



Estimation of genetic distance among genotypes of caraway (*Carum carvi* L.) using RAPD-PCR

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ABSTRACT. In order to estimate genetic diversity among starting materials and breeding strains of caraway, a collection of 17 accessions from botanical gardens in Europe, two cultivars 'Rekord' and 'Kończewicki', and four own breeding strains were analyzed by RAPD-PCR. The representative samples, each of five individual plants of accession, cultivar or strain, were taken from young rosette leaves. Forty of Genset Oligos RAPD primers were used for analysis and eight of them produced clear and reproducible banding patterns. In total, 62 banding patterns were obtained revealing 23 polymorphic bands, whereas the number of polymorphic bands ranged from two to four for one primer. The GS12 and GS43 primers generated two polymorphic bands, while each of the GS8, GS21, GS22, GS41 and GS53 primers generated three bands. The GS53 primer was the most informative one, revealing 60% of the estimated polymorphism. The estimated value of genetic distance ranged from 0.22 to 0.67. The lowest genetic distance was found between accessions from Cluj and Lusanne (0.22). The highest genetic distance was estimated between accession from Berlin and the strain no. 6 of cultivar 'Kończewicki' (0.67). UPGMA cluster analysis, based on eight RAPD primers, categorized the analyzed genotypes into four groups.

Keywords: *Carum carvi* L., genetic distance, genotype, RAPD.

Determinação da distância genética entre os genótipos de cominho (*Carum carvi* L.) por RAPD-PCR

RESUMO. Dezessete acessos oriundos de hortos botânicos na Europa, dois cultivares Rekord e Kończewicki, e quatro cepas foram analisados por RAPD-PCR para determinar a diversidade genética entre os materiais incipientes e as cepas do cominho. As amostras com cinco plantas individuais de acessos, cultivares ou cepas derivaram-se de jovens folhas em roseta. Quarenta Genset Oligos RAPD primers foram usados para análise, dos quais oito produziram padrões claros e reproduzíveis de banda. Sessenta e dois padrões de bandas foram obtidos, revelando 23 bandas polimórficas, enquanto o número de bandas polimórficas chegou entre dois a quatro para um primer. Os primers GS12 e GS43 produziram duas bandas polimórficas, enquanto cada um dos primers GS8, GS21, GS22, GS41 and GS53 gerou três bandas. O primer mais informativo foi GS53, revelando 60% do polimorfismo estimado. A distância genética estava entre 0,22 e 0,67. A menor distância genética encontrou-se entre os acessos oriundos de Cluj e Lusanne (0,22) e a maior distância genética foi estimada entre o acesso oriundo de Berlin e a cepa 6 do cultivar 'Kończewicki' (0,67). A análise de agrupamento UPGMA, baseada em oito RAPD primers, colocou os genótipos analisados em quatro grupos.

Palavras-chave: *Carum carvi* L., distância genética, genótipo, RAPD.

Introduction

Caraway (*Carum carvi* L.), a biennial herb of the family Apiaceae is widely cultivated in northern Europe, Russia, India, Canada and the United States and the world production of fruit may reach 15-20 thousand tones a year. In Poland caraway is the one of the most important cultivated medicinal plant which is cultivated on the area of 5000 ha and the fruit production is five thousand tones a year. The dried

fruits of caraway (*Carvi Fructus*) are used mainly as a spice in food industry (SEIDLER-ŁOŻYKOWSKA; BOCIANOWSKI, 2012). In medicine caraway is used for its digestive, carminative, spasmolytic and stimulant properties. Limonene and carvone are the most important compounds of essential oil which is the main active substance of caraway fruit. Caraway is also the source of essential oil for cosmetic industry. Moreover, caraway oil is used as an alternative sprouting inhibitor during potatoes or flower bulb

storage (HARTMANS et al., 1995). Moreover, caraway could be a component of animal forage to enhance their well-being (SADOWSKA; OBIDOWSKA, 1998). Development of high yielding cultivars characterized by high content of active substances and better adapt to abiotic and biotic stress is the main aim of caraway breeding programs. The detailed evaluation of starting materials with respect to their genetic diversity is necessary for identifying the sources of variation. The assessment of genetic diversity with the use of molecular markers enables to define specific markers associated with, or linked to important agronomic and phenotypic traits which could be further used for selection.

