Brazilian Journal of Chemical Engineering

ISSN 0104-6632 Printed in Brazil www.abeq.org.br/bjche

Vol. 26, No. 02, pp. 299 - 306, April - June, 2009

COMPARATIVE STUDY OF HYDRODYNAMIC BEHAVIOR OF LIQUID EXPANDED BED ADSORPTION: MATHEMATICAL AND SHORT-CUT METHODS

M. Jahanshahi^{1*}, A. A. Ghoreishi¹, E. Vasheghani-F², M. Khavarpour³ and A. Abedijaber⁴

¹Nanobiotechnology Research Lab., Faculty of Chemical Engineering,
Phone/Fax: +98(0)111-3234204, Babol University of Technology, Iran.
E-mail: mmohse@yahoo.com, mjahan@nit.ac.ir

²Department of Chemical Engineering, Faculty of Engineering, Tarbiat Modarres University, Tehran, Iran.

³Islamic Azad University, Department of Chemical Engineering, Ayatollah Amoli Branch, Iran.

⁴Department of Chemical & Biomolecular Engineering, University of Tennessee, Knoxville, USA.

(Submitted: December 18, 2007; Revised: June 30, 2008; Accepted: August 13, 2008)

Abstract - Liquid fluidized/expanded bed adsorption (LF/EBA) is a novel chromatography technique for capturing target biomolecules directly from crude feedstock in downstream processing. To widely extend the LF/EBA technology in biological industries, a better understanding of the expanded-bed behavior is necessary to maximize its efficient applications. In this study, streamline adsorbent was loaded into the column and the bed was subjected to physical and hydrodynamical experiments. Hydrodynamic characteristics of the bed (e.g., D_{axl} and B_o) with different settled bed height (SBH=5,6,7,8,9,10,11,12cm) along with a variety of column diameters (1,1.6,2.5 cm) have been investigated based on exact mathematical as well as approximate short-cut methods. It was found that the increasing column diameter promoted bed mixing, which was undesirable, and the optimal values for the bed expansion and SBH based on minimal liquid mixing were 80%-130% and 8-10 cm, respectively. However, exploiting the short-cut method for the bed hydrodynamic assessments also demonstrated results as good as the mathematical method. Hydrodynamic behavior of liquid fluidized bed adsorption and the generic application of the short-cut method and its potential as a simple method for study of hydrodynamic performance is discussed.

Keywords: Fluidized bed adsorption; Bed mixing; Settled bed height; Bed expansion; Mathematical method; bioseparation.

INTRODUCTION

Generally, standard purification techniques for protein recovery involve the use of a packed bed column after broth clarification, concentration, and filtration steps. The requirement of several sequential operations prior to the initial capturing of proteins increases processing time and cost. The overall yield of purified proteins is also reduced. Therefore, a reduction in the number of purification steps is

beneficial for the biotechnology and pharmaceutical industries. Expanded-bed adsorption is a relatively new technique for the primary recovery of proteins starting from unclarified broths (Amersham, 1998., Jahanshahi et al, 2003., Jahanshahi et al, 2004).

Due to its high bed voidage and good column efficiency, expanded bed adsorption (EBA) has been widely employed to recover target bioproducts directly from cell culture broths, cell disruptates and other unclarified feedstocks (Camprubi et al, 2006.,

^{*}To whom correspondence should be addressed

Clemmitt et al, 2002., Ferreira et al, 2000., Jahanshahi, 2004., Jahanshahi et al, 2005., Jahanshahi et al, 2006., Thommes et al, 1995., Ujam et al, 2003). This process combines clarification, concentration, and initial capturing in a single step. This technique offers the potential advantages of both higher packed bed and fluidized bed. The upward flow through the bed of adsorbents provides the higher void fraction within the bed, which allows the particulate materials to pass through whilst the target bioproduct is adsorbed onto the solid phase (Tong and Sun, 2002).

