

## ***In Vitro* Culture of *Vriesea gigantea* and *Vriesea philippocoburgii*: Two Vulnerable Bromeliads Native to Southern Brazil**

Annette Droste<sup>1,2\*</sup>, Anelise Machado da Silva<sup>2</sup>, Adriana Vieira Matos<sup>2</sup> and Júlia Winck de Almeida<sup>2</sup>

<sup>1</sup>Programa de Pós-Graduação em Biologia; <sup>2</sup>Laboratório de Cultura de Tecidos Vegetais; UNISINOS; C. P. 275; 93001-970; São Leopoldo - RS - Brasil

### **ABSTRACT**

*Micropropagation studies were carried out using the seeds for establishing an in vitro culture of Vriesea gigantea and Vriesea philippocoburgii. Germination rate of V. gigantea was higher than of V. philippocoburgii. Plantlets of V. philippocoburgii gave rise to many adventitious shoots when cultivated in Knudson basal medium. In contrast, for V. gigantea, a higher salts-concentration was needed, so that the number of shoots was increased by Murashige and Skoog medium. Addition of activated charcoal and naphthaleneacetic acid in regeneration medium allowed the growth of shoots and the formation of roots, confirming the success of in vitro culture. The differences in expression of the genotypes reinforce the need of more research in order to set up the conditions that could offer the best response of the specific tissues.*

**Key words:** *In vitro* culture, conservation, propagation, Bromeliaceae, *Vriesea*

### **INTRODUCTION**

The damaging effect of human activities on natural habitats has resulted in a great number of fragile plants, including species of the Bromeliaceae family. In addition to this, plants with ornamental qualities are collected intensively and illegally, leading to altered plant communities (Fay, 1994). *Vriesea gigantea* Gaudich. and *Vriesea philippocoburgii* Wawra are two native bromeliads from the state of Rio Grande do Sul, in Southern Brazil, which belong to the category of epiphytic tank species. Like other species of the genus *Vriesea*, they produce seeds with low capacity of germination (Mekers, 1977; Mercier and Kerbauy, 1995). After flowering, only one or

two offshoots are formed and develop very slowly. Because their highly ornamental qualities, these bromeliads are collected intensively from natural habitat and the conventional methods of propagation cannot supply the marked. Thus, *V. gigantea* and *V. philippocoburgii* may be viewed as threatened species (Rio Grande do Sul, 2003). Different *in vitro* methods have been used to achieve the conservation or multiplication of specific genotypes of other bromeliads (Mekers, 1977; Hosoki and Asahira, 1980; Mercier and Kerbauy, 1992, 1994, 1995; Carneiro et al., 1999; Arrabal et al., 2002). When initiation of the culture is done through seeds obtained from a wide range of capsules, many different genotypes can be conserved *in vitro* (Fay, 1994; Pence, 1999). Otherwise, through the multiplication of seedlings

\* Author for correspondence

obtained *in vitro*, a large number of genetically homogeneous plants can be obtained.

Due to the great commercial importance and the status of conservation of *V. gigantea* and *V. philippocoburgii*, there is a special interest in *in vitro* culture as an alternative for the production of plants. For both species, tissue culture has been used as a tool to investigate physiological aspects of nitrogen nutrition and endogenous hormones (Mercier et al., 1997; Mercier and Kerbauy, 1998; Endres et al., 2002). In the reports, the medium used was Knudson (1946), based on studies with other species of the genus *Vriesea*. In the present study, we compared the preference of these bromeliads for different medium-compositions in order to evaluate the potential of tissue culture as an alternative to produce plants.