Only a few species of medicinal plants were applied for analysis with respect to determination of genetic diversity using molecular markers, i.e., marjoram (*Origanum majorana* L.), St. John's Wort (*Hypericum perforatum* L.), artichoke (*Cynara scolymus* L.), sweet basil (*Origanum basilicum* L.), chamomile (*Chamomila recutita* (L.) Rausch.) (ARNHOLDT-SCHMITT, 2002; KLÖCKE et al., 2002; MESSMER et al., 2002; WETZEL et al., 2002). Frequently, the selected molecular markers (random amplified polymorphic DNA, RAPD) are used for the detection of plant adulterants in some spice i.e. black pepper, chili or turmeric (DHANYA; SASIKUMER, 2010). Within the family of Apiaceae, genotypes of carrot (*Daucus carota* L.) lines and accessions were characterized with the use of molecular markers. Bradeen et al. (2002) analyzed 124 carrot genotypes with 140 AFLP markers revealing high genetic diversity among wild type and population varieties. Grzebelus et al. (2001) used 33 RAPD and 88 AFLP markers for evaluation of genetic diversity among 31 carrot lines and accessions, as well as heterosis effect of three F₁ hybrids, in comparison with their parental lines. Based on the polymorphism level revealed by RAPD and AFLP analyzes, dendrograms showing genetic distance among the tested lines and accessions were constructed (GRZEBELUS et al., 2002; BARAŃSKI et al., 2004). As to our knowledge no results on molecular analysis of caraway genetic diversity were published yet. Although there were done some research in caraway doubled haploid (DH) lines by Ferrie et al. (2011) who investigated agronomic performance of 25 caraway DH lines in the field trials.

Here we present our results concerning assessment of genetic diversity among starting materials and breeding strains of caraway.

Material and methods

Plant material

A collection of 24 caraway accessions (Table 1) was established at the Institute of Medicinal Plants in Poznan.

Table 1. Origin of caraway collection (2007-2008).

Objects	Source
17 accessions	European botanical gardens of Bayreuth, Berlin, Bonn, Cluj, Cracow, Gottingen, Jena, Lousanne, Nantes, Poznan, Prague, Reykiavik, Riga, Salzburg, Ulm, Warsaw, Wroclaw
Cultivar 'Rekord'	Czech Republic
Cultivar 'Kociczewicki' strain no. 6 and no. 7	The maintenance breeding of cv. 'Kociczewicki' Institute of Medicinal Plants, Poland
Strains no. 9/10, no. 9/12, no. 9/13, no. 60/8	The breeding program Institute of Medicinal Plants, Poland

In 2007 and 2008 seeds of the collected accessions were sown in a greenhouse and 5-8-leaves plantlets were planted into a field at the beginning of May. In autumn, every year young rosette leaves of five plants from each accession were taken. Then these five leaves were mixed to make the bulk samples for DNA isolation.

RAPD-PCR

Amplification was performed in a reaction volume 25 µL containing 35 ng of primer, 1.5-4.5 ng DNA, 10 mM Tris-HCL, pH 8.3, 2 mM MgCl₂, 2.5 µg BSA, 100 µM of each dNTP, and 1.5 U of Taq DNA polymerase (Fermentas). PCR reaction, with the use of forty RAPD primers (Genset Oligos), was carried out in a Applied Biosystem thermocycler according to the following protocol: at 95°C for 5 min., 45 cycles: 95°C for 1 min., 35°C for 1 min., 72°C for 2 min., and final extension at 72°C for 5 min.

Amplification products were resolved by 1.5% agarose gel electrophoresis at 100 V by 2.5h. The 100-bp DNA Ladder Plus (GeneRuler™) was used as DNA fragment length determination.

Statistical analysis

The coefficients of genetic distance (D) for all caraway collection accessions were calculated according to the formula given by Nei (1972):

$$D_{N,AB} = \frac{2N_{AB}}{N_A + N_B},$$

where:

N_A is the number of bands present in accession A,

N_B is the number of bands present in accession B,

N_{AB} is the number of bands present in accessions A and B.

Based on calculated coefficients the accessions were grouped hierarchically using the unweighted pair group method of arithmetic means (UPGMA). The relationship among the accessions was presented as a dendrogram. The reliability of the similarity tree was assessed by the bootstrap method (FELSENSTEIN, 1985) with 1000 replications. We used 95% as the statistically significant values. Bootstrap analysis was performed by using the program MEGA 3.1.

