

Comparison between Winkler's extractor and pitfall traps to estimate leaf litter ants richness (Formicidae) at a rainforest site in southeast Brazil

Orsolon-Souza, G.^{a*}, Esbérard, CEL.^b, Mayhé-Nunes, AJ.^b, Vargas, AB.^c, Veiga-Ferreira, S.^a, and Folly-Ramos, E.^d

^aPrograma de Pós-graduação em Biologia Animal, Instituto de Biologia, Universidade Federal Rural do Rio de Janeiro – UFRRJ, Rod. BR 465, Km 7, CEP 23890-000, Seropédica, RJ, Brazil

^bDepartamento de Biologia Animal, Instituto de Biologia, Universidade Federal Rural do Rio de Janeiro – UFRRJ

^cPrograma de Pós-graduação em Ciências Ambientais e Florestais, Instituto de Florestas, Universidade Federal Rural do Rio de Janeiro – UFRRJ

^dDepartamento de Engenharia e Meio Ambiente, Universidade Federal da Paraíba – UFPB, R. Manoel Gonçalves, s/n, CEP 58297-000, Rio Tinto, PB, Brazil

*e-mail: guilhermeorsolon@yahoo.com.br

Received July 19, 2010 – Accepted November 11, 2010 – Distributed November 30, 2011

(With 2 figures)

Abstract

The aim of this study was to compare in the same site the efficiency of the two most used techniques for sampling ant diversity, Winkler's extractors and pitfalls. We studied communities of leaf litter ants from the Brazilian Atlantic Forest, compared richness estimates for genera and species, and built species accumulation curves. These two methods resulted in a satisfactory sampling of richness; 21.3% of the genera and 47.6% of the species were collected exclusively with Winkler's extractors, whereas 6.4% of the genera and 9.5% of the species were collected exclusively with pitfalls. Winkler's extractor had proven to be the most efficient single sampling technique to estimate richness. However, pitfalls also recorded a significant portion of the total richness. Despite differences in efficiency, species accumulation curves for both techniques were similar, as well as the curve obtained with both methods combined. We noticed that Winkler's extractors were c. 74.0% more efficient than pitfalls in the Atlantic Forest. Therefore, sampling techniques must be used with a well-structured sampling design in order to advance knowledge on the ant fauna of Brazilian biomes, especially in the leaf litter, allowing more complete environmental analyses.

Keywords: Atlantic rainforest, sampling techniques, species accumulation curves.

Comparaç o entre extrator de Winkler e armadilha de queda para estimar a riqueza em esp cies de formigas (Formicidae) de serapilheira em floresta tropical do sudeste do Brasil

Resumo

O objetivo deste estudo foi comparar, em um mesmo ambiente, a efici ncia das duas t cnicas de coletas mais utilizadas para amostrar a diversidade de formigas, o extrator de Winkler e a armadilha de queda (*pitfall traps*). Para a comunidade de formigas de serapilheira da Mata Atl ntica, comparamos as estimativas de riqueza de g neros, de esp cies e as curvas de acumula o de esp cies. Os dois m todos resultaram em uma amostragem de riqueza satisfat ria, sendo 21,3 e 47,6% exclusivamente g neros e esp cies coletadas pelo extrator de Winkler e 6,4 e 9,5% pela armadilha de queda. O extrator de Winkler mostrou ser a t cnica mais eficiente para amostrar a riqueza de uma  rea quando apenas uma das t cnicas possa ser utilizada. No entanto, o uso da armadilha de queda registrou uma parcela n o t o desprez vel da riqueza. Quanto   efici ncia das t cnicas de coleta, as curvas de acumula o de esp cies para os duas t cnicas mostraram-se similares com a obtida com os dois m todos simultaneamente. Notou-se, portanto, que o extrator de Winkler apresenta uma efici ncia de amostragem da riqueza na ordem de 74,0% maior que a armadilha de queda para ambiente de Mata Atl ntica. Vale ressaltar que as t cnicas de coleta devem estar associadas a um delineamento amostral bem estruturado para que se possa ampliar ainda mais o conhecimento sobre a mirmecofauna dos biomas brasileiros, principalmente daquelas com serapilheira, permitindo an lises ambientais mais completas.

Palavras-chave: Mata Atl ntica, t cnicas de coleta, curva de acumula o de esp cies.

1. Introduction

Species richness is a central topic in community ecology (Longino et al., 2002) and a fundamental component of biodiversity (Gaston, 1996). However, it is difficult to estimate the species richness of a given area, since the available techniques do not always provide a representative sample of the total richness (Agosti et al., 2000; Agosti and Alonso, 2001).