The hydrodynamic behavior of fluidized beds/expanded beds applied to chromatographic adsorption is different from that of conventional liquid-solid fluidized beds. From the conventional chemical engineering viewpoint, a fluidized bed is one in which there is a significant degree of mixing, in both the solid and fluid phases, e.g., in gasfluidized systems (Levenspiel, 1972). In many applications, mixing of the solid phases is desirable, but the mixing in liquid fluidized systems is not as severe as in gas-fluidized systems. In a packed bed, the adsorbent beads are stationary and liquid flow through the bed approximates to plug flow. Thus, the equilibration of theoretical number (conventionally referred to as plates) is maximized, which results in good adsorption chromatographic performance. It is highly desirable to minimize the degree of mixing in order to mimic the adsorption characteristics found in a packed bed contactor with respect to capacity and resolution (Jahanshahi et al, 2002., Thommes, 1997). Therefore, in order to maximize the adsorption efficiency of EBA, it is necessary to investigate its hydrodynamic behavior properly.

The work herein presents comparative tests of physical and hydrodynamical performance of

LF/EBA. For the purpose of demonstration of the principle, the effect of column diameter, settled bed height and bed expansion are investigated by RTD mathematical method and compared with the result of a short-cut method.

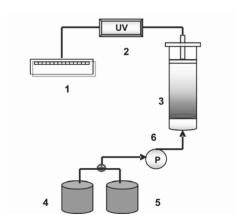
EXPERIMENTAL

Material

Streamline Quartz Base Matrix, adsorbent, was purchased from Amersham Biosciences. StreamlineTM 40 columns of 1-cm i.d were also purchased from Amersham Biosciences. A dilute acetone solution (1% v/v) was used as the input tracer to the column. 10 mM Tris/HCl at pH 7.5 was used to expand the bed. Figure 1 shows the expanded bed setup.

Measurement of the Hydrodynamic Performance

The bed expansion characteristics of the adsorbent were determined in an expanded bed operated with buffer A (10 mM Tris/HCl containing 0.2% (w/v) sodium azide at pH 7.5). The adsorbent was repeatedly washed prior to use with equilibration buffer A and filtered until pH and conductivity reached equilibration values. A given amount of adsorbent particles was transferred to the column (in an Amersham Bioscience contactor; 1 cm inner diameter) and allowed to sediment uniformly. Bed expansion of adsorbent was measured and recorded with increasing liquid superficial velocity through the inlet of the column. The superficial flow velocity was subsequently plotted against bed expansion defined as the percentage ratio of expanded and settled bed heights.



Key:

- 1. Chart recorder;
- 2. On-line UV spectrophotometer;
- 3. Fluidized bed column;
- 4. Buffer reservoir, 10 mM Tris/HCl containing 0.2% (w/v) sodium azide at pH 7.5;
- 5. Tracer reservoir containing 1% (v/v) acetone in buffer;
- 6. Peristaltic pump.

Figure 1: The experimental configurations for the study of RTD experiments

Residence Time Distribution (RTD) measurements were performed by using a negative step signal method (Amersham Bioscience, 1998, Levenspiel,1972). A bed of the adsorbent particles was fully expanded using buffer A at the test flow rate. A dilute acetone solution (1% v/v) was used as the input to the column in a system fluidised with buffer A. The UV absorbance of the acetone was measured spectrophotometrically at 280 nm in the exit stream from the column using a UV monitor.

Theory and Method

The relationship between liquid superficial velocity (U) and bed voidage (ε) in a fluidised bed can be described by the classical correlation first postulated by Richardson and Zaki (1954):

$$U = U_t \varepsilon^n \tag{1}$$

The Richardson – Zaki coefficient (n) can be calculated from correlations available in the literature. The model of Shiller and Naumann is commonly used for the prediction of terminal velocity of a spherical particle:

$$G_a = 18 \text{ Re}_t + 2.7 \text{ Re}_t^{1.687} \quad 3.6 < G_a < 10^5$$
 (2)

where

$$G_{a} = \frac{\rho \cdot (\rho_{p} - \rho) \cdot g \cdot d_{p}^{3}}{\mu^{2}}$$
 (3)

and

$$Re_{t} = \frac{\rho.d_{p}.U_{t}}{\mu} \tag{4}$$

The model of Shiller and Naumann has been successfully used to estimate the particle terminal velocity.