Results and discussion

DNA of caraway collection accessions was tested by 40 RAPD primers and eight of them (GS8, GS12, GS21, GS22, GS38, GS41, GS43, GS53) revealed polymorphism (Table 2).

Table 2. Amplification products of RAPD primers (2007-2008).

Primer	Sequence 5' - 3'	Number of bands	Size of DNA (bp) min-max	Number and percentage of polymorphic bands
GS 8	AAAGCTGCGG	10	400-1500	3 (30.0)
GS 12	TCGGCGATAG	7	200-1300	2 (28.6)
GS 21	GGTGACGCAA	7	300-1500	3 (42.9)
GS 22	GTCTGACGGT	9	400-2000	3 (33.3)
GS 38	CAGGGGACGA	7	200-1500	4 (57.1)
GS 41	GTGGCTTGGG	8	200-1200	3 (37.5)
GS 43	GTGGCCGATG	9	200-1300	2 (22.2)
GS 53	ACGCCAGAG	5	300-1500	3 (60.0)
total		62		23

In total, 62 of amplified DNA fragments were obtained, whereas 23 were polymorphic (37.1%). The

number of polymorphic bands revealed by one primer ranged from two (GS12, GS43) to four (GS38). The size of amplification products ranged from 200 – 1500 base pair (bp). The GS53 primer revealed the highest polymorphism (60.0%), while the smallest one the GS43 (22.2%).

The genetic distance value estimated on the basis of RAPD markers ranged from 0.22 to 0.67 (Table 3). The lowest genetic distance was found between accessions from Cluj and Lousanne (0.22). Also low genetic distance values were found for accessions from Cluj and Bayreuth (0.26), Wrocław and Jena (0.26) and Cluj and Prague (0.27). The highest genetic distance was revealed between accession from Berlin and strain no. 6 of cultivar 'Kończewicki' (0.67). High genetic distances were found between accession from Berlin and strain no. 9/13 (0.66) and between accession from Warsaw and strain no. 9/12 (0.66).

The dendrogram constructed on the basis of genetic distance sorted the tested accessions into four groups, with full statistical reliability – bootstrap value equal to 100% (Figure 1):

1. strain 9/12;
2. Cracow, cv. 'Kończewicki strain no. 6, strains 9/13, 60/8;
3. Riga, Bonn, strain 9/10;
4. Reykiavik, Berlin, Salzburg, Warsaw, cv. 'Rekord', Jena, Wrocław, Poznań, Göttingen, cv. 'Kończewicki strain no. 7, Nantes, Bayreuth, Cluj, Lousanne, Ulm, Prague.

Table 3. Genetic differentiation between 24 accessions of caraway collection (2007-2008).