Studies on communities of leaf litter arthropods (e.g. Romero and Jaffé, 1989; Parr and Chown, 2001) point out the need for well-structured sampling protocols to properly estimate richness and evenness. In the case of ants, foraging area, nest dispersion, and activity patterns are important factors that must be considered in a sampling design (Wang et al., 2001). Researchers of ant communities suggest the combination of different techniques as the best way to estimate richness and abundance of ants (Delabie et al., 2000; Longino et al., 2002).

Pitfall traps, Winkler's extractors, food baits and hand collecting are the most efficient techniques, and, hence, the most frequently used in ant community studies. Both Winkler's extractors and pitfalls have proven to be efficient in sampling abundance and richness of leaf litter ant communities (Romero and Jaffé, 1989; Wang et al., 2001). However, Parr and Chown (2001) compared both techniques and suggested that species sampled with Winkler's extractors are more abundant and smaller than those sampled with pitfalls.

It is also possible to use data obtained with both techniques to estimate species richness. Species accumulation curves allow estimating sample completeness (Colwell and Coddigton, 1994), assessing the efficiency of the method used, and comparing different inventories (Soberón and Llorente, 1993).

Therefore, it is important to compare in the same site the efficiency of the two most used techniques for sampling ant diversity. We studied communities of leaf litter ants from the Brazilian Atlantic Forest, compared richness estimates for genera and species, and built species accumulation curves with data from pitfall traps and Winkler's extractors.

2. Materials and Methods

2.1. Study area

This study was carried out in Dense Ombrophylous Forest (IBAMA, 1996), in the "Reserva Biológica do Tinguá" (ReBio Tinguá, 22° 28' -22° 39' S and 43° 13' -43° 34' W), Nova Iguaçu municipality, state of Rio de Janeiro, in November 2003. Sampling was carried out in submontane rainforest (Veloso et al., 1991) at 400 m asl on average.

2.1.1. Characterisation of submontane rainforest

Submontane rainforest presents canopy height varying from 15 to 20 m on average, discontinuous in some places and where Rubiaceae, Mimosoideae, Myrtaceae and Lauraceae are the most speciose families (Rodrigues, 1996). Understory is relatively dense with high frequency of small trees and bushes. Herbaceous climbers and lianas are common, particularly at the forest edge. Some epiphytes like Orchidaceae, Bromeliaceae, Araceae and Cactaceae species are relatively frequent close to streams (IPJBRJ, 2002) (Figure 1).

2.2. Sampling

Winkler's Extractors (Wke): We marked 25 points along a 1,200 m transect, and at each point we stretched two perpendicular 25 m lines; one to the left and the other to the right. At the end of each line we delimited a 1 m² plot, totalling 50 plots. Sifted samples from each plot were separated and placed into Winkler's extractors for 48 hours. This procedure was adapted from Delabie et al. (2000).

Pitfall traps (Pft): We use 50 pitfalls that were distributed along the previously described transect, between the plots with Winkler's extractors. Pitfalls consisted of 300 mL plastic glasses with a diameter of 7.0 cm, containing 3% formalin. Pitfalls were buried in the soil with the upper border parallel to the ground, and remained open for 48 hours.

2.3. Identification of ants

Ant genera were identified following Bolton (1994), and subfamilies following Bolton (2003). Whenever



Figure 1. Partial vision of Dense Ombrophylous Forest in "Reserva Biológica do Tinguá". a) The circle shows local study in submontane rainforest; b) Understory relatively dense with high frequency of small trees and bushes; c) Discontinuous canopy in some places is noted. The arrows indicate transects. November/2003.

possible, identification at species level was carried out with keys from taxonomical reviews or by comparison with identified specimens from the Entomological Collection Ângelo Moreira da Costa Lima (CECL), at the Institute of Biology of the Universidade Federal Rural do Rio de Janeiro, Brazil. Vouchers were deposited in the same collection.

2.4. Data analysis

Species accumulation curves were built using the number of samples (Soberón and Llorente, 1993). According to Moreno and Halffter (2000) and Medellín (1993), this method is better for sampling units that grow linearly.

For each technique, the total of species per sample was randomised 1,000 times in the program EcoSim® (Gotelli and Entsminger, 2009) to estimate confidence intervals (adapted from Lourenço et al., 2010). Accumulation curves were built from observed and randomised data. Logarithmic functions for the randomised curves of each technique were also presented.

