

Dideoxy Single Allele-Specific PCR - DSASP New Method to Discrimination Allelic

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ABSTRACT

Gastric cancer (GC) is a multifactorial disease with a high mortality rate in Brazil and worldwide. This work aimed to evaluate single nucleotide polymorphisms (SNP) rs1695, in the Glutathione S-Transferase Pi (GSTP1) gene in GC samples by comparative analysis Specific PCR - ASP and Dideoxy Single Allele-Specific PCR - DSASP methods. The DSASP is the proposed new method for allelic discrimination. This work analyzed 60 GC samples, 26 diffuse and 34 intestinal types. The SNP rs1695 of the GSTP1 gene was significantly associated with GC analyzed by DSASP method ($\chi^2 = 9.7$, $P < 0.05$). A comparative analysis of the data obtained from both methods did not differ significantly ($\chi^2 = 0.08$, $P > 0.05$). These results suggest that the SNP rs1695 of the GSTP1 gene was a risk factor associated with gastric carcinogens is and the DSASP method was a new successfully low-cost strategy to study allelic discrimination.

Key words: DSASP method, genotyping, Gastric Cancer

INTRODUCTION

The gastric cancer (GC) is a multifactorial disease involving genetic, epigenetic and environmental factors, including diet, chronic atrophic gastritis, radiation exposure and infection by *Helicobacter pylori*. It is the fourth most common type of cancer and the second most common cancer related cause of death in the world population (Khan et al. 2005; Crew and Neugut 2006; Jemal et al. 2011).

With the completion of the Human Genome Project, a large number of subtle variations (polymorphisms) among the population have been found (Sachidanandam et al. 2001; Collins et al. 2003). The most abundant type of these variations is single nucleotide polymorphisms (SNPs), with more than nine million reported in public databases. The SNPs are a single nucleotide variation at a specific location in the genome that is by definition found in more than 1% of the population (Brookes 1999; Rocha et al. 2006).

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SNPs have been described as responsible for genetic predisposition for several complex diseases (Nunokawa et al. 2010; Giusti et al. 2010; Heinzen et al. 2010). There are studies on establishing the association of certain SNPs with different behaviors of cancer cells in various types of tumors in an attempt to determine molecular markers (Gao et al. 2014; Lui et al. 2014; Tengström et al. 2014).

SNPs in the genes that encode enzymes involved in the biotransformation of carcinogens have also been linked to GC development. The genes of the family of glutathione S-transferase (GST) seem relevant to cancer susceptibility (Miller et al. 2002).

The Glutathione S-Transferase Pi (*GSTP1*) gene, mapping of chromosomal region 11q13, and encoding the isoenzyme P1 participates in the metabolism of halogenated compounds, molecules of low molecular weight reactive epoxides of some with polycyclic aromatic hydrocarbons (PAHs) (Islami et al. 2012).

Due to the importance of the SNPs analysis, methods for analyzing allelic discrimination, in particular methods based on the primer extension, have been developed, which include Allele-Specific PCR (ASP) method involving PCR amplification of genomic DNA using allele-specific primers that are labeled differently and a common reverse primer (Gibbs et al. 1989).

Single Base Extension (SBE) method, consisting in designing the primers before the polymorphic position of interest and an embedded ddNTP specific such as, Mass spectrometry-based detection, MassEXTENDtm (Haff and Smirnov 1997; Ross et al. 1998), and fluorescence detection, which uses a primer that anneals one base upstream of the SNP site followed by its extension with ddNTPs that are labeled with different fluorescent tags. Products are detected by fluorescence after capillary electrophoresis and the color of dye indicates incorporated base(s) and hence the SNP genotype (Sokolov 1990; Le et al. 2002).

The DSASP method, which is based on the work of Sanger et al. (1977), is a new strategy for the analysis of allelic discrimination. DSASP method is based upon incorporation of ddNTP predicted in specific positions relative to the SNP of interest. In this study, the SNP rs1695 of the *GSTP1* gene in GC samples previously analyzed by ASP method (Araújo et al. 2014) was analyzed by DSASP method.