The Bodenstein number (B_o) and axial dispersion coefficient (D_{axl}) , which expresses the state of liquid dispersion and fluidization behavior, can be calculated according to the following Eqs. (5), (6), (7) and (8).

$$\frac{\delta^2}{\overline{t^2}} = \frac{2}{B_0} - 2(\frac{1}{B_0})^2 \cdot [1 - \exp(B_0)]$$
 (5)

$$\varepsilon = 1 - \frac{(1 - \varepsilon_0).H_0}{H} \tag{6}$$

$$N = \frac{\overline{t^2}}{\delta^2} \tag{7}$$

$$B_0 = \frac{U.H}{\varepsilon.D_{axl}} \tag{8}$$

The axial dispersion in the liquid phase can also be interpreted in a form more familiar to researchers in chromatography based on the theoretical plate number (Pai et al, 2000). For small derivations from plug flow, the axial dispersion of the liquid phase can be expressed by Eq. (9) (Bilerau et al, 2001; Lan, 2000; Mullick and Flickinger, 1999).

$$D_{axl} = \frac{U.H}{2.\epsilon.N}$$
 (9)

In this article, two methods for calculation of the mean residence time t and variance σ are used. The first is a mathematical method (exact method), where t is called the centroid of the distribution and is an important parameter for RTD analysis. It is calculated as follows:

$$\bar{t} = \frac{\int_{0}^{\infty} tCdt}{\int_{0}^{\infty} Cdt} \approx \frac{\sum_{i} t_{i}C_{i}\Delta t_{i}}{\sum_{i} C_{i}\Delta t_{i}}$$
(10)

where C_i is the concentration of tracer at time t_i . Variance (σ^2) is the next most important parameter of the RTD curve, which is given by:

$$\delta^{2} = \frac{\int_{0}^{\infty} (t - \bar{t})^{2} C dt}{\int_{0}^{\infty} C dt} = \frac{\int_{0}^{\infty} t^{2} C dt}{\int_{0}^{\infty} C dt}$$
(11)

The second one is called the short-cut method (Figure 2). In this method, the time interval from switching the feed to the buffer until the signal reached 50% of the full deflection is taken as the mean residence time (t). The time interval of the signal between 84.15% and 15.85% yields the spread of the distribution (2σ) .

The curve in Figure 2 is representative of a typical experiment to determine theoretical plate numbers. A dilute tracer solution is used as the input to the contactor. The tracer concentration of the exit stream from the expanded bed contactor is measured and recorded against elapsed time (t).

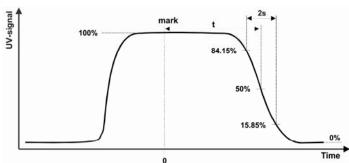


Figure 2: UV signal recording during the test procedure for the determination of the theoretical number using the step-input method

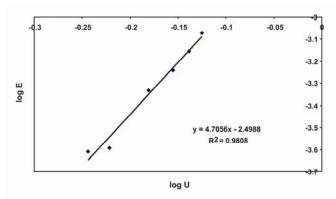


Figure 3: Bed expansion of adsorbent operated at settled bed height of 10 cm as a function of linear flow velocity.

RESULT AND DISCUSSION

Bed Expansion Characteristics and Richardson- Zaki Parameters

The bed expansion contributes to the adsorption efficiency as a composite function of liquid distribution, liquid properties (e.g., viscosity, density), particle characteristics and the configuration of the column in terms of wall and distributor effects (Thommes, 1997). In order to estimate the variation of the bed expansion as a function of flow velocity throughout the bed, the Richardson – Zaki equation was used .The experiments were performed in a clean system (buffer) and bed expansions are shown in Figure 3. The theoretical terminal velocity was derived from the Shiller and Naumann model.

The experimental values of terminal velocity can be determined by fitting to the Richardson – Zaki equation. The summary of bed expansion characteristics is presented in Table1.The experimental value of Richardson – Zaki coefficient determined here is close to the value of 4.8

commonly used in the laminar flow regime.

The expansion characteristics of adsorbent operated at a settled bed height of 10 cm was determined .The adsorbent was expanded in buffer A in an Amersham Bioscience contactor (1 cm inner diameter) with respect to increasing linear flow velocity.

