

# Comparison of multiallelic distances for the quantification of genetic diversity in the papaya

Helaine Christine Cancela Ramos<sup>1</sup>, Messias Gonzaga Pereira<sup>1</sup>, Leandro Simões Azeredo Gonçalves<sup>1\*</sup>, Antonio Teixeira do Amaral Júnior<sup>1</sup> and Carlos Alberto Scapim<sup>2</sup>

<sup>1</sup>Laboratório de Melhoramento Genético Vegetal, Centro de Ciências Tecnológicas Agropecuária, Universidade Estadual do Norte Fluminense "Darcy Ribeiro", Av. Alberto Lamego, 2000, 28013-600, Campos dos Goytacazes, Rio de Janeiro, Brazil.

<sup>2</sup>Departamento de Agronomia, Centro de Ciências Agrárias, Universidade Estadual de Maringá, Maringá, Paraná, Brazil.

\*Author for correspondence. E-mail: Isagrural@yahoo.com.br

**ABSTRACT.** This study aimed to compare multiallelic distances to quantify genetic diversity in papaya. We evaluated forty-three individuals in the  $S_2$  generation, from the backcross between  $F_1$  (Cariflora x SS783) and Cariflora, and four accessions from the UENF/Caliman Germplasm Bank. Genetic distances used were Smouse and Peakall (1999), Kosman and Leonard (2005) and weighted index. Clustering among genotypes was performed using the hierarchical unweighted pair-group method with arithmetic mean analysis (UPGMA) and projection of the distance on the bidimensional plan. A high correlation between genetic distances was observed; however, through UPGMA group analysis, the distance determined by the weighted index provided the complete separation of 52BC<sub>1</sub>S<sub>2</sub>-08, 52BC<sub>1</sub>S<sub>2</sub>-29 and 52BC<sub>1</sub>S<sub>2</sub>-34 inbred lines. Through projection of distances in the plan, Kosman and Leonard (2005) coefficients and weighted allowed the differentiation of individuals in the  $S_2$  generation (52BC<sub>1</sub>S<sub>2</sub>-08, 52BC<sub>1</sub>S<sub>2</sub>-29 and 52BC<sub>1</sub>S<sub>2</sub>-34), the progenitor ('Cariflora' and 'SS783'), and the four germplasm bank accessions in a different manner than the Smouse and Peakall (1999) index, which did not provide this discrimination among the accessed genotypes. We conclude that the Kosman and Leonard (2005) coefficient and weighted index are more efficient than the Smouse and Peakall (1999) algorithm on the disposition of the accessed genotypes in dendrograms and in the Cartesian axis displaying genetic similarity.

**Keywords:** *Carica papaya* L., microsatellite markers, cultivar characterization, cluster analysis.

**RESUMO.** Comparação de distâncias multi-alélicas sobre a quantificação da diversidade genética em mamão. O presente trabalho visou à comparação de distâncias multi-alélicas sobre a quantificação da diversidade genética em mamão. Para tanto, foram avaliados 43 indivíduos da geração  $S_2$ , oriunda do retrocruzamento entre  $F_1$  ('Cariflora' x 'SS783') e 'Cariflora', e quatro acessos do Banco de Germoplasma da UENF/Caliman. As distâncias genéticas utilizadas foram: Smouse e Peakall (1999), Kosman e Leonard (2005) e índice ponderado. Posteriormente foi realizado o agrupamento entre os genótipos utilizando *unweighted pair-group method with arithmetic means analysis* (UPGMA) e projeção de distância no plano bi-dimensional. Observou-se elevada correlação entre as distâncias genéticas, entretanto, pela análise de agrupamento UPGMA, a distância utilizando o índice ponderado proporcionou a completa separação das linhagens 52RC<sub>1</sub>S<sub>2</sub>-08, 52RC<sub>1</sub>S<sub>2</sub>-29 e 52RC<sub>1</sub>S<sub>2</sub>-34. Pela projeção das distâncias no plano, os coeficientes Kosman e Leonard (2005) e índice ponderado permitiram a separação dos indivíduos da geração  $S_2$  (52RC<sub>1</sub>S<sub>2</sub>-08, 52RC<sub>1</sub>S<sub>2</sub>-29 e 52RC<sub>1</sub>S<sub>2</sub>-34) para com os progenitores ('Cariflora' e 'SS783') e em relação aos quatro acessos do banco de germoplasma, diferentemente do índice de Smouse e Peakall (1999), que não proporcionou essa distinção entre os genótipos avaliados. Conclui-se, pois, que o coeficiente Kosman e Leonard (2005) e o índice ponderado foram mais eficientes que o algoritmo de Smouse e Peakall (1999) na disposição dos genótipos avaliados em dendrogramas e eixos cartesianos representativos da similaridade genética.