The DSASP method has some peculiarity, as in PCR only one strand of DNA is amplified, since only one primer is used. Also in the PCR, three different deoxynucleotides and one specific dideoxynucleotide are added. After amplification, the PCR product is hybridized to its complementary sequence previously defined and synthesized.

The design of primer in the DSASP method takes into account the position of the SNP of interest in relation to the preceding nucleotides and the position of incorporation of the specific dideoxynucleotides before and after the SNP of interest. The PCR amplification product is hybridized to its complementary sequence to perform allelic discrimination by amplification of fragments of different sizes or different melting temperatures (T_m), each fragment or T_m corresponds to its specific allele.

MATERIAL AND METHODS

Samples

The present study analyzed 60 samples of GC from the samples in the Laboratory of Molecular and Structural Biology Oncogenetics (LBMEO). The samples were composed of 26 cases of diffuse-types GC and 34 intestinal types according to the classification of Laurén (1965) and were obtained with the patients' consent. The study was previously approved by the University Hospital João of Barros Barreto and Hospital of Clinics of Faculty of Medicine of Ribeirão Preto ethics committee. All tissues were frozen in liquid nitrogen immediately after tumor resection and stored at -80°C . Genomic DNA isolated by proteinase K digestion and phenol-chloroform extraction methods and stored at -20°C according to the method described by Rey et al. (1992).

Method Dideoxy Single Allele-Specific PCR - DSASP

To detect the SNP rs1695 (A>G) of the *GSTP1* gene DSASP method involving specific primer with specific characteristics, and sequence complementary to the hybridization step. These sequences were obtained by *in silico* validation (Software GeneRunner).

The primer and the complementary sequence for DSASP genotyping were designed following the manufacturers' recommendations. For genotyping the SNP rs1695 (A>G) of the *GSTP1* gene by

DSASP, the primer designed considered the position of incorporation of the specific dideoxynucleotide (ddGTP) upstream seven nucleotides and downstream 23 nucleotides in relation to *SNP* of interest. Only one primer was used in the PCR and a set of nucleotides comprising ddGTP and dATP, dCTP, and dTTP. The complementary sequence corresponded to 45 nucleotides used in the hybridization step. Then,

the PCR amplification product was hybridized with its complementary sequence. As a result, the G allele corresponded to the amplification of a 38-nucleotides fragment and T_m : 52°C and A allele corresponded to the amplification of a 45-nucleotides fragment and T_m : 65°C. Therefore, the alleles were discriminated specifically by presenting different T_m , which could melting curve analysis by qPCR (Fig.1).

