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# Biodegradation of Stored jet Fuel by a *Nocardia sp.* Isolated from Contaminated Soil

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#### **ABSTRACT**

The aim of this study was to investigate the potential of degradation of an autochthonous bacterial strain, isolated from petroleum derivatives contaminated soil samples against jet fuel hydrocarbons. The autochthonous bacterial strain was characterized as Nocardia sp. Evaluation of their degrading abilities was carried out by presumptive assays as redox indicator test and by observations of surface tension decreases in aqueous medium. Degradation of jet fuel hydrocarbons was evaluated by chromatographic methods. Experiments were performed in flasks at two biostimulation rates. A bacterial strain of Pseudomonas aeruginosa UFPEDA 39 was utilized as a reference microorganism. The bacterial strain, identified as Nocardia sp, demonstrate high ability to degrade jet fuel compounds as well as to produce surface active compounds when compared to the reference microorganism.

**Key words**: biodegradation, *Nocardia*, jet fuel

#### INTRODUCTION

Contamination of soils and aquatic environments by petroleum hydrocarbons as a consequence of accidental spillage can cause serious damages to living organisms. Since the spills implies in economical and environmental losses, authorities and petrochemical industry have been forced to reduce the risk of spillage through the modifications in technical standards and operational procedures (Milne et al, 1998; Oudot et al, 1994). Referred as a cost–effective, natural and efficient clean-up treatment of hazardous waste, bioremediation technologies have been

more intensively studied in the last three decades (Atlas, 1995a). Studies on petroleum hydrocarbon biodegradation have focused on biodegradability of crude oil, petroleum derivatives or isolated hydrocarbons.

Microbial action is directly influenced by several biotic and abiotic parameters, amongst these, special attention must be given to temperature, pH, nitrogen and phosphorous sources, as well as microelements (Atlas 1984; Atlas, 1995b; Ruberto et al, 2003).

Petroleum derivatives are a complex mixture of hydrocarbons and their biodegradability depends on their bioavailability, which is directly related to

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composition, chain length, ramification and steric and electronic effects. As a rule, n-alkanes are more degradable than branched alkanes; polycyclic aromatics hydrocarbons are less degradable than monocyclic aromatics. Jet fuel composition includes, predominantly, n-alkanes and branched-alkanes, ranging from 15 to 30 carbon atoms and few amounts of polycyclic aromatics hydrocarbons (PAH) are also observed in trace levels (Bernabei et al, 2003).

As reported in literature as hydrocarbon-degrading microorganisms, Nocardioforms actinomycetes have been frequently isolated from hydrocarbon-contaminated sites. The present work aimed at the characterization of a *Nocardia* bacterial strain as a jet fuel-degrading microorganism.

#### MATERIAL AND METHODS

# Jet fuel samples

The jet fuel samples used in this work were harvested from the tanks of the Suape Terminal, Pernambuco, Brazil, with the consent of the Brazilian Oil Company (Petrobrás SA). analysis **Preliminary** of total petroleum hydrocarbons (TPH) produced a profile of hydrocarbons varying from nC<sub>9</sub> to nC<sub>31</sub>, with predominance of hydrocarbons ranging from nC<sub>11</sub> to nC<sub>17</sub>. Polycyclic aromatic hydrocarbons were detected in trace levels.

### Microorganisms

The autochthonous bacterial strain was isolated from petroleum derivatives contaminated soil samples, by utilizing pour plate technique. Jet fuel was nebulized onto plate dishes containing Büshnell-Haas solid mineral medium (KH2PO4 1.0g; K<sub>2</sub>HPO<sub>4</sub> 1.0g; NH<sub>4</sub>NO<sub>3</sub> 1.0g; MgSO<sub>4</sub>.7H<sub>2</sub>O 0.2g; FeCl<sub>3</sub> 0.05g; CaCl<sub>2</sub>.2H<sub>2</sub>O 0.02g; Agar 15.0g and H<sub>2</sub>O 1000 mL). Colonies were transferred to the complex medium Tryptic Soy Agar - TSA<sup>TM</sup> (Oxoid). The selected culture was identified on the basis of its composition in terms of Fatty Acids profile Methyl Ester (FAME) by gas chromatography using a CG 5890 (Hewlett-Packard, series II) equipped with a flame ionization detector, employing the microbial identification system (MIDITM).