**Palavras-chave:** *Carica papaya* L., marcador microssatélite, caracterização de cultivares, análise de agrupamento.

## Introduction

The analysis of variance for DNA sequences is of great importance to genetic analysis in plants. Molecular markers are considered important tools to

trace variations in the genome (OLIVEIRA et al., 2010; VARSHNEY et al., 2005). A variety of molecular markers have been developed, including restriction fragment length polymorphisms (RFLPs), simple

sequence repeats (SSRs), random amplified polymorphic DNA (RAPD), sequence tagged sites (STS), expressed sequence tags (EST), sequence characterized amplified regions (SCAR), inter simple sequence repeats (ISSR), amplified fragment length polymorphism (AFLP), and single nucleotide polymorphisms (SNPs) (SCHLÖTTERER, 2004; SHULMAN, 2007; LEAL et al., 2010).

SSRs, or microsatellites, as molecular markers for plant genome analysis are gaining importance and replacing other markers in genetic studies, mainly due to reproducibility, multiallelic traits, codominant inheritance, relative abundance, and wide genome coverage (SQUIRREL et al., 2003; VARSHNEY et al., 2005; ZANE et al., 2002). In genetic breeding programs, SSRs have been used for different purposes, such as assisted selection, genetic variance characterization, linkage disequilibrium analysis, and quantitative trait loci (QTLs) mapping (BERNARDO, 2008; EATHINGNTON et al., 2007; JENA; MACKILL, 2008; SCHULMAN, 2007).

In order to quantify genetic variance among individuals, different similarity and dissimilarity coefficients were described, aiming at building a similarity or dissimilarity matrix from all the possible pairs of genotype combinations in order to trace the population structure based on the affinities of each individual in the set of individuals tested (GONÇALVES et al., 2008, 2009; KOSMAN; LEONARD, 2005). According to Reif et al. (2005), the choice of a coefficient will depend on some factors, such as the properties of molecular markers used (whether codominant or dominant), germplasm genealogy (estimated through covariance among related individuals), operational taxonomic unity (OTU) (e.g., open-pollinated populations, inbred lines, hybrids, and other genetic structures), the objective of the study (e.g., genetic diversity quantification, QTL identification) and adequacy of studies using multivariate analysis.

For codominant markers, coefficients that quantify genetic variance among individuals inside each locus (KOSMAN; LEONARD, 2005; PEAKALL et al., 1995; SMOUSE; PEAKALL, 1999) become favorable for dichotomous coefficients (e.g., Jaccard, Sorensen-Dice and Simple Matching) due to lack of violation of the independence among loci in addition to preventing the loss of information, primarily if there is high heterozygosity, which is more commonly found in some allogamous species (KOSMAN; LEONARD, 2005; LAURENTIN, 2009).

Kosman and Leonard (2005) validated this inadequacy of dichotomous coefficients using a

locus composed of four alleles (A, B, C and D), where there is a specific band, in that 0 and 1 correspond to absence and presence of a band, respectively as follows: A = (1000), B = (0100), C = (0010), and D = (0001). Through this reasoning, the dissimilarities between the genetic states AA and AB are equally distant to AB and AC, with respect to different coefficients. Using the Jaccard or Sorensen-Dice algorithms, for instance, the distance between AA and AB is 0.50 and 0.67 and the distances between AB and AC is 0.33 and 0.50, respectively.

We aim to compare the different algorithms in the composition of divergent groups of papaya. Forty three individuals of the  $S_2$  generation, from a backcrossing between  $F_1$  (Cariflora x SS783) and Cariflora, were evaluated with four accessions from the UENF Germplasm Bank in order to verify the genetic diversity of selections of  $S_2$  as to the parents, as well as to quantify the genetic diversity of the bank accessions of the parent 'Cariflora' using microsatellite molecular markers.

## Material and methods

### Plant materials and DNA extraction

Forty-three hermaphroditic genotypes of the  $S_2$  generation, corresponding to 17, 14 and 12 genotypes from the families 52BC<sub>1</sub>S<sub>2</sub>-34, 52BC<sub>1</sub>S<sub>2</sub>-29II and 52BC<sub>1</sub>S<sub>2</sub>-08, respectively, originating from the plant number 52 from the first backcross (BC<sub>1</sub>) between an  $F_1$  plant and the recurrent Cariflora parent, were evaluated. In addition to the parents and the  $S_2$  generation, four accessions from the germplasm bank (BAG's 1, 2, 3 and 4) were also evaluated. These materials were obtained from the experiment conducted at Caliman Agrícola S.A. Company (Linhares, Espírito Santo State, Brazil) and carried out at the UENF Vegetal Genetic Breeding Laboratory, where the DNA extraction and molecular analysis were performed.

Total DNA cellular was extracted from young leaves of inbred lines using the CTAB method (DOYLE; DOYLE, 1990), with some modifications suggested by Daher et al. (2002). After DNA extraction, DNA was quantified via agarose gel analysis (0.8%) with the High DNA Mass Ladder (Invitrogen, USA). The gel was stained with ethidium bromide, and the image was captured using the Eagle-Eye II photo documentation system.

