

Biology and Thermal Requirements of *Utetheisa ornatrix* (L.) (Lepidoptera: Arctiidae) Reared on Artificial Diet

André Gustavo Corrêa Signoretti¹, Dori Edson Nava^{2*}, José Maurício Simões Bento¹ and José Roberto Postali Parra¹

¹Departamento de Entomologia, Fitopatologia e Zoologia Agrícola; Escola Superior de Agricultura "Luiz de Queiroz"; Universidade de São Paulo; 13418-310; Piracicaba - SP - Brasil. ²Embrapa Clima Temperado; nava@cpact.embrapa.br; 96001-970; Pelotas - RS - Brasil

ABSTRACT

A study on the biology of *Utetheisa ornatrix* reared on the artificial diet, was conducted to determine the thermal requirements in each development stage. The aim was to find out the thermal regions in the São Paulo state where the pest could develop on *Crotalaria* spp. The insects were reared on an artificial diet based on the white beans and yeast. *U. ornatrix* thermal requirements were tested at 18, 20, 22, 25, 28, 30, and 32°C 70±20% RH, and 14-h photophase. Lower threshold temperatures (TT) and thermal constants (K) for the egg, larval and pupal stages were 12.7°C and 51.2 GDD, 13.5°C and 290.9 GDD, 13.8°C and 108.4 GDD, respectively; TT and K values for the biological cycle were 13.8°C and 436.3 GDD.

Key words: *Crotalaria*, Rattlebox moth, rearing technique

INTRODUCTION

The Rattlebox moth, *Utetheisa ornatrix* (L., 1758) (Lepidoptera: Arctiidae), is considered to be the most important *Crotalaria* spp. (*Crotalaria anagyroides*, *C. falsajuncea*, *C. juncea*, *C. stipularia*, *C. usaramohensis*, *C. vittelina*) (Leguminosae) pest in Brazil (Gallo et al. 2002). It occurs on four other plant species (Silva et al. 1968), and has been found from the North America (east of the Rocky Mountains) to South America (Argentina, Brazil and Chile) (Pease 1968; Silva et al. 1968). The larvae feed on the green pods and on the developing seeds, and they may reduce the crop yields when the infestation is high (Ferro 2001).

In the past few years, the area planted to the *crotalaria* has increased significantly, reaching about 3,000 ha in Brazil (José Aparecido Donizete Cardoso, personal communication). The *crotalaria* is used for the green manure and cover crop (Rosa et al. 2004; Castro et al. 2005) as an controlling nematode agent and as a fiber for the tobacco industry (Nogueira et al. 1992).

The definition of the pest thermal requirements has helped the pest management programs to predict the pest occurrence (Nava and Parra 2003) and to mass-rear the natural enemies for the applied biological control programs (Parra et al. 2002). In this study on the biology of *U. ornatrix* reared on the artificial diet in laboratory the thermal requirements for each insect's development stages in the laboratory were determiner. Results aimed

* Author for correspondence

to predict the insect occurrence based on the number of annual generations and to facilitate the basic laboratory studies to develop alternative pest control methods.

MATERIAL AND METHODS

Biology of *U. ornatrix* in artificial diet

U. ornatrix adults were obtained from *Crotalaria* fields in Piracicaba, SP and were taken to the laboratory. They were maintained in PVC tube cages (10 × 20 cm) until the oviposition and for the preliminary studies, the cages were covered with the multipurpose office paper (as the egg laying substrate). The adults were fed a 10% honey solution and the eggs were collected daily by cutting the containing egg paper.

The first-generation caterpillars (n=150) were inoculated and individually placed into glass vials (2.5 × 8.5 cm) for the biological study on the artificial diet. The glass vials were filled to one third with the artificial diet containing the white bean (130.0 g), carrot (30.0 g), yeast extract (20.0 g), vitamin solution (40.0 ml), Vitagold® (B complex vitamins) (2.0 ml), ascorbic acid (1.6 g), sorbic acid (0.6 g), methyl parahydroxybenzoate (nipagin) (1.1 g), propionic acid (0.4 ml), Tetrex® (tetracycline) (0.3 mg), forlamdehyde 40% (1.0 ml), agar (7.0 g), and distilled water (500.0 ml). The vitamin solution were compose of: niacinamide (1.00 mg), calcium pantothenate (1.00 mg), thiamine (1.00 mg), riboflavin (0.25 mg), pyridoxine (0.25 mg), folic acid (0.25 mg), biotin (0.02 mg), vitamin B12 (0.002 mg) and inositol (20.00 mg).