A known bacterial strain of *Pseudomonas* aeruginosa DAUFPE 39 from the Department of Antibiotics of the Federal University of

Pernambuco (UFPE), Brazil, was utilized as the alochthonous reference microorganism.

# Presumptive assays

The redox indicator 2,6 dichlorophenol indophenol (DCPIP) was utilized in aqueous phase as an oxidation indicator as indicated by Hanson et al, (1993) and adapted by Miranda et al, (2007). Cultures were allowed to grow in Büshnell-Hass mineral medium (BH) containing 0.02mL of DCPIP and 1% of jet fuel as the sole carbon source, in 250mL conical flasks at 30±1°C and 200rpm. Abiotic control was utilized in both experiments. The required time that promote the changes in colour of medium was registered.

Alochthonous and autochthonous cultures were acclimated in BH modified media by increasing jet fuel concentrations (from 1 to 15%) at two C:N ratio (95:1 coded N1 and 23:1 coded N2), at the same temperature and agitation conditions described previously. Cultures were allowed to grow by 12 days in each jet fuel concentration and, afterwards were re-inoculated in a fresh medium containing a new jet fuel concentration (different from the previous one). Initial inoculum population was ca. 10<sup>8</sup> CFU.mL<sup>-1</sup>.

# **Biodegradation Assays**

After acclimatation period, biodegradation assays were performed in 2,800mL conical flasks, containing 100 mL aliquot of inoculum from the previuous acclimated culture in the series of acclimatation experiments, corresponding approximately 17% (v/v); 380 milliliters of modified BH media, corresponding to 68% (v/v) and 80 mL of jet fuel, corresponding to 14% (v/v) of total volume. Modified BH media were adjusted at two C:N ratios: 95:1 and 23:1, by adding ammonium nitrate to the media. approximated carbon mass was calculated considering the range of hydrocarbons in the jet fuel. The majority of hydrocarbons in the jet fuel samples were straight-chain (≅92.15%) and branched-alkanes ( $\cong 7.8\%$ ). Since the general formula of these compounds is  $C_nH_{2n+2}$  the pondered average of carbon in the samples was 0.8495, and the approximated carbon mass was given as:  $mC = \rho vf$ , were, mC was the approximated carbon mass;  $\rho$  was the jet fuel density; V was the jet fuel volume and f was the factor considered as the average of all Ccompounds ranges proportion (0.8495).

Ammonium nitrate mass was adjusted to compose the desired C:N ratio as follows:

$$mNH_4NO_3 = \frac{\left(mC/mCd\right) \times 80{,}05}{28}$$
, where,  $mNH_4NO_3$ 

was the ammonium nitrate utilized mass to compose the C:N ratio; mC was the approximated carbon mass; mCd was the carbon desired proportion to compose the C:N ratio. The calculated NH<sub>4</sub>(NO<sub>3</sub>) mass to 95:1 C:N ratio, was 1.71g, and to the 23:1 C:N ratio, was 6.99g.

During biodegradation experiments, pH, surface tension and microbial growth were measured at five-day intervals. Surface tension was measured in an analogical tensiometer (DüNoy<sup>TM</sup>), as recommended by ASTM (1999); pH was measured in a digital pH-meter and microbial growth was quantified by counting Colonies Forming Unities (CFU) in plates containing TSA<sup>TM</sup> medium. Samples of 1 mL were withdrawn from biodegradation conical flasks and serially diluted from 10<sup>-1</sup> to 10<sup>-6</sup>. Thereafter, 0.1 mL of an appropriate dilution was spread onto the medium surface (spread plate technique) and incubated by 24 h. Biodegradation assays, microbial growth and all analytical measurements were made in triplicate.