## MATERIALS AND METHODS

*Plant material and culture conditions.* Mature capsules of *V. gigantea* and *V. philippocoburgii* were surface sterilized for 30 sec in 70% ethanol and 15 min in 4% sodium hypochlorite plus 0.1% Tween-20, followed by four rinses in sterile distilled water. Seeds were extracted from capsules and placed on semi-solid media after cutting their appendices off. Two independent experiments were conducted. In Experiment I, seeds of both species were germinated on basal K medium (Knudson, 1946), testing modifications in salts and the addition of vitamins, as following: (K) K medium; (K/MS micro) K macro-nutrients with MS (Murashige and Skoog, 1962) micro-nutrients; (K/MS vit) K medium added by MS vitamins. All media were supplemented with 2% sucrose, 1.2% agar, at pH 6.4 (before autoclaving). For Experiment II, seeds of *V. gigantea* were used. Part of the capsules was submitted to cold pre-treatment. Three capsules were maintained for 7, 14 and 21 days at 4°C, when seeds were placed on germination medium. As control, seeds from three capsules were placed on media immediately after sterilization. Germination was made on MS medium with different concentrations of macro-nutrients and vitamin-compositions: (MS) MS medium full strength; (1/2 MS) MS medium with macro-nutrients at half strength; (MSB5) MS salts with B5 (Gamborg et al., 1968) vitamins. MS and 1/2 MS were supplemented with 2% sucrose, whereas MSB5 was supplemented with 3% sucrose. All media were gelified with 1.2% agar,

at pH 6.4 (before autoclaving). The number of seeds tested for each treatment in Experiments I and II is shown in Tables 1 and 2, respectively.

The cultures from both experiments were maintained on the media for 45 days at  $26 \pm 1^\circ\text{C}$  in a 16 h light regime at a light intensity of  $22.5 \mu\text{Em}^{-2}\text{s}^{-1}$ . After this period, the germinated seeds were counted and the seedlings were transferred to media with hormones to allow the production of adventitious shoots.

*Multiplication of adventitious shoots.* After 45 days of culture, seedlings were transferred individually to test tubes containing 8 ml of medium. Tissues obtained from Experiment I were placed on K, K/MS micro and K/MS vit, respectively, containing 2 mg/L BAP, 0.5 mg/L NAA, 2% sucrose, at pH 6.4 (before autoclaving). Seedlings obtained from the three different media tested in Experiment II were randomly transferred to MS and 1/2 MS, containing 2 mg/L BAP, 0.5 mg/L NAA, 2% sucrose, at pH 6.4 (before autoclaving). The concentrations of hormones used in the present study were based on positive results for shoot multiplication in other species of the genus *Vriesea* (Mercier and Kerbauy, 1992, 1994). Subcultures to media with the same composition were performed at monthly intervals, over four months. The number of tubes/medium varied from 20 to 23. Multiplication efficiency was calculated as the percentage of seedlings that gave rise to adventitious shoots as well as the mean number of shoots/seedling.

Data of percentage of seedlings that produced adventitious shoots were analyzed using the chi-square test. The number of adventitious shoots/seedling was analyzed using the one-way analysis of variance (*V. philippocoburgii*) and Kruskal-Wallis non parametric analysis of variance (*V. gigantea*) and the comparison among species were made using the Mann-Whitney test.

*Plant development from adventitious shoots.* After six months of culture, the adventitious shoots obtained from Experiment I and II were transferred to K medium and MS medium, respectively, containing 0.2 mg/L NAA, 2% sucrose, 0.5% activated charcoal, 1.2% agar at pH 6.4 (before autoclaving). After three months, a subculture to medium with the same composition was made for further three months. Whole plants were transplanted to plastic cups containing vermiculite, covered with plastic film. Plants were gradually exposed to ambient humidity over one week and then transferred to plastic pots with soil.

## RESULTS AND DISCUSSION

*In vitro* germination. Seed germination initiated eight-then days after sowing and occurred on all media tested, resulting in the production of one plant per seed. In Experiment I, both *V. gigantea* and *V. philippocoburgii* showed high germination rates (99 and 89%, respectively) (Table 1). For *V. gigantea*, cold pre-treatment did not cause any increase of germination rate (Table 2). Nevertheless, a shorter time was necessary to induce germination when compared with seeds not exposed to cold pre-treatment (data not shown). The low germination percentage obtained in 1/2 MS was due to contamination of seeds from one locus of the capsule. The results showed that capsules could be stocked in freezer, which could be interesting when a high number of samples were collected and a longer time was needed to process and place all material in culture.

The percentages of germination obtained here were similar to that obtained previously in the literature. Mercier and Kerbaux (1994) registered up to 90% of germination for *V. hieroglyphica* using 1/2 K and 1/4 K media, with each seed giving rise to 3-7 plantlets. For *V. splendens*, Mekers (1977) observed that good germination of seeds was achieved on K medium supplemented with MS micro-nutrients, although, without hormones, the percentage of germinating seeds was only 62%. On the other hand, on MS medium, seeds germinated, but seedlings died after some weeks. The author attributed this to the sensitivity of the species to high salt-concentrations present in MS medium. When using MS media containing only 1/2 or 1/4 of the normal salt-concentration, Mekers (1976) obtained a greater surviving rate of plantlets.

*Multiplication of adventitious shoots and development of plants.* Adventitious shoot multiplication occurred in response to all media tested. The potential of multiplication, expressed by the percentage of seedlings that produced adventitious shoots ranged from 40 to 68% for *V. gigantea* and did not vary significantly among media ( $p = 0.17$ ) (Table 3). For *V. philippocoburgii*, all seedlings gave rise to adventitious shoots (Table 3). Species showed different genotypic responses to culture conditions. In Experiment I, the average number of shoots/seedling did not vary significantly among media within each species, ranging from 0.55 to 1.05 for *V. gigantea* ( $p = 0.22$ ) and from 3.60 to

5.05 for *V. philippocoburgii* ( $p = 0.25$ ). Comparing the species, *V. gigantea* and *V. philippocoburgii* showed an average of 0.72 and 4.3 shoots/seedling, respectively. These differences were statistically significant ( $p < 0.001$ ) (Table 3). While showing low capacity of multiplication on the different modifications of K medium, plantlets of *V. gigantea* gave rise to higher numbers of adventitious shoots on MS and 1/2 MS (Table 3). MS medium proved to be the best medium for this species (3.14 shoots/seedling;  $p = 0.047$ ). Some earlier reports showed that for other species of the genus *Vriesea*, the surviving and multiplication rates were higher on low salt-concentration media. Mekers (1977) investigating the *in vitro* culture of some bromeliads, including three species of *Vriesea*, concluded that for multiplication of plantlets using seedlings as explants, a low salt-concentration was needed, like 1/3 to 1/2 of MS medium, or normal to half strength of K medium. In two other reports, twenty two shoots were formed per seedling for *V. fosteriana* after three months on K medium containing 2 mg/L BAP and 0.5 mg/L NAA (Mercier and Kerbaux, 1992), while for *V. hieroglyphica*, seven shoots/plantlet were produced after six months in K medium with the same concentrations of hormones (Mercier and Kerbaux, 1994). Recent reports that focused physiological aspects of *V. philippocoburgii* and *V. gigantea*, utilized K medium, even normal or diluted (Mercier et al., 1997; Endres and Mercier, 2001; Endres et al., 2002; Endres and Mercier, 2003).

At the last stage of the culture, a lower concentration of NAA (0.2 mg/L) was used. Initially, adventitious shoots proliferation continued. It seems that cytokinins have a prolonged residual activity even after transferring the shoots to a cytokinin-free medium (Mekers, 1977). After three months, root and apical shoot growth occurred. Mercier and Kerbaux (1992) obtained an increase in length and in the number of roots for *V. fosteriana*, using this concentration of NAA, when compared with other levels and combinations of growth regulators.

To increment the development of the plantlets, activated charcoal was added to the medium at the last stage. Addition of activated charcoal have been reported for other species such as *Anemone canadensis*, *Anemone hephehensis*, *Nicotiana tabacum* (Johansson, 1983), *Allium cepa*, *Daucus carota* and *Haplopappus gracilis* (Fridborg and Erikson, 1975; Fridborg et al., 1978).

**Table 1** - Seed germination of *Vriesea gigantea* and *Vriesea philippocoburgii* on three medium- compositions, based on Knudson medium.

Medium	<i>Vriesea gigantea</i>			<i>Vriesea philippocoburgii</i>		
	Total number of seeds	Germinated seeds		Total number of seeds	Germinated seeds	
		Number	%		Number	%
K <sup>1</sup>	621	614	98.9	140	124	88.6
K/MS micro <sup>2</sup>	621	614	98.9	199	176	88.4
K/MS vit <sup>3</sup>	654	648	99.1	204	183	89.7
Total	1896	1876	98.9	543	483	88.9

<sup>1</sup> K medium, 2% sucrose, 1.2 % agar, <sup>2</sup>K macro-nutrients, MS micro-nutrients, 2% sucrose, 1.2 % agar, <sup>3</sup>K salts, MS vitamins, 2% sucrose, 1.2% agar.

**Table 2** - Seed germination of *Vriesea gigantea* on three MS medium-compositions, with different times of cold pre-treatment.

Pre-treatment (days)	Medium	Total number of seeds	Germinated seeds	
			Number	%
0	MS <sup>1</sup>	263	247	93.9
	1/2 MS <sup>2</sup>	247	169	64.4
	MSB5 <sup>3</sup>	228	224	98.2
7	MS	254	237	93.3
	1/2 MS	251	239	95.2
	MSB5	269	264	98.1
14	MS	252	225	89.3
	1/2 MS	240	232	96.7
	MSB5	271	263	97.0
21	MS	259	240	92.7
	1/2 MS	247	235	95.1
	MSB5	280	271	96.8
Total		3061	2846	93.0

<sup>1</sup> MS full strength, 2 % sucrose, 1.2 % agar, <sup>2</sup>MS with macro-nutrients at half strength, 2 % sucrose, 1.2 % agar, <sup>3</sup>MS salts, B5 vitamins, 3 % sucrose, 1.2 % agar.

**Table 3** - Effect of different medium-compositions on adventitious shoot multiplication of *Vriesea gigantea* and *Vriesea philippocoburgii* after four months on multiplication medium.

Experiment	Medium <sup>1</sup>	<i>V. gigantea</i>		<i>V. philippocoburgii</i>		P value
		Responsive explants (%)	Number of shoots/explant (mean ± S.D.)	Responsive explants (%)	Number of shoots/explant (mean ± S.D.)	
I	K	55	0.55 ± 0.76b <sup>2</sup>	100	3.60 ± 1.70	p < 0.001 <sup>3</sup>
	K/MS micro	60	1.05 ± 1.10ab	100	5.05 ± 2.26	
	K/MS vit	40	0.55 ± 0.76b	100	4.25 ± 2.53	
Mean ± S.D.			0.72 ± 0.90		4.30 ± 2.23	
II	MS	68	3.14 ± 5.10 a			
	1/2 MS	65	1.61 ± 2.59 ab			
Mean ± S.D.			2.38 ± 3.85			
p value		0.17 <sup>4</sup>	0.047 <sup>5</sup>	0.25 <sup>6</sup>		

<sup>1</sup> All media contained 2 mg/L BAP, 0.5 mg/L NAA, 2% sucrose, 1.2 % agar, <sup>2</sup>Multiple comparison test: different letters in the same column indicate significant differences at 0.05, <sup>3</sup>p value obtained according to Mann-Whitney, <sup>4</sup>p value obtained according to Chi-square, <sup>5</sup>p value obtained according to Kruskal-Wallis.

Favourable effects of its addition to the medium were probably due the adsorption of inhibiting substances derived from the agar (Kohlenbach and Wernicke, 1978), auxins and cytokinins (Weatherhead et al., 1978), chelates (Heberle-Bors, 1980) and phenolic substances (Fridborg et al., 1978). After transplanting to vermiculite and later to soil, all regenerated plants grew and showed normal phenotypes over more than ten months.

Seed of *V. gigantea* and *V. philippocoburgii* could be cultured *in vitro* on Knudson and Murashige and Skoog media, giving rise to adventitious shoots and plants. Seedlings of *V. gigantea* were capable to produce only few adventitious shoots on K medium. Otherwise, a higher salt-concentration, specially of micro-nutrients, present in MS medium led to higher shoot multiplication rates, being indicated for *in vitro* culture of this species.

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#### RESUMO

A importância das bromélias como plantas ornamentais e o fato de várias espécies nativas do Rio Grande do Sul estarem ameaçadas de extinção leva ao interesse pela sua micropropagação. Uma cultura *in vitro* de *Vriesea gigantea* e *Vriesea philippocoburgii* foi estabelecida a partir de sementes. A taxa de germinação de *V. gigantea* foi superior à taxa de germinação de *V. philippocoburgii*. Plântulas de *V. philippocoburgii* deram origem a vários brotos adventícios quando cultivadas em meio basal de Knudson. Por outro lado, para *V. gigantea*, foi necessária uma maior concentração de sais, sendo que o número de brotos foi aumentado com o meio Murashige and Skoog. A adição de carvão ativado e ácido naftalenoacético ao meio de regeneração permitiu o desenvolvimento de brotos e a formação raízes,

confirmando o sucesso da cultura *in vitro*. A expressão diferencial dos genótipos justifica a necessidade de continuadas pesquisas para que se estabeleçam as condições ideais para os tecidos específicos.

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