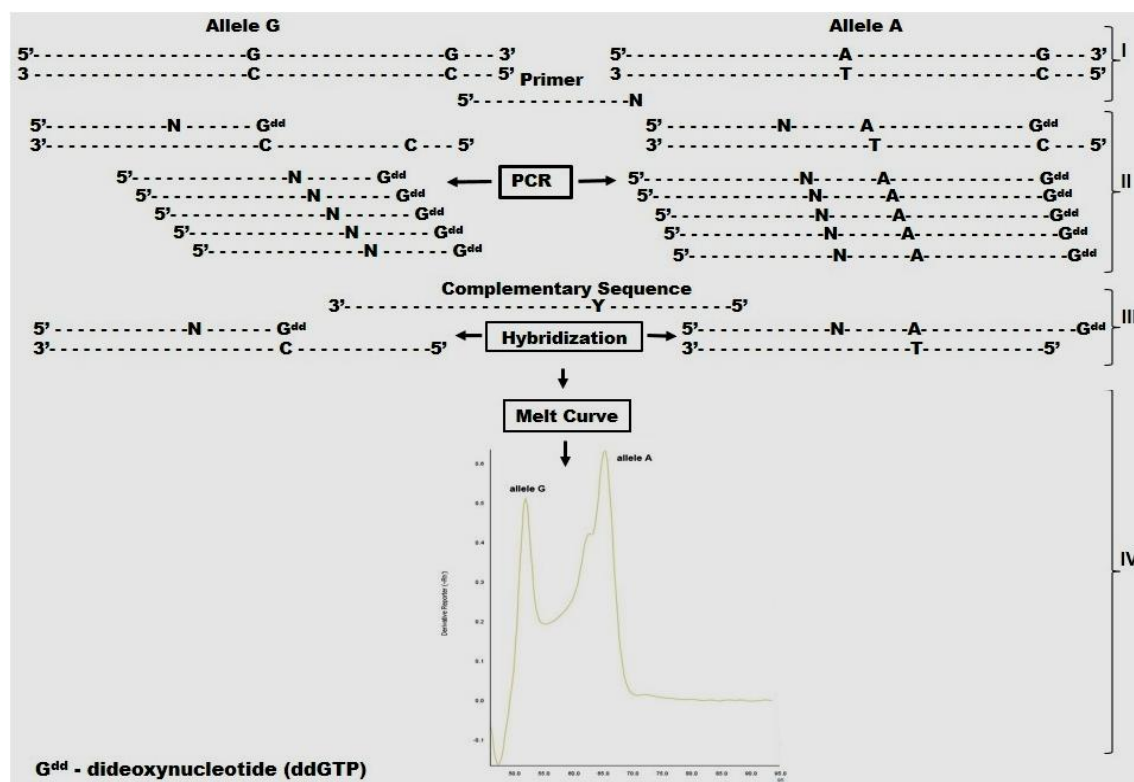


Figure 1 - The stages of DSASP method for analysis of *SNP* rs1695 in *GSTP1* gene: I - the *SNP* selection and design of the primer; II - PCR with one primer and dideoxynucleotide of interest; III - hybridization reaction with the specific complementary sequence and IV - melting curve analysis by qPCR.

Validation in silico

The primers used for DSASP of the *SNP* rs1695 (A>G) of the *GSTP1* gene were designed based on the database Ensembl Genome Browser and using

the Gene Runner program (Table 1) to evaluate the annealing temperature, the formation of secondary structure and size of the amplified fragments.

Table 1 - Primer and complementary sequence for genotyping of *SNP* rs1695 of the *GSTP1* gene by DSASP method.

SNP	Primer / Complementary Sequence – CS
rs1695	F1: 5'-GGTGAATGACGGCCTGGAGGACCTCCGCTG-3' CS: 5'-AGGGAGAYGTATTTGCAGCGGAGGTCTCCAYGCCGTCATTACC-3'

PCR Conditions

PCR was performed in a final volume of 25 μ L containing 200 μ M dNTP (dATP, dCTP, dTTP

and ddGTP), 2.0 mM $MgCl_2$, 20 ng/ μ L DNA, 200 μ M primer and 0.5 U AmpliTaq Gold (Life Technologies - Carlsbad, CA). PCR conditions for

amplification of single stranded DNA were as follows: a pre-denaturation for 3 min at 94°C and 80 cycles of 94°C for 20 s of 50°C for 45 s and 72°C for 30 s with a final extension of 5 min at 72°C.

Hybridization conditions

The product of the PCR amplification of each sample was subjected to hybridization step under the following conditions: 200 pM of the complementary sequence at 4°C for 10 min.

Melting Curve Analysis

The melting curve were analysed to determine the T_m performed by Real Time PCR equipment 7500 Fast Real-Time PCR System (Life Technologies - Carlsbad, CA) following the conditions: preheat starting at 25°C to 95°C for 1 min, folding up to 45°C for 5 min and gradual heating (1°C min) until a temperature of 95°C for 5 min. For melting curve analysis, SYBR® Green Mix (Life Technologies - Carlsbad, CA) was used.

Statistical Analysis

To evaluate the prognostic and predictive values of the *SNP* rs1695 of the *GSTP1* gene in gastric cancer samples obtained by DSASP method analyzes of Hardy-Weinberg and Chi-square test were carried out using the statistical program R. The results were considered statistically significant $P \leq 0.05$.

The results obtained by DSASP method were validated by comparison with results obtained by Allele Specific PCR - ASP (Araújo et al. 2014) for the same *SNP* and the same sample using the Chi-square test using the R statistical program considered non-significant $P > 0.05$.

