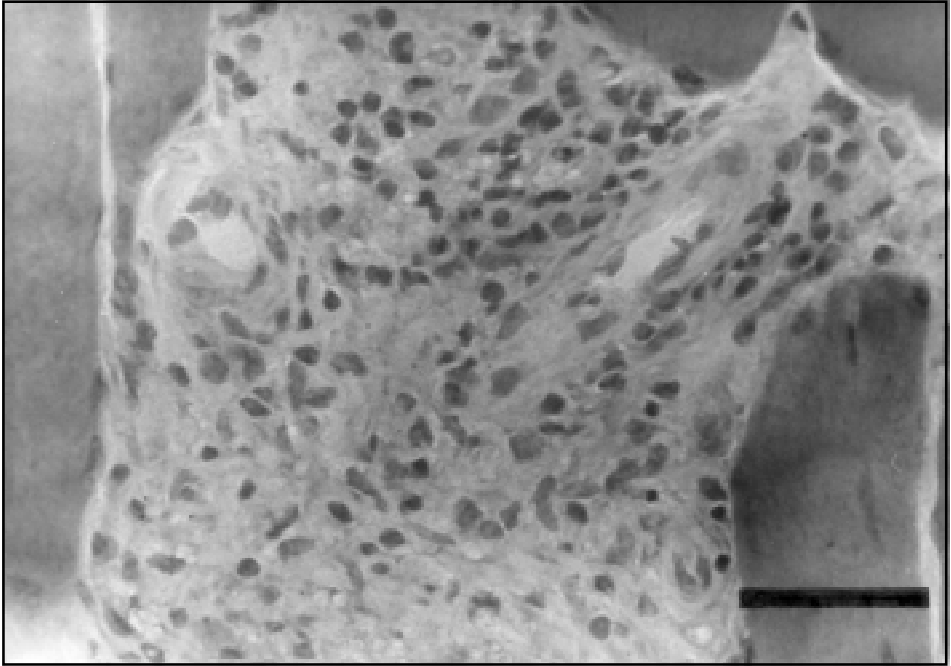
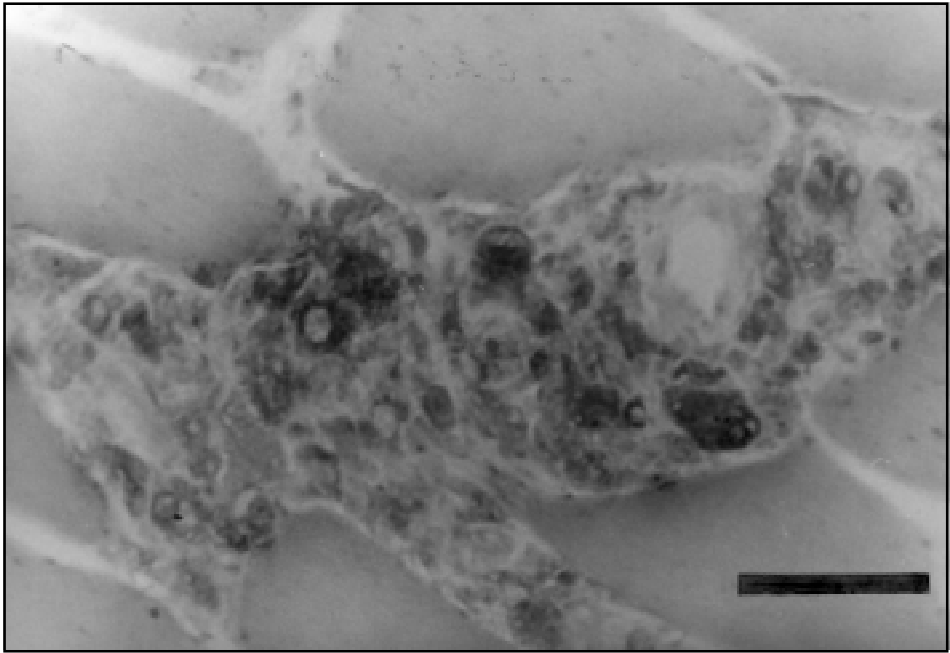


Fig 2. Inflammatory infiltrate in the perimysium with mononuclear cells. Haematoxylin & Eosin. Bar 50  $\mu$ .