Accession	Jena	9/2	Krakow	Poznan	Reykiavik	'Rekord'	Bayreuth	Riga	Salzburg	Bonn	Ulm	Prague	Wrocław	Göttingen	Warsaw	9/13	60/8	Kończewicki 6	Kończewicki 7	Berlin	Nantes	9/10	Cluj	Lousanne	
Jena	0																								
Str. 9/12	0.56	0																							
Krakow	0.40	0.54	0																						
Poznan	0.32	0.49	0.31	0																					
Reykiavik	0.56	0.45	0.48	0.44	0																				
'Rekord'	0.46	0.47	0.47	0.37	0.52	0																			
Bayreuth	0.32	0.50	0.44	0.35	0.39	0.41	0																		
Riga	0.45	0.55	0.53	0.42	0.38	0.53	0.40	0																	
Salzburg	0.37	0.53	0.49	0.42	0.48	0.46	0.41	0.46	0																
Bonn	0.36	0.57	0.49	0.42	0.57	0.56	0.37	0.32	0.39	0															
Ulm	0.39	0.46	0.37	0.34	0.35	0.43	0.43	0.45	0.40	0.45	0														
Prague	0.43	0.51	0.50	0.40	0.45	0.50	0.38	0.47	0.46	0.59	0.31	0													
Wrocław	0.26	0.47	0.49	0.29	0.45	0.58	0.35	0.46	0.37	0.36	0.32	0.46	0												
Göttingen	0.35	0.50	0.46	0.30	0.48	0.32	0.38	0.46	0.32	0.40	0.38	0.44	0.43	0											
Warsaw	0.41	0.66	0.35	0.41	0.49	0.42	0.42	0.61	0.32	0.50	0.38	0.45	0.53	0.33	0										
Str. 9/13	0.45	0.62	0.39	0.42	0.61	0.59	0.55	0.64	0.48	0.48	0.48	0.49	0.52	0.39	0.43	0									
Str. 60/8	0.41	0.56	0.42	0.47	0.47	0.45	0.42	0.62	0.52	0.41	0.56	0.58	0.54	0.42	0.40	0.40	0								
Kończewicki str. 6	0.48	0.64	0.37	0.36	0.51	0.49	0.40	0.53	0.58	0.48	0.42	0.46	0.48	0.43	0.44	0.45	0.54	0							
Kończewicki str. 7	0.48	0.37	0.43	0.42	0.42	0.37	0.35	0.56	0.46	0.48	0.34	0.37	0.40	0.38	0.44	0.48	0.51	0.42	0						
Berlin	0.34	0.59	0.57	0.55	0.53	0.57	0.50	0.47	0.38	0.43	0.46	0.44	0.50	0.44	0.45	0.64	0.59	0.67	0.50	0					
Nantes	0.39	0.49	0.48	0.31	0.38	0.45	0.45	0.45	0.42	0.54	0.36	0.33	0.45	0.37	0.49	0.51	0.56	0.38	0.33	0.40	0				
Str. 9/10	0.40	0.61	0.47	0.49	0.52	0.47	0.35	0.43	0.43	0.30	0.46	0.56	0.43	0.44	0.54	0.49	0.48	0.52	0.43	0.44	0.48	0			
Cluj	0.38	0.41	0.49	0.35	0.39	0.44	0.26	0.43	0.41	0.49	0.32	0.27	0.38	0.33	0.50	0.48	0.54	0.40	0.35	0.41	0.40	0.35	0		
Lousanne	0.36	0.38	0.44	0.33	0.43	0.39	0.29	0.47	0.39	0.44	0.36	0.33	0.44	0.34	0.45	0.52	0.43	0.44	0.41	0.42	0.32	0.36	0.22	0	

