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Quantification of 5-methylcoumarin-4-glucoside and 11,12-dihydroxy-5-methylcoumestan in Six Peruvian Species of the Genus *Mutisia* (Asteraceae)

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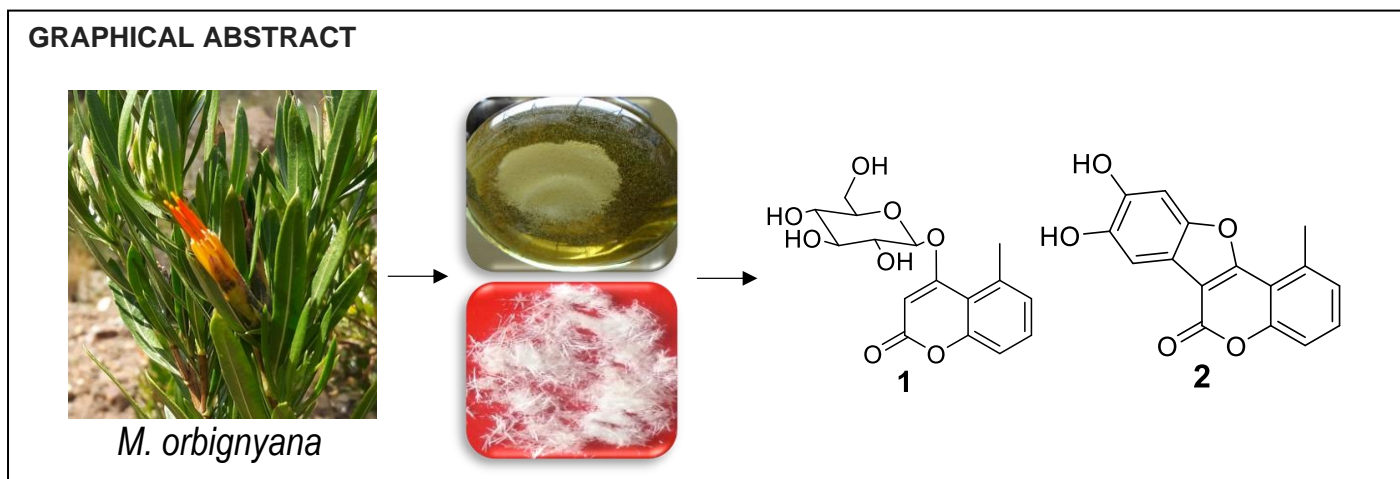
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HIGHLIGHTS

- A 5-methylcoumarin and a coumestan were isolated from *M. orbignyana*.
- An HPLC method was established for the quantification of both compounds.
- The concentrations of both compounds were assessed in six samples of *Mutisia*.
- *Mutisia orbignyana* displayed the highest contents of both isolated compounds.

Abstract: *Mutisia* L. f. (Mutisiinae, Mutisieae, Mutisioideae, Asteraceae) is a South American genus that contains several medicinal species. *Mutisia acuminata* Ruiz & Pav. and *M. orbignyana* Wedd. are known sources of the 12-dihydroxy-5-methylcoumestan and 5-methylcoumarin-4-glucoside respectively. The 12-dihydroxy-5-methylcoumestan exhibited notable antihepatotoxic activity in a previous study. In the present work, 5-methylcoumarin-4-glucoside and 12-dihydroxy-5-methylcoumestan were isolated from a Peruvian collection of *M. orbignyana*. Additionally, the relative concentrations of those two isolated compounds in six Peruvian species of *Mutisia* (*M. acuminata*, *M. cochabambensis* Hieron., *M. lanata* Ruiz & Pav., *M. orbignyana*, *M. venusta* S.F. Blake and *M. wurdackii* Cabrera) were determined by an HPLC method. The highest concentrations of both constituents were observed in the sample of *M. orbignyana*.

Keywords: *Mutisia*; Asteraceae; 5-methylcoumarin; coumestan.