### SSR analysis

For the SSR marker amplification, 32 primers were selected, as previously reported by Santos et al. (2003) in addition to 45 clone sequences from Pérez et al. (2006) that are publicly available at GenBank

(www.genbank.atlanta.org.edu). Clones containing DNA sequences that flank the microsatellite region were used to design primer pairs (Forward and Reverse) using the sequence analysis programs Genamics Expression version 1.0.0.0 and Oligo version 6.68. In order to obtain a greater primer specificity, a few criteria for primer design were established, such as the minimum primer size (> 14 bp) and T<sub>m</sub>'s ("melting temperatures (T<sub>m</sub>) ranging from 35 to 45°C. Besides these criteria, sequences with high G and C content at their extremities (mainly at the 3'-OH end) were avoided to prevent nonspecific amplification.

DNA from the parents ('Cariflora' and 'SS783') was initially used to optimize the reaction and screen 77 synthesized primers. Amplifications were performed with a final reaction volume of 20 µL, containing 10 ng of DNA, 10 mM Tris-HCl (pH 8.3), 50 mM KCl, 2 mM of MgCl<sub>2</sub>, 100 µM dNTP, 2 µM of each primer, and 1 U of Taq DNA polymerase. Amplifications to optimize annealing temperature were performed on a Eppendorf gradient thermal cycler, according to the following program: denaturation for 4 min. at 94°C, followed by 32 cycles of 94°C for 30 sec, 53°C for 1 min., and 72°C for 1 min.). After 32 cycles, a final extension of 7 min. at 72°C was performed. Amplification products were separated on a non-denatured 8%

polyacrylamide gel, stained with ethidium bromide, and imaged using the Eagle-Eye II system. After optimizing the reaction conditions, 17 primers were selected due to greater complementarity, reproducibility, and presence of polymorphisms among parents (Table 1).

#### Data analysis

Data obtained from the amplification of microsatellite markers were converted to a number code for each allele in the locus. For example, if the locus presented three alleles, the representation was 11, 22 and 33 to homozygous forms (A<sub>1</sub>A<sub>1</sub>, A<sub>2</sub>A<sub>2</sub> and A<sub>3</sub>A<sub>3</sub>) and 12, 13 and 23 to heterozygous forms (A<sub>1</sub>A<sub>2</sub>, A<sub>1</sub>A<sub>3</sub>, and A<sub>2</sub>A<sub>3</sub>). Genetic distances among genotypes were estimated using the Smouse and Peakall (1999), Kosman and Leonard (2005) and weighted index (CRUZ, 2008) coefficients. The grouping among genotypes was performed through the Hierarchical Unweighted Pair-group Method with Arithmetic Means Analysis (UPGMA) and through the projection of the distance in the bidimensional plane (CRUZ; VIANNA, 1994). All analyses were processed through the GENALEX 6 (PEAKALL; SMOUSE, 2006), GENES (CRUZ, 2008) and R (www.r-project.org) software programs.

**Table 1.** Microsatellite primers used in the molecular analysis of the 49 genotypes and the number of alleles found per primer.

Name	Repetition	Sequence (5' → 3')	T <sub>a</sub> (°C)	Number of alleles found
mCpCIR01	CT <sub>(18)</sub> GA <sub>(3)</sub>	F: ATCGTCTCCTTTTTCTGGTT R: TCTGCCCTCCCAATACACTAAT	57	2
mCpCIR02	TC <sub>(24)</sub>	F: AGCCACAACCTACGGGAAAT R: AGTAAACGGAGGAAAATGAGT	57	3
mCpCIR05	TC <sub>(18)</sub>	F: ATCGTCTCCTTTTTCTGGTT R: TTCTGCCCTCCCAATACACTA	57	2
mCpCIR08	CT <sub>(20)</sub> AC <sub>(5)</sub>	F: ACCCAACCAAGCAATCTCCAT R: AGCAAACCACTCACTCTCATA	59	3
mCpCIR09	CT <sub>(9)</sub>	F: TGACCATATAAACCCCTAACGA R: TAAGAAACAGCGAAACCCCTA	59	2
mCpCIR16	CT <sub>(9)</sub>	F: TACACTGCCTAACACCCATT R: AACCAACCATAACTGCCTTT	59	3
mCpCIR17	GA <sub>(14)</sub>	F: ACAACAAGTCCCCAAATCT R: TACACTGCCTAACACCCATT	59	2
mCpCIR23	TC <sub>(8)</sub>	F: ACCATTACTTCCCCCTATTT R: ACTCACTTTCCCTTTCTTTCCA	56	2
mCpCIR28	TC <sub>(8)</sub>	F: ATCAAGGAAGTGCAAATTT R: ATGAGCCAATGAGAAGAGGGA	56	2
mCpCIR35	TC <sub>(20)</sub>	F: ACATACAAAACACTTACCACCA R: TCAGACATACTGCATCTCAA	56	3
mCpCIR39	CT <sub>(10)</sub>	F: ATAGCAAACAGAAAAACCCA R: ATAGAAAAGAGAAAGCGA	59	2
mCpCIR40	TC <sub>(13)</sub> TC <sub>(21)</sub>	F: TCGGTTCTCAGGTTTCTTCTAA R: ACAATCACAGGCACACAT	57	2
mCpCIR45	GA <sub>(14)</sub>	F: AAAAGGACGAAAAGGAGACT R: TTTGAACTACCTACGGAAT	56	3
S285	GAT <sub>(3)</sub>	F: AATGTGTGAGAAATAGGTT R: AATCTATCCTCCTCATGTA	59	3
S414	AC <sub>(7)</sub>	F: ATTCTTAGCCAGATGATGT R: ATTGCATGTACACATACCGT	59	2
S422	GAT <sub>(8)</sub>	F: ACGCATCACACGTATATCTA R: ATAACCTCGCTACATCCTCT	56	3
S552	GAT <sub>(4)</sub>	F: ACAAGTGGAACTCCTATA R: CAATGGAACTTCTGCTACTA	52	2