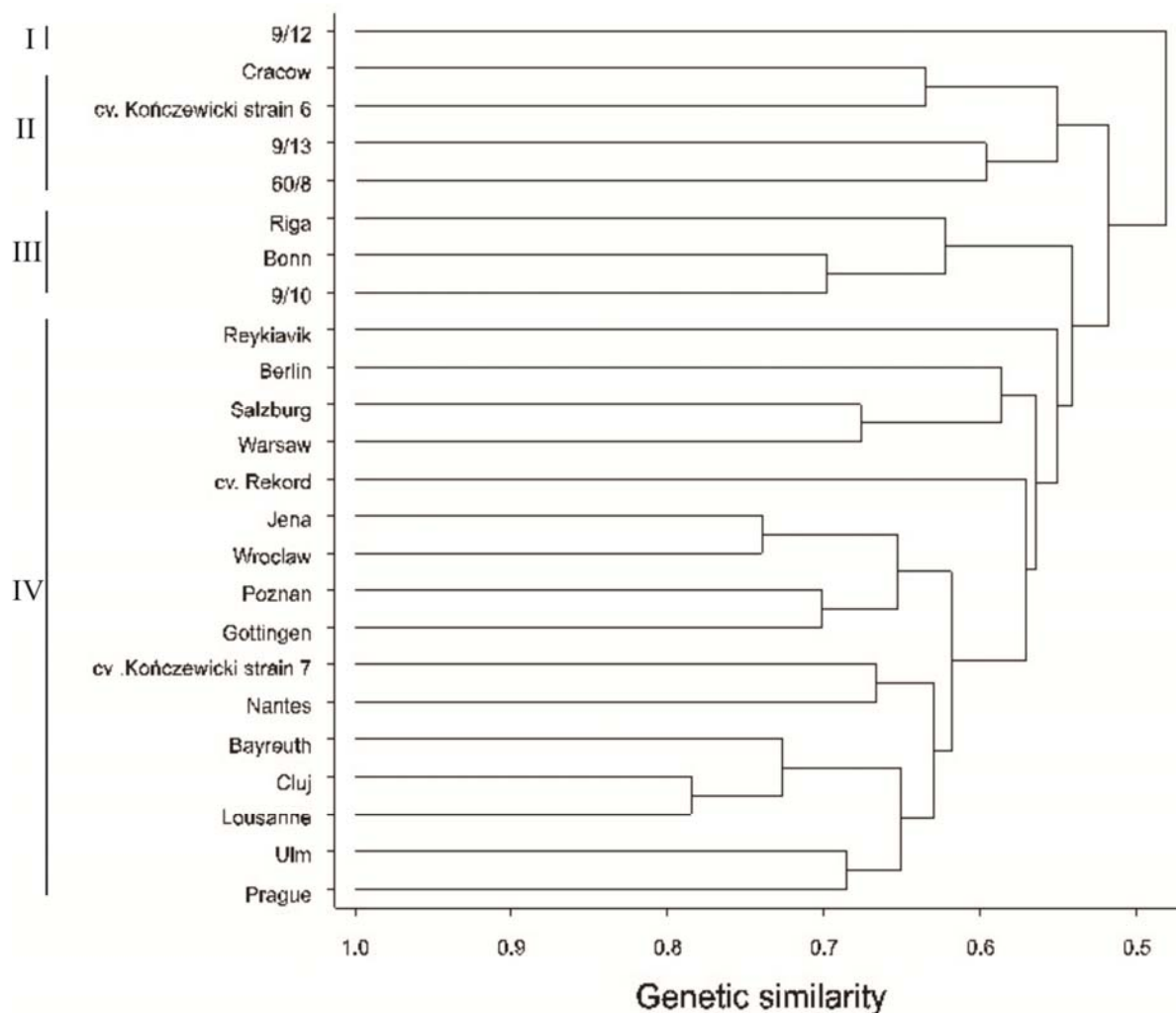


Figure 1. Cluster dendrogram of 24 caraway objects basing on eight RAPD primers (2007-2008).

RAPD can be applied for detection of genetic variation in many species and the estimated genetic distance value can be used for selection of starting materials in breeding programs (GRZEBELUS et al., 1997, 2001; BARAŃSKI et al., 2001; BOCIANOWSKI et al., 2003; NOWAKOWSKA et al., 2004; KUCZYŃSKA et al., 2007; BOCIANOWSKI et al., 2011; IRZYKOWSKA et al., 2012, 2013a and b; LIERSCH et al., 2013). In the presented research, a collection of 24 accessions, cultivars and breeding strains of caraway was analyzed using RAPD-PCR. The obtained results revealed significant differentiation of banding patterns among the collected caraway plants, thus enabling evaluation of genetic distance among them.

Up to now, no results have been published on caraway DNA analysis by RAPD-PCR, as well as by other molecular markers. Nevertheless, some medicinal plants species were analyzed by this method. Arnholdt-Schmitt (2002) determined genetic variability among St. John's wort (*Hypericum*

perforatum L.) populations, using 44 RAPD primers. The 11 of St. John's wort populations in which determined 23 polymorphic bands were used for genetic distance evaluation (from 0.1054 to 1.0986). Moreover, RAPD markers linked to hypericine and flavonols content were identified. Sixteen RAPD primers were used for determining DNA polymorphism among marjoram (*Origanum majorana* L.) accessions (KLÖCKE et al., 2002) and the detected 128 polymorphic bands were applied for evaluation of genetic distance among the analyzed accessions. Twenty of Iranian black cumin (*Bunium persicum* Boiss.) accessions were analyzed with the use of 15 RAPD and 17 AFLP primers revealing 192 polymorphic bands by RAPD. The genetic distance ranged from 0.40 to 0.82 and it did not correlate with geographical distance of places of genotype origin (PEZHMANMEHR et al., 2009). Pirkhezri et al. (2010) reported the RAPD analysis of chamomile (*Chamomilla recutita* (L.) Rausch.) populations and cultivars determining 205

polymorphic bands by the selected 18 primers. The range of similarity varied from 0.15 to 0.78 and the populations were divided into two main groups. Lately, genetic variability of 32 cumin (*Nigella sativa* L.) genotypes was detected with the use of 58 RAPD primers, generating 164 (66%) polymorphic fragments. UPGMA cluster analysis indicated seven distinct groups (IQBAL et al., 2011). All the authors mentioned above pointed out that their studies provided important information for management of germplasm resources and could be used in breeding programs for crop improvement.

Conclusion

The presented in this work evaluation of genetic variation of the caraway collection will be useful for selection of appropriate plant materials in breeding programs and will be applied for future research on designing genetic markers linked to the important quality traits, as: carvone and limonene content.