The diet preparation followed the methodology developed by Parra (2001). During the pupation, the insects were separated and placed in the plastic cups on a tray covered with the filter paper which was daily moistened. The emerging adults (25 pairs) were separated by the sex (based on the abdomen tip) and placed under the air-conditioned room at 25 ± 2°C, 70 ± 10% RH, and 14-h photophase.

The biological parameters measured were the duration and viability of the egg, larval and pupal stages, and of the biological cycle (egg to adult); number of instars; male and female pupal weight, pre-oviposition and oviposition periods, sex ratio, fecundity, and longevity. The number of the instars was obtained by daily measuring the head capsule width of 20 caterpillars, with a Wild MM

5235 ocular micrometer attached to a stereoscopic microscope and by adopting the multimodal frequency curve for these measurements. The hypotheses were tested using the linear model for Dyar's rule and the MOBAE software (Parra and Haddad 1989). The sex ratio (sr) was obtained by the formula $sr = \frac{\text{♀}}{\text{♀} + \text{♂}}$.

Thermal requirements of *U. ornatrix*

The insects were reared on the artificial diets in incubators maintained at 18, 20, 22, 25, 28, 30, and 32 ± 1°C. The embryonic development was observed in 120 eggs/temperature. The eggs were distributed on the acrylic dishes (6.0 × 2.0 cm) containing the partially moistened (bottom part) filter paper. The larval development was observed in 150 larvae/temperature; the insects were individualized in the glass vials (2.0 × 7.0 cm) containing the artificial diet. The pupal development was studied using 100 insects and each one was placed in the acrylic dishes (6.0 × 2.0 cm). The dishes were kept moistened by the humid cotton pads placed in the dishes.

The duration and viability of the embryonic period and the larval and pupal stages were assessed. After determining the duration of the development stages and of the biological cycle (egg to adult) at different temperatures from 18 to 30°C, the threshold temperature (TT) and thermal constant (K) were calculated using the hyperbole method (Haddad et al. 1999). At 32°C, the duration was not taken into account since it was not statistically different from the duration at 30°C (P≥0.05), thus avoiding the linearity of the hyperbole curve that represented the duration and temperature.

Data analysis

The tests were organized in a completely randomized design and the data submitted to ANOVA. The means were compared by the Tukey test at 5% probability level. The viability data were transformed to arc sine $\sqrt{x + 4}$. The probable number of *U. ornatrix* generations per year for the isotherms within the observed range was calculated via the general thermal constant formula $K = D(T - T_b)$, based on the temperatures that enhanced the insect development (Silveira Neto et al. 1976). The number of the annual generations was obtained by dividing 365 days by the duration value obtained for each temperature.

RESULTS AND DISCUSSION

Biology of *U. ornatrix* on artificial diet

The artificial diet containing the bean and yeast extract led to an adequate development of the immature and adult *U. ornatrix* stages. The duration and viability of the egg, larval, and pupal stages were 4.5 and 84.2, 26.5 and 85.3, and 10.3 days and 91.4%, respectively, for a total duration of 42 days and a 65.6% viability of the biological cycle (egg to adult) (Table 1).

The larval head capsules grew in a geometric progression at each ecdysis, at a constant rate of about 1.4, following Dyar's rule (1890). All the larvae had five instars (Fig. 1). The sex ratio was 0.49, or 1M:1F. The pupal weight was 204.1 mg for the males and 184.9 mg for the females (Table 2). The pre-oviposition period lasted 4.1 days and the oviposition period 18.6 days (Table 2). The fecundity mean was 239.5 eggs, with the longevity values of 24.0 and 22.8 days for the males and females, respectively (Table 2). The daily fecundity mean was 12.9 eggs per day per female, which was higher than the nine eggs mentioned by Johnson et al. (1985), when rearing *U. ornatrix* on the natural diet (*Crotalaria*).