## Results and discussion

Thirty nine alleles were detected and related to 16 analyzed loci among genotypes composed of a minimum of two and a maximum of three alleles per locus, providing an average of 2.4 alleles per locus (Table 1). However, when only the  $S_2$  population was analyzed, this number was reduced to one to two alleles per locus, providing an average of 1.7 alleles per locus. This low polymorphism level in  $S_2$  is derived from autogamy and favors allele fixation, despite the fact that  $S_2$  generation individuals are derived from a single  $BC_1$  plant. Nevertheless, the frequency of this allele is in accordance with the expected value to the backcrossed progeny, due to the heterozygous character of the recurrent progenitor used in this work.

Analysis of distant correlation estimates, obtained through Smouse and Peakall (1999) and Kosman and Leonard (2005) coefficients and weighted index (CRUZ, 2008), showed high association among genetic distance estimates (Table 2), probably due to the low number of alleles per locus verified in this work. Simulation results based on the increase of the number of alleles for a locus showed that the correlation between the Kosman and Leonard (2005) distance and the Smouse and Peakall (1999) weighted index corresponded to the equation  $\hat{y} = 1.0426 - 0.0278x$ , where  $R^2 = 0.9638$  ( $p < 0.01$ ), which indicated a decrease in Pearson correlation estimates for nearly 2.78% for each allele added to the locus.

**Table 2.** Correlation among matrices of distances from the Smouse and Peakall (1999), Kosman and Leonard (2005) coefficients and Weighted Index (CRUZ, 2008).

Matrices	Smouse and Peakall (1999)	Kosman and Leonard (2005)	Weighted Index (CRUZ, 2008)
Smouse and Peakall (1999)	-		
Kosman and Leonard (2005)	0.9338**	-	
Weighted Index (CRUZ, 2008)	0.9187**	0.9725**	-

\*\*Significant correlation at 1% probability, using the Mantel test, with 1000 permutations.

Using these results, it is possible to validate that in a population composed of high variability and a great number of alleles per locus, the correlation among the Kosman and Leonard (2005) coefficient and the Smouse and Peakall (1999) weighted index will tend not to reveal a high magnitude of association.

The high correlation among the Kosman and Leonard (2005) and the weighted index are mainly due to the coefficient traits because Kosman and Leonard (2005) assert that the total number of alleles that are common in each combination of individuals divided by the number of evaluated loci. Thus, if a locus is composed of three alleles (A, B and C) and

the five possible combinations of diploid individuals (AA, AB, BB, BC and CC), there will be 100% identity and a value 0 for the distance between AA and AA, or between AB and AB. Additionally, between AA and AB or AB and AC, there will be 50% identity, exhibiting a  $\frac{1}{2}$  value of distance. Finally, between the AA and AB individuals, there will be 0% identity and distance will have a value of 1. Weighted index presents the same characteristics on the Kosman and Leonard (2005) coefficient; however, it shows an average number of alleles in each evaluated locus in the population. In this work, the low frequencies of alleles per locus explain the high estimates of correlations.