3. Results and Discussion

In total, we collected ants from ten subfamilies, 37 genera and 105 species (Table 1). Those two methods resulted in a satisfactory sampling of richness; 21.3% of the genera and 47.6% of the species were collected exclusively with Winkler's extractors, whereas 6.4% of the

genera and 9.5% of the species were collected exclusively with pitfalls (Table 2).

Myrmicinae was the most frequent ant subfamily sampled with both techniques, and this result is probably related to the high richness and abundance of this group in the leaf litter fauna of the Neotropics (Silva and Brandão, 1999). Veiga-Ferreira et al. (2005) and Vargas et al. (2007) recorded a high number of species of Myrmicinae in the Atlantic Forest and the Restinga of Rio de Janeiro, respectively. For each of the subfamilies Cerapachyinae and Ecitoninae, only one species was sampled with Winkler's extractors and pitfalls. This result is probably related to habitat conditions that can affect the structure of the ant community (Andersen, 2000; Hölldobler and Wilson, 1990; Nakamura et al., 2003), and consequently the frequency of each species. According to Lassau and Hochuli (2004), the structural complexity of the environment directly affects species richness.

Although pitfall traps tend to capture larger ants (Olson, 1991), such as the specimens of Ectatomminae and Ponerinae recorded in the present study, they can also sample abundance and richness of the leaf litter ant community (Romero and Jaffé, 1989; Wang et al., 2001). Winkler's extractors captured smaller and cryptic leaf litter ants, such as the Myrmicinae, or even ants that are rarely collected with other techniques (Bestelmeyer et al., 2000; Castilho et al., 2007). For the Atlantic Forest,

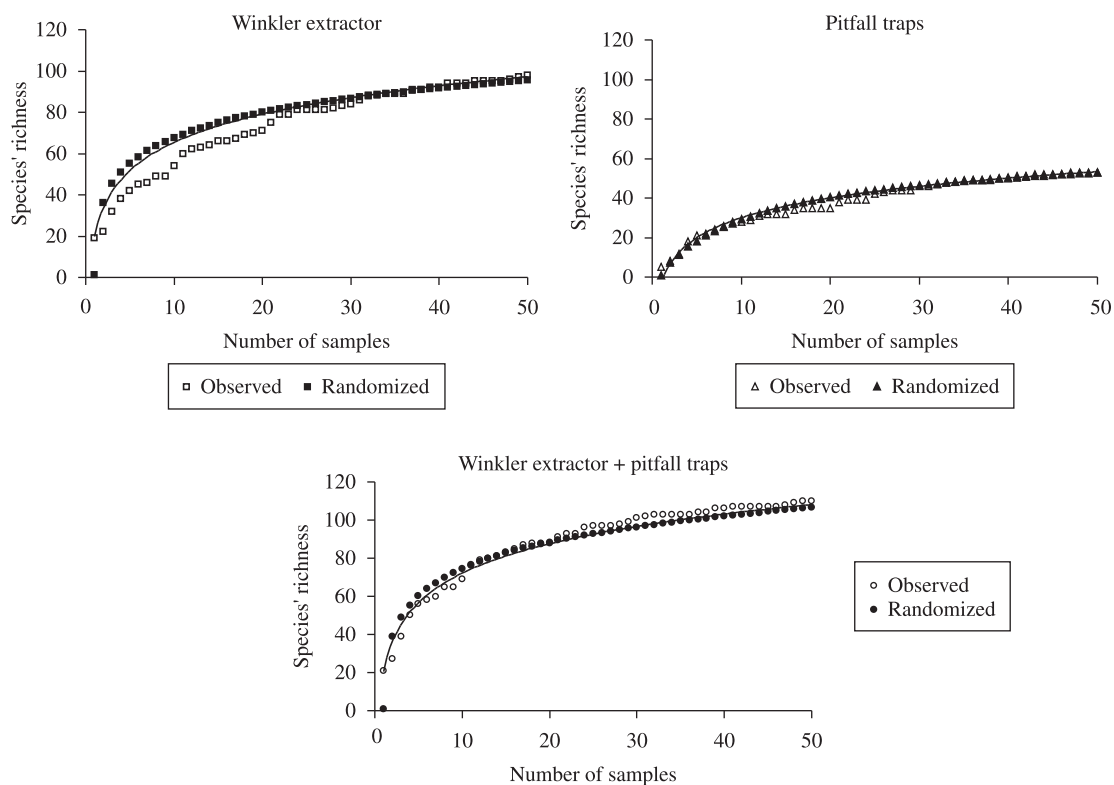


Figure 2. Species accumulation curve observed and using randomised samples. a) Winkler's extractor; b) Pitfall traps; c) Winkler's extractor and Pitfall traps concomitantly. "Rebio Tingua", Brazil, November 2003.