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References

- ARNHOLDT-SCHMITT, B. Characterization of *Hypericum perforatum* L. plants from various accessions by RAPD fingerprinting. **Journal of Herbs Spices Medicine Plants**, v. 9, n. 2/3, p. 163-170, 2002.
- BARAŃSKI, R.; SZKLARCZYK, M.; GRZEBELUS, D.; JAGOSZ, B. Wykorzystanie markerów molekularnych w hodowli warzyw. **Biotechnologia**, v. 1, n. 52, p. 47-50, 2001.
- BARAŃSKI, R.; GRZEBELUS, D.; KOTLIŃSKA, T.; MICHALIK, B. Wykorzystanie markerów DNA do oszacowania zmienności w polskiej kolekcji zasobów genowych marchwi (*Daucus carota* L.). **Zeszyty Problemowe Postępów Nauk Rolniczych**, v. 497, n. 1, p. 201-207, 2004.
- BOCIANOWSKI, J.; CHEŁKOWSKI, J.; KUCZYŃSKA, A.; WIŚNIEWSKA, H.; SURMA, M.; ADAMSKI, T. Assessment of RAPD markers for barley doubled haploid lines resistant and susceptible to *Fusarium culmorum* at seedling and adult plant growth stages. **Journal of Applied Genetics**, v. 44, n. 3, p. 355-360, 2003.
- BOCIANOWSKI, J.; KOZAK, M.; LIERSCH, A.; BARTKOWIAK-BRODA, I. A heuristic method of searching for interesting markers in terms of quantitative traits. **Euphytica**, v. 181, n. 1, p. 89-100, 2011.
- BRADEEN, J. M.; BACH, I. C.; BRIARD, M.; CLERC, V.; GRZEBELUS, D.; SENALIK, D. A.; SIMON, P. W. Molecular diversity analysis of cultivated carrot (*Daucus carota* L.) and wild *Daucus* populations reveals a genetically nonstructured composition. **Journal of the American Society for Horticultural Science**, v. 127, n. 3, p. 383-391, 2002.
- DHANYA, K.; SASIKUMAR, B. Molecular marker based adulteration detection in traded od and agricultural commodities of plant origin with special reference to spices. **Current Trends in Biotechnology and Pharmacy**, v. 4, n. 1, p. 454-489, 2010.
- FELSENSTEIN, J. Confidence limits on phylogenies: an approach using bootstrap. **Evolution**, v. 39, n. 4, p. 783-791, 1985.
- FERRIE, A. M. R.; BETHUME, T. D.; ARGANOSA, G. C.; WATERER, D. Field evaluation of doubled haploids plants in the Apiaceae: dill (*Anethum graveolens* L.), caraway (*Carum carvi* L.), and fennel (*Foeniculum vulgare* Mill.). **Plant Cell, Tissue and Organ Culture**, v. 104, n. 3, p. 407-413, 2011.
- GRZEBELUS, D.; SZKLARCZYK, M.; MICHALIK, B. The use of RAPD markers for genotype identification of carrot lines and F₁ hybrids. **Journal of Applied Genetics**, v. 38A, n. 1, p. 33-41, 1997.
- GRZEBELUS, D.; SZKLARCZYK, M.; BARAŃSKI, R.; JAGOSZ, B.; SIMLAT, M.; MICHALIK, B. Przykłady wykorzystania markerów molekularnych w polskiej hodowli roślin warzywnych. **Folia Horticulturae**, v. 13, n. 1A, p. 15-23, 2001.
- GRZEBELUS, D.; BARAŃSKI, R.; KOTLIŃSKA, T.; MICHALIK, B. Assessment of genetic diversity in a carrot (*Daucus carota* L.) germplasm collection. **Plant Genetic Resources Newsletter**, v. 130, p. 51-53, 2002.
- HARTMANS, K. J.; DIEPENHORST, P.; BAKKER, W.; GORRIS, L. G. M. The use of carvone in agriculture: sprout suppression of potatoes and antifungal activity against potato tuber and other plant diseases. **Industrial Crops and Products**, v. 4, n. 1, p. 3-13, 1995.
- IQBAL, M. S.; NADEEM, S.; MEHBOOB, S.; GHAFOR, A.; RAJOKA, M. I.; QURESHI, A. S.; NIAZ, B. Exploration of genotype specific fingerprinting of *Nigella sativa* L. using RAPD markers. **Turkish Journal of Agriculture and Forestry**, v. 35, n. 6, p. 569-578, 2011.
- IRZYKOWSKA, L.; BOCIANOWSKI, J.; BATURO-CIEŚNIEWSKA, A. Association of mating-type with mycelium growth rate and genetic variability of *Fusarium culmorum*. **Central European Journal of Biology**, v. 8, n. 7, p. 701-711, 2013.
- IRZYKOWSKA, L.; WERNER, M.; BOCIANOWSKI, J.; KAROLEWSKI, Z.; FRUZYŃSKA-JÓZWIĄK, D. Genetic variation of horse chestnut and red horse chestnut and trees susceptibility to *Erysiphe flexuosa* and *Cameraria ohridella*. **Biologia**, v. 68, n. 5, p. 851-860, 2013b.
- IRZYKOWSKA, L.; WEBER, Z.; BOCIANOWSKI, J. Comparison of *Claviceps purpurea* populations originated from experimental plots or fields of rye. **Central European Journal of Biology**, v. 7, n. 5, p. 839-849, 2012.