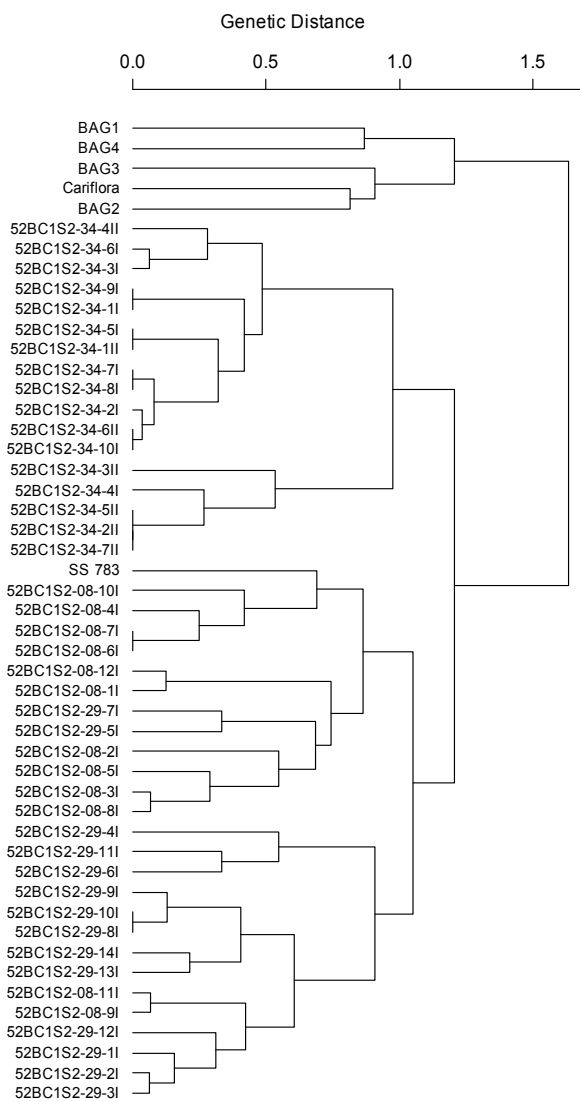
Kosman and Leonard (2005) report that the Smouse and Peakall (1999) proposal to evaluate dissimilarities with respect to a single locus of multiallelic diploid genotypes is rather mechanistic (geometric) and does not possess a strong genetic relationship.

Through UPGMA group analysis, using the Smouse and Peakall (1999) and Kosman and Leonard (2005) coefficients as well as the weighted index, all dendrograms differentiated the germplasm bank accessions (BAG1, BAG2, BAG3 and BAG4) as well as the 'Cariflora' genotype from the other inbred lines (52BC<sub>1</sub>S<sub>2</sub>-08, 52BC<sub>1</sub>S<sub>2</sub>-29 and 52BC<sub>1</sub>S<sub>2</sub>-34) and from the 'SS783' parent (Figures 1, 2 and 3, respectively).

However, in the  $BC_1$  population derived from the crossing between contrasting inbred lines, the expectation is that an average of 75% of the progeny genome can be derived from the recurrent 'Cariflora' parent. When observing the dendrograms, it is possible to verify a greater similarity of inbred lines with respect to the 'SS783' parent. This observation is mainly due to the selection of plants carried out for the first backcrossing generation, which was based only in phenotypic observations. Therefore, quantitative traits favor the similarity of the  $BC_1$  descent with the donor parent.

Genotypes from the UENF/Caliman germplasm bank were included in this work in order to help the identification of a hermaphrodite genotype, which was present among the accessions pertaining to the 'Cariflora' genotype. Previously, in the Cariflora genotypes, only dioic genetic material was known. In this context, two genotypes containing only female flowers (BAG1 and BAG4), one male (BAG3) and the hermaphrodite genotypes under study (BAG2), all of which represented 'Cariflora' (underwent), were submitted for molecular analysis. Analysis of the 16 loci allowed the identification of 33 alleles among the aforementioned genotypes, confirming an average of 2.06 alleles per locus. Using the analysis of the dendrogram, we can verify

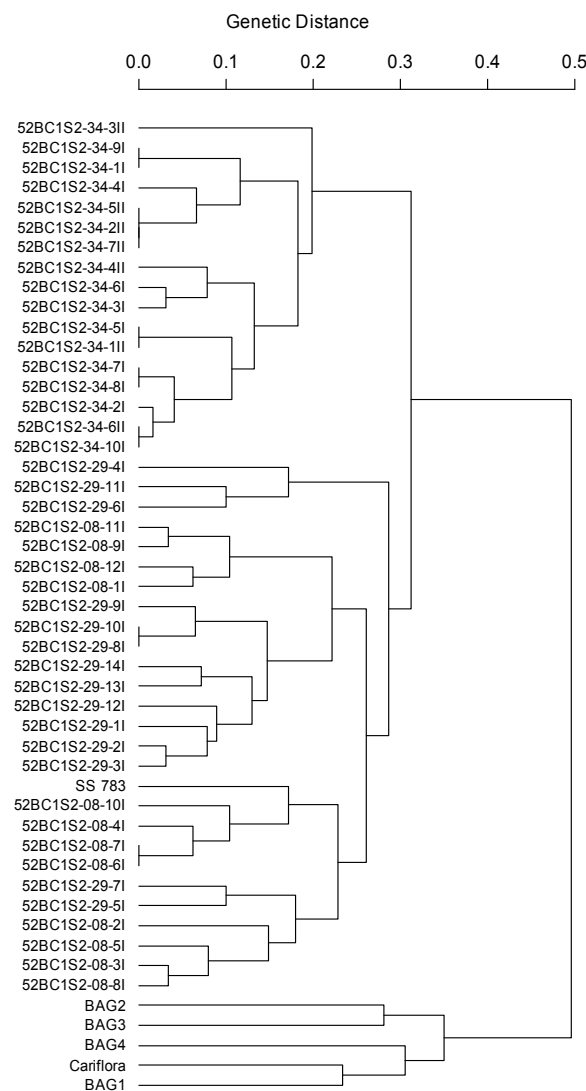
that these four genotypes were grouped next to the 'Cariflora' parent.