Table 1. List of species collected by Winkler's extractor and pitfall traps in submontane rainforest in "Reserva Biológica do Tinguá", Rio de Janeiro state, Brazil. November/2003.

Subfamilies	Species/Morphospecies	Winkler's extractor	Pitfall traps	
Amblyoponinae	<i>Amblyopone armigera</i> Mayr, 1897	#	#	
	<i>Amblyopone elongata</i> (Santschi, 1912)	#	#	
	<i>Prionopelta punctulata</i> Mayr, 1866	#	-	
Ceraphachyinae	<i>Cerapachys splendens</i> Borgmeier, 1957	#	-	
Dolichoderinae	<i>Azteca</i> sp.1	-	#	
	<i>Linepithema pulex</i> Wild, 2007	#	-	
	<i>Linepithema</i> sp.1	#	#	
Ecitoninae	<i>Labidus</i> sp.1	-	#	
Ectatominae	<i>Ectatomma permagnum</i> Forel, 1908	#	-	
	<i>Ectatomma edentatum</i> Roger, 1863	#	-	
	<i>Ectatomma brunneum</i> F. Smith, 1858	-	#	
	<i>Gnamptogenys cf. horni</i>	#	-	
	<i>Gnamptogenys horni</i> Santschi, 1929	#	#	
	<i>Gnamptogenys rastrata</i> (Mayr, 1866)	#	#	
	<i>Gnamptogenys cf. porcata</i>	#	-	
	<i>Gnamptogenys cf. lunaris</i>	#	-	
	<i>Gnamptogenys</i> sp.1	-	#	
Formicinae	<i>Brachymyrmex</i> sp.1	#	-	
	<i>Brachymyrmex</i> sp.2	#	#	
	<i>Brachymyrmex</i> sp.3	#	#	
	<i>Camponotus novogranadensis</i> Mayr, 1870	#	-	
	<i>Camponotus punctulatus</i> Mayr, 1868	#	-	
	<i>Camponotus</i> sp.1	#	-	
	<i>Camponotus</i> sp.2	-	#	
Myrmicinae	<i>Acromyrmex aspersus</i> (F. Smith, 1858)	#	-	
	<i>Acromyrmex coronatus</i> (Fabricius, 1804)	#	-	
	<i>Acromyrmex niger</i> (F. Smith, 1858)	#	-	
	<i>Apterostigma</i> sp.1	#	#	
	<i>Apterostigma</i> sp.2	#	#	
	<i>Cyphomyrmex</i> gr. <i>strigatus</i> sp.1	#	#	
	<i>Cyphomyrmex</i> gr. <i>strigatus</i> sp.2	#	#	
	<i>Cyphomyrmex</i> gr. <i>rimnosus</i> sp.1	#	-	
	<i>Mycocepurus smithii</i> Forel, 1893	#	#	
	<i>Mycetarotes carinatus</i> Mayh�-Nunes, 1995	#	-	
	<i>Sericomyrmex</i> sp.1	#	#	
	<i>Sericomyrmex</i> sp.2	#	#	
	<i>Sericomyrmex</i> sp.3	#	#	
	<i>Trachymyrmex</i> sp.1	#	#	
	Myrmicinae	<i>Basiceros bruchi</i> Santschi, 1922	#	-
		Non-Attini	<i>Basiceros disciger</i> (Mayr, 1887)	#
	<i>Basiceros iheringi</i> (Emery, 1887)		#	-
<i>Basiceros rugiferum</i> (Mayr, 1887)	#		#	
<i>Carebara urichi</i> (Wheeler, 1922)	#		#	
<i>Crematogaster nigropilosa</i> Mayr, 1870	#		-	
<i>Hylomyrma balzani</i> (Emery, 1894)	#		-	
<i>Hylomyrma reitteri</i> (Mayr, 1887)	#		#	

= collected by the technique; - = non-collected by the technique.

Table 1. Continued...