INTRODUCTION

Among the main antihepatotoxic agents of natural origin silymarin stands out for its great commercial impact and wedelolactone due to its high dissemination because it comes from *Eclipta alba* (Asteraceae), a common weed that occurs in the lowlands of India subcontinent [1]. In 1988, Daily and coauthors [2] reported the presence of 11,12-dihydroxy-5-methylcoumestan among other substances in the methanolic extract of *Mutisia acuminata* (Mutisieae, Mutisioideae, Asteraceae). Furthermore, in 2009, Flores and coauthors [3], established the presence of the 5-methylcoumarin-4-glucoside (**1**) and 11,12-dihydroxy-5-methylcoumestan (**2**) in *M. orbignyana* (Figure 1). The chemical structure of compound **2** is related to wedelolactone and a remarkable antihepatotoxic effect has also been demonstrated for it [2]. Regarding compound **1**, a high concentration of it has recently been reported in *Vernonia glaberrima* (Vernoniaeae, Cichoroideae, Asteraceae) [4] showing cytotoxic activity. The presence of 5-methylcoumarins in member of *Mutisioideae* subfamily has also been extensively reviewed by Vestena and coauthors 2022 [5], and members of *Mutisia* genus stands out as potential sources of 5-methylcoumarins with potential antihepatotoxic activity. Therefore, in the present work, both 5-methylcoumarin-4-glucoside (**1**) and 11,12-dihydroxy-5-methylcoumestan (**2**) were purified from a Peruvian assession of *M. orbignyana*. Additionally, the relative concentrations of both substances in six Peruvian species of *Mutisia* genus (*M. acuminata*; *M. cochabambensis*; *M. lanata*; *M. orbignyana*, *M. venusta* and *M. wurdackii*) were determined by an HPLC method as part of our program for the search of new source of 5-methylcoumarins.

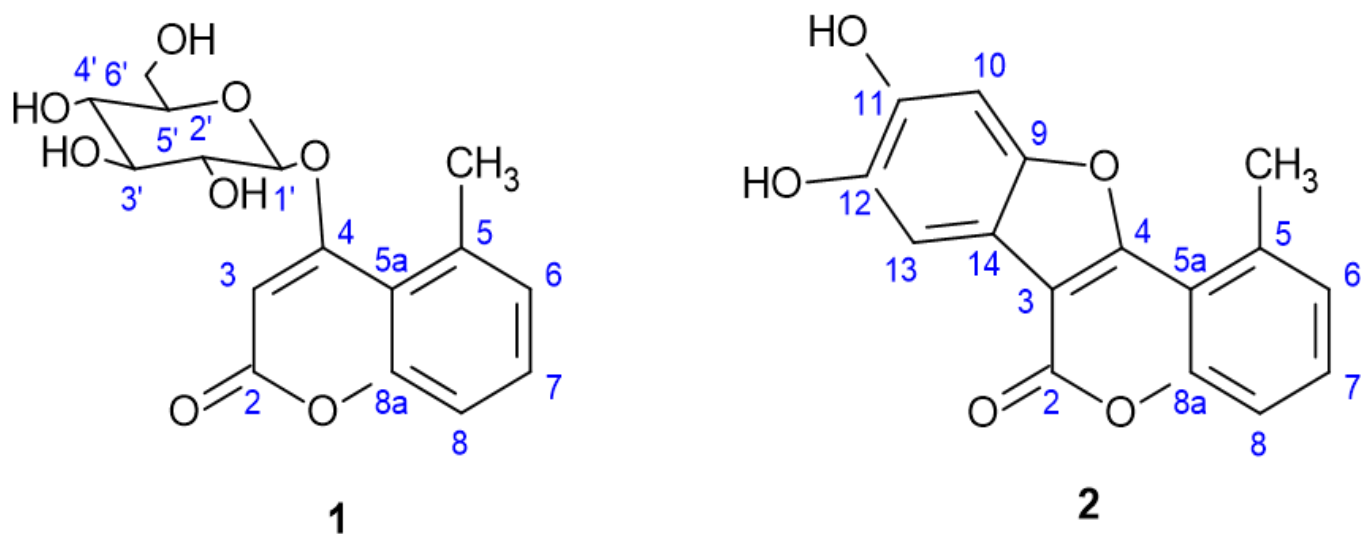


Figure 1. Structures of 5-methylcoumarin-4-glucoside (**1**) and 11,12-dihydroxy-5-methylcoumestan (**2**)

MATERIAL AND METHODS

Plant material

Aerial parts (leaves) of *M. acuminata* Wedd., *M. cochabambensis* Hieron., *M. lanata* Ruiz & Pav. and *M. venusta* S.F. Blake were collected in Cusco region, Perú. *Mutisia orbignyana* was collected in Moquegua region, Perú whereas *M. wurdackii* Cabrera was collected in Amazonas region, Perú. The plants were translated to the phytochemistry laboratory (Chemistry Department, UNSAAC), and dried at room temperature.