**Figure 1.** UPGMA dendrogram based on the analysis of 49 papaya genotypes using the Smouse and Peakall (1999) distance. (Co-phenetic correlation = 0.73).

Comparing dendrograms obtained from the Smouse and Peakall (1999), Kosman and Leonard (2005) coefficients and weighted index, it was verified that the last coefficient provided the complete separation of inbred lines 52BC<sub>1</sub>S<sub>2</sub>-08, 52BC<sub>1</sub>S<sub>2</sub>-29 and 52BC<sub>1</sub>S<sub>2</sub>-34. Furthermore, the Smouse and Peakall (1999) and Kosman and Leonard (2005) coefficients provided dendrograms with some disturbances between 52BC<sub>1</sub>S<sub>2</sub>-08 and 52BC<sub>1</sub>S<sub>2</sub>-29. Concerning the cophenetic correlation, which estimates the association between matrices obtained through grouping and the distance matrix, values of 0.85, 0.83 and 0.73 were observed for the weighted index, the Kosman and

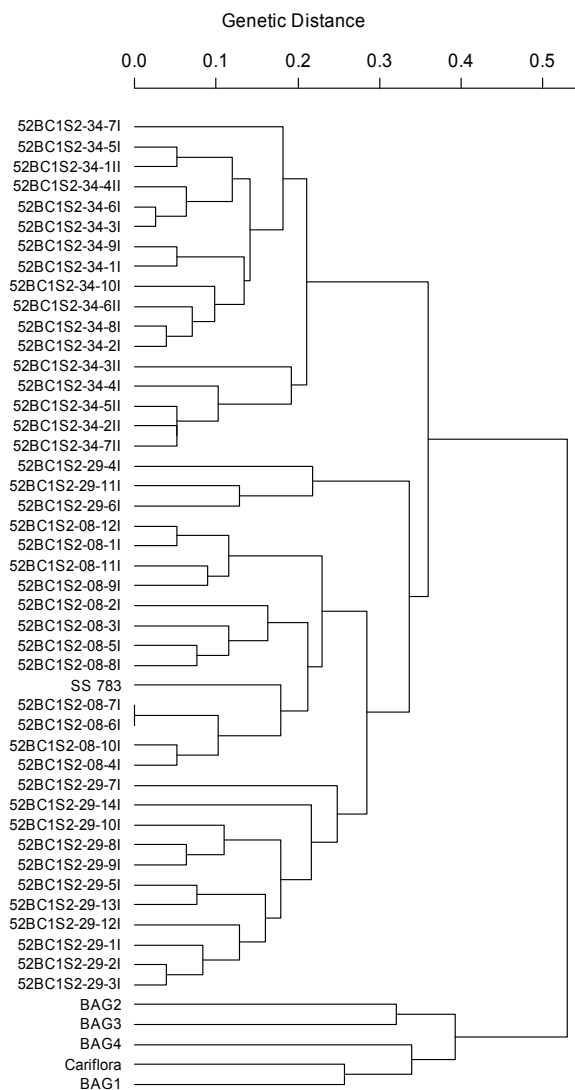
Leonard (2005) and the Smouse and Peakall (1999) coefficients, respectively, with respect to the UPGMA grouping. These results show greater consistency with the weighted index and the Kosman and Leonard (2005) coefficient for quantification of genetic diversity in this study.



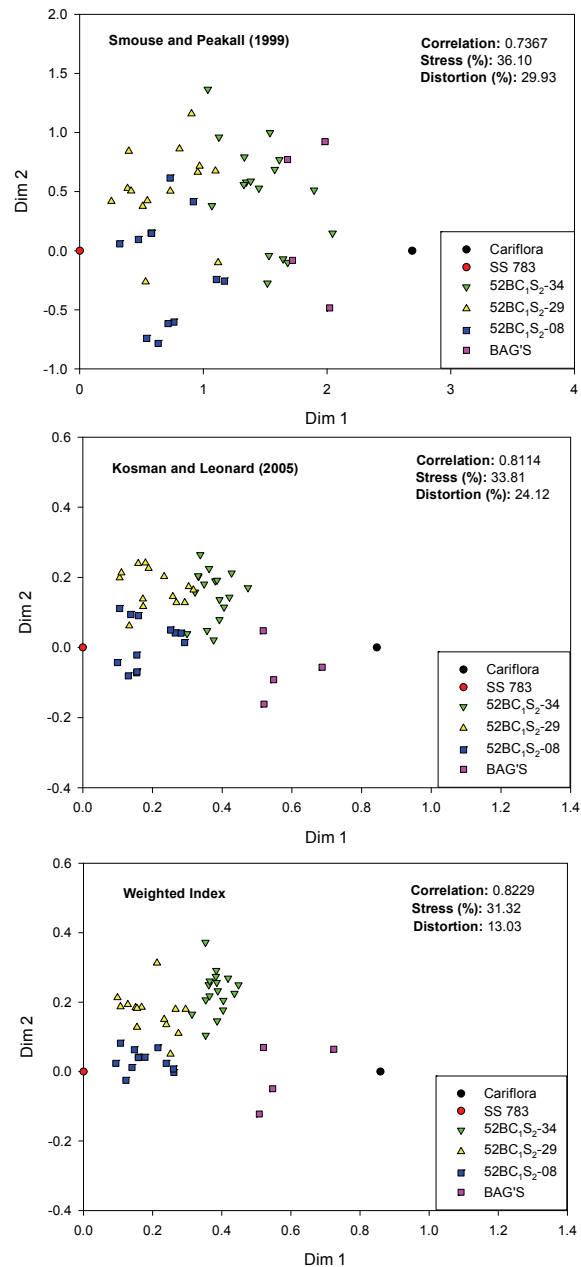
**Figure 2.** UPGMA dendrogram based on the analysis of 49 papaya genotypes using the Kosman and Leonard (2005) distance. (Co-phenetic correlation = 0.83).