- KLÖCKE, E.; LANGBEHN, J.; GREWE, C.; PANK, F. DNA fingerprinting by RAPD on *Origanum majorana* L. **Journal of Herbs, Spices, and Medicinal Plants**, v. 9, n. 2/3, p. 171-176, 2002.
- KUCZYŃSKA, A.; SURMA, M.; KACZMAREK, Z.; ADAMSKI, T. Relationship between phenotypic and genetic diversity of parental genotypes and the frequency of transgression effects in barley (*Hordeum vulgare* L.). **Plant Breeding**, v. 126, n. 4, p. 361-368, 2007.
- LIERSCH, A.; BOCIANOWSKI, J.; KOZAK, M.; BARTKOWIAK-BRODA, I. Comparison of isozyme, RAPD and AFLP markers in genetic similarity assessment of CMS *ogura* F1 hybrids of winter oilseed rape (*Brassica napus* L.) parental lines. **Acta Biologica Cracoviensia. Series: Botanica**, v. 55, n. 1, p. 49-57, 2013.
- MESSMER, M.; SCHEIDER, E.; STEKLY, G.; BUTER, B. Determination of the progenitors and the genetic stability of artichoke cultivar 'Saluschocke' using molecular markers. **Journal of Herbs, Spices, and Medicinal Plants**, v. 9, n. 2/3, p. 177-182, 2002.
- NEI, M. Genetic distance between populations. **American Naturalist**, v. 106, n. 949, p. 283-292, 1972.
- NOWAKOWSKA, J.; MIKOŁAJCZYK, K.; KRÓTKA, K.; BARTKOWIAK-BRODA, I. Ocena dystansu genetycznego linii rodzicielskich mieszańców F₁ rzepaku ozimego (*Brassica napus* L.) za pomocą metody RAPD. **Rośliny Oleiste**, v. 25, n. 2, p. 353-370, 2004.
- PEZHMANMEHR, M.; HASSANI, M. E.; JAHANSOOZ, F.; NAJAFI, A. A.; SEFIDKON, F.; MARDI, M.; PIRSEIEDI, M. Assessment of genetic diversity in some Iranian populations of *Bunium persicum* using RAPD and AFLP markers. **Iranian Journal of Biotechnology**, v. 7, n. 2, p. 93-100, 2009.
- PIRKHEZRI, M.; HASSANI, M. E.; HADIAN, J. Genetic diversity in different populations of *Matricaria chamomilla* L. growing in southwest Iran, based on morphological and RAPD markers. **Research Journal of Medicinal Plant**, v. 4, n. 1, p. 1-13, 2010.
- SADOWSKA, A.; OBIDOSKA, G. Pharmacological uses and toxicology of caraway. In: NÉMETH, E., (Ed.). **Caraway. The genus Carum**. London: Harwood Academic Publishers, 1998, p. 165-174.
- SEIDLER-ŁOŻYKOWSKA, K.; BOCIANOWSKI, J. Evaluation of variability of morphological traits of selected caraway (*Carum carvi* L.) genotypes. **Industrial Crops and Products**, v. 35, n. 1, p. 140-145, 2012.
- WETZEL, S. B.; KRUGER, H.; HAMMER, K.; BACHMANN, K. Investigations on morphological, biochemical and molecular variability of *Ocimum* L. species. **Journal of Herbs, Spices, and Medicinal Plants**, v. 9, n. 2/3, p. 183-188, 2002.

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