Experimental values of U_t and n were derived from bed expansion profiles and settled bed voidage of 0.4 and these data were applied in all experiments.

The Effect of Different Settled Bed Height (SBH) on Hydrodynamic Behavior

The effect of different settled bed height (SBH) on hydrodynamic behavior of the expanded bed was investigated. The results are shown in Table 2 and, in order to better understand the variation of axial dispersion with SBH, the axial dispersion coefficients as a function of SBH are shown in Figure 4. Comparison of axial dispersion confirms that the 8-10 cm SBH is the optimal value (in both methods) that can be used in expanded beds.

Table 1: Experimental and theoretical values of expansion coefficient (n_{exp} and n_t respectively) and terminal velocities (U_t exp and U_{tt} , respectively) for adsorbent

n _{exp}	n _t	$U_{t exp}$	U _{tt}
4.7056	4.7454	3.171×10 ⁻³	3.2526×10 ⁻³

Table 2: B_0 and D_{axl} for different settled bed heights obtaining with the exact (a) and approximation (b) methods. These experiments are carried out in a column with a 1.6 cm diameter.

SBH	EBH	σ	t	B_0	$\mathbf{D}_{\mathbf{axl}} \times 10^{5}$
5	10	1.95011	2.60057	3.5565	1.3315
6	12	2.13387	2.66059	3.1090	1.82787
7	14	2.14326	3.13497	4.2790	1.54940
9	18	3.53547	5.03721	4.0599	2.09962
10	20	2.04085	2.95353	3.0076	3.14916

(a)

SBH	EBH	3	U (ml/min)	$U\times10^4$ (m/s)	B _o	$\mathbf{D_{axl}} \times 10^6$
5	10	0.7	4	3.315	8.78940	5.3800
6	12	0.7	4	3.315	14.1414	4.0180
7	14	0.7	4	3.315	16.5514	4.0057
8	16	0.7	4	3.315	24.9738	3.0340
9	18	0.7	4	3.315	9.18600	9.2796
10	20	0.7	4	3.315	9.78100	9.6835
11	22	0.7	4	3.315	10.9490	1.0508
12	24	0.7	4	3.315	9.99800	1.6700

(b)

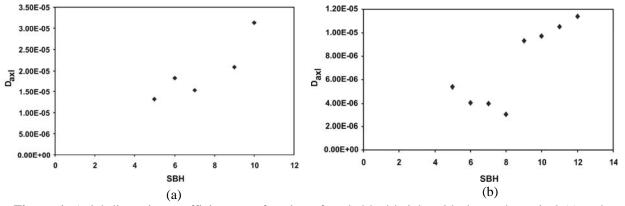


Figure 4: Axial dispersion coefficient as a function of settled bed height with the mathematical (a) and short-cut (b) methods. Axial dispersion measurements were performed using a negative step signal method. Dilute acetone (1% (v/v) in buffer A was used as the input to the contactor and the UV absorbance of the bed effluent was monitored, quantified and recorded. These experiments were carried out in a column with a 1.6 cm diameter.

The Effect of Bed Expansion on Hydrodynamic Behavior

The effect of different bed expansions (30%, 80%, 130%, 180%) was investigated by both methods to obtain the optimum bed expansion for an

acceptable axial dispersion coefficient. The result for different bed expansions are shown in Table 3. Comparison of axial dispersion values confirms that bed expansion between 100%-130% is the optimal value based on the minimum liquid mixing that can be used in expanded beds.

Table 3: B_0 and D_{axl} for different bed expansions obtained by the exact (a) and approximation (b) methods. These experiments were carried out in column with a 1 cm diameter.