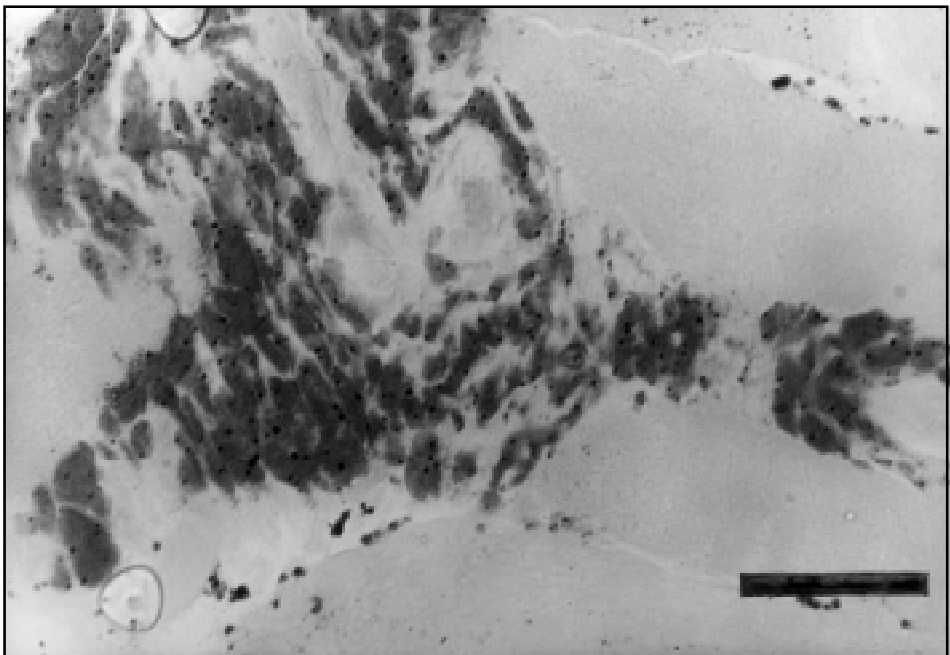
Table 5. Histological diagnosis of anterior tibial muscle biopsy.

|                              | Types of leprosy |             |             |            |               |
|------------------------------|------------------|-------------|-------------|------------|---------------|
|                              | All types        | Lepromatous | Tuberculoid | Borderline | Indeterminate |
| Normal                       | 3                | 1           | 1           | -          | 1             |
| Inflammatory myopathy        | 5                | 2           | 2           | 1          | -             |
| Inflammatory myopathy + AFB  | 6                | 6           | -           | -          | -             |
| Active denervation           | 3                | 1           | 1           | -          | 1             |
| Active denervation + AFB     | 1                | 1           | -           | -          | -             |
| Chronic denervation          | 8                | 5           | 2           | 1          | -             |
| Chronic denervation + AFB    | 1                | 1           | -           | -          | -             |
| Active + chronic denervation | 4                | 2           | 2           | -          | -             |
| Mixed                        | 4                | 2           | 2           | -          | -             |
| Mixed + AFB                  | 1                | 1           | -           | -          | -             |
| Non specific                 | 4                | 1           | 3           | -          | -             |
| Total                        | 40               | 23          | 13          | 2          | 2             |

AFB, acid fast bacillus; Non specific (Inflammatory neuropathy, type 1 fiber atrophy, type 2 fiber atrophy, borderline for myopathy); Mixed (Denervation + Myopathy).



*Fig 3. Inflammatory cells (macrophages) in the interstitial, spreading into the endomysium through the vasculo-nervous space. Acid phosphatase. Bar 50  $\mu$ .*



*Fig 4. Macrophages filled with lipids, in the interstitial space. Oil red O. Bar 50  $\mu$ .*