Although 75% eggs-adult viability has been considered adequate for the insects reared on artificial diets (Singh 1983), in the present study the viability was below that percentage. However

U. ornatrix gradually could adapt to the artificial diet in future generations, given that the number of insects reared on the diet for one generation was not different to those on the natural diet, thus showing the adequate nutritional value of the artificial medium. As mentioned earlier, one of the objectives of this work was to study *U. ornatrix* biology when reared on the artificial diet for the mass-rearing production. Apparently the insect can be reared on such medium allowing to study the alternative biological control methods, such as with the *Cotesia* sp. parasitoid, frequently found as a parasite on the caterpillar.

This study is the first to record the rearing of *U. ornatrix* on artificial diet, although the data line refers to one generation only. It has been hypothesized in previous studies that continued rearing of *U. ornatrix* requires that the insect feeds on *Crotalaria* sp. because of chemical compound, which are required for the of the plant insect to mate, such as hydroxydanoidal, the precursor of its sex pheromone (Conner et al. 1990). The females select males that sequester the largest amount of the alkaloid as a natural selection strategy because the larger males will probably have more sperm cells to fertilize the eggs. Further studies should be conducted to determine the possibility of rearing the insect on the artificial diet for the successive generations.

Table 1 - Mean (\pm SD) duration and viability of *Utetheisa ornatrix* eggs, larvae, and pupae reared on artificial diet. Temperature $25 \pm 2^\circ\text{C}$, relative humidity $70 \pm 10\%$, and 14-hour photophase.

Stage/Period	Duration (days)	Viability (%)
Egg	4.5 ± 0.1	84.2 ± 12.34
Larva	26.5 ± 2.52	85.3 ± 12.89
Pupa	10.3 ± 0.96	91.4 ± 9.15
Life cycle (egg-adult)	42.0	65.6

Table 2 - Number of instar, duration of pre-oviposition and oviposition periods, sex ratio, fecundity, longevity of male e female, and pupal weight of male e female of *Utetheisa ornatrix* reared on artificial diet. Temperature $25 \pm 2^\circ\text{C}$ temperature, relative humidity $70 \pm 10\%$, and 14-hour photophase.

Biological parameters	Mean \pm SD
Number of instars	5.0
Pre-oviposition (days)	4.1 ± 1.74
Oviposition (days)	18.6 ± 5.18
Sex ratio	0.49
Mean fecundity (number of eggs)	239.5 ± 128.4
Longevity (days)	
Male	24.0 ± 4.35
Female	22.8 ± 6.74
Pupal weight (mg)	
Male	204.1 ± 031.1
Female	184.9 ± 024.5

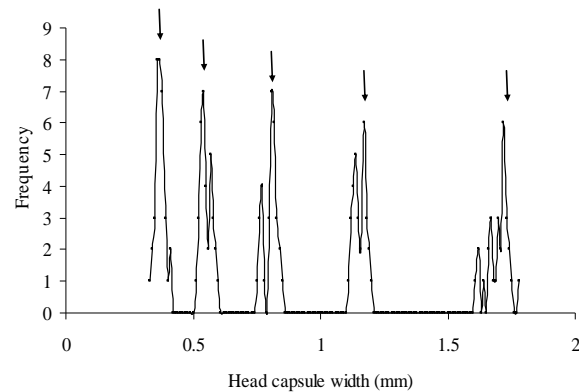


Figure 1 - Frequency distribution of head capsule widths of *Utetheisa ornatrix* reared on artificial diet. Arrows indicate the number of instars (n=5).