Through the projection of distances in the plane, based on a multidimensional scale analysis (Figure 4), it is possible to conceive the greater similarity on individuals of the S<sub>2</sub> generation with the SS783 parent, confirming the groupings based on the Kosman and Leonard (2005) coefficient and the weighted index through UPGMA grouping. These algorithms also reveal the greatest correlation estimates with projection of matrices of distances in the plane, as well as the

lowest value of stress and deviations when compared with the Smouse and Peakall (1999) index. Furthermore, these algorithms allowed the separation of individuals in the S<sub>2</sub> generation (52BC<sub>1</sub>S<sub>2</sub>-08, 52BC<sub>1</sub>S<sub>2</sub>-29 and 52BC<sub>1</sub>S<sub>2</sub>-34) as to progenitors (Cariflora and SS783) as well as the four germplasm bank accessions (Figure 4) in a different manner from the Smouse and Peakall (1999) index, which did not provide this differentiation among the evaluated genotypes (Figure 4). We can conclude, therefore, that the Kosman and Leonard (2005) coefficient and weighted index were more efficient than the Smouse and Peakall (1999) algorithm on displaying the evaluated genotypes in dendrograms and Cartesian axis representing the genetic similarity.



**Figure 3.** UPGMA dendrogram based on the analysis of 49 papaya genotypes using the Weighted index (CRUZ, 2008) (Cophenetic correlation = 0.85).



**Figure 4.** Projection of distances in the plane, considering 49 papaya genotypes using the Smouse and Peakall (1999) dissimilarity coefficient, the Kosman and Leonard (2005) dissimilarity coefficient and the Weighted Index (CRUZ, 2008).

According to Reif et al. (2005) and Laurentin (2009), selection of the appropriate similarity index is highly important for a reliable understanding of the genetic dispersion of genotypes, primarily when procedures improve yields on selective processes are desired. These studies on the Cariflora genotype sex conversion from the dioic to a ginoic-andromonoic population demonstrate that the evaluation and selection of individuals in the segregant populations is not only focused on recovering the

Cariflora, but also to provide the selection of segregant genotypes exhibiting desirable agronomic qualities aimed at the short or medium period of attainment of superior endogamic inbred lines.

### Conclusion

Kosman and Leonard (2005) coefficient and the weighted index were more efficient than the Smouse and Peakall (1999) algorithm for the disposition of the accessed genotypes in dendrograms and the Cartesian axis for genetic similarity.