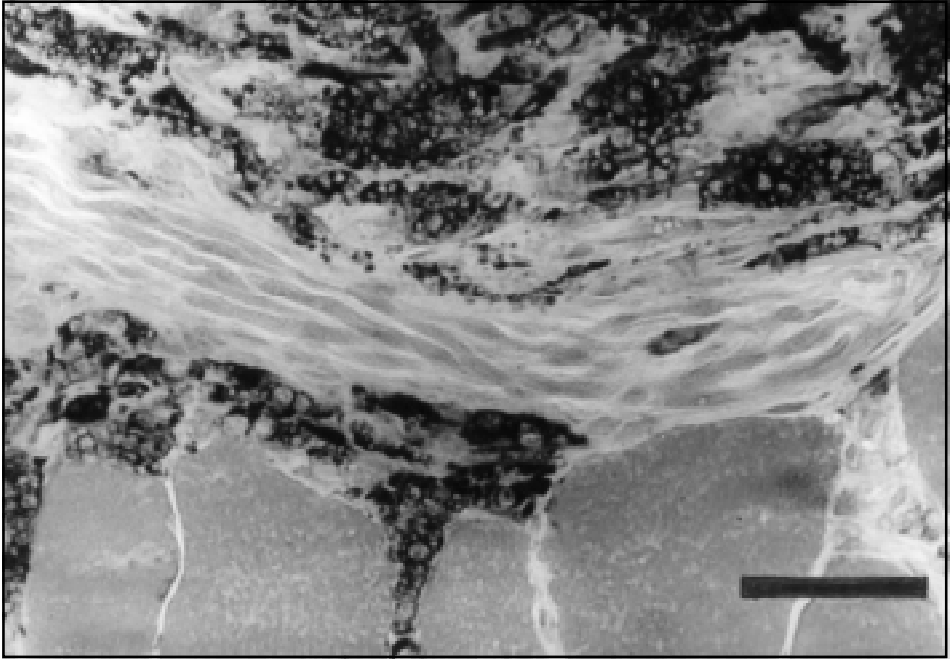


Fig 5. Lymphomononuclear cells near a nervous branch in the interstitial space. Non specific esterase. Bar 50  $\mu$ .

Table 6. Relation between anterior tibial muscle biopsy and electromyography.

| Histological diagnosis             | EMG         |            |        |         |                    |
|------------------------------------|-------------|------------|--------|---------|--------------------|
|                                    | Denervation |            |        |         |                    |
|                                    | Normal      | Borderline | Active | Chronic | Active and Chronic |
| Normal                             | 1           | -          | -      | 2       | -                  |
| Interstitial inflammatory myopathy | -           | 1          | 1      | 8       | 1                  |
| Active denervation                 | 1           | -          | 1      | 2       | -                  |
| Chronic denervation                | 1           | -          | -      | 5       | 3                  |
| Active and chronic denervation     | 1           | -          | -      | 3       | -                  |
| Mixed                              | 1           | 1          | 1      | -       | 2                  |
| Non specific                       | 1           | 1          | -      | 1       | 1                  |
| Total                              | 6           | 3          | 3      | 21      | 7                  |

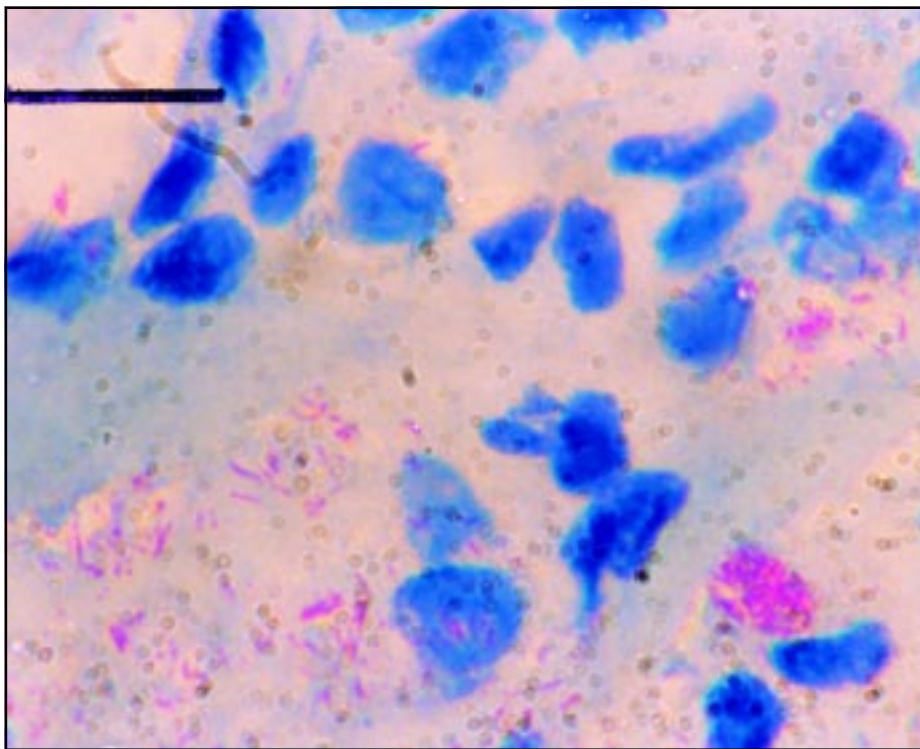


Fig 6. Acid fast bacilli (red) inside macrophages and interstitial tissue in the vasculo-nervous space. Ziehl-Neelsen (Fite-Faraco). Bar 20  $\mu$ .