Thermal requirements

All *U. ornatrix* stages developed within the temperature range chosen for the study (18 to 32°C), although at the highest temperature (32°C), the viability was significantly lower than the smaller (Table 3). The duration of *U. ornatrix* development stages decreased as the temperature increased (Table 3). The egg stage lasted eight days at 18°C and three days at 32°C. The larval stage duration was 56.5 days at 18°C and 17 days at 32°C. At the pupal stage, the duration decreased from 24.1 days at 18°C to 6.3 days at 32°C. The duration of the biological cycle (egg to adult) varied from 90.6 days at 18°C to 26.9 days at 32°C. At 28, 30 and 32°C, all stage durations were similar, which indicated that the development might be hindered at the temperatures higher than 32°C.

The temperature did not affect the *U. ornatrix* egg viability, which was above 89.3% (Table 3). For the larval and pupal stages, the viability was significantly lower at 32°C. The viability was 78.1 at 70.0% and 98.8 at 77.9% within the thermal range of 18 to 30°C (Table 3). Likewise, 32°C was harmful for the development of the biological cycle of *U. ornatrix*; at the other temperatures, the viability varied from 56.8 to 75.3% (Table 3). Therefore, it could be attributed that within the temperature range under the study, the best temperatures for *U. ornatrix* development were from 18 to 30°C. Also, apparently, the insect prefer lower temperatures because all the stages

were similar at 28°C and higher temperatures. This adverse situation was demonstrated from the decrease in the viability for the larval and pupal stages at 32°C (Table 3).

The threshold temperature (TT) for the embryonic period was 12.7°C and for the larval and pupal stage, they were 13.5 and 13.8°C, respectively. The thermal constants (K) for the egg, larval, and pupal stages were 51.2, 290.9, and 108.4 GDD, respectively. The TT and K values for the biological cycle (egg to adult) were 13.8°C and 436.3 GDD. As reported by Honek (1996), in general, the species adapted to the tropical conditions had a TT of 13.7°C, which indicated that the 13.8°C TT value for the biological cycle of *U. ornatrix* was close to this value (Table 4). The insect populations of different regions can have different thermal requirements and the factors other than the latitude (such as food) can affect the insect development (Zeiss et al. 1996; Mihsfeldt 1998). The regions with the isotherms within the 18 to 30°C range, which were appropriate temperatures for the insect development (Table 3) could have 3.5 to 13.5 generations annually (Fig. 2). In the situations where crotalaria has been the host specially *C. juncea*, cultivated predominantly in the northern part of the São Paulo state (São José do Rio Preto), monthly temperatures varied from 25.6 to 20.1 and the insect might go up to 10 generations/year (Fig. 2).

Table 3 - Mean duration and viability (\pm SD) and variation interval (values in parentheses) for egg, larval, and pupal stages, and for the biological cycle (egg-adult) of *Utetheisa ornatix* reared on artificial diet at different temperatures, relative humidity $70 \pm 10\%$, and 14-h photophase.