SBH	ЕВН	σ	t	\mathbf{B}_0	$\mathbf{D_{axl}} \times 10^5$	
10	13	2.73169	3.23369	2.7997	1.1677	
10	18	3.20752	4.42120	3.8000	2.7416	
10	23	3.39944	4.62144	3.6655	5.3979	
10	28	1.44565	2.61388	6.5371	4.6614	
(a)						

U $U\times10^4$ SBH **EBH** Bo $\mathbf{D_{axl}} \times 10^6$ 3 (ml/min) (m/s)0.54 5.3923 10 13 0.64 1.358 6.0628 10 18 0.66 1.8 3.821 6.1803 1.6857 3.9046 10 23 0.74 3 6.366 5.0673 28 0.78 1.4684 2.0751 10 8.488 4 (b)

2.50E-04 6.00E-04 5.00E-04 2.00E-04 4.00E-04 1.50E-04 3.00E-04 1.00E-04 2.00E-04 1.00E-04 5.00E-05 0.00E+00 0.00E+00 50% 100% 150% 200% 50% 100% 150% 200% 0% Bed expansion Bed expansion (b)

Figure 5: Axial dispersion coefficient as a function of bed expansion with the mathematical (a) and short-cut (b) methods. Axial dispersion measurements were performed using a negative step signal method. Dilute acetone (1% (v/v) in buffer A was used as the input to the contactor and the UV absorbance of the bed effluent was monitored, quantified and recorded. These experiments were carried out in a column with a 1 cm diameter.

The Effect of Column Diameter on Hydrodynamic Performance

A step input of a tracer solution (acetone) was used to characterize the overall degree of mixing and to investigate the effect of column diameter. It is clear that an increase of the column diameter causes an increase in the liquid mixing, but this prediction is qualitative, and, in order to have a quantitative prediction for designing the large scale expanded bed columns, comparison of the Bodenstein Number (B_o) and Axial Coefficient (D_{axl}) for different column

diameters is vital. Here these values for three different column diameters (1, 1.6 and 2.5 cm) were calculated by the two mentioned methods. Comparison of $B_{\rm o}$ demonstrated our expectation that an increase of column diameter causes an increase in mixing. Because larger $B_{\rm o}$ represents lower mixing and lower mixing is itself a sign of higher efficiency, scaling up somehow disturbs the efficiency of the columns .The results obtained with the exact and approximate methods are shown in Table 4. The settled bed height was 10 cm and the bed was expanded to 23 cm.

Table 4: B_0 , and D_{axl} for columns with different diameters obtained with the exact (a) and approximation (b) methods.

ID(cm)	σ	t	\mathbf{B}_{0}	$D_{axl} \times 10^5$
1	3.399442	4.602144	3.6655	3.2388
1.6	3.722655	4.167832	2.5069	4.3162
2.5	3.127859	3.638553	2.7064	5.8487

ID (cm)	3	U (ml/min)	U×10 ⁴ (m/s)	$\mathbf{B}_{\mathbf{o}}$	D _{axl} ×10 ⁵
1	0.74	1.8	3.8197	10.1867	1.1654
1.6	0.74	4.2	3.4815	6.9147	1.5649
2.5	0.74	15	5.0929	7.04586	2.2466

(b)

CONCLUSIONS

The hydrodynamic characteristics of a liquid fluidized bed adsorption were investigated herein. The streamline adsorbent was subjected to bed expansion assessment and conformed to the Richardson – Zaki correlation. The terminal velocity of the adsorbent beads and bed expansion behavior could be approximated using the Shiller and Naumann model. The mass transfer between liquid and solid phases in a liquid fluidized bed in the low Reynolds number region has often been reported to be affected by axial dispersion; therefore, experimental data concerning the degree of liquid-solid mixing was obtained here from step-input studies.

The results indicated that, upon increasing column diameter, $B_{\rm o}$ decreased. However, RTD analysis showed reduced $D_{\rm axl}$ values with decreasing bed expansion. It also has been found that a settled bed height of 8-10 cm and a bed expansion between 80%-130% are suitable with respect to the operation of fluidized bed systems. All calculations were carried out by the mathematical as well as the approximate short-cut method.

To the best of our knowledge, the current study is the first one that has demonstrated the potential for exploiting the short-cut method side-by-side with the commercial mathematical method. If the large scale hydrodynamic investigation of fluidized bed adsorption is successful, optimization of hydrodynamic performance with good adsorption will contribute to the de-bottlenecking of current biopharmaceutical manufacture.

ACKNOWLEDGMENTS

The authors acknowledge support of Babol University of Technology and Tehran University for various aspects of this collaborative work.