RESULTS AND DISCUSSION

The allelic frequencies the *SNP* rs1695 of in *GSTP1* gene obtained by the DSASP were 0.74 and 0.26 for the A and G alleles, respectively, with the following genotype distribution: 46.7% (n = 28) A/A (Ile/Ile), 53.3% (n = 32) A/G (Ile/Val) and 0% (n = 0) G/G (Val/Val) (Table 2) with $\chi^2 = 9.70$. Second statistical analysis shows this sample is not in Hardy-Weinberg equilibrium ($P < 0.05$). Samples showed only genotypes A/A and A/G. A melting temperature of 65°C for A allele (Fig.2) and 52°C for G allele (Fig.3) was observed.

Table 2 - Results of the allelic frequencies and genotype distribution of *SNP* rs1695 of the *GSTP1* gene obtained by DSASP method.

SNP	Genotypes		Allele frequency		χ^2	P-value
	A/A	A/G	A	G		
rs1695	28 (46.7%)	32 (53.3%)	0.74	0.26	9.7	0.01
	G/G					
	0 (0.0%)					

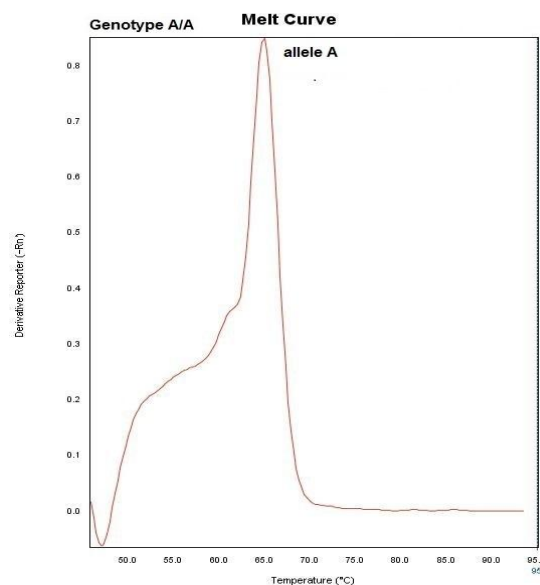


Figure 2 - Genotype A/A of *SNP* rs1695 of the *GSTP1* gene determined by DSASP method by melting curve analysis with T_m 65°C for A allele.

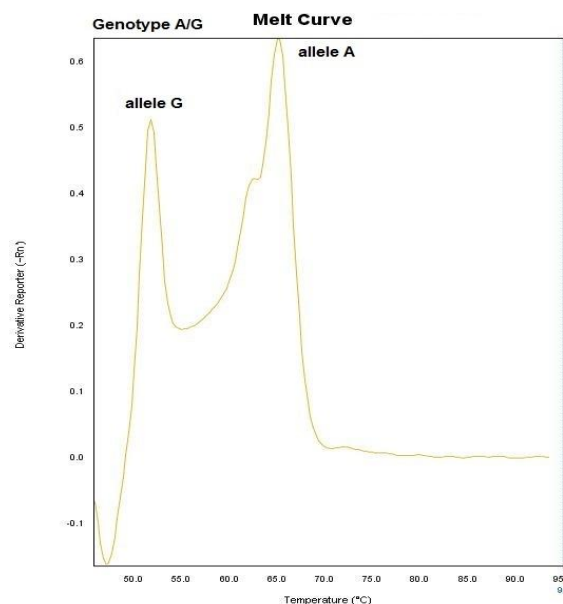


Figure 3 - Genotype A/G of *SNP* rs1695 of the *GSTP1* gene determined by DSASP method by melting curve analysis with T_m 52°C for G allele and T_m 65°C for A allele.

The data obtained by Araújo et al. (2014) by ASP method for allelic frequencies of the *SNP* rs1695 of in *GSTP1* gene were 0.73 and 0.27 for the A and G alleles, respectively, with the following genotype distribution: 45% (n = 27) A/A (Ile/Ile), 55% (n = 33) A/G (Ile/Val) and 0% (n = 0) G/G (Val/Val) (Table 3).

The *SNP* rs1695 of in *GSTP1* gene were significantly associated with GC in the samples analyzed ($\chi^2 = 8.73$, $P < 0.05$).

Table 3 - Results of the allelic frequencies and genotype distribution of *SNP* rs1695 of the *GSTP1* gene obtained by ASP method by.