Temperatura (°C)	Egg stage	Larval stage	Pupal stage	Life cycle
18	8.0 \pm 0.00 a (8.0-8.0)	56.5 \pm 1.96 a (46.0-74.0)	24.1 \pm 1.23 a (20.0-27.0)	90.6 \pm 0.89 a (84.0-93.0)
20	7.0 \pm 0.00 ab (7.0-7.0)	45.9 \pm 2.25 b (40.0-60.0)	18.2 \pm 0.98 b (15.0-21.0)	71.9 \pm 2.54 b (65.0-77.0)
22	6.3 \pm 0.33 b (6.0-7.0)	40.7 \pm 1.52 b (36.0-60.0)	14.9 \pm 1.05 c (14.0-17.0)	63.2 \pm 1.89 c (56.0-70.0)
25	4.7 \pm 0.67 c (4.0-6.0)	24.0 \pm 0.91c (24.0-34.0)	9.1 \pm 0.75 d (8.0-10.0)	38.3 \pm 1.64 d (29.0-42.0)
28	3.0 \pm 0.00 d (3.0-3.0)	18.6 \pm 0.59 d (14.0-24.0)	7.3 \pm 1.00 e (6.0-9.0)	28.8 \pm 0.81 e (21.0-34.0)
30	3.0 \pm 0.00 d (3.0-3.0)	18.5 \pm 0.36 d (15.0-22.0)	7.0 \pm 0.65 e (6.0-9.0)	27.9 \pm 0.63 e (20.0-32.0)
32	3.0 \pm 0.00 d (3.0-3.0)	17.0 \pm 0.59 d (14.0-22.0)	6.3 \pm 0.49 e (5.0-7.0)	26.9 \pm 0.91 e (18.0-32.0)
Viability (%)				
18	98.1 \pm 1.93 a (94.2-100.0)	70.0 \pm 7.69 a (54.5-84.9)	93.6 \pm 4.72 a (80.0-100.0)	64.3 \pm 6.32 a (51.2-75.0)
20	98.2 \pm 1.19 a (95.9-100.0)	70.4 \pm 11.16 a (56.7-85.6)	82.2 \pm 2.95 a (77.8-90.9)	56.8 \pm 4.08 a (50.5-71.3)
22	98.6 \pm 1.35 a (95.9-100.0)	77.3 \pm 9.64 a (62.2-90.9)	98.8 \pm 1.25 a (95.0-100.0)	75.3 \pm 3.57 a (63.9-82.0)
25	100.0 \pm 0,00 a (100.0-100.0)	77.7 \pm 11.9 a (63.6-95.5)	82.4 \pm 6.83 a (72.9-95.3)	64.0 \pm 6.44 a (56.8-73.2)
28	100.0 \pm 0,00 a (100.0-100.0)	78.1 \pm 6.88 a (63.6-95.5)	77.9 \pm 6.08 a (61.1-80.0)	60.8 \pm 3.91 a (49.1-72.4)
30	89.3 \pm 9.99 a (69.4-100.0)	78.1 \pm 1.25 a (76.2-81.8)	90.9 \pm 4.01 a (81.3-100.0)	63.4 \pm 6.49 a (56.4-75.3)
32	95.8 \pm 2.72 a (90.3-98.7)	60.7 \pm 6.64 b (54.6-80.9)	62.2 \pm 8.32 b (40.0-75.0)	30.2 \pm 2.50 b (17.2-43.1)

Mean values followed by the same letter in the columns are not statistically different according to Tukey test ($P \geq 0.05$).

Table 4 - Lower threshold temperature (TT), thermal constant (K), regression analysis (1/D), and determination coefficient (R^2) for the egg, larval, and pupal stages and for the life cycle (egg-adult) of *Utetheisa ornatix* in the laboratory.

Stages/Period	TT (°C)	K (GDD ¹)	Regression equation	R ²	P
Egg	12.7	51.2	1/D = 0.019524t _i - 0.247656	0.9336	≤ 0.05
Larva	13.5	290.9	1/D = 0.003438t _i - 0.046346	0.9598	≤ 0.05
Pupa	13.8	108.4	1/D = 0.009227t _i - 0.127699	0.9781	≤ 0.05
Egg-adult	13.8	436.3	1/D = 0.002292t _i - 0.031721	0.9704	≤ 0.05

⁽¹⁾GDD: Growing Degree-days.