Reduction on jet fuel (most significantly hydrocarbons) was estimated by means of comparing the peaks of abiotic control to those of the treated samples, and results were reported in terms of percentage of reduction of hydrocarbons. Samples were centrifuged at 10,000rpm at 5°C, followed by heptane extraction. Chromatographic and mass-spectrometric analyses were performed by Gas Chromatograph coupled to the mass (Shimadzu<sup>TM</sup>, GC-MS spectrometer 17A/QP5050) utilizing a 30m x 0.35μm x 0.25μm chromatographic column DB-5 (5% diphenyl and 95% dimethylpolysiloxane). Oven temperature was programmed from 60 to 280°C at 10°C.min<sup>-1</sup>, and held for 35min. Helium was utilized as the carrier gas. The Thermal Conductivity Detector's (TCD) temperature was adjusted to 280°C. Areas of the most significantly peaks of chromatograms were automatically calculated and peaks were integrated considering the height. Individual compounds were identified by mass spectrometry utilizing electron impact ionization system (EI).

### **RESULTS AND DISCUSSION**

#### **Selected culture**

High pH and surface tension reduction were observed to the autochthonous culture of *Nocardia sp*, when compared to those reported by Neufeld et al (1980), who have worked with other species of actinomycetes.

As reported by Ron and Rosemberg (2001), biosurfactants allowed the uptake and utilization of hydrocarbons, leading to an increase in microbial growth and a decreasing in surface tension and pH in aqueous media. Surface tension and pH are indirect parameters to investigate the biodegradability of hydrocarbons. Reduction in surface tension and pH observed to the cultures indicated the presence of biosurfactants and production of acid metabolites as an evidence of biodegradation occurrence. Evaluations of these parameters associated to the DCPIP screening technique are important tools in the investigation of degrading capabilities of microorganisms.

# Biodegradation

Nine most significantly compounds were selected based on their chromatographic peaks. Mass spectrometric analysis identified the straight-chain alkanes: n-decane, n-undecane, n-dodecane, ntridecane. and the branched alkanes: metildecane, 3-etil 2,7-dimetiloctane, 6metildodecane, 2,6,10,14-tetrametilheptadecane, 2,6,11-trimetildodecane (Table 1).

Alochthonous culture eliminated the straight-chain alkane n-decane and the branched-alkanes 4metildecane and 3-etil 2.7-dimetiloctane, after 60 days in the 23:1 C:N ratio (Table 1). Several reports have showed that straight-chain alkanes have often been found to be readily degraded in field and laboratory studies, by a large diversity of bacteria (De Jong et al, 1997). The most evidenced mechanism to describe the straight-chain alkanes biodegradation is the terminal oxidation which leads to the conversion of hydrocarbons into the corresponding more stable compounds, following the sequence of oxidation of the terminal carbon, from the more reduced to the more oxidized compound, such as alcohols, aldehydes, carboxylic acids and fat acids (Alexander, 1994; Atlas 1984; Glazer and Nikaido, 1995; Seeger et al, 1997). Although it is generally true that branched alkanes are more recalcitrant than their equivalent straightchain alkanes, studies have demonstrated that many of these branched compounds were more degradable than they have previously been reported (Geerdink et al, 1996). Present results showed that this occurrence was shown by autochthonous culture.

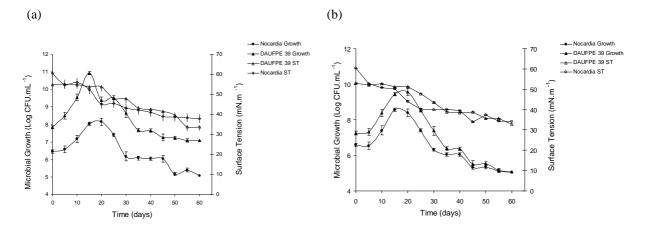
**Table 1 -** Degradation in jet fuel hydrocarbons after 60-day process by pure and mixed cultures in both 95:1 and 23:1 C: N ratio.