Subfamilies	Species/Morphospecies	Winkler's extractor	Pitfall traps
	<i>Lachnomyrmex plaumanni</i> Borgmeier, 1957	#	-
	<i>Megalomyrmex drifti</i> Kempf, 1961	#	#
	<i>Megalomyrmex goeldii</i> Forel, 1912	#	#
	<i>Megalomyrmex silvestrii</i> Wheeler, 1909	#	-
	<i>Oxyopocus</i> sp.1	#	#
	<i>Oxyopocus</i> sp.2	#	-
	<i>Pheidole</i> sp.1	#	#
	<i>Pheidole</i> sp.2	#	#
	<i>Pheidole</i> sp.3	#	#
	<i>Pheidole</i> sp.4	#	#
	<i>Pheidole</i> sp.5	#	#
	<i>Pheidole</i> sp.6	#	#
	<i>Pheidole</i> sp.7	#	#
	<i>Pheidole</i> sp.8	#	#
	<i>Pheidole</i> sp.9	#	-
	<i>Pheidole</i> sp.10	#	-
	<i>Pyramica</i> sp.1	#	#
	<i>Pyramica</i> sp.2	#	#
	<i>Pyramica</i> sp.3	#	-
	<i>Pyramica</i> sp.4	-	#
	<i>Rogeria</i> sp.1	#	-
	<i>Rogeria</i> sp.2	#	-
	<i>Rogeria</i> sp.3	#	-
	<i>Solenopsis</i> sp.1	#	#
	<i>Solenopsis</i> sp.2	#	#
	<i>Solenopsis</i> sp.3	#	-
	<i>Solenopsis</i> sp.4	#	#
	<i>Solenopsis</i> sp.5	#	-
	<i>Solenopsis</i> sp.6	#	#
	<i>Solenopsis</i> sp.7	#	-
	<i>Strumigenys elongata</i> Roger, 1863	#	-
	<i>Wasmannia auropunctata</i> (Roger, 1863)	#	#
	<i>Wasmannia lutzi</i> Forel 1908	#	#
Ponerinae	<i>Anochetus mayri</i> Emery, 1884	#	-
	<i>Centromyrmex</i> sp.1	-	#
	<i>Hypoponera</i> sp.1	#	#
	<i>Hypoponera</i> sp.2	#	-
	<i>Hypoponera</i> sp.3	#	-
	<i>Hypoponera</i> sp.4	#	-
	<i>Hypoponera</i> sp.5	#	-
	<i>Hypoponera</i> sp.6	#	-
	<i>Leptogenys</i> sp.1	#	-
	<i>Odontomachus chelififer</i> (Latreille, 1802)	#	#
	<i>Odontomachus haematodus</i> (Linnaeus, 1758)	-	#
	<i>Odontomachus meinerti</i> Forel, 1905	#	-
	<i>Odontomachus</i> sp.1	#	-
	<i>Pachycondyla bucki</i> (Borgmeier, 1928)	#	-

= collected by the technique; - = non-collected by the technique.

Table 1. Continued...

Subfamilies	Species/Morphospecies	Winkler's extractor	Pitfall traps
	<i>Pachycondyla ferruginea</i> (F. Smith, 1858)	#	-
	<i>Pachycondyla harpax</i> (Fabricius, 1804)	#	#
	<i>Pachycondyla stigma</i> (Fabricius, 1804)	#	#
	<i>Pachycondyla striata</i> F. Smith, 1858	#	#
	<i>Pachycondyla venusta</i> Forel, 1912	#	#
	<i>Thaumatomyrmex mutilatus</i> Mayr, 1887	#	-
Proceratiinae	<i>Discothyrea sexarticulata</i> Borgmeier 1954	#	#
	<i>Proceratium brasiliense</i> Borgmeier, 1959	#	-
Pseudomyrmecinae	<i>Pseudomyrmex</i> sp.1	#	-
	<i>Pseudomyrmex</i> sp.2	#	-
	<i>Pseudomyrmex</i> sp.3	-	#
	<i>Pseudomyrmex</i> sp.4	-	#

= collected by the technique; - = non-collected by the technique.

Table 2. Distribution of genera and species richness of each Formicidae subfamilies sampled by Winkler's extractor (Wke) and pitfall traps (Pft) at "Rebio Tingua", Brazil, November/2003.

Subfamily	Genera			Species		
	Wke	Pft	Common both techniques	Wke	Pft	Common both techniques
Amblyoponinae	2 (1)	1 (-)	1	3 (1)	2 (-)	2
Cerapachyinae	1 (1)	-	-	1 (1)	-	-
Dolichoderinae	1 (-)	2 (1)	1	2 (1)	2 (1)	1
Ecitoninae	-	1 (1)	-	-	1 (1)	-
Ectatomminae	2 (-)	2 (-)	2	7 (5)	4 (2)	2
Formicinae	2 (-)	2 (-)	2	6 (4)	3 (1)	2
Myrmicinae	20 (4)	9 (-)	14	54 (23)	32 (1)	31
Ponerinae	6 (3)	4 (1)	3	20 (12)	8 (2)	6
Proceratiinae	2 (1)	1 (-)	-	2 (1)	1 (-)	1
Pseudomyrmecinae	1 (-)	1 (-)	1	2 (2)	2 (2)	-
Total	37 (10)	23 (3)	24	97 (50)	55 (10)	45

Exclusive genera and species for each technique are within parenthesis.