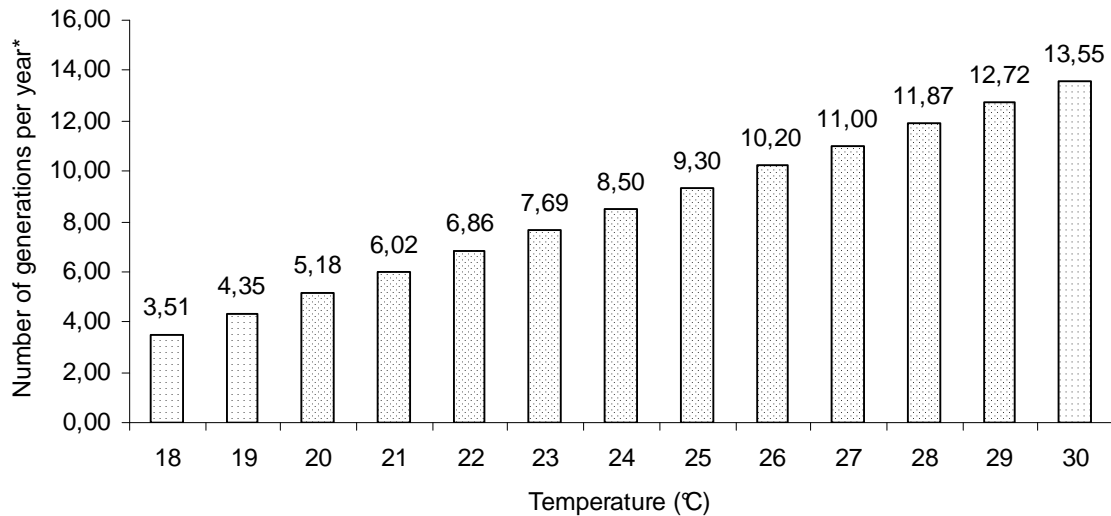


Figure 2 - Number of *Utetheisa ornatrix* generations in regions with same isotherms for the appropriate temperature ranges for insect development. Obtained with the formula $K = D(T - T_b)$, where $K = 436.28\text{GDD}$; $T = \text{variable}$, 18 to 30°C; $T_b = 13.8^\circ\text{C}$ and $D = \text{variable}$ to be determined. The number of generations per year was obtained by dividing 365 days by D .

ACKNOWLEDGMENTS

We thank Prof. Marinéia de Lara Haddad for help with the data analysis, Neide Graciano Zério, for help with insect rearing and to Arione da Silva Pereira and Maria do Carmo Bassols Raseira for revision of the English.

RESUMO

Estudou-se a biologia de *Utetheisa ornatrix* em dieta artificial, visando a determinação das exigências térmicas dos diferentes estágios de desenvolvimento, numa tentativa de avaliar as regiões do estado de São Paulo em que a praga tem maiores condições de se tornar problema para *Crotalaria* spp. Foram criados insetos em dieta artificial à base de feijão e levedura de cerveja. Para a determinação das exigências térmicas de *U. ornatrix* foram utilizadas as temperaturas de 18, 20, 22, 25, 28, 30 e 32°C, umidade relativa de $70 \pm 20\%$ e fotofase de 14 horas. Os limiares térmicos inferiores de desenvolvimento (T_b) e as constantes térmicas (K) para os estágios de ovo, lagarta e pupa foram de 12,7°C e 51,2 GD, 13,5°C e 290,9 GD, 13,8°C e 108,4 GD, respectivamente,

resultando em valores de T_b e K para o ciclo biológico (ovo-adulto) de 13,8°C e 436,3 GD. O inseto pode dar até 10 gerações anuais nas regiões em que *Crotalaria* spp. é mais importante para o estado de SP.

REFERENCES

- Dyar, H.G. (1890), The number of molts of lepidopterous larvae. *Psyche*, **5**, 420-433.
- Castro, C. M.; Almeida, D. L.; Ribeiro, R. L. D. and Carvalho, J. F. (2005), Plantio direto, adubação verde e suplementação com esterco de aves na produção orgânica de berinjela, *Pesq. Agropec. Bras.*, **40**, 495-502.
- Conner, W. E.; Roach, B.; Benedict, E.; Meinwald, J. and Eisner, T. (1990), Courtship pheromone production and body size as correlates of larval diet in males of the arctiid moth, *Utetheisa ornatrix*, *J. Chem. Ecol.*, **16**, 543-552.
- Ferro, V. G. (2001), Padrões de utilização de *Crotalaria* spp. (Leguminosae, Papilionoidea, Crotalariae) por larvas de *Utetheisa ornatrix* (Lepidoptera, Arctiidae). Dissertação de Mestrado, Universidade Estadual de Campinas, Campinas-SP, Brasil.