## DISCUSSION

The neurophysiological evaluation of these patients demonstrated that the study of nervous conduction in the peroneal nerves brought out meaningful results. Some patients had no conduction or very large latencies due to the severity of the disease. These abnormalities were asymmetric, according to the nature of the disease. These findings are in agreement with the literature, which points to the peroneal nerve as the most frequently compromised one, after the ulnar nerve<sup>24-27</sup>.

The electromyographic study of the tibialis anterior muscle showed alterations in most of the cases; 77.5% showed signs of denervation. These denervation potentials are due to peroneal nerve or its branch involvement<sup>2</sup>. The electromyographic examination allows the detection of precocious alterations in leprosy, even when clinical examination gives no evidence of it<sup>18</sup>. Electromyographic alterations can be detected in up to 98% of the cases when several muscles are studied<sup>25,28</sup>. We did not find electromyographic potentials compatible with primary muscle involvement (myopathic), as it had rarely been described<sup>18,29-31</sup>. The scope of this work was the study of the involvement of the anterior tibial muscle in leprosy and for this reason we did not study other muscles with EMG, even knowing that we could find abnormalities in other muscles, if a complete examination was done. Also, even studying only one muscle, as in this paper, a good correlation can be obtained, according previously published papers<sup>21,32</sup>.

As expected, we found a histological pattern of denervation, with angulated atrophic fibers, piknotic nuclear clumps, large agglomerations of atrophic fibers and type grouping<sup>21,23</sup>. We were able to record the presence of an inflammatory infiltrate superimposed to the denervation, preponderant

in the interstitial and peri-fascicular area. This infiltration was of mononuclear cells, monocytes, lymphocytes and macrophages. Sometimes, it was very severe, with the formation of a granuloma with giant cells. These findings were described previously, with some variation as "Leprous Interstitial Myositis"<sup>12-14,16-19,33,34</sup>.

The inflammatory infiltration was present in the perimysium, near the blood vessels, and infiltrated the muscle following the vascular tree inside de muscle fascicle. In reality, it does not damage the muscle fibers and fiber necrosis was rare. The inflammatory infiltration among the striated muscle fibers, muscular fascicle atrophy as well as the presence of AFB was previously called "Myositis Interstitialis Leprosa"<sup>16,19</sup>. Most of our patients who presented inflammatory interstitial myopathy were carriers of leprosy in its lepromatous form, as it had already been observed in the literature. The fact that, in this group of patients, there is anergy to *M. leprae*, with systemic dissemination of the bacillus, somehow explains the characterization of a real bacillary septicemia<sup>13,14</sup>.

The presence of AFB was observed in 7 cases with inflammatory reaction, being detected in the interstitial area of muscle fascicles, together with inflammatory cells, granulomas, inside the macrophages, around vessels, nerves, in the arterioles, in nerve fascicles and inside foam cells. The presence of AFB, compatible with *M. leprae* in the smooth muscle and in the striated muscle of patients with leprosy has been demonstrated<sup>4,11-14,16,17,19,20,34,35</sup>.

Job et al. described alterations in the striated muscle, defining three levels of lesions: on the first, there is an invasion and proliferation of *M. leprae* in the muscular fibers; on the second there is a degeneration of the muscular fibers, an infiltration of lymphocytes, macrophages, polymorphonuclear cells and bacillus fragmentation; on the third, a destruction of the muscle fibers occurs, with the presence of fibrous tissues, but with absence of bacilli<sup>17</sup>.

Pearson et al. propose that Hansen's bacillus can survive and multiply in the striated muscle more easily than it does in the skin, and that it is less attacked by the body's immune system, disseminating itself systemically afterwards. The muscle would, then, have an exponential role in the pathogenesis of the leprosy, functioning as a bacillary reservoir<sup>14</sup>.

We did not find AFB inside muscle fibers in our material, even after a long search and several serial slides section for each case, as it was previously described by several authors<sup>14-18,36,37</sup> and in experimental animal studies<sup>6,7,9,10</sup>. Also, some authors doubt the muscle fiber invasion by AFB<sup>12,38</sup>, and the absence of AFB in some of our cases could be due to treatment.

In concluding, we found, besides the denervation process due to peroneal nerve involvement, an interstitial inflammatory myopathy with AFB, the so-called "Leprous Interstitial Myositis" in 45% of the cases. This interstitial myositis was predominantly observed in patients with the lepromatous form. The muscle inflammatory reaction has no electromyographic representation, since the examination only detected denervation, suggesting that the lesion takes place in the perivascular, perineural and interstitial area between the muscle fibers.

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