Isolation of 5-methylcoumarin-4-glucoside (1) and 11,12-dihydroxy-5-methylcoumestan (2)

50 g of dried *M. orbignyana* were exhausted with 96% ethanol. The dried ethanolic extract was suspended in hot water (85 °C) and vacuum filtered to remove waxes and chlorophylls. The aqueous extract was partitioned with ethyl acetate. The ethyl acetate extract was evaporated and vacuum liquid chromatographed with a gradient of hexane and ethyl acetate, the fraction of hexane:ethyl acetate (7:3) yielded by ethanol recrystallization 15.4 mg of **2** (0.034%) and the ethyl acetate fraction yielded by ethanol recrystallization 27 mg of **1** (0.054%). The structures of the isolated compounds were established based on NMR spectroscopy, high-resolution MS and comparison with literature data as 5-methylcoumarin-4- β -glucoside (**1**) and 11,12-dihydroxy-5-methylcoumestan (**2**) [2,3].

5-Methylcoumarin-4- β -glucoside (**1**): ¹HNMR (DMSO-d₆, 400MHz) δ 5.98 (s, 1H, H-3), δ 7.16 (d, 1H, H-6), δ 7.51 (t, 1H, H-7), δ 7.25 (d, 1H, H-8), δ 2.71 (s, 3H, C₅-CH₃), δ 5.20 (d, 1H, H-1'), δ 3.41 (t, 1H, H-2'), δ 3.39 (t, 1H, H-3'), δ 3.20 (t, 1H, H-4'), δ 3.49 (m, 1H, H-5'), δ 3.72 (t, 1H, H-6'), δ 4.63 (m, 1H, -OH-6'), δ 5.11, 5.20, 5.49 (3d, 3H, -OH- 2',3',4'). ¹³CNMR (DMSO-d₆, 100 MHz) δ 161.7 (C-2), δ 93.4 (C-3), δ 167.0 (C-4), δ 114.2 (C-5a), δ 137.5 (C-5), δ 128.2 (C-6), δ 132.4 (C-7), δ 115.3 (C-8), δ 154.8 (C-8a), δ 23.6 (CH₃), δ 100.3 (C-1'), δ 73.5 (C-2'), δ 77.0 (C-3'), δ 70.0 (C-4'), δ 77.8 (C-5'), δ 61.0 (C-6'). LC-HRESIMS negative mode *m/z* 383.0984 [M - H]⁻ (calcd for C₁₆H₁₈O₈+CH₂O₂, 383.0978).

11,12-Dihydroxy-5-methylcoumestan (**2**): ¹HNMR (DMSO-d₆, 400MHz) δ 9.60 (br, 2H, 11,12-OH), δ 7.53 (t, 1H, H-7), δ 7.40 (d, 1H, H-6) δ 7.30 (s, 1H, H-10), δ 7.30 (d, 1H, H-8), δ 7.23 (s, 1H, H-13), δ 2.85 (s, 3H, C₅-CH₃). ¹³CNMR (DMSO-d₆, 100 MHz) δ 158.8 (C-2), δ 104.7 (C-3), δ 157.5 (C-4), δ 113.4 (C-5), δ 111.8 (C-5a), δ 134.1 (C-6), δ 130.5 (C-7), δ 126.7 (C-8), δ 153.0 (C-8a), δ 105.5 (C-9), δ 114.7 (C-10), δ 149.4 (C-11), δ 146.4 (C-12), δ 98.8 (C-13), δ 144.7 (C-14), δ 20.8 (-CH₃). LC-HRESIMS negative mode *m/z* 281.0445 [M - H]⁻ (calcd for C₁₆H₁₀O₅, 383. 281.0450).