C:N ratio	Hydrocarbon		Degradation (%	<b>b)</b>
95:1		Sterile control	Nocardia sp	DAUFPE 39
	n-decane	0	93,80	98,60
	4-metildecane	0	92,00	98,40
	3-etil-2,7-dimetiloctane	0	88,90	99,10
	n-undecane	0	99,50	96,40
	Dodecane	0	97,00	88,60
	6-metil-dodecane	0	96,30	92,40
	2,6,10,14-tetrametilheptadecane	0	84,60	97,00
	n-tridecane	0	94,30	87,20
	2,6,11-trimetildodecane	0	80,80	88,60
23:1		Sterile control	Nocardia sp	DAUFPE 39
	n-decane	0	100	99,00
	4-metildecane	0	100	99,90
	3-etil-2,7-dimetiloctane	0	100	99,99
	n-undecane	0	99,60	99,10
	Dodecane	0	99,80	97,20
	6-metil-dodecane	0	99,99	99,40
	2,6,10,14-tetrametilheptadecane	0	99,90	98,20
	n-tridecane	0	99,80	96,30
	2,6,11-trimetildodecane	0	99,80	97,90

Branched alkanes, such as 2.6.10.14tetrametilheptadecane and 2,6,11 trimetildodecane, were the most recalcitrant compounds (Table 1). This low biodegradability could be associated to the structure of these compounds. Generally, all kinds of bacteria oxidize compounds with a lower molecular mass more easily. Amongst the hydrocarbons, aliphatic compounds are likely to be more easily degraded. Also, between branched and straight-chain alkanes, bacteria preferentially degrade straight-chain alkanes hydrocarbons. Bacteria will utilize compounds that are more difficult to degrade only if they are exposed previously to such compounds to allow the development of an adapted population. The autochthonous pure culture presented highest reduction in all the investigated compounds either with 95:1 and 23:1 C:N ratios. The 23:1 C:N ratio promoted higher biodegradation efficiency than 95:1 for both autochthonous and alochthonous cultures (Table 1). The influence of carbonnitrogen ratio (C:N) and carbon-nitrogenphosphorous ratio (C:N:P) on biodegradation has been well reported (Walworth et al 1997). Decreases in hydrocarbon concentration are expected when nitrogen supply is augmented.

The alochthonous bacterial strain *Pseudomonas aeruginosa* DAUFPE 39 demonstrated high adaptation to the carbon source, which could be confirmed by the microbial growth curve (Fig. 1a), and the percentage of remaining hydrocarbons (as summarized in Table 1). Owing to acclimatation period, using increasing concentrations of jet fuel, the lag phase was diminished (Fig. 1b). High surface tension and pH reduction were observed for autochthonous culture (50.7 and 30.4%, respectively) as showed in Table 2. These results confirmed the production of biosurfactants by the degrading bacteria. In general, bacteria growing in hydrocarbon environments produce biosurfactants

although some bacteria are not capable to produce these molecules, and others are capable to reduce the surface tension by means of cell surface hydrophobicity, acting the cell itself as a biosurfactant. This feature depends on the mechanism of assimilation of hydrocarbons utilized by each bacterial group (Alexander, 1994; Ron and Rosemberg, 2001).



**Figure 1** - Microbial growth and surface tension (ST) of *Nocardia* sp and *Pseudomonas* aeruginosa (DAUPE 39) - (a) 95:1 C:N ratio (b) 23:1 C:N ratio.

**Table 2 -** Surface tension and pH reduction percentage of aqueous media after 60 days by cultures in both 95:1 and 23:1 C:N ratio.

Culture	pH reduction (%)		Surface tension reduction (%)	
	95:1	23:1	95:1	23:1
Nocardia sp	31.4	30.4	51.7	50.7
DAUFPE 39	26.8	21.3	40.0	37.9

# **CONCLUSIONS**

The studied bacterial strain of *Nocardia* sp demonstrated high abilities to degrade the analyzed constituents of jet fuel. No significant difference were observed between the C:N ratios and further investigations must be conduced towards the optimization of this parameter. *Pseudomonas aeruginosa* DAUFPE 39 demonstrate high adaptation to the carbon source, as shown by the cell growth and reduction of the surface tension.

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# **RESUMO**

O presente estudo objetivou a investigação da capacidade degradadora de uma linhagem bacteriana autóctone (isolada de amostras de solo contaminadas com derivados de petróleo) contra hidrocarbonetos de querosene de aviação. A linhagem foi caracterizada como Nocardia sp. A avaliação do seu potencial degradador deu-se realizada mediante testes com indicador redox e observações na redução da tensão superficial na fase aquosa. A degradação do querosene foi avaliada por métodos cromatográficos. experimentos foram realizados utilizando-se duas taxas de bioestímulo. Uma linhagem bacteriana Pseudomonas aeruginosa UFPEDA utilizada como referência. A linhagem autóctone demonstrou alta eficiência na degradação de hidrocarbonetos do querosene bem como para produzir compostos ativos de superfície quando comparada com a linhagem de referência.