Winkler's extractor had proven to be the most efficient single sampling technique to estimate richness. However, pitfalls also recorded a significant portion of the total richness. Lassau and Hochuli (2004) collected ants using only pitfalls in complex environments, and, although those environments were different from the one studied here, ant richness was higher in environments with more complex structures. Therefore, pitfalls must be used together with other sampling techniques in the field, especially in tropical forests.

On the one hand, our results are consistent with Parr and Chown (2001), who, when comparing sampling techniques, suggested that species collected with Winkler's extractors are more abundant and smaller than the ones collected in pitfalls. On the other hand, Olson (1991) showed that Winkler's extractors and pitfalls together can sample c. 75.0% of the species richness estimated for the

leaf litter ant fauna, a percentage considered reasonable for ecological studies on leaf litter.

Despite differences in efficiency, species accumulation curves for both techniques were similar ($y = 19.743 \ln(x) + 19.962$; $r^2 = 0.9688$; $p < 0.001$ for Wke, $y = 14.487 \ln(x) - 3.348$; $r^2 = 0.995$; $p < 0.001$ for Pft), as well as the curve obtained with both methods combined ($y = 22.378 \ln(x) + 20.575$; $r^2 = 0.9739$; $p < 0.001$ for Wke + Pft) (Figure 2). To obtain the total of 105 species recorded with both techniques combined, it would be necessary to take 64.07 samples with Wke and 111.84 with Pft. Therefore, we noticed that Winkler's extractors were c. 74.0% more efficient than pitfalls in the Atlantic Forest, corroborating results from Parr and Chown (2001).

Sampling techniques must be used with a well-structured sampling design in order to advance knowledge on the

ant fauna of Brazilian biomes, especially in the leaf litter (Castilho et al. (2007) and Veiga-Ferreira et al. (2010) for new records of ant's species), allowing more complete environmental analyses. The influence of ants on edaphic processes, flux of energy and matter in ecosystems (Brown, 1997), predation, and seed dispersal (Folgarait, 1998; Passos and Oliveira, 2002, 2003; Leal, 2003) makes this insect group a good study model for environmental surveys and monitoring (Silva and Brandão, 1999).

Acknowledgements – We are grateful to the team at the Reserva Biológica do Tinguá (Nova Iguaçu office) and to IBAMA for the study license. This study was supported by grants from the Brazilian governmental to GOS (CAPES), to AJMN and CELS (CNPq).