Quantification of 5-methylcoumarin-4-glucoside (1) and 11,12-dihydroxy-5-methylcoumestan (2)

For quantification purposes, 500 mg of each dried plant material was macerated for 24 hours with 20 mL of 70% ethanol, this operation was repeated 3 times and the filtrates were made up to 60mL. Five mL of each extract were evaporated at 40°C, weighed and redissolved in one mL of methanol and filtrated by 0.22 μ m PTFE membrane for conditioning in chromatographic vials. The chromatographic method was developed in a Thermo Scientific Ultimate 3000 UHPLC chromatograph with automatic injection and DAD detector. A Zorbax Eclipse Plus C₁₈ Column (1.8 μ m particle size, 4.6 x 100 mm) was used. Column temperature was set at 35°C. Mixtures of 0,1% formic acid in water (A) and acetonitrile (B) were used as mobile phase with the following gradient program (minutes, %B): 0,2; 5,30; 10,80; 12,80; 15,100; 17,100; 19,2; 20,2. DAD: 200-400 nm: 280, 345, 254, 330 nm. The total run time was 20 minutes and a flow rate of 0.4 mL/min. The relative concentrations of both compounds were quantified using calibration curves prepared with standard solutions of **1** and **2** dissolved in methanol with six data points. The calibrations curves were obtained by potting the peak area signals as a function of the concentration. The equation curves were obtained (Y=64.55X-2.71 for **1**, Y=115.84X-1.43 for **2**) and linearity was evaluated by least-squares regression analysis (*r*² = 0.9996 for **1**, and *r*² = 0,9995 for **2**). For quantification of **1** the chromatograms at 280 nm were used, whereas **2** was quantified at 345nm.

RESULTS AND DISCUSSION

In the Figure 2, typical UHPLC chromatograms of *M. orbignyana* are displayed. Compound **1** eluted at a retention time of 8.20 min whereas compounds **2** eluted at 12.09 min. In addition to the isolated constituent additional chromatographic peaks were observed, highlighting the occurrence of two peaks with retention times of 6.99 (peak **a**) and 8.89 (peak **b**) which displayed the characteristic UV spectra of the phenylpropanoids. Although phenylpropanoid are ubiquitous constituent in members of Asteraceae family,

there is no previous report on the occurrence of phenylpropanoids in *M. orbignyana*. The phenylpropanoids 3-O-caffeoylquinic acid and 5-O-caffeoylquinic acid were previously describe in *M. friesiana* [6].