- Gallo, D.; Nakano, O.; Silveira Neto, S.; Carvalho, R. P. L.; Batista, G. C.; Berti Filho, E.; Parra, J. R. P.; Zucchi, R. A.; Alves, S. B.; Vendramim, J. D.; Marchini, L. C.; Lopes, J. R. S. and Omoto, C. (2002), *Entomologia Agrícola*, Piracicaba: FEALQ, pp. 920, (Biblioteca de Ciências Agrárias Luiz de Queiroz, 10).
- Haddad, M. L.; Parra, J. R. P. and Moraes, R. C. B. (1999), Métodos para estimar os limites térmico inferior e superior de desenvolvimento de insetos. Piracicaba: FEALQ, pp. 29
- Honék, A. (1996), Geographical variation in thermal requirements for insect development. *Eur. J. Entomol.*, **93**, 303-312.
- Johnson, D. F.; Molyneux, R. J. and Merrill, G. B. (1985), Chemistry of toxic range plants. Variations in pyrrolizidine alkaloid content of *Senecio*, *Amsinckia*, and *Crotalaria* species. *J. Agric. Food Chem.*, **33**, 50-55.
- Mihsfeldt, L. H. (1998), *Biologia e exigências térmicas de Tuta absoluta* (Meyrick, 1917) em dieta artificial. Tese de Doutorado, Escola Superior de Agricultura "Luiz de Queiroz", Piracicaba-SP, Brasil.
- Nava, D. E. and Parra, J. R. P. (2003), Biology of *Cerotoma arcuatus* (Coleoptera: Chrysomelidae) and field validation of a laboratory model for temperature requirements. *J. Econ. Entomol.*, **96**, 609-614.
- Nogueira, F. D.; Paula, M. B.; Gontijo, P. T. G. and Tanaka, T. (1992), Adubação verde, fosfato natural e gesso para a cultura da mandioca em latossolo roxo textura argilosa. *Pesp. Agropec. Bras.*, **27**, 357-372.
- Parra, J. R. P. (2000), O controle biológico e o manejo de pragas: passado, presente e futuro, In: Guedes, J. V. C., Costa I. D. and Castiglioni E. Bases e técnicas do manejo de insetos. Santa Maria: UFSM/CCR/DFS, pp. 59-69.
- Parra, J. R. P. (2001), Técnicas de criação de insetos para programa de controle biológico. 6º ed. Piracicaba: FEALQ, pp. 134.
- Parra, J. R. P. and Haddad, M. L. (1989), Determinação do número de ínstares de insetos. Piracicaba: FEALQ, pp 49.
- Parra, J. R. P.; Botelho, P. S. M; Corrêa-Ferreira, B. S. and Bento, J. M. S. (2002), Controle biológico no Brasil. São Paulo, Manole, pp. 609
- Pease, R. W. (1968). Evolution and hybridization in the *Utetheisa ornatrix* complex (Lepidoptera: Arctiidae). I. Inter and intrapopulation variation and its relation to hybridization. *Evolution*, **22**, 719-735.
- Rosa, R. C.; Moura, R. M.; Pedrosa, E. M. R. (2004), Efeitos do uso de *Crotalaria juncea* e carbofuran em fitonematóides ectoparasitos de cana-de-açúcar. *Fit. Brasil.*, **29**, 447-449.
- Silva, A. G. d'A.; Gonçalves, C. R.; Galvão, D. M.; Gonçalves, A. J. L.; Gomes, J.; Silva, M.N. and Simoni, L. (1968), Quarto catálogo dos insetos que vivem nas plantas do Brasil; seus parasitos e predadores. Tomo 1, pt. 2. Rio de Janeiro, Min. Agric, pp. 622.
- Silveira Neto, S.; Nakano, O.; Barbin, D. and Villa Nova, N. A. (1976), Manual de ecologia dos insetos. Piracicaba: Agronômica Ceres, pp. 419.
- Singh, P. (1977), Artificial diets for insects, mites, and spiders. New: York: Plenum, pp. 594.
- Singh, P. (1983). A general purpose laboratory diet mixture for rearing insects. *Insect Sci. Appl.*, **4**, 357-362.
- Zeiss, M. R.; Koehler, K. J. and Pedigo, L. P. (1996), Degree-day requirements for development of the bean leaf beetle (Coleoptera: Chrysomelidae) under two rearing regimes. *J. Econ. Entomol.*, **89**, 111-118.

Received: February 17, 2006;
Revised: August 09, 2008;
Accepted: May 05, 2008.