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### References

- BERNARDO R. Molecular markers and selection for complex traits in plants: learning from the last 20 years. **Crop Science**, v. 48, n. 5, p. 1649-1664, 2008.
- CRUZ, C. D. **Programa GENES**: diversidade genética. Viçosa: UFV, 2008.
- CRUZ, C. D.; VIANA, J. M. S. A methodology of genetic divergence analysis based on sample unit projection on two-dimensional space. **Revista Brasileira de Genética**, v. 17, n. 1, p. 69-73, 1994.
- DAHER, R. F.; PEREIRA, M. G.; TUPINAMBA, E. A.; AMARAL JÚNIOR, A. T.; ARAGÃO, W. N.; RIBEIRO, F. E.; OLIVEIRA, L. O.; SAKIYAMA, N. S. Assessment of coconut tree genetic divergence by compound sample RAPD marker analysis. **Crop Breeding and Applied Biotechnology**, v. 2, n. 3, p. 431-438, 2002.
- DOYLE, J. J.; DOYLE, J. L. Isolation of plant DNA from fresh tissue. **Focus**, v. 12, n. 27, p. 13-15, 1990.
- EATHINGTON, S. R.; CROSBIE, T. M.; EDWARDS, M. D.; REITER, R. S.; BULL, J. K. Molecular markers in a commercial breeding program. **Crop Science**, v. 47, n. 3, p. 154-163, 2007.
- GONÇALVES, L. S. A.; RODRIGUES, R.; AMARAL JÚNIOR, A. T.; KARASAWA, M.; SUDRÉ, C. P. Comparison of multivariate statistical algorithms to cluster tomato heirloom accessions. **Genetics and Molecular Research**, v. 7, n. 4, p. 1289-1297, 2008.
- GONÇALVES, L. S. A.; RODRIGUES, R.; AMARAL JÚNIOR, A. T.; KARASAWA, M.; SUDRÉ, C. P. Heirloom tomato gene bank: assessing genetic divergence based on morphological, agronomic and molecular data using a Ward-modified location model. **Genetics and Molecular Research**, v. 8, n. 1, p. 364-374, 2009.
- JENA, K. K.; MACKILL, D. J. Molecular markers and their use in marker-assisted selection in rice. **Crop Science**, v. 48, n. 4, p. 1266-1276, 2008.
- KOSMAN, E.; LEONARD, K. J. Similarity coefficients for molecular markers in studies of genetic relationships between individuals for haploid, diploid, and polyploidy species. **Molecular Ecology**, v. 14, n. 2, p. 415-424, 2005.
- LAURENTIN, H. Data analysis for molecular characterization of plant genetic resources. **Genetic Resource and Crop Evolution**, v. 56, n. 2, p. 277-292, 2009.
- LEAL, A. A.; MANGOLIN, C. A.; AMARAL JÚNIOR, A. T.; GONÇALVES, L. S. A.; SCAPIM, C. A.; MOTT, A. S.; ELOI, I. B. O.; CORDOVÉS, V.; SILVA, M. F. P. Efficiency of RAPD versus SSR markers for determining genetic diversity among popcorn lines. **Genetics and Molecular Research**, v. 9, n. 1, p. 9-18, 2010.
- OLIVEIRA, E. C.; AMARAL JÚNIOR, A. T.; GONÇALVES, L. S. A.; PENA, G. F.; FREITAS JÚNIOR, S. P.; RIBEIRO, R. M.; PEREIRA, M. G. Optimizing the efficiency of the touchdown technique for detecting inter-simple sequence repeat markers in corn (*Zea mays*). **Genetics and Molecular Research**, v. 9, n. 2, p. 835-842, 2010.
- PEAKALL, R.; SMOUSE, P. E.; HUFF, D. R. Evolutionary implications of allozyme and RAPD variation in diploid populations of Buffalograss (*Buchloë dactyloides* (Nutt.) Engelm.). **Molecular Ecology**, v. 4, n. 2, p. 135-147, 1995.
- PEAKALL, R.; SMOUSE, P. E. GENALEX 6: genetic analysis in Excel. Population genetic software for teaching and research. **Molecular Ecology Notes**, v. 6, n. 1, p. 288-295, 2006.
- PÉREZ, O. J.; DAMBIER, D.; OLLITRAULT, P.; COPPENS, D. G.; BROTTIER, P.; FROELICHER, Y.; RISTERUCCI, A. M. **Sequências de clones com região microssatélite**. Available from: <<http://www.ncbi.nlm.nih.gov>>. Access on: 20 Jan. 2006.
- REIF, J. C.; MELCHINGER, A. E.; FRISCH, M. Genetical and mathematical properties of similarity and dissimilarity coefficients applied in plant breeding and seed bank management. **Crop Science**, v. 45, n. 1, p. 1-7, 2005.
- SANTOS, S. C.; RUGGIERO, C.; SILVA, C. L. S. P.; LEMOS, G. M. A microsatellite library for *Carica papaya* L. cv Sunrise Solo. **Revista Brasileira de Fruticultura**, v. 25, n. 2, p. 263-267, 2003.
- SCHLÖTTERER, C. The evolution of molecular markers – just a matter of fashion? **Nature Reviews**, n. 5, v. 1, p. 63-69, 2004.
- SCHULMAN, A. H. Molecular markers to assess genetic diversity. **Euphytica**, v. 158, n. 3, p. 313-321, 2007.
- SMOUSE, P. E.; PEAKALL, R. Spatial autocorrelation analysis of individual multiallele and multilocus genetic structure. **Heredity**, v. 82, n. 5, p. 561-573, 1999.
- SQUIRRELL, J.; HOLLINGSWORTH, P. M.; WOODHEAD, M.; RUSSEL, J.; LOWE, A. J.; GIBBY, M.; POWELL, W. How much effort is required to isolate nuclear microsatellites from plants? **Molecular Ecology**, v. 12, n. 6, p. 1339-1348, 2003.

VARSHNEY, R. K.; GRANER, A.; SORRELL, M. E. Genic microsatellite markers in plants: features and applications. **Trends in Biotechnology**, v. 23, n. 1, p. 48-55, 2005.

ZANE, L.; BARGELLONI, L.; PATARNELLO, T. Strategies for microsatellite isolation: a review. **Molecular Ecology**, v. 11, n. 1, p. 1-16, 2002.

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