SNP	Genotypes		Allele frequency		χ^2	P-value
	A/A	A/G	A	G		
rs1695	27 (45%)	33(55%)	0.73	0.27	8.73	0,01
	G/G	0 (0.0%)				

The comparative analysis between the data obtained by Araújo et al. (2014) by the ASP method with data obtained by the DSASP method did not significantly differ ($\chi^2 = 0.08$, $P > 0.05$), as expected.

Carcinogenesis may be due to the defects in the mechanisms associated with the protection of cellular damage caused by xenobiotics. Among these mechanisms, there is population variability in the frequency of polymorphisms in the genes for detoxification / metabolism. The *GSTP1* gene encodes an enzyme that acts as a carcinogen detoxication and has been described as a "guardian" gene because it actively protects the cell from oxidative damage mediated by carcinogenic components (Suvakov et al. 2013).

Dunna et al. (2012) studied the SPN rs1695 in 290 patients with acute leukemia, 147 with acute lymphocytic leukemia (ALL) and 143 with acute myeloid leukemia (AML) and 248 healthy people. The observed genotypes differed significantly among the three groups, with 51 and 10.9% of the genotypes A/G and G/G in LLA, in AML 44.8 and 18.9% of the genotypes A/G and G/G, while the control group showed 42.3 and 1.2% of the genotypes A/G and G/G, respectively. These results suggested that the G/G genotype was a risk for the development of ALL and AML factor was also associated with poor prognosis (Dunna et al. 2012).

Ibarrola-Villava et al. (2012) analyzed the samples from 562 Spanish patients with sporadic malignant

melanoma (MM) and 338 cancer-free control. This was the first time that the *GSTP1* rs1695 polymorphism was reported to be associated with MM. In addition, this study was one of the largest GST polymorphism studies undertaken in the Spanish population and the first time that copy number variants were scrutinized in relation to MM (Ibarrola-Villava et al. 2012).

The analysis of SNPs rs1695 and rs2853676 of *GSTP1* and Telomerase Reverse Transcriptase (*TERT*) genes respectively in 301 samples and 302 control samples Gliomas in Chinese patients showed a positive association for susceptibility to develop this neoplasm, therefore, could be considered risk factors for development gliomas (Li et al. 2012).

Gu et al. (2014) analyzed the *SNP* rs7208693 of the Myeloperoxidase - *MPO* gene and haplotype SNPs rs1695, rs4891, rs762803 and rs749174 of the *GSTP1* gene in 266 samples of lung câncer and 307 controls without personal history of the disease were recruited in this study in Chinese Han population. Haplotype SNPs rs1695, rs4891, rs762803 and rs749174 of the *GSTP1* gene exhibited an association with lung cancer susceptibility in smokers in the overall population and in the studied subgroups but not the *MPO* *SNP* rs7208693 (Gu et al. 2014).

Xu et al. (2012) studied 929 diagnostic GC patients in China and found a positive association between the SNPs rs1695 of the *GSTP1* gene and rs4880 of the *SOD2* gene to gastric carcinogenesis in this population. However, García-González et al. (2012), who studied the *SNP* rs1695 in 557 patients originating from southern Europe as GC, observed no association of this *SNP* with susceptibility to gastric cancer in this population (García-González et al. 2012).

The analysis of *SNP* rs1695 in 60 samples of GC patients from Northern Brazil by ASP method showed a positive association between the polymorphism in question and the development of intestinal-type gastric cancer, corroborating the findings of Xu et al. (2012) and Araújo et al. (2014).

The same samples analyzed by Araújo et al. (2014) using the ASP method were used to validate the DSASP method. The results obtained by the method DSASP were statistically different. Therefore, the results obtained by the method DSASP ratified the findings of Araújo et al. (2014).

CONCLUSION

The DSASP method could be a low-cost strategy for the studies on allelic discrimination, and theoretically could analyze any SNP of interest. In relation to the ASP method, it costed about half per sample. The DSASP method compared with other methods such as SBE, Fluorescence detection or MassEXTENDtm methods, appeared advantageous because it was performed with the reagentes and equipments of low cost. Therefore, DSASP method could be a new low-cost strategy to study allelic discrimination.

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