REFERENCES

Amersham Bioscience: Expanded Bed Adsorption – principles and methods, Pharma Biotech, Uppsala, Sweden (1998).

Bilerau, H., Zhang, Z., Lyddiatt, A., Direct process

integration of cell disruption and fluidized bed adsorption for the recovery of intracellular proteins. Chem. Tech. Biotechol., 74, 208 (2001).

Camprubi, S., Bruguera, M., Canalias, F., Purification of recombinant histidine-tag streptolysin O using immobilized metal affinity expanded bed adsorption (IMA-EBA), J. Biological Macromolecules, 38, 134–139 (2006).

Clemmitt, R. H., Chase, H. A., Facilitated downstream processing of a histidine-tagged protein from unclarified *E. coli* homogenates using immobilized metal affinity expanded-bed adsorption. Biotechnol, Bioeng, 67, 206-216 (2000).

Ferreira G. N. M., Cabral, J. M. S., Prazeres, D. M. F., Anion exchange purification of plasmid DNA using expanded bed adsorption, Bioseparation, 9, 1-6 (2000).

Jahanshahi, M., Re-design of downstream processing techniques for nanoparticulate bioproducts. Iranian Journal of Biotechnol, 2, (2004).

Jahanshahi, M., Ling, T. C., Ghoreyshi, A., Khavarpour, M., Analysis of performance of the anion exchange and pseudo-affinity chromatography for intracellular enzyme purification, Iranian Journal of Chem. Eng. 3, 92-107 (2006).

Jahanshahi, M., Pacek, A. W., Nienow, A. W., Lyddiatt, A., Fabrication by three-phase emulsification of pellicular adsorbents customized for liquid fluidized bed adsorption of bioproducts, J. Chem. Tech. Biotechnol., 78, 111-1120 (2003).

Jahanshahi, M., Sun, Y., Santos, E., Pacek, A. W., Franco, T. T., Nienow, A. W., Lyddiatt, A., Operational intensification by direct products sequestration from cell disruptates: Application of a pellicular adsorbent in a mechanically integrated disruption-fluidized bed adsorption process, Biotechnol. Bioeng., 80, 201-212 (2002).

Jahanshahi, M., Williams, S., Lyddiatt, A., Shojaosadati, S. A., Preparation and purification of synthetic protein nanoparticulates, IET Nanobiotechnology Journal, 151, 176-182 (2004).

Jahanshahi, M., Zhang, Z., Lyddiatt, A., Subtractive chromatography for purification and recovery of nano-bioproducts, IET Nanobiotechnology Journal, 152, 12-126 (2005).

- Lan, J. C. W., Protein purification using fluidized bed chromatography, Physical and biochemical characterization of a simple adsorbent contactor, University of Birmingham, PhD Thesis (2000).
- Levenspiel, O., Chemical Reaction Engineering, Wiley & Sons, New York (1972).
- Mullick, A., Flickinger, M. C., Expanded bed adsorption of human serum albumin from very dense Saccharomyces cerevisiae suspensions on fluoride-modified zirconia, Biotechnol. Bioeng., 65, 282 (1999).
- Pai, A., Gondkar, S., Lali, A., Enhanced performance of expanded bed chromatography on rigid superporous adsorbent matrix, J. Chromatogr., A., 867, 113-130 (2000).
- Thommes, J., Fluidized bed adsorption as a primary

- recovery step in protein purification, Advances in Biochem. Engine./Biotechnol., 58, 185,(1997).
- Thommes, J., Halfar, M., Lenz, S., Kula, M. R., purification of monoclonal antibodies from whole hybridoma fermentation broth by fluidized bed adsorption, Biotechnol. Bioeng., 45, 205-211 (1995).
- Tong, X., Sun, D. Y., Particle size and density distributions of two dense matrices in an expanded bed system, J. Chromatogr A. 977, 173-183 (2002).
- Ujam, L. B., Clemmitt, R. H., Clarke, S. A., Brooks, R. A., Rushton, N., Chase, H. A., Isolation of monocytes from human peripheral blood using immuno-affinity expanded-bed adsorption, Biotechnol. Bioeng., 83, 554-566 (2003).