References

- ANDERSEN, AN., 2000. A global ecology of rain forest ants: Functional groups in relation to environmental stress and disturbance. In AGOSTI, D., MAJER, JD., ALONSO, LE. and SCHULTZ, TR. (Eds.). *Ants: Standard methods for measuring and monitoring biodiversity*. Washington: Smithsonian Institutions Press. p. 25-34
- AGOSTI, D. and ALONSO, LE., 2001. The all Protocol: a standard protocol for the collection of ground-dwelling ants. *Anet Newsletter*, vol. 3, p. 8-11.
- AGOSTI, D., MAJER, JD., ALONSO, LE. and SCHULTZ, TR. (Eds.), 2000. *Ants: standard (Hymenoptera: Formicidae) methods for measuring and monitoring biodiversity*. Washington: Smithsonian Institutions Press. 280 p.
- BESTELMEYER, BT., AGOSTI, D., ALONSO, LE., BRANDÃO, CRF., BROWN JUNIOR, WL., DELABIE, JHC. and SILVESTRE, R., 2000. Field techniques for the study of ground-living ants: An overview, description, and evaluation. In AGOSTI, D., MAJER, JD., ALONSO, LT. and SCHULTZ, TR. (Eds.). *Ants: Standard methods for measuring and monitoring biodiversity*. Washington: Smithsonian Institutions Press. p.122-144
- BOLTON, B., 1994. *Identification Guide to the Ant Genera of the World*. London: Harvard University Press. 222 p.
- , 2003. Synopsis and classification of Formicidae. *Memoirs of the American Entomological Institute*, vol. 71, p. 1-370.
- BRAZ, DM., MOURA, MVLP. and ROSA, MMT., 2004. Chave de identificação para as espécies de Dicotiledôneas arbóreas da Reserva Biológica do Tinguá, RJ, com base em caracteres vegetativos. *Acta Botanica Brasílica*, vol. 18, no. 2, p. 225-250.
- BROWN, KS., 1997. Diversity, disturbance and sustainable use of Neotropical forests: insects as indicators for conservation monitoring. *Journal of Insect Conservation*, vol. 1, no. 1, p. 1-18. <http://dx.doi.org/10.1023/A:1018432508944>
- CASTILHO, ACC., DELABIE, JHC., MARQUES, MI., ADIS, J. and MENDES, LF., 2007. Registros Novos da Formiga Criptobiótica *Creightonidris scambognatha* Brown (Hymenoptera: Formicidae). *Neotropical Entomology*, vol. 36, no. 1, p. 150-152. PMID:17420875. <http://dx.doi.org/10.1590/S1519-566X2007000100020>
- COLWELL, RK. and CODDINGTON, JA., 1994. Estimating terrestrial biodiversity through extrapolation. *Philosophical Transactions of the Royal Society of London, Series B*, vol. 345, p. 101-118. PMID:7972351. <http://dx.doi.org/10.1098/rstb.1994.0091>
- DELABIE, JHC., AGOSTI, D. and NACIMEJNTO, IC., 2000. Litter ant communities of the Brazilian Atlantic rain forest region. In AGOSTI, D., MAJER, JD., ALONSO, LT. and SCHULTZ, TR. (Eds.). *Sampling ground-dwelling ants: case studies from the world's rain forests*. Perth, Australia: Curtin University School of Environmental Biology. p. 1-17. (Bulletin, vol. 18).
- FOLGARAIT, PJ., 1998. Ant biodiversity and its relationship to ecosystem functioning: a review. *Biodiversity and Conservation*, vol. 7, p. 1221-1244. <http://dx.doi.org/10.1023/A:1008891901953>
- GASTON, KJ., 1996. Species richness: measure and measurement. In GASTON, KL. (Ed.). *Biodiversity: a biology of numbers and difference*. Cambridge: Blackwell Science. p. 77-113.
- GOTELLI, N.J. and ENTSMINGER, GL., 2009. *EcoSim*: Null models software for ecology. Version 7. Jericho: Acquired Intelligence Inc. & Kesey-Bear. Available from: <<http://garyentsminger.com/ecosim.htm>>. Access in: 05 jul. 2010.
- HÖLDOBLER, B. and WILSON, EO., 1990. *The Ants*. Cambridge: The Belknap Press of Harvard University. 733 p.
- Instituto Brasileiro do Meio Ambiente e dos Recursos Naturais Renováveis - IBAMA, 1996. *Plano de ação emergencial - Reserva Biológica do Tinguá*. Brasília: IBAMA, 114 p.
- Instituto de Pesquisas Jardim Botânico do Rio de Janeiro, 2002. *Projeto Tinguá: Paisagem e flora da reserva biológica do Tinguá: subsídios ao monitoramento da vegetação*. Rio de Janeiro: Instituto de Pesquisas Jardim Botânico do Rio de Janeiro, Universidade Federal Rural do Rio de Janeiro. 55 p. Relatório técnico-científico.
- LASSAU, SA. and HOCHULI, DF., 2004. Effects of habitat complexity on ant assemblages. *Ecography*, vol. 27, no. 2, p. 157-164.
- LEAL, IR., 2003. Dispersão de Sementes por Formigas. In LEAL, IR., TABARELLI, M. and SILVA, JMC. *Ecologia e Conservação da Caatinga*. Recife: Ed. Univ. UFPE. p. 593-624.
- LONGINO, JT., CODDINGTON, J. and COLWELL, RK., 2002. The ant fauna of a tropical rain forest: estimating species richness three different ways. *Ecology*, vol. 83, no. 3, p. 689-702. [http://dx.doi.org/10.1890/0012-9658\(2002\)083\[0689:TAF0AT\]2.0.CO;2](http://dx.doi.org/10.1890/0012-9658(2002)083[0689:TAF0AT]2.0.CO;2)
- LOURENÇO, EC., COSTA, LM., SILVA, RM. and ESBÉRARD, CEL., 2010. Bat diversity of Ilha da Marambaia, Southern Rio de Janeiro State, Brazil (Chiroptera, Mammalia). *Brazilian Journal of Biology*, vol. 70, no. 3, p. 511-519. <http://dx.doi.org/10.1590/S1519-69842010000300007>
- MEDELLÍN, RA., 1993. Estructura y diversidad de una comunidad de murciélagos en el trópico húmedo mexicano. In MEDELLÍN, RA. and CEBALLOS, TG. (Eds.). *Avances en el estudio de los mamíferos de México*. México: Asociación Mexicana de Mastozoología. vol. 1, p 333-354.
- MORENO, CE. and HALFFTER, G., 2000. Assessing the completeness of bat biodiversity inventories using species accumulation curves. *Journal of Applied Ecology*, vol. 37, p. 149-158. <http://dx.doi.org/10.1046/j.1365-2664.2000.00483.x>
- NAKAMURA, A., PROCTOR, H. and CATTERALL, CP., 2003. Using soil and litter arthropods to assess the state of rainforest restoration. *Ecological Management & Restoration*, vol. 4, p. 20-28. PMID:17923455. <http://dx.doi.org/10.1046/j.1442-8903.4.s.3.x>
- OLSON, DM., 1991. A comparison of the efficacy of litter sifting and pitfall traps for sampling leaf litter ants (Hymenoptera:

- Formicidae) in a Tropical wet forest, Costa Rica. *Biotropica*, vol. 23, no. 2, p. 166-172.
- PARR, CL. and CHOWN, SL., 2001. Inventory and bioindicator sampling: testing pitfall and Winkler methods with ants in a South African savanna. *Journal of Insect Conservation*, vol. 5, p. 27-36. <http://dx.doi.org/10.1023/A:1011311418962>
- PASSOS, L. and OLIVEIRA, PS., 2002. Ants affect the distribution and performance of seedlings of *Clusia criuva*, a primarily bird-dispersed rain forest tree. *Journal of Ecology*, vol. 90, p. 517-528. <http://dx.doi.org/10.1046/j.1365-2745.2002.00687.x>
- PASSOS, L. and OLIVEIRA, PS., 2003. Interactions between ants, fruits and seeds in a restinga forest in south-eastern Brazil. *Journal of Tropical Ecology*, vol. 19, p. 261-270. <http://dx.doi.org/10.1017/S0266467403003298>
- RODRIGUES, HC. 1996. *Composição florística e fitossociológica de um trecho de Mata Atlântica na Reserva Biológica do Tinguá, Nova Iguaçu, Rio de Janeiro*. Rio de Janeiro: Museu Nacional, Universidade Federal do Rio de Janeiro. 77 p. Dissertação de Mestrado.
- ROMERO, H. and JAFFE, K., 1989. A comparison of methods for sampling ants (Hymenoptera:Formicidae) in savannas. *Biotropica*, vol. 21, no. 4, p. 348-352. <http://dx.doi.org/10.2307/2388285>
- SILVA, RR. and BRANDÃO, CRF., 1999. Formigas (Hymenoptera: Formicidae) como indicadores da qualidade ambiental e da biodiversidade de outros invertebrados terrestres. *Biotemas*, vol. 12, no. 2, p. 55-73.
- SOBERÓN, J. and LLORENTE, J., 1993. The use of species accumulation functions for the prediction species richness. *Conservation Biology*, vol. 7, no. 3, p. 480-488. <http://dx.doi.org/10.1046/j.1523-1739.1993.07030480.x>
- VARGAS, AB., MAYHÉ-NUNES, AJ., QUEIROZ, JM., SOUZA, GO. and FOLLY-RAMOS, E., 2007. Efeitos de Fatores Ambientais sobre a Mirmecofauna em Comunidade de Restinga no Rio de Janeiro, RJ. *Neotropical Entomology*, vol. 36, no. 1, p. 28-37. PMID:17420859. <http://dx.doi.org/10.1590/S1519-566X2007000100004>
- VEIGA-FERREIRA, S., MAYHÉ-NUNES, AJ. and QUEIROZ, JM., 2005. Formigas de serapilheira na Reserva Biológica do Tinguá, Estado do Rio de Janeiro, Brasil (Hymenoptera: Formicidae). *Revista Universidade Rural, Série Ciências da Vida*, vol. 25, no. 1, p. 49-54.
- VEIGA-FERREIRA, S., ORSOLON-SOUZA, G. and MAYHÉ-NUNES, AJ., 2010. Hymenoptera, Formicidae Latreille, 1809: New records for Atlantic Forest in the state of Rio de Janeiro. *Check List*, vol. 6, no. 3, p. 442-444.
- VELOSO, HP., RANGEL-FILHO, ALR. and LIMA, JCA., 1991. *Classificação da Vegetação brasileira adaptada a um sistema universal*. Rio de Janeiro: IBGE. 124 p.
- WANG, C., STRAZANAC, J. and BUTLER, L., 2001. A comparison of Pitfall Traps with Bait Traps for studying leaf litter ant communities. *Journal Economic Entomology*, vol. 94, no. 3, p. 761-765. PMID:11425034. <http://dx.doi.org/10.1603/0022-0493-94.3.761>