#### REFERENCES

- Alexander, M.: (1994), Biodegradation and Bioremediation. academic Press, 2ed.
- ASTM D 971. 99<sup>th</sup>, Standard tests Method for Interfacial Tension of Oil against Water by the Ring *in*: Method *American Society for testing and Materials*, 1999.
- Atlas, R. M., 1984. *Petroleum Microbiology*. New York. Macmillan Publishing Company.
- Atlas, R. M., (1995a), Bioremediation of Petroleum Pollutants. *International Biodeterioration and Biodegradation*. pp 317-327.
- Atlas, R. M., (1995b), *Handbook of Media for Environmental Microbiology*. London. CRC Press.
- Bernabei, M.; Reda, R.; Galiero, R.; Bocchinfuso, G., 2003. Determination of total and polycyclic aromatic hydrocarbons in aviation jet fuels. *Journal of Chromatography* A, **985**, 197-203.
- De Jonge, H., Freijer, J.I., Verstraten, J.M., Westerveld, J., Van Der Wielen, F.W.M., (1997), Relation between bioavailability and fuel oil hydrocarbon composition in contaminated soils. *Environmental Science and Technology*, **31**, 771±775.
- Geerdink, M.J., Kleijntjens, R.H., van Loosdrecht, M.C.M., Luyben, K.C.A.M., (1996), Microbial decontamination of polluted soil in aslurry process. *Journal of Environmental Engineering*, **11**, 975±982.
- Glazer, A. N.; Nikaido, H., (1995), *Microbial Biotechnology Fundamentals of Applied Microbiology*. New York. W. H. Freeman and Company Editors.
- Hanson, K. G.; Desai, D.; Desai, A. J., (1993), A rapid and simple screening technique for potential crude oil degrading microorganisms. *Biotechnology Techniques* 7, 745-748.

- Neufeld, R. J.; Zajic, J. E.; Gerson, D. F., (1980), Cell surface Measurements in Hydrocarbon and Carbohydrate Fermentetions. *Applied and Environmental Microbiology.* **39**. 511-517.
- Milne, B.J.; Baheri, H.R.; Hill, G. A., (1998), Composting of a heavy oil refinery sludge. *Environmental Progress*, 17, 24-27.
- Miranda, R.C.M; Souza, C.S.; Gomes, E.B.; Lovaglio, R.B.; Lopes, C.E.; Sousa, M. F. V. Q., (2007), Biodegradation of Diesel Oil by Yeasts Isolated from the Vicinity of Suape Port in the State of Pernambuco Brazil. *Brazilian Archives of Biology and Technology*. **50** (1), 147-152.
- Oudot, J., (1994), Soil pollution by petroleum products and treatments. *Analysis Magazine*, **22**, 16 18.
- Ron, E. Z.; Rosenberg, E., (2001), Natural roles of biosurfactants. *Environmental Microbiology* 3, 229-236
- Ruberto, L.; Vasquez, S., C.; Cormack, W. P. M. (2003), Effectiveness of the Natural Bacterial flora, Bioestimulation and Bioaugmentation on the Bioremediation of Hydrocarbon Contaminated Antarctic Soil. *Internat. Biodeteriorat. and Biodegradat.*, **52**, 115–125
- Seeger, M.; Timmis, K., N.; Hofer, B., (1997), Bacterial Pathways for the Degradation of Polychlorinated Biphenyls. *Marine chemistry* **58**, 327-33.
- Walworth, J. L.; Woolard, C. R.; Braddock, J. F.; Reynolds, C. M., (1997), The Role of Soil Nitrogen Concentration in Bioremediation. Fourth International *in situ* and On-site Bioremediation Symposium, New Orleans, April 28 May 1, Battelle. *Bioremediation*, **9**, 283-288

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