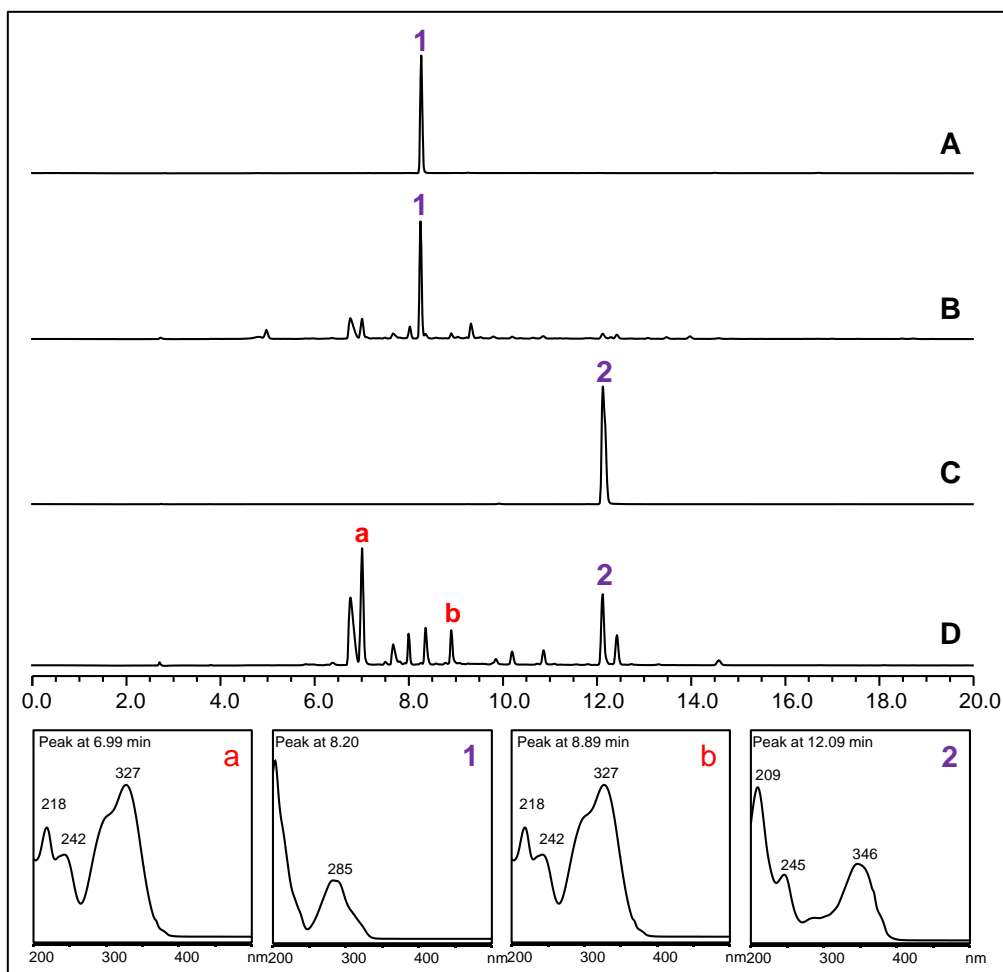


Figure 2. Typical chromatograms of standard compounds and *Mutisia orbignyana*. (A) Chromatogram of a standard solution of 5-methylcoumarin-4-β-glucoside (**1**) monitored at 280nm. (B) Chromatogram of the ethanolic extract of *M. orbignyana* monitored at 280nm. (C) Chromatogram of a standard solution of 11,12-dihydroxy-5-methylcoumestan (**2**) monitored at 345 nm. (D) Chromatogram of the ethanolic extract of *M. orbignyana* monitored at 345nm. In the bottom, four selected UV spectra of unidentified phenylpropanoid derivatives (**a** and **b**) and the standard compounds **1** and **2**.

The identity of those phenylpropanoids in *M. orbignyana* as well as their occurrence in other member of *Mutisia* genus need to be elucidated in further investigations. The Table 1 displays the content of **1** and **2** in six Peruvian species from *Mutisia*. *Mutisia orbignyana* is the plant with the highest content of both substances while in *M. cochabambensis* neither of those compounds were present. According to Cabrera [7], who divided the genus *Mutisia* into six sections based on morphological aspects, *M. orbignyana* belongs to the *Isantha* section, the most basal. *Mutisia orbignyana* has a particularly high concentration of **1** comparable to the African plant *Vernonia glaberrima* (*Cichoroideae*). On the other hand, to our knowledge there is no other genus apart from *Mutisia* that displays the presence of **2**.

Table 1. Content of 5-methylcoumarin-4-glucoside (**1**) and 11,12-dihydroxy-5-methylcoumestan (**2**) in six *Mutisia*¹.

Species	1	2
<i>M. acuminata</i>	3.6	0.78
<i>M. cochabambensis</i>	-	-
<i>M. lanata</i>	0.45	-
<i>M. orbignyana</i>	47.6	6.83
<i>M. venusta</i>	0.61	-
<i>M. wurdackii</i>	0.31	0.34

¹ mg of compound/g of dry plant. Concentrations were obtained with the following calibration curves: Y=64.55X-2.71 for **1**, Y=115.84X-1.43 for **2**.

CONCLUSION

Mutisia acuminata and especially *M. orbignyana* are candidate species to extract both 5-methylcoumarin-4-glucoside (1) and 11,12-dihydroxy-5-methylcoumestan (2). The other Peruvian *Mutisia* have none or small amounts of these compounds. However, further analysis of samples from different geographical origin and collection times are required.

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Conflicts of Interest: The authors declare no